

Grant County Public Utility District Implementation Plan 2018-19 Priest Rapids Hatchery Monitoring and Evaluation

Introduction

The Public Utility District No. 2 of Grant County (GCPUD) has guiding principles and approaches for the monitoring and evaluation (M&E) of all of its hatchery programs that are provided in an overarching M&E plan that encompasses all of its programs (Pearsons and Langshaw 2009, Hillman et al. 2013, Hillman et al. 2017). The first comprehensive sampling for Monitoring and Evaluation of Priest Rapids Hatchery fall Chinook salmon production began in the fall of 2010. Though the comprehensive M&E programs are relatively recent, the Washington Department of Fish & Wildlife (WDFW) has been conducting monitoring and evaluation of URB fall Chinook in the Hanford Reach dating back to the early 1980's monitoring the sport fishery, hatchery returns, and escapement for run reconstruction and coded wire tag (CWT) returns. This implementation plan summarizes the Tasks and Objectives specified in the M&E Plan for GCPUD hatchery programs. The M&E Plan approach is also included in Section 11 of the Priest Rapids Hatchery (PRH) Hatchery and Genetic Management Plan (HGMP). Meeting the Objectives as well as accomplishing the Tasks listed in the M&E Plan requires the assemblage of data and analysis from numerous Programs. This implementation plan identifies all Tasks that must be accomplished to meet the M&E Objectives as well as the Agency responsible for funding, staffing, supervision, and data collection.

Project Coordination

WDFW M&E staff dedicated to PRH will work in conjunction with fish culture staff from PRH, the Columbia River Coded Wire Tag Recovery Program (CRCWTP), Region 3 Fish Management, the WDFW District 4 Fish Biologist, UCR Steelhead Monitoring and Evaluation, and the GCPUD Research Science team and the United States Army Corps of Engineers (USACE) to complete all tasks included in the M&E Plan. In addition, samples collected at PRH and in the field will be transported and analyzed by WDFW Labs including the WDFW Scale Reading Lab, WDFW Genetics Lab, and the WDFW Otolith Lab. PRH M&E staff will process all coded-wire tag samples. Data collection and analysis associated with the PRH M&E and Hanford Reach population monitoring is incorporated into the WDFW Traps, Weirs, and Spillways (TWS) data base for use in M&E analyses, forecasting and managing fall Chinook salmon. WDFW will secure and hold all environmental permits necessary for work that is described in this statement of work. Data collection design, review, analysis, and reporting will be completed as a combined effort between GCPUD and WDFW Biologist(s). The performance period of this implementation plan is from July 1, 2018 – June 30, 2019.

Objectives

The objective of the PRH M&E plan is to evaluate the performance of the PRH program relative to the goals and objectives of the PRH program. The overarching goal of the PRH program is to meet GPUD's hatchery mitigation by producing fish for harvest while keeping genetic and

ecological impacts within acceptable limits.

Objective 1: Determine if the PRH program has affected abundance and productivity of the Hanford Reach Population.

Objective 2: Determine if the run timing, spawn timing, and spawning distribution of both the natural and PRH components of the Hanford Reach population are similar.

Objective 3: Determine if genetic diversity, population structure, and effective population size have changed in natural spawning populations as a result of the PRH program. Additionally, determine if PRH programs have caused changes in phenotypic characteristics of the Hanford Reach population.

Objective 4: Determine if the PRH adult-to-adult survival (i.e., hatchery replacement rate) is greater than the Hanford Reach adult-to-adult survival (i.e., natural replacement rate) and equal to or greater than the program specific hatchery replacement rate (HRR) expected value based on survival rates listed in the BAMP (1998).

Objective 5: Determine if the stray rate of PRH fish is below the acceptable levels to maintain genetic variation between stocks.

Objective 6: Determine if PRH fish were released at the programmed size and number.

Objective 7: Determine if harvest opportunities have been provided using PRH returning adults.

Objective 8: Determine if the PRH has increased pathogen type and/or prevalence in the Hanford Reach population.

Objective 9: Determine if ecological interactions attributed to PRH fish affect the distribution, abundance, and/or size of non-target taxa of concern that were deemed to be at sufficient risk.

Methods

The PRH M&E plan is primarily organized in tables to facilitate review and provide clear direction for implementation. This plan was designed to be consistent with M&E plans that were designed and are currently being implemented by Chelan and Douglas Public Utility Districts. A variety of field methods will be used to collect the data necessary to achieve M&E objectives. Methods include redd surveys, carcass surveys, adult trapping, data collection at the hatchery during spawning, data collected at the hatchery during rearing, juvenile collection and tagging in the natural environment, disease monitoring, and NTTOC monitoring in the natural environment, if identified as necessary in a risk assessment.

Tagging and marking will be an essential component of apportioning hatchery and natural origin production and stray rate. All of the hatchery origin fish will be marked prior to release.

Two mass marking techniques have been proposed, otolith marking and adipose fin clipping. Currently, all hatchery origin fish will continue to be otolith marked. Otolith marking was initiated during brood year 2007.

WDFW and GPUD will refine subsample size estimates for submission of otolith samples to the WDFW Otolith Lab after the ages of the fish sampled have been determined by the WDFW Scale Lab (typically during the winter). The size of the subsamples by age and gender is based on number of samples collected and the relationship between sample size and deviation of the estimated variable from the cumulative estimate of a variable.

Passive Integrated Transponder (PIT) tag observations will be inventoried at the PIT-tag arrays located at Bonneville, McNary, Ice Harbor, and Priest Rapids dams and the PRH discharge channel.

The following Tasks are necessary to meet the Objectives of the M&E Plan (some of these tasks are funded wholly or in part by other organizations):

- Task 1.** PRH sampling of adult returns at the trap
- Task 2.** PRH sampling of adult returns during spawning
- Task 3.** Compilation of PRH origin URB fall Chinook salmon in the sport harvest including the Hanford Reach, Yakima River, Wanapum Tribal Fishery, ocean, and lower Columbia commercial and tribal harvest
- Task 4.** Redd surveys in the PRH Discharge Channel, and Yakima River;
- Task 5.** Adult counts at dams and hatcheries
- Task 6.** Carcass surveys in the Hanford Reach, Hatchery Discharge Channel, Priest Rapids Dam Pool, and Yakima River
- Task 7.** Juvenile marking and tagging at PRH
- Task 8.** Juvenile marking and tagging of the Hanford Reach natural population
- Task 9.** Sampling at the fish ladder trap (OLAFT) for run reconstruction and potentially for broodstock collection
- Task 10.** Sampling and analysis to determine potential coded wire tag bias and carcass recovery bias
- Task 11.** Operation and evaluation of PIT tag detections at the PRH discharge channel and derived estimates from dams.
- Task 12.** Collection of natural origin brood stock by gaff and line fishing in the Hanford Reach.

Task 1. Sampling Information, Methods, and Metrics for Data Collected at the Priest Rapids Hatchery Volunteer Trap (HGMP, Attachment 5, Table 7)

Objectives: 1, 2, 3, 4, & 5

Frame: Population of fish collected at the PRH trap

Sample Unit: Fish surplused. Fish transported from the trap and ponded for broodstock are included in Task 2.

Sample Size: Sample all Chinook salmon regardless of external marks or fin clips that are not used for broodstock to identify the presence of CWT. The sample rate for detailed biological data collection will be set in September after WDFW updates the run forecast for fall Chinook salmon returns to PRH to accommodate a sample goal of 1,000 fish surplused from the PRH volunteer trap.

Schedule: September 1 through December 15

Methods: All Chinook salmon surplused or found as a mortality will be scanned for the presence of a CWT.

All in-sample Chinook salmon will be sampled for age (scale sample), gender, fork length and post orbital hypural plate (POHP) length. In-sample fish will also be sampled for origin by either the collection of CWT if present or otoliths. The CWT fish from the in-sample group will have their snout removed, bagged and labeled for processing at either the PRH wet lab or the WDFW Pasco office. Non CWT fish will have their otoliths removed, placed in pre-labeled vials filled with a preservative, and cataloged for processing. Select otoliths will be sent to the WDFW Otolith Lab to determine if they are PRH origin.

The collection of scales from in-sample fish in conjunction with CWT recoveries is necessary to validate age reading and to provide a cross reference for data collections. To reduce the workload of processing “out-of-sample” CWT fish, scale samples will not be collected. These fish will still be sampled for gender, fork length, and the snout removed, bagged, and labeled for processing.

Data Collected: Scale (age), otolith (PRH origin), fork length, POHP length, gender, and CWT (origin).

Personnel and Equipment: A team of M&E technicians with oversight by a lead M&E technician and the M&E biologist, transport vehicles, and standard sampling gear. Staff will sample PRH returns from the volunteer trap. Staff will be responsible for biological sampling, data entry, and reading of CWT. These activities will occur at both the PRH wet lab and the WDFW Pasco office. The WDFW Otolith Lab will process otoliths to determine if they are PRH origin. The WDFW Scale Lab will read all scales to age including years in freshwater.

Task 2. Sampling Information, Methods, and Metrics for Data Collected at the Priest Rapids Hatchery during Holding and Spawning (HGMP, Attachment 5, Table 8)

Objectives: 1, 2, 3, 4, 5, & 8

Frame: Spawning population of PRH

Sample Unit: PRH Broodstock

Sample Size: Sample all Chinook salmon for origin and CWTs. The sample rates (in-sample) for detailed biological data collection will be set in September after WDFW updates the run forecast for fall Chinook salmon returns to PRH to accommodate an in-sample goal of 1,000 ponded fish originating from the PRH volunteer trap. In-sample rates for other sources of broodstock (e.g., OLAFT and ABC fishery) will be set after the collection sizes are determined.

Schedule: October 26 through December 10, 2 days per week

Methods: All fish ponded, regardless of source will be scanned for the presence of CWTs after they are spawned, surplused, or found as a mortality. All in-sample Chinook salmon will be sampled for age (scale sample), gender, fork length and POHP length. In-sample fish will also be sampled for origin by either the collection of CWT if present or otoliths. The CWT fish from the in-sample group will have their snout removed, bagged and labeled for processing at either the PRH wet lab or the WDFW Pasco office. Non CWT fish will have their otoliths removed, placed in pre-labeled vials filled with a preservative, and cataloged for processing. Select otoliths will be sent to the WDFW Otolith Lab to determine if they are PRH origin.

The collection of scales from in-sample fish in conjunction with CWT recoveries is necessary to validate age reading and to provide a cross reference for data collections. To reduce the workload of processing out-of-sample CWT fish, scales will not be collected. These fish will only be sampled for gender, fork length, and the snout removed, bagged, and labeled for processing. Fecundity will be collected during sampling of spawned fish. Ovarian fluid will be drained from the egg takes of females sub-sampled for fecundity, eggs will then be weighed, and the requisite number of eggs based on 2013 sampling (e.g., 100 eggs) will be collected and weighed to estimate fecundity for the female. Scales, otoliths, and fork length will be collected for each female in the fecundity sample. Attempts will be made to measure fecundity on presumptive hatchery origin (collected from volunteer trap) and natural origin fish (collected from fish captured in the ABC fishery or OLAFT possessing no marks or tags such as an adipose clip or CWT). The goal will be to get 100 samples from each of the two origins (total 200) and to cover the full range of female size.

Data Collected: Scale (age), otolith (PRH origin), fecundity, egg weight, fork length, POHP length, gender, and CWT (origin).

Personnel and Equipment: Up to six WDFW M&E technicians with M&E Biologist oversight, one vehicle. The WDFW Otolith Lab will process otoliths to determine if they are PRH origin. The PRH M&E staff will process CWT samples to determine origin. The WDFW Scale Lab will

read all scales to age including years in freshwater. The WDFW Fish Health Specialist will follow disease testing protocols established for WDFW hatcheries.

Task 3. Sampling Information, Methods & Metrics for Harvest Sampling (HGMP, Attachment 5, Table 9)

Objectives: 1, 2, 4, 5, & 7

Frame: Harvest of natural origin Hanford Reach fall Chinook salmon and fall Chinook salmon released from PRH (e.g., Wanapum Tribal Harvest, Yakima River Fall Chinook salmon Sport Harvest, Hanford Reach fall Chinook salmon sport harvest)

Sample Unit: All Chinook salmon observed during the Hanford Reach salmon fishery.

Task 3.1 Hanford Reach Sport Fishery, Phenotypic Metrics

Sample size: All Chinook salmon sampled are scanned for the presence of CWTs. A minimum of 350 Chinook salmon from the sport harvest will be sub-sampled at the appropriate rate (e.g., every 10th carcass) to determine origin and other phenotypic metrics; age, gender, and length at age.

Schedule: Daily from August 16 through October 22

Methods: Staff will be stationed at primitive boat launches throughout the Hanford Reach including Vernita Bridge, Waluke, and Ringold. All anglers encountered will be interviewed to determine catch and estimate harvest. Harvested Chinook salmon from these anglers will be sampled to determine origin (CWT), age (scales), gender, and length. Methods are fully described in the WDFW Annual Report (Hoffarth, 2008).

Data Collected: Species harvested and released, location, number of boats, number of anglers, effort (angler hours and trailer index counts), catch per unit effort, harvest per unit effort, incidental catch. Biological data will include age (scale), fork length, gender, fecundity, and origin (CWT).

Personnel and Equipment: WDFW CRCWTP will provide three staff for monitoring the sport harvest and Region 3 will provide one additional staff. PRH M&E will provide one additional staff for sport harvest monitoring. Staff start dates are staggered to match angler effort and reduce costs. First staff member starts August 16, second staff member begins September 1, third staff member begins September 15, and the final staff member starts when the Hanford Reach opens for steelhead, typically October 1. The CRCWTP enters all data collected during the sport fishery. The WDFW District 4 Fish Biologist is responsible for analyzing the data, generating weekly harvest and ESA impact estimates, and evaluating if current harvest is within the harvest guidelines of the Hanford Reach Fall Chinook Harvest Management Plan. PRH M&E staff will process CWT samples to origin and age. The WDFW Scale Lab will read all scale to age including years in freshwater.

Task 3.2. Yakima River Fall Salmon Sport Fishery

Sample Size: All Chinook salmon observed in the harvest are scanned for the presence of a CWT and sampled for run reconstruction. No otolith sampling has been designated at this time.

Schedule: Five days per week from September 1 through October 22

Methods: Staff monitors bank and boat anglers from the Duportail Access Area upstream to Prosser. All anglers encountered will be interviewed to determine catch and estimate harvest. Harvested Chinook salmon from these anglers will be sampled to determine origin (CWT), age (scales), gender, and length. Methods are fully described in the WDFW Annual Report (Hoffarth, 2008).

Data Collected: Species harvested and released, numbers of boats, number of anglers (bank & boat), effort (angler hours), catch per unit effort, harvest per unit effort, incidental catch, scale (Chinook, coho, and steelhead), fork length, gender, and origin (CWT).

Personnel and Equipment: WDFW, Region 3, provides one technician for monitoring the sport. The WDFW District 4 Fish Biologist is responsible for entering all data, analyzing the data, and generating weekly harvest and ESA impact estimates. The PRH M&E staff will process CWT samples to origin and age. The WDFW Scale Lab will read all scale to age including years in freshwater

Task 3.3 Wanapum Tribal Fall Chinook Salmon Fishery

Sample Size: All Chinook salmon harvested are sampled. No otolith sampling has been designated at this time.

Schedule: September 1 through October 15

Methods: All Chinook salmon and Coho harvested are sampled to determine origin (CWT), age (scales), gender, and length. All Chinook salmon and coho harvested will be scanned for the presence of a CWT. All snouts with a positive signal will be transported to the WDFW District 4 Office for processing. Methods are fully described in the WDFW Procedures for Sampling the Wanapum Fishery (Hoffarth, 2009).

Data Collected: Species harvested, incidental catch, number of nets, mesh size, age (scale), fork length, gender, and origin (CWT).

Personnel and Equipment: GCPUD Cultural staff will sample the fishery and provide the data and samples to the WDFW District 4 Fish Biologist. The WDFW District 4 Fish Biologist and staff from the Columbia River CWT Recovery Program (CRCWTRP) enter all data from the fishery. The PRH M&E staff will process CWT samples to origin and age. The WDFW Scale Lab will read all scale to age including years in freshwater

Task 4. Sampling Information, Methods, and Metrics for Redd Surveys (HGMP, Attachment 5, Table 4)

Objectives: 1 & 4

Frame: Redds in the Yakima River below Prosser

Sample Unit: Visible redds located in the Yakima River

Sample size: Total count of visible redds

Schedule: Weekly between October 22 and November 30

Methods: Foot and boat surveys will be conducted as generally described by Gallagher et al. (2007) and Murdoch et al. (2008). Redds will be identified based upon their relatively clean substrate and a bowl and tail spill morphology.

Data Collected: Redd counts, fish counts (live, dead). Biological data will be collected from carcasses recovered during these surveys. Biological data will include age (scale), fork length, gender, egg retention and origin (CWT in the Yakima).

Task 4.1. Yakima River Redd Surveys (RM 8-46)

Two WDFW staff persons funded by CRCWTRP, all four sections of river completed weekly, one section per day, boat survey (cataraft).

Task 5. Sampling Information, Methods, and Metrics for Adult Counts at Dams and Hatcheries

Objectives: 1, 2, 3, 4

Frame: Fall Chinook salmon in the Hanford Reach

Sample Unit: Fall Chinook salmon counted at dams or weirs (McNary, Priest Rapids, Ice Harbor, Prosser, PRH trap, Ringold Hatchery trap)

Sample Size: Total count or subsample

Schedule: Daily from August 9 through November 15

Methods: Dam counts using observers or video as generally described by Wagner (2007).

Data Collected: Fall Chinook salmon are recorded into two categories based on fork length, adults and jacks. Adults are all Chinook salmon greater than 22 inches in fork length.

Personnel and equipment: PRH M&E and RSH M&E Staff hatchery trap counts. Adult fish passage counts at dams are provided by the USACE, YKFP, and GCPUD.

Task 5.1. Adult Chinook Salmon Counts at Mainstem Hydroelectric and Diversion Projects

Fish counts at mainstem Projects including the Columbia, Snake, and Yakima Rivers necessary to meet PRH M&E objectives are funded by non-related programs by GCPUD, WDFW, the USACE, and the Yakama Indian Nation. These data are readily available on the internet and will be downloaded at frequent intervals by the WDFW District 4 Fish Biologist.

Task 5.2 Adult Chinook Salmon Counts at Priest Rapids Hatchery

Hatchery returns to Priest Rapids are enumerated by hatchery staff in coordination with M&E staff.

Task 5.3. Adult Chinook Salmon Counts at Ringold Springs Hatchery

Hatchery returns to Ringold Springs Hatchery are enumerated by hatchery staff in coordination with the Ringold Springs Hatchery M&E Program.

Task 6. Sampling Information, Methods, and Metrics for Carcass Surveys in the Natural Environment (HGMP, Attachment 5, Table 6)

Objectives: 1, 2, 3, 4, & 5

Frame: Hanford Reach, Hatchery Discharge Channel, Priest Rapids Pool, and Yakima River

Task 6.1. Hanford Reach Stream Surveys

Sample Unit: Salmon carcasses partitioned by reach, five river reaches have been established in the Hanford Reach:

- 1 - Vernita Bridge to Priest Rapids Dam
- 2 - Island #2 to Vernita Bridge
- 3 - Wooden Powerline Towers to Island #2
- 4 - Wooded Island to Wooden Powerline Towers
- 5 – Richland to Wooded Island

Carcasses recovered in the Columbia River immediately downstream of the PRH discharge channel will be included with those Chinook salmon recovered in the discharge channel but will be tracked separately should additional analysis of these fish be necessary. Stream survey crews will also sample in the Priest Rapids Pool two days per week.

Sample Size: All carcasses observed in the surveys will be sampled for the presence of a CWT. Approximately 2,500 carcasses will be sampled in the Hanford Reach for origin based on CWT and otoliths. These sampled fish will be used to determine other phenotypic metrics; age, gender, fork length, egg retention, in addition to the determination of origin.

Schedule: October 27 through December 6

Methods: All carcasses that are encountered will be collected with a gaff or by hand. Surveys will occur by boat or foot. Methods will generally follow Crawford et al. (2007), Murdoch et al. (2008); and Hoffarth et al. (2008). All Chinook salmon will be wanded for the presence of a

CWT. All Chinook salmon with a CWT present will be sampled for age (scale sample), gender, fork length, and the snout will be bagged and labeled for processing at the WDFW CWT Lab. Otoliths will be collected from in-sample fish, placed in a vial, and stored with an appropriate index number. Otoliths will be sent to the WDFW Otolith Lab for decoding to determine if they are PRH origin. Chinook salmon sub-sampled for run reconstruction will be sampled for age (scale samples), gender, fork length, and spawning success. Fish will be identified to gender based on morphology. Female Chinook salmon in the sub-sample will be dissected to determine spawn success based on the percentage of egg retention (e.g., 0.0-10%, 11-37%, 38-62%, 63-87%, 88-100%). The percent of egg retention will be first visually estimated and then calculated by dividing the count of eggs retained by an estimated fecundity based on length versus fecundity regressions. In addition, retained eggs will be quantified by counting in the field or bagged and later estimated in the office. Carcasses will be cut in half to avoid duplicate sampling in future surveys.

Data Collected: Scale (age), otolith (PRH origin), fork length, gender, spawn success, CWT (origin), location.

Personnel and Equipment: Three boat crews with a three person crew operating seven days per week. PRH M&E staff, RSH M&E staff, and CRCWTRP staff to perform field sampling, collect CWTs and otoliths, and enter data into the data base. This will require a total of 13 (three CRWTRP staff, three RSH M&E staff, and seven PRH M&E staff), three boats, three vehicles and standard sampling equipment. The WDFW Otolith Lab will process otoliths to determine if they are PRH origin. The PRH M&E staff will process CWT samples to origin and age. The WDFW Scale Lab will read all scale to age including years in freshwater.

Task 6.1.a. Evaluation of Carcass Drift for Post-spawn Fall Chinook Salmon

Sample Unit: Post spawn carcasses encountered in the Hanford Reach River Surveys

Sample Size: A minimum of 1,000 carcasses will be tagged in the Hanford Reach to evaluate carcass recovery bias and drift.

Schedule: October 30 through December 15

Methods: Collect carcasses during normal carcass recovery efforts described in Task 6.1. Include only fish from outside of the demographic sample but included in the CWT sample. Transport select carcasses (intact and relatively fresh) to main spawning areas, staple numbered tag inside of each operculum, measure length, and record gender. Release fish in areas with redds and record dates and UTM coordinates for release location of each tagged fish. Smaller and larger sized fish should be included in the tag group to insure that the full range of fish sizes are included. Fish should be released in a way to simulate a fish that dies after spawning (e.g., release in proximity to redds and at the depth of redd. Collect length, gender, and record dates and UTM coordinates for recovery location from recovered tagged fish during normal recovery efforts described in Task 6.1. Plot tag and recovery locations to estimate distance of drift (Km).

Personnel and Equipment: This work will coincide with carcass recovery. It will require use of manufactured numbered plastic tags, three staple guns and staples, 3 GPS devices, three jet boats and three vehicles. Two crews are funded by PRH M&E and RSH M&E.

Task 6.2. Priest Rapids Hatchery Discharge Channel Stream Surveys

Sample Unit: PRH Discharge Channel, approximately one-quarter mile in length from the Columbia River to the PRH trap.

Sample Size: Sample all carcasses recovered in the survey for CWT. In addition, carcasses will be sub-sampled at the same rate as Chinook salmon in the Hanford Reach stream survey listed above to determine other demographic; age, gender, presence of adipose fin, length, spawn success (including counts of retained eggs).

Schedule: October 25 through December 8

Methods: All carcasses that are encountered will be collected with a gaff or by hand. Surveys will occur by foot. Methods will generally follow Crawford et al. (2007), Murdoch et al. (2008); and Hoffarth et al. (2008). All fish will be wanded for the presence of CWT.

All Chinook salmon with a CWT present will be sampled for age (scale sample), gender, fork length, and the snout will be bagged and labeled for processing at the WDFW CWT Lab. Chinook salmon sub-sampled for run reconstruction will be sampled for age (scale samples), gender, fork length, CWT, otoliths, and spawning success. Otoliths will be collected at the same sample rate as Hanford Reach natural origin otolith collections. Otoliths will be sent to the WDFW Otolith Lab for decoding to determine if they are PRH origin. Fish will be identified to gender based on morphology. Female Chinook salmon in the sub-sample will be dissected to determine spawn success based on the percentage of egg retention (i.e., 0.0-10%, 11-37%, 38-62%, 63-87%, 88-100%). The percent of egg retention will be first visually estimated and then calculated by dividing the count of eggs retained by an estimated fecundity based on length versus fecundity regressions. . In addition, retained eggs will be quantified by counting in the field or bagged and later estimated in the office. Carcasses will be cut in half to avoid duplicate sampling in future surveys.

Data Collected: Scale (age), otolith (PRH origin), fork length, gender, spawn success, CWT (origin), otoliths (origin) and location.

Personnel and Equipment: Two PRH M&E staff and standard sampling equipment.

Task 6.3. Priest Rapids Pool Stream Surveys

Sample Unit: Priest Rapids Dam upstream to Wanapum Dam

Sample Size: All Chinook salmon recovered in the survey will be sampled for a CWT. The in-sample rate for biological data collection will be set in October 31 based on escapement size

Schedule: November 1 through December 15, two days per week

Methods: All carcasses that are encountered will be collected with a gaff or by hand. Surveys will occur by boat and foot. All Chinook salmon will be wanded for the presence of a CWT. Chinook salmon with a CWT present will be sampled for age (scale sample), gender, fork length, and the snout will be bagged and labeled for processing at the WDFW CWT Lab. Biological data collected from in-sample collections will include scale samples (age), gender, fork length, CWT or otoliths, and spawning success. Fish will be identified to gender based on morphology. Otoliths will be collected, placed in a vial, and stored with an appropriate index number. Otoliths will be sent to the WDFW Otolith Lab for decoding to determine if they are PRH origin. Female Chinook salmon in the sub-sample will be dissected to determine spawn success based on the percentage of egg retention (i.e., 0.0-10%, 11-37%, 38-62%, 63-87%, 88-100%). The percent of egg retention will be first visually estimated and then calculated by dividing the count of eggs retained by an estimated fecundity based on length versus fecundity regressions. In addition, retained eggs will be quantified by counting in the field or bagged and later estimated in the office. Carcasses will be cut in half to avoid duplicate sampling in future surveys.

Data Collected: Scale (age), otolith (PRH origin), fork length, gender, spawn success, CWT (origin), location.

Personnel and Equipment: Two field staff dedicated to the Hanford Reach stream surveys will accomplish this task as a component of the scope of work for technicians assigned to M&E for the natural environment funded by GCPUD and USACE (Listed in this Task above).

Task 6.4. Yakima River Stream Surveys

Sample Unit: Yakima River from the Duportail Access Area upstream to the I82 Bridge located immediately downstream Prosser Dam

Sample Size: All Chinook salmon recovered in the survey will be sampled for CWT. The in-sample rate for biological data collection will be set in October 31 based on escapement size.

Schedule: October 22 through November 30, all four sections of river completed weekly, one section per day, boat survey (cataraft)

Methods: All carcasses that are encountered will be collected with a gaff or by hand. Surveys will occur by boat. Methods will generally follow Crawford et al. (2007), Murdoch et al. (2008); and Hoffarth et al. (2008). All Chinook salmon will be wanded for the presence of CWTs. Chinook salmon with a CWT present will be sampled for age (scale sample), gender, fork length, and the snout will be bagged and labeled for processing at the WDFW CWT Lab. All Chinook salmon will be sampled for run reconstruction. Sampling will include scale samples (age), gender, fork length, and spawning success. Fish will be identified to gender based on morphology. Female Chinook salmon in the sub-sample will be dissected to determine spawn success based on the percentage of egg retention (i.e., 0.0-10%, 11-37%, 38-62%, 63-87%, 88-

100%). The percent of egg retention will be first visually estimated and then calculated by dividing the count of eggs retained by an estimated fecundity based on length versus fecundity regressions. Carcasses will be cut in half to avoid duplicate sampling in future surveys.

Data Collected: Scale (age), fork length, gender, spawn success, CWT (origin), location.

Personnel and Equipment: One boat (cataraft) with a two person crew funded by CWCWTRP standard sampling equipment (funded by CRCWTRP)

Task 7. Sampling Information, Methods, and Metrics for Data Collected to Monitor Fish Culture of Juveniles

Objectives: 6 & 8

Frame: Abundance and size (individual length and weight) of smolts at PRH at the time of release

Sample Unit: Representative sample of juveniles prior to release

Sample Size: Approximately 300 fish from each rearing vessel to determine size metrics

Schedule: Prior to release in late May and mid June

Methods: Estimate abundance and size at each life stage: egg, transfer to raceways, transfer to ponds, and release. Estimates of abundance will be generated by subtracting mortalities at subsequent life stages from estimates of green eggs.

Size and length data will be collected by M&E staff to calculate CVs for both matrixes. Each fish will be weighed (grams) and measured to fork length (millimeters).

The fish health specialist will respond to all fish disease outbreaks at the request of the fish hatchery staff and will visit PRH at least once a month. Water temperatures will be recorded hourly from egg to release.

Personnel and Equipment: Two PRH M&E staff

Data Collected: Abundance, fish per pound, length of individual fish, weight of individual fish, and fish health records

Task 8. Sampling Information, Methods, and Metrics to Monitor Natural Origin Juvenile Fish Abundance and Size

Objective(s): 1

Frame: Naturally produced juveniles in the Hanford Reach

Sample Unit: All Chinook salmon collected by beach seine in the Hanford Reach during the CWT tagging program.

Sample Size: Appropriate samples will be taken from both the marked and unmarked Chinook salmon.

Schedule: Late May to Early June (typically 10 day marking program)

Methods: Chinook salmon collected during the CWT marking program will be routinely sampled by length. Mark groups and tag codes will be enumerated. Goal of the marking program is to tag and adipose clip 200,000 of the natural production of fall Chinook salmon in the Hanford Reach.

Data Collected: Total numbers of Chinook salmon collected size at marking, mark numbers.

Personnel and Equipment: CWT trailer, technicians, supervision by both Columbia River Intertribal Fish Commission (CRITFC) and WDFW, collection is conducted by the Yakama Indian Nation, Umatilla Indian Nation, and CRITFC. This project is funded by the Bonneville Power Administration.

Task 9. Estimate Juvenile CWT Mark Rate and Adipose Clip Rates at Time of Release

Objectives: 4 and 7

Frame: PRH Juvenile at Time of Release

Sample Unit: All Chinook salmon collected at PRH.

Sample Size: 1,000 juveniles from each of five channel ponds at time of release.

Schedule: May and June

Methods: Sample lots of 1,000 PRH juveniles from rearing ponds. Scan sample fish with a V detector to determine proportion of CWT fish in the sample. Visually inspect sample fish for adipose clip to determine proportion of adipose clipped fish in the sample. Calculate proportions for both CWT and adipose clipped juveniles sampled. Compare the estimates to both the CWT and adipose clip proportions reported by hatchery staff and report comparisons in proportions.

Data Collected: Total number of juvenile sampled, total number of CWT fish detected, CWT loss rates, and total number of adipose clipped (non CWT) fish observed.

Personnel and Equipment: Two PRH M&E staff, CWT V-detectors, Tricaine, fish holding vessels and collection nets

Task 10. Sampling Information, Methods, and Metrics to Monitor URB Fall Chinook Salmon at the Off Ladder Adult Fish Trap (OLAFT)

Objectives: 5

Frame: Run of the River URB Fall Chinook salmon at the OLAFT at Priest Rapids Dam

Sample Unit: All Chinook salmon collected at the OLAFT.

Sample Size: Sample size will vary dependent upon returns and days available to sample. Goal is to collect 500 samples.

Schedule: September 1 to no later than November 15 (typically Tuesdays and Thursdays) Schedule may vary dependent upon staff availability and fish passage. Program will end if insufficient numbers of Chinook salmon are passing through the fish ladders.

Methods: Chinook salmon will be collected and sampled in conjunction with the UCR Steelhead Monitoring and Evaluation Program. All Chinook salmon encountered during operation of the OLAFT will be diverted to a separate holding and handling location dedicated to the fall Chinook salmon sampling. Chinook salmon will not be anesthetized during handling. Fork lengths, gender and the presence of fin clips will be recorded. Scales will be collected to determine age. Chinook salmon will be scanned for the presence of CWTs but no lethal sampling will occur. All data will be entered into a data base to be used for run reconstruction of the above Priest URB fall Chinook salmon population. Methods and protocols may be modified with the approval of the appropriate Agencies/Committees to accommodate additional Programs, such as, adult broodstock collection, PIT tagging/PIT tag detection.

Data Collected: Total numbers of Chinook salmon collected, marks (fin clips), CWT presence, fork length, age (scales), and gender.

Personnel and Equipment: WDFW staff will sample Chinook salmon for age composition. The trap will be operated by WDFW Steelhead M&E Program. Additional staff will be provided by the host Agency if sampling is expanded to accommodate special Projects. All equipment will be provided by monitoring/evaluation Programs.

Task 11. Operation and Data Analysis of PIT tag Detections at the Priest Rapids Hatchery Discharge Channel

Frame: PRH Juvenile Releases and Adult Returns

Sample Size: Up to 43,500 PIT juvenile fall Chinook salmon released from PRH. The number and origin of adult returns likely to be detected at the array is unknown.

Schedule: May 10 to December 15

Methods: The PIT-tag antennae arrays provide PIT tag detection of juveniles at release as well as returning adults. The PRH M& E staff will monitor PIT detections and analyze results to determine the abundance of both juvenile and adult PRH fall Chinook salmon, travel time and speed of juvenile fish, re-ascension of mini-jacks, and identification of non-PRH origin fish. PIT tag detections will be compared against adult detections at hydroelectric projects in the Columbia and Snake rivers to estimate interdam loss, smolt to adult survival to the hatchery, juvenile downstream survival, as well as adult re-ascension and fall back rates at McNary, Ice Harbor and Priest Rapids dams. WDFW maintenance crews will conduct routine maintenance of the PIT tag arrays as needed. An in depth inspection of the arrays will occur during May prior to the hatchery release of juvenile Chinook salmon and again during late August for preparation for the adult Chinook salmon return.

The volitional releases of juveniles from between holding ponds needs to be staggered by several days or the PIT-tag antennae array system will be overwhelmed by the high numbers of out-migrants and the data lost. The juvenile release strategy will be coordinated with hatchery staff.

Data Collected: Total numbers and timing of Chinook salmon PIT detections collected.

Personnel and Equipment: The PRH M&E staff.

Task 12. Angler Broodstock Collection

Objective(s): 3, 4, and 9

Frame: The Hanford Reach fall Chinook salmon

Sample Unit: All Chinook salmon collected in post season fishery.

Sample Size: Our goal is to collect up to 500 natural origin fish.

Schedule: October 26, 27, 28.

Methods: The angler broodstock collection (ABC) will occur post season of the established fall Chinook salmon fishery in the Hanford Reach. The following is a general description of the methods employed for the ABC. Volunteers will register each morning of the fishery; receive instructions on proper fish handling of live fish; and be advised of pertinent information regarding the fishery such as recording incidental catch of other fish species. Anglers will use conventional hook and line fishing techniques to capture fall Chinook salmon. There will be two GCPUD boat crews to shuttle fish from the river to the transport truck at Vernita Launch. Only fall Chinook in good physical condition with an intact adipose fin will be retained and used for broodstock. Fish collection sites will be established at the Vernita Bridge and Waluke boat launches. Tanker trucks and crews will receive fish and transport them to PRH. Fish collected from the ABC fishery will be held separately from the other broodstock so that they can be monitored and sampled independently. All Chinook collected during this program will be sampled at the hatchery. Otoliths will be collected to determine origin from a suitable

proportion of fish captured in the ABC fishery.

Data Collected: Chinook salmon collected will be sampled for marks (fin clips), CWTs; otoliths, fork length, age (scales), gender, and fecundity.

Personnel and Equipment: M&E staff, PRH staff, District Fish Biologist, GCPUD staff, and volunteer anglers. Two jet boats, fish holding tubes, two live tanks, one WDFW 1,500 gallon truck, and one GCPUD 1,500 gallon tank truck. *The second collection location at the Waluke boat launch will only occur if GCPUD provides the 1500 gallon tank truck and operator for each day of the collection.*

Expected Results and Applicability

Data collected from the 2018 return will be reviewed to determine if URB fall Chinook salmon production from PRH met all of the Objectives/Goals of the M&E Plan. In 2018, all PRH brood year returns will possess an otolith mark which will allow for determination of pNOB, pHOS, and PNI at a high level of confidence

Data Compilation and Analysis

Data will be maintained at the WDFW District office in Pasco and backed up on the WDFW Network (S:drive) and a dedicated thumb drive. The data is backed up weekly during the field season and each day the database is proofed. Data will be routinely entered into an Access or Excel data base in-season. The TWS Access database is the primary means of organizing and storing biological data. Some data associated with carcass recovery is entered into Excel spreadsheets. Age, CWT, and otolith information will be entered into the data base when it returns from the WDFW Labs. Scale cards are the primary means of recording data in the field. The scale cards will be proofed for obvious errors by staff at the end of each field day and again prior to entering data into the TWS which occurs routinely September through December. Staff will proof the data entered into the TWS mid-December and again in early January as scale age and CWT results becomes available. The otolith data is entered into the TWS and proofed as results become available generally in March and April. Proofing the TWS generally requires two staff; one to read aloud the data on scale cards and data sheets while another verifies the data in the TWS.

The Historical data as well as current data will be incorporated into the analysis and reporting for the PRH M&E Program. Historical data will be mined and presented to the maximum extent possible so that the longest data set can be evaluated. This will include mining data from other WDFW sources and from Battelle.

Summary of New Activities

Brief descriptions of alterations to existing Tasks or new tasks associated with PRH M&E during 2017-18 are given in Table 1.

Table 1. New activities for Priest Rapids Monitoring and Evaluation for 2018.

Activity	Associated Task
None	

Table 2. Summary of project timelines for data entry, analysis, and reporting July 1, 2018 – June 30, 2019

Activity	Dates
Priest Rapids Hatchery	
Trapping, Broodstock Collection, and Surplus Operations	September 1 – December 15
Spawning	October 15 – December 10
Hatchery Discharge Channel Stream Surveys	October 15 – December 15
PIT tag Array Maintenance and Evaluation	July 1 – June 30
Hanford Reach	
Hanford Reach Fall Chinook salmon Fishery	August 1 – October 22
Hanford Reach Stream Survey	November 1 – December 15
Priest Rapids Pool	
Priest Pool Stream Surveys	November 1 – December 15
Data Entry & Analysis	
Data Entry Return Year	September 1 – March 31
Data Review and Analysis	January 1 – June 30
Reporting	
Draft Monitoring and Evaluation Report	May 15
Final Monitoring and Evaluation Report	June 30

Summary of Sample Sizes

Sample size of each Task associated with PRH M&E during the performance period is given in Table 3.

Table 3. Sample size goals by Task. The sample size for otolith decoding will be determined after the age distribution data is available.

Task	Sample Size
Task 1. Priest Rapids Hatchery sampling of adult returns at the trap	100% for CWT, 1,000 for other
Task 2. Priest Rapids Hatchery sampling of adult returns during spawning	100% for CWT, 1,000 for other
Task 3. Compilation of Priest Rapids Hatchery origin URB fall Chinook salmon in the sport harvest including the Hanford Reach, Yakima River, Wanapum Tribal Fishery, ocean, and lower Columbia commercial and tribal harvest	All Encountered
Task 3.1 Hanford Reach Sport Fishery, Phenotypic Metrics	350
Task 3.2. Yakima River Fall Salmon Sport Fishery	All Encountered
Task 3.3 Wanapum Tribal Fall Chinook Salmon Fishery	All Encountered
Task 4. Redd surveys in the Priest Rapids Hatchery Discharge Channel, and Yakima River;	
Task 4.1 Hatchery Discharge Channel Redd Surveys	All Observed
Task 4.2 Yakima River Redd Surveys (RM 8-46)	All Observed
Task 5. Adult counts at dams and hatcheries	
Task 5.1. Adult Chinook Salmon Counts at Mainstem Hydroelectric and Diversion Projects	All Returns
Task 5.2 Adult Chinook Salmon Counts at Priest Rapids Hatchery	All Returns
Task 5.3. Adult Chinook Salmon Counts at Ringold Springs Hatchery	All Returns
Task 6. Carcass surveys in the Hanford Reach, Hatchery Discharge Channel, and Yakima River	
Task 6.1. Hanford Reach Stream Surveys	100% for CWT, 2,500 for other demographic data.
Task 6.1.a. Evaluation of Carcass Drift Post-spawn Fall Chinook Salmon	Minimum 1,000
Task 6.2. Hatchery Discharge Channel Stream Surveys	All Encountered
Task 6.3. Priest Rapids Pool Stream Surveys	All Encountered
Task 6.4. Yakima River Stream Surveys	All Encountered
Task 7. Sample Information, Methods, and Metrics for data collected to monitor fish culture of Juveniles	1,500 (300 x 5 Ponds) juveniles individually weighed and measured
Task 8. Juvenile marking and tagging of the Hanford Reach natural population	200,000 natural origin juvenile fall Chinook
Task 9. Estimate Juvenile CWT Mark Rate and Adipose Clip Rates at Time of Release	5,000 (1,000 x 5 ponds) juveniles for CWT tag rate and adipose clip rate estimate.
Task 10. Sampling at the fish ladder trap (OLAFT) for run reconstruction and potentially for broodstock collection	500 adult Chinook
Task 11. Operation and evaluation of PIT tag detections at the Priest Rapids Hatchery discharge channel and derived estimates from dam observations.	43,000 PIT Juvenile Release All PIT Adult Returns

References

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