

---

# MONITORING AND EVALUATION OF THE CHELAN AND GRANT COUNTY PUDs HATCHERY PROGRAMS

## 2014 ANNUAL REPORT

June 1, 2015



**T. Hillman**  
**M. Miller**  
*BioAnalysts*

**C. Willard**  
*Chelan PUD*

*Prepared by:*  
**M. Johnson**  
**C. Moran**  
**M. Tonseth**  
**A. Murdoch**  
*WDFW*

**B. Ishida**  
**C. Kamphaus**  
*Yakama Nation*

**T. Pearsons**  
**P. Graf**  
*Grant PUD*

*Prepared for:*  
**HCP Hatchery Committees and the PRCC Hatchery Sub-Committee**  
**Wenatchee and Ephrata, WA**

---

---

**Citation:** Hillman, T., M. Miller, M. Johnson, C. Moran, M. Tonseth, A. Murdoch, C. Willard, B. Ishida, C. Kamphaus, T. Pearsons, and P. Graf. 2015. Monitoring and evaluation of the Chelan and Grant County PUDs hatchery programs: 2014 annual report. Report to the HCP and PRCC Hatchery Committees, Wenatchee and Ephrata, WA.

# TABLE OF CONTENTS

<b>SECTION 1: INTRODUCTION.....</b>	<b>1</b>
<b>SECTION 2: SUMMARY OF METHODS.....</b>	<b>7</b>
2.1 Broodstock Collection and Sampling .....	7
2.2 Within Hatchery Monitoring.....	9
2.3 Juvenile Sampling.....	10
2.4 Spawning/Carcass Surveys .....	12
<b>SECTION 3: WENATCHEE STEELHEAD .....</b>	<b>17</b>
3.1 Broodstock Sampling.....	18
3.2 Hatchery Rearing .....	23
3.3 Disease Monitoring.....	33
3.4 Natural Juvenile Productivity .....	33
3.5 Spawning Surveys.....	40
3.6 Life History Monitoring.....	43
3.7 ESA/HCP Compliance.....	58
<b>SECTION 4: WENATCHEE SOCKEYE SALMON .....</b>	<b>61</b>
4.1 Broodstock Sampling.....	61
4.2 Hatchery Rearing .....	66
4.3 Disease Monitoring.....	72
4.4 Natural Juvenile Productivity .....	72
4.5 Spawning Escapement .....	76
4.6 Carcass Surveys .....	78
4.7 Life History Monitoring.....	80
4.8 ESA/HCP Compliance.....	94
<b>SECTION 5: WENATCHEE (CHIWAWA) SPRING CHINOOK.....</b>	<b>95</b>
5.1 Broodstock Sampling.....	95
5.2 Hatchery Rearing .....	102
5.3 Disease Monitoring.....	108
5.4 Natural Juvenile Productivity .....	109
5.5 Spawning Surveys.....	121
5.6 Carcass Surveys .....	127
5.7 Life History Monitoring.....	132
5.8 ESA/HCP Compliance.....	148
<b>SECTION 6: NASON CREEK SPRING CHINOOK.....</b>	<b>153</b>
6.1 Broodstock Sampling.....	153
6.2 Hatchery Rearing .....	155
6.3 Disease Monitoring.....	157
6.4 Natural Juvenile Productivity .....	158
6.5 Spawning Surveys.....	162
6.6 Carcass Surveys .....	164
6.7 Life History Monitoring.....	165
6.8 ESA/HCP Compliance.....	172

<b>SECTION 7: WHITE RIVER SPRING CHINOOK .....</b>	<b>175</b>
7.1 Captive Brood Collection .....	175
7.2 Hatchery Spawning and Release.....	177
7.3 Disease Monitoring.....	182
7.4 Natural Juvenile Productivity .....	182
7.5 Spawning Surveys.....	186
7.6 Carcass Surveys .....	188
7.7 Life History Monitoring.....	190
7.8 ESA/HCP Compliance.....	198
<b>SECTION 8: WENATCHEE SUMMER CHINOOK .....</b>	<b>201</b>
8.1 Broodstock Sampling.....	201
8.2 Hatchery Rearing .....	207
8.3 Disease Monitoring.....	213
8.4 Natural Juvenile Productivity .....	214
8.5 Spawning Surveys.....	215
8.6 Carcass Surveys .....	219
8.7 Life History Monitoring.....	224
8.8 ESA/HCP Compliance.....	235
<b>SECTION 9: METHOW SUMMER CHINOOK .....</b>	<b>237</b>
9.1 Broodstock Sampling.....	237
9.2 Hatchery Rearing .....	244
9.3 Disease Monitoring.....	250
9.4 Natural Juvenile Productivity .....	250
9.5 Spawning Surveys.....	251
9.6 Carcass Surveys .....	255
9.7 Life History Monitoring.....	259
9.8 ESA/HCP Compliance.....	271
<b>SECTION 10: OKANOGAN/SIMILKAMEEN SUMMER CHINOOK .....</b>	<b>273</b>
10.1 Broodstock Sampling.....	273
10.2 Hatchery Rearing .....	274
10.3 Disease Monitoring.....	280
10.4 Spawning Surveys.....	280
10.5 Carcass Surveys .....	282
10.6 Life History Monitoring.....	285
10.7 ESA/HCP Compliance.....	296
<b>SECTION 11: CHELAN FALLS SUMMER CHINOOK .....</b>	<b>297</b>
11.1 Broodstock Sampling.....	297
11.2 Hatchery Rearing .....	298
11.3 Spawning Surveys.....	305
11.4 Carcass Surveys .....	307
11.5 Life History Monitoring.....	310
11.6 ESA/HCP Compliance.....	320
<b>SECTION 12: REFERENCES.....</b>	<b>323</b>
<b>SECTION 13: APPENDICES.....</b>	<b>327</b>

## LIST OF APPENDICES

---

- Appendix A:** Abundance and Total Numbers of Chinook Salmon and Trout in the Chiwawa River Basin, Washington, 2014.
- Appendix B:** Fish Trapping at the Chiwawa and Wenatchee Smolt Traps during 2014.
- Appendix C:** Summary of CSS PIT-Tagging Activities in the Wenatchee River Basin, 2014.
- Appendix D:** Wenatchee Steelhead Spawning Escapement Estimates, 2014.
- Appendix E:** Examining the Genetic Structure of Wenatchee River Basin Steelhead and Evaluating the Effects of the Supplementation Program.
- Appendix F:** NPDES Hatchery Effluent Monitoring, 2014.
- Appendix G:** Steelhead Stock Assessment at Priest Rapids Dam, 2014.
- Appendix H:** Wenatchee Sockeye Salmon Spawning Escapement, 2014.
- Appendix I:** Genetic Diversity of Wenatchee Sockeye Salmon.
- Appendix J:** Genetic Diversity of Natural Chiwawa River Spring Chinook Salmon.
- Appendix K:** Fish Trapping at the Nason Creek Smolt Trap during 2014.
- Appendix L:** Fish Trapping at the White River Smolt Trap during 2014.
- Appendix M:** Genetic Diversity of Upper Columbia Summer Chinook Salmon.
- Appendix N:** Summer Chinook Spawning Ground Surveys in the Methow and Chelan Rivers, 2014.



## PREFACE

---

This annual report is the result of coordinated field efforts conducted by Washington Department of Fish and Wildlife (WDFW), the Confederated Tribes and Bands of the Yakama Nation (Yakama Nation), Chelan County Public Utility District (Chelan PUD), and BioAnalysts, Inc. An extensive amount of work was conducted in 2006 through 2014 to collect the data needed to monitor the effects of the Chelan and Grant County PUD Hatchery Programs. This work was directed and coordinated by the Habitat Conservation Plan (HCP) Hatchery Committees, consisting of the following members: Bill Gale, U.S. Fish and Wildlife Service (USFWS); Rob Jones, Craig Busack, and Lynn Hatcher, National Marine Fisheries Service (NMFS); Catherine Willard and Alene Underwood, Chelan PUD; Tom Scribner and Keely Murdoch, the Yakama Nation; Mike Tonseth, WDFW; Kirk Truscott, Confederated Tribes of the Colville Reservation (Colville Tribes), and Mike Schiewe, Anchor QEA (Chair). This report also includes monitoring efforts funded by Grant County Public Utility District (Grant PUD). Grant PUD helps fund the spring and summer Chinook monitoring programs. Work funded by Grant PUD was directed and coordinated by the Priest Rapids Coordinating Committee (PRCC) Hatchery Sub-Committee, which consists of the same agency and tribal representatives listed for the HCP Hatchery Committee and replaces Chelan PUD representatives with Grant PUD representatives, Todd Pearsons and Peter Graf.

The approach to monitoring the hatchery programs was guided by the updated monitoring and evaluation plan for PUD hatchery programs (Hillman et al. 2013). Technical aspects of the monitoring and evaluation program were developed by the Hatchery Evaluation Technical Team (HETT), which consisted of the following scientists: Carmen Andonaegui, WDFW; Matt Cooper, USFWS; Steve Hays, Chelan PUD; Tracy Hillman, BioAnalysts; Tom Kahler, Douglas PUD; Russell Langshaw, Grant PUD; Greg Mackey, Douglas PUD; Joe Miller, formerly Chelan PUD; Josh Murauskas, formerly Chelan PUD; Andrew Murdoch, WDFW; Keely Murdoch, Yakama Nation; Todd Pearsons, Grant PUD; and Mike Tonseth, WDFW. The updated plan also directs the analyses of hypotheses developed by the HETT. Most of the analyses outlined in the updated plan will be conducted in the five-year comprehensive reports.

Most of the work reported in this paper was funded by Chelan and Grant PUDs. Bonneville Power Administration purchased some of the Passive Integrated Transponder (PIT) tags that were used to mark juvenile Chinook and steelhead captured in tributaries and also helped fund a portion of the screw trap efforts in Nason Creek. We thank Charlie Paulsen for analyzing PIT-tag data for each program. This is the ninth annual report written under the direction of the HCP.

*“I often say that when you can measure something and express it in numbers, you know something about it. When you cannot measure it, when you cannot express it in numbers, your knowledge is of a meager and unsatisfactory kind. It may be the beginning of knowledge, but you have scarcely in your thoughts advanced to the stage of science, whatever it may be.”*

*Lord Kelvin*





## SECTION 1: INTRODUCTION

Chelan and Grant PUDs implement hatchery programs as part of their respective agreements related to the operation of Rocky Reach, Rock Island, Wanapum, and Priest Rapids Hydroelectric Projects. The fish resource management agencies developed the following general goal statements for the hatchery programs, which were adopted by the HCP Hatchery Committees and PRCC Hatchery Sub-Committee (hereafter, Hatchery Committees):

1. *Support the recovery of ESA-listed species by increasing the abundance of the natural adult population, while ensuring appropriate spatial distribution, genetic stock integrity, and adult spawner productivity.*

Includes the Wenatchee spring Chinook, Wenatchee summer steelhead, and Methow spring Chinook programs.

2. *Increase the abundance of the natural adult population of unlisted plan species, while ensuring appropriate spatial distribution, genetic stock integrity, and adult spawner productivity. In addition, provide harvest opportunities in years when spawning escapement is sufficient to support harvest.*

Includes the Wenatchee sockeye, Wenatchee summer/fall Chinook, Methow summer/fall Chinook, Okanogan summer/fall Chinook, and Okanogan sockeye programs.

3. *Provide salmon for harvest and increase harvest opportunities, while segregating returning adults from natural tributary spawning populations.*

Includes the Chelan Falls summer Chinook program.

Following the development of the Hatchery and Genetic Management Plans (HGMPs), artificial propagation programs are now characterized into three categories. The first type, integrated conservation programs, are intended to support or restore natural populations. These programs focus on increasing the natural production of targeted fish populations. A fundamental assumption of this strategy is that hatchery fish returning to the spawning grounds are reproductively similar to naturally produced fish. The second type, safety-net programs, are extensions of conservation programs, but are intended to function as reserve capacity for conservation programs in years of low returns. The safety-net provides a demographic and genetic reserve for the natural population. That is, in years of abundant returns, they function like segregated programs, and in years of low returns, they can be managed as conservation programs. Lastly, harvest augmentation programs are intended to increase harvest opportunities while limiting interactions with wild-origin counterparts.

Monitoring is needed to determine if the hatchery programs are meeting the intended management objectives of conservation, safety-net, or harvest augmentation programs. Objectives for hatchery programs are generally grouped into three categories of performance indicators:

1. In-Hatchery Indicators: Are the programs meeting the hatchery production objectives?
2. In-Nature Indicators: How do hatchery fish from the programs perform after release?

- a. Conservation Programs:
    - How do the programs affect target population abundance and productivity?
    - How do the programs affect target population long-term fitness?
  - b. Safety-Net Programs:
    - How do the programs affect target population long-term fitness?
  - c. Harvest Augmentation Programs:
    - Do the programs provide harvest opportunities?
3. Risk Assessment Indicators: Do the programs pose risks to other populations?

The specific objectives identified in the updated monitoring and evaluation plan are as follows:

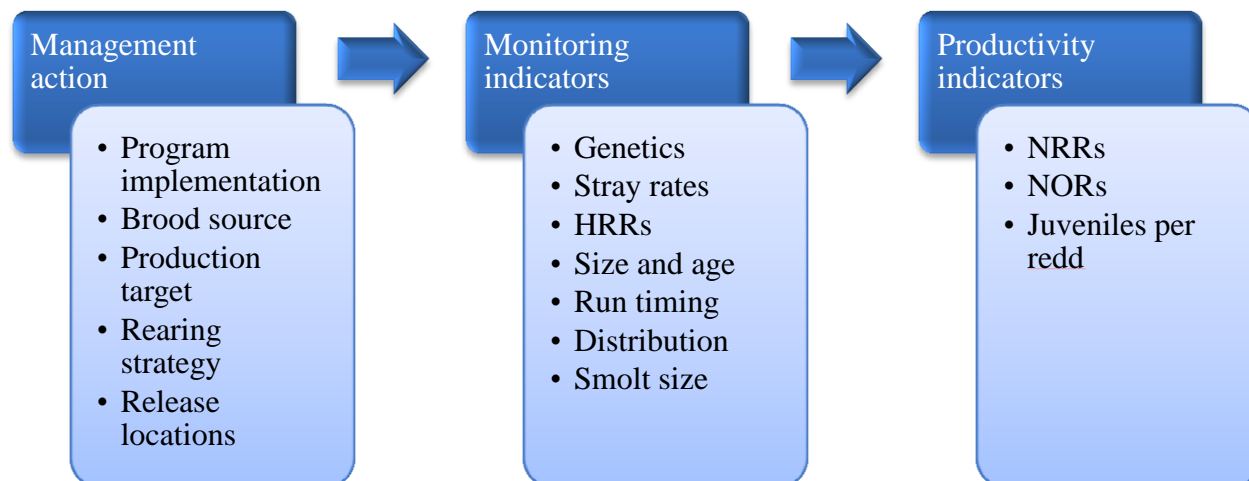
1. *Determine if conservation programs have increased the number of naturally spawning and naturally produced adults of the target population and if the program has reduced the natural replacement rate (NRR) of the supplemented population.*
2. *Determine if the proportion of hatchery fish on the spawning grounds affects the freshwater productivity of supplemented stocks.*
3. *Determine if the hatchery adult-to-adult survival (i.e., hatchery replacement rate, HRR) is greater than the natural adult-to-adult survival (i.e., natural replacement rate, NRR) and the target hatchery survival rate.*
4. *Determine if the proportion of hatchery-origin spawners (pHOS or PNI) is meeting management target.*
5. *Determine if the run timing, spawn timing, and spawning distribution of both the hatchery component is similar to the natural component of the target population or is meeting program-specific objectives.*
6. *Determine if stray rate of hatchery fish is below the acceptable levels to maintain genetic variation among stocks.*
7. *Determine if genetic diversity, population structure, and effective population size have changed in natural spawning populations as a result of the hatchery program.*
8. *Determine if hatchery programs have caused changes in phenotypic characteristics of natural populations.*
9. *Determine if hatchery fish were released at the programmed size and number.*
10. *Determine if appropriate harvest rates have been applied to conservation, safety-net, and segregated harvest programs to meet the HCP/SSSA goal of providing harvest opportunities while also contributing to population management and minimizing risk to natural populations*

Two additional regional objectives that were not explicit in the goals specified above but were included in the updated monitoring and evaluation plan because they relate to goals and concerns of all artificial production programs include:

11. Determine if the incidence of disease has increased in the natural and hatchery populations.
12. Determine if the release of hatchery fish affects non-target taxa of concern (NTTOC) within acceptable limits.

Objective 12 was completed using an extensive risk assessment that concluded risks from the PUD hatchery programs were within containment objectives approved by the Hatchery Committees (Mackey et al. 2014; Pearsons et al. 2012).

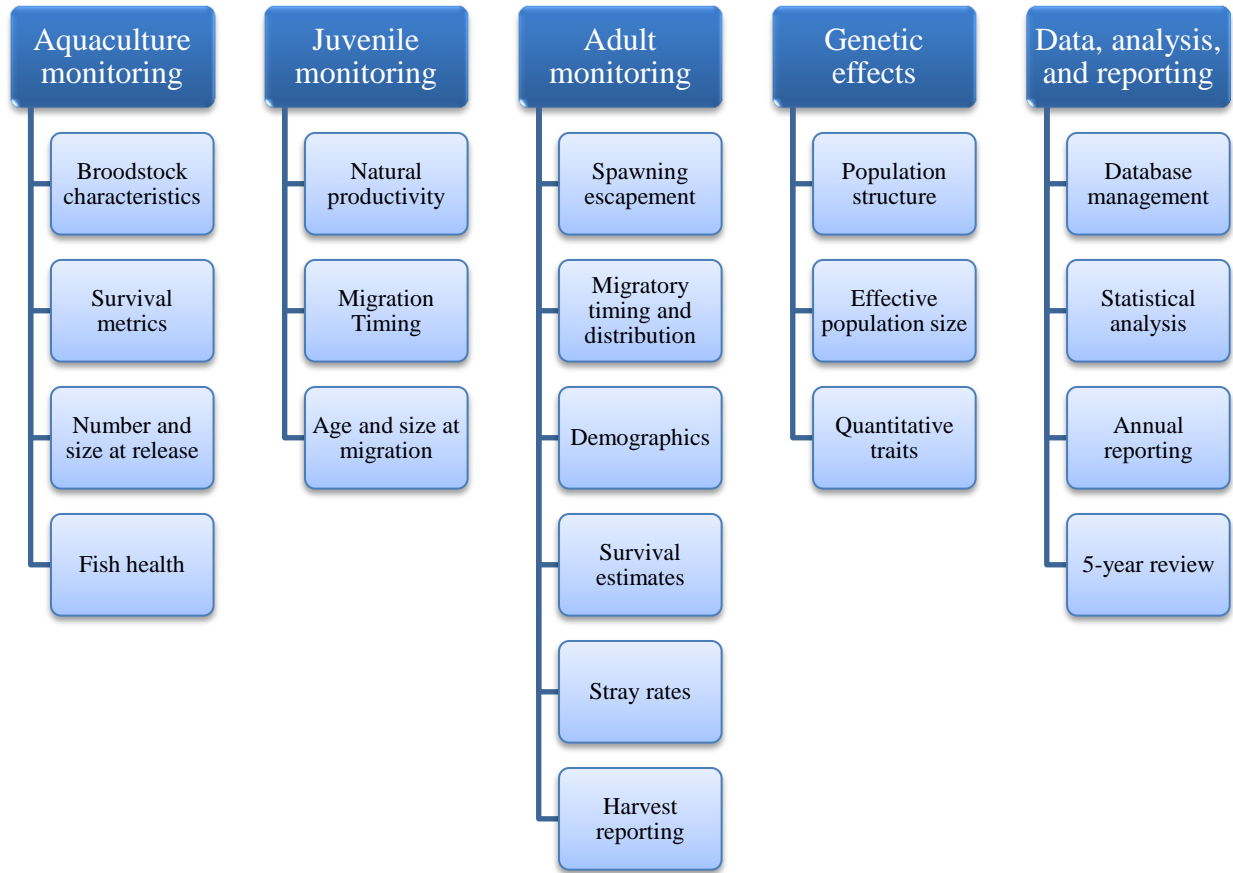
Objectives in the updated plan have been organized in a hierarchy where productivity indicators are the primary metrics used to assess if conservation and safety-net program goals have been met; harvest rates and effects on non-targeted populations are used for harvest programs. In cases where productivity indicators are not available, or results are equivocal, monitoring indicators may be used to help evaluate the performance of the program. Evaluations of monitoring indicators may not provide sufficiently powerful conclusions on which to base management actions; although they may provide insight as to why a productivity indicator did or did not meet the program goal. Therefore, the relationship between hatchery programs and indicators can be viewed in a chain-of-causation: management actions within the hatchery programs affect the status of monitoring indicators, which in turn influence productivity indicators (Figure 1.1).



**Figure 1.1.** Relationship of indicators to the assessment of propagation programs. Management actions affect monitoring indicators, which influence productivity indicators. Monitoring indicators may be used to hypothesize the magnitude of influence on productivity.

Attending each objective is one or more testable hypotheses (see Hillman et al. 2013). Each hypothesis will be tested statistically following the routines identified in the updated monitoring and evaluation plan. Most of these analytical routines will be conducted at the end of five-year monitoring blocks, as outlined in the updated plan.

Both monitoring and productivity indicators will be used to evaluate the success of the hatchery programs. In the event that the statistical power of tests that involve productivity indicators is insufficient to inform sound management decisions, some of the monitoring indicators may be used to guide management. Figure 1.2 shows the categories of indicators associated with each component of monitoring.



**Figure 1.2.** Overview of monitoring and evaluation plan categories and components (not including regional objectives).

Throughout each five-year monitoring period, annual reports will be generated that describe the monitoring and evaluation data collected during a specific year. This is the ninth annual report developed under the direction of the Hatchery Committees. The purpose of this report is to describe monitoring activities conducted in 2014. Activities included broodstock collection, collection of life-history information, within hatchery spawning and rearing activities, juvenile monitoring within streams, and redd and carcass surveys. Data from reference areas are not included in this annual report (reference data are in the five-year reports). To the extent currently possible, we have included information collected before 2014.

This report is divided into several sections, each representing a different species, stock, or spawning aggregate (i.e., steelhead, sockeye salmon, spring Chinook salmon, and summer Chinook salmon). For all species we provide annual broodstock information; hatchery rearing history, release data, and survival estimates; disease information; juvenile migration and

productivity estimates; redd counts, distribution, and spawn timing; spawning escapements; and life-history characteristics. For salmon species, we also provide information on carcasses. Brood year 2011 was the final sockeye salmon hatchery release, and beginning in 2013, only natural adult and juvenile sockeye productivity monitoring results are reported. Beginning in 2013, we added a separate section on Nason Creek spring Chinook salmon and in 2014 we added a separate section on White River spring Chinook salmon. The Colville Tribes began conducting monitoring of Okanogan summer Chinook in 2013 (in lieu of Chelan PUD); however, we retained the Okanogan summer Chinook section in this report. The Okanogan summer Chinook section includes monitoring information up to 2013. Monitoring results for 2013 and beyond can be found in annual reports prepared by the Colville Tribes to Bonneville Power Administration (BPA). Monitoring results of Grant PUD's fall Chinook salmon mitigation produced at Priest Rapids Hatchery can be found in annual reports written by WDFW and Grant PUD.

Finally, we end each section by addressing compliance issues with ESA/HCP mandates. For each Hatchery Program, WDFW and the PUDs are authorized annual take of ESA-listed spring Chinook and steelhead through Section 10 of the Endangered Species Act (ESA), including:

1. ESA Section 10(a)(1)(A) Permit No. 1395, which authorizes the annual take of adult and juvenile endangered upper Columbia River (UCR) spring Chinook and endangered UCR steelhead associated with implementing artificial propagation programs for the enhancement of UCR steelhead. The authorization includes takes associated with adult broodstock collection, hatchery operations, juvenile fish releases, monitoring and evaluation activities, and management of adult returns related to UCR steelhead artificial propagation programs in the UCR region (NMFS 2003a).
2. ESA Section 10(a)(1)(A) Permit No. 18121, which authorizes the annual take of adult and juvenile endangered UCR spring Chinook and endangered UCR steelhead associated with implementing artificial propagation programs in the Chiwawa River for the enhancement of UCR spring Chinook. The authorization includes takes associated with adult broodstock collection, hatchery operations, juvenile fish releases, and monitoring and evaluation activities supporting UCR spring Chinook artificial propagation programs in the UCR region (NMFS 2004).
3. ESA Section 10(a)(1)(A) Permit No. 18118, which authorizes the annual take of adult and juvenile endangered UCR spring Chinook and endangered UCR steelhead associated with implementing artificial propagation programs in Nason Creek for the enhancement of UCR spring Chinook. The authorization includes takes associated with adult broodstock collection, hatchery operations, juvenile fish releases, and monitoring and evaluation activities supporting UCR spring Chinook artificial propagation programs in the UCR region (NMFS 2004).
4. ESA Section 10(a)(1)(A) Permit No. 18120, which authorizes the annual take of adult and juvenile endangered UCR spring Chinook and endangered UCR steelhead associated with implementing artificial propagation programs in the White River for the enhancement of UCR spring Chinook. The authorization includes takes associated with adult broodstock collection, hatchery operations, juvenile fish releases, and monitoring and evaluation activities supporting UCR spring Chinook artificial propagation programs in the UCR region (NMFS 2004).

5. ESA Section 10(a)(1)(A) Permit No. 1347, which authorizes the annual incidental take of adult and juvenile endangered UCR spring Chinook and endangered UCR steelhead through actions associated with implementing artificial propagation programs for the enhancement of non-listed anadromous fish populations in the UCR. The authorization includes incidental takes associated with adult broodstock collection, hatchery operations, juvenile fish releases, and monitoring and evaluation activities associated with non-listed summer Chinook, fall Chinook, and sockeye salmon artificial propagation programs in the UCR region (NMFS 2003b).

## SECTION 2: SUMMARY OF METHODS

Sampling in 2014 followed the methods and protocols described in Hillman et al. (2013). In this section we only briefly review the methods and protocols. More detailed information can be found in the updated monitoring and evaluation plan (Hillman et al. 2013).

### 2.1 Broodstock Collection and Sampling

Methods for collecting broodstock are described in the Annual Broodstock Collection Protocols (Appendix A in WDFW 2014). Generally, broodstock were collected over the migration period (to the extent allowed in ESA-permit provisions) in proportion to their temporal occurrence at collection sites, with in-season adjustments dictated by 2014 run timing and trapping success relative to achieving weekly and annual collection objectives. Pre-season weekly collection objectives are shown in Table 2.1 and assumptions associated with broodstock trapping are provided in Table 2.2.

**Table 2.1.** Weekly collection objectives for steelhead and Chinook in 2014.

Collection week beginning day	Chiwawa/Nason Spring Chinook <sup>a</sup>		Hatchery Chelan Falls Summer Chinook	Wild Wenatchee Summer Chinook	Methow Summer Chinook	Wenatchee Steelhead	
	Hatchery	Wild				Hatchery	Wild
29 June			90	120	18	1	1
6 Jul			70	12	16	1	1
13 Jul			70	30	16	2	2
20 Jul			40	66	12	3	3
27 Jul			36	30	10	3	3
3 Aug			22	10	10	3	3
10 Aug				6	8	2	3
17 Aug				4	6	2	2
24 Aug					6	2	2
31 Aug					4	2	3
7 Sep						3	3
14 Sep						5	5
21 Sep						5	5
28 Sep						5	5
5 Oct						6	6
12 Oct						6	6
19 Oct						6	8
26 Oct						6	6
<b>Total</b>	<b>66</b>	<b>138</b>	<b>328</b>	<b>278</b>	<b>106</b>	<b>63</b>	<b>67</b>

<sup>a</sup> Chiwawa/Nason Spring Chinook were collected from the Chiwawa Weir, Nason Creek (via tangle netting), and Tumwater from the week of June 22 through the week of August 10. No specific weekly objectives were generated.

**Table 2.2.** Biological and trapping assumptions associated with collecting broodstock for the Chelan and Grant PUD Hatchery Programs.<sup>1</sup>

Assumptions	Wenatchee Steelhead	Chiwawa Spring Chinook	Nason Spring Chinook (Conservation)	Nason Spring Chinook (Safety Net)	Wenatchee Summer Chinook	Methow Summer Chinook
Production level	247,300 yearling smolts	144,026 yearling smolts	125,000 yearling smolts	98,670 yearling smolts	500,001 yearling smolts	200,000 yearling smolts
Broodstock required	130 adults (not to exceed 33% of population)	74 adults (not to exceed 33% of population)	64 adults (not to exceed 33% of population)	66 adults	278 adults (not to exceed 33% of the population)	100 adults (not to exceed 33% of the population)
Trapping period	1 July-15 Nov	1 May – 15 July (Tumwater) 15 June-1 Aug (Chiwawa Weir)	23 June – 17 Aug	1 May – 15 July	1 July – 15 Sept	29 June – 30 Aug
# days/week	5	7 (Tumwater) Not to exceed 15 cumulative trapping days (Chiwawa Weir)	5	7	5	3
# hours/day	24	24 (Tumwater) 24 up/24 down (Chiwawa Weir)	24	24	24	16
Broodstock composition	50% HxH; 50% WxW	Sliding scale; minimum 33% wild (depends on the number of wild fish)	100% wild	100% hatchery	100% wild	100% wild
Trapping site	Dryden Dam for HxH; Tumwater for WxW. (Tumwater will be used if weekly quota not achieved for HxH at Dryden Dam; Dryden Dam will be used if weekly quota not achieved for WxW at Tumwater)	Tumwater Dam and Chiwawa Weir	Nason Creek Tangle Netting	Tumwater Dam	Dryden Dam (Tumwater will be used if weekly quota not achieved at Dryden Dam)	Wells Dam east or west ladder

Several biological parameters were measured during broodstock collection at adult collection sites. Those parameters included the date and start and stop time of trapping; number of each

<sup>1</sup> Throughout this document, “HxH” refers to hatchery by hatchery crosses and “WxW” refers to wild by wild crosses.



species collected for broodstock; origin, size, and sex of trapped fish; age from scale analysis; and pre-spawn mortality. For each species, trap efficiency, extraction rate, and trap operation effectiveness were estimated following procedures in Hillman et al. (2013). In addition, a representative sample of most species trapped but not taken for broodstock were sampled for origin, sex, age, and size (stock assessment).

## 2.2 Within Hatchery Monitoring

Methods for monitoring hatchery activities are described in Hillman et al. (2013). Biological information collected from all spawned adult fish included age at maturity, length at maturity, spawn time, and fecundity of females. In addition, all fish were checked for tags and females were sampled for disease.

Throughout the rearing period in the hatchery, fish were sampled for growth, health, and survival. Each month, lengths and weights were collected from a sample of fish and rearing density indices were calculated. In addition, fish were examined monthly for health problems following standard fish health monitoring practices for hatcheries. Various life-stage survivals were estimated for each hatchery stock. These estimates were then compared to the “standard” survival rates identified in Table 2.3 to provide insight as to how well the hatchery operations were performing. Failure to achieve a survival standard could indicate a problem with some part of the hatchery program. However, failure to meet a standard may not be indicative of the overall success of the program to meet the goals identified in Section 1.

**Table 2.3.** Standard life-stage survival rates for fish reared within the Chelan PUD hatchery programs (from Hillman et al. 2013).

Life stage	Standard survival rate (%)
Collection-to-spawning (females)	90
Collection-to-spawning (males)	85
Unfertilized egg-to-eyed	92
Unfertilized egg-to-ponding	98
30 d after ponding	97
100 d after ponding	93
Ponding-to-release	90
Transport-to-release	95
Unfertilized egg-to-release	81

Nearly all hatchery fish from each stock were marked (adipose fin clip) or tagged (coded-wire tag) in 2014. Different combinations of marks and tags were used depending on the stock. In addition, in 2014, Chelan PUD personnel PIT tagged 10,114 juvenile hatchery Chiwawa spring Chinook and 20,234 juvenile Nason Creek spring Chinook in August; 15,180 Wenatchee steelhead (5,051 WxW steelhead and 10,129 HxH steelhead) during September; and 10,000 Chelan River summer Chinook in March, 10,159 Methow (Carlton) summer Chinook in August, and 20,641 Wenatchee summer Chinook in September, November, and December. PIT tags will be used to estimate migration timing and survival rates (e.g., smolt-to-adult) outside the hatchery.

Lastly, the size and number of fish released were assessed and compared to programmed production levels. The goal of the program is that numbers released and their sizes should fall within 10% of the programmed targets identified in Table 2.4. However, because of constraints due to run size and proportions of wild and hatchery adults, production levels may not be met every year.

**Table 2.4.** Targets for fish released from the PUD hatchery programs; CV = coefficient of variation.

Hatchery stock	Release targets	Size targets		
		Fork length (CV)	Weight (g)	Fish/pound
Wenatchee Summer Chinook	500,001	163 (9.0)	45.4	10 <sup>a</sup>
Methow Summer Chinook	200,000	163 (9.0)	45.4	15
Chelan Falls Summer Chinook (yearlings)	576,000	161 (9.0)	45.4	10 <sup>b</sup>
Chiwawa Spring Chinook	144,026	155 (9.0)	37.8	18
Nason Spring Chinook	223,670	155 (9.0)	37.8	24
Wenatchee Steelhead	247,300	191 (9.0)	75.6	6

<sup>a</sup>An experimental release size of 30-45 grams (10-15 FPP) is in place for brood years 2012-2014.

<sup>b</sup>An experimental release size of 20-45 grams (10-22 FPP) is in place for brood years 2012-2014.

## 2.3 Juvenile Sampling

Juvenile sampling within streams included operation of rotary smolt traps, snorkel observations, and PIT tagging. Methods for sampling juvenile fish are described in Hillman et al. (2013).

A smolt trap was located on the Wenatchee River near the town of Cashmere at RM 8.3 (Lower Wenatchee Trap), in Nason Creek about 0.6 miles upstream from the mouth, in the White River, and in the Chiwawa River about 0.7 miles upstream from the mouth (Chiwawa Trap). All traps operated throughout the smolt migration period. The Chiwawa River, White River, and Nason Creek traps operated throughout most of the year (March through November), but not during icing or extreme high flow conditions. The following data were collected at each trap site: water temperature, discharge, number and identification of all species captured, degree of smoltification for anadromous fish, presence of marks and tags, size (fork lengths and weights), and scales from smolts. Trap efficiencies at each trap site were estimated by using mark-recapture trials conducted over a wide range of discharges. Linear regression models relating discharge and trap efficiencies were developed to estimate daily trap efficiencies during periods when no mark-recapture trials were conducted. The total number of fish migrating past the trap each day was estimated as the quotient of the daily number of fish captured and the estimated daily trap efficiency. Summing the daily totals resulted in the total emigration estimate.

Snorkel observations were used to estimate the number of juvenile spring Chinook salmon, juvenile rainbow/steelhead, and bull trout within the Chiwawa River basin. The focus of the study was on juvenile spring Chinook salmon. Sampling followed a stratified random design with proportional allocation of sites among strata. Strata were identified based on unique combinations of geology, land type, valley bottom type, stream state condition, and habitat types. A total of 161 randomly selected sites were surveyed during August (Table 2.5). Counts of fish within each sampling site were adjusted based on detection efficiencies, which were related to water temperature. That is, non-linear models that described relationships between water

temperatures and detection efficiencies (Hillman et al. 1992) were used to estimate total numbers of fish within sampling sites. These numbers were then converted to densities by dividing total fish numbers by the wetted surface area and water volume of sample sites. Total numbers within a stratum were estimated as the product of fish densities times the total wetted surface or water volume for the stratum. The sum of fish numbers across strata resulted in the total number of fish within the basin. The calculation of total numbers, densities, and degrees of certainty are fully explained in Hillman and Miller (2004).

Working in collaboration with the Comparative Survival Study (CSS) funded by BPA, crews PIT tagged juvenile wild Chinook, wild and hatchery steelhead, and wild sockeye salmon collected at the smolt traps and collected within the Chiwawa River and Nason Creek using electrofishing techniques. The proposed number of wild spring Chinook and steelhead to be tagged at each location is provided in Table 2.6. The goal of this tagging program is to estimate freshwater juvenile productivity, better understand life-history characteristics, overwinter movement and survival of salmonids, and to calculate SARs of Chinook salmon in the Wenatchee River basin. The PIT tagging effort funded by the PUDs in the Chiwawa River and Nason Creek is specifically directed at addressing uncertainties of estimating abundance using screw traps (e.g., fish passage during times when trapping is not possible).

**Table 2.5.** Location of strata and numbers of randomly sampled snorkel sites within each stratum that were sampled in the Chiwawa River Basin in 2014.

Reach/stratum	River miles (RM)	Number of randomly selected sites
<b>Chiwawa River</b>		
1	0.0-3.8	11
2	3.8-5.5	5
3	5.5-7.9	8
4	7.9-8.9	6
5	8.9-10.8	5
6	10.8-11.8	6
7	11.8-20.0	28
8	20.0-25.4	24
9	25.4-28.8	11
10	28.8-31.1	11
<b>Phelps Creek</b>		
1	0.0-0.4	1
<b>Chikamin Creek (includes Minnow Creek)</b>		
1	0.0-1.5	15
<b>Rock Creek</b>		
1	0.0-0.7	14
<b>Unnamed stream on USGS map</b>		
1	0.0-0.1	1
<b>Big Meadow Creek</b>		
1	0.0-1.0	7
<b>Alder Creek</b>		

Reach/stratum	River miles (RM)	Number of randomly selected sites
1	0.0-0.1	4
<b>Brush Creek</b>		
1	0.0-0.1	2
<b>Clear Creek</b>		
1	0.0-0.1	2

**Table 2.6.** Number of wild spring Chinook and steelhead proposed for tagging at different locations within the Wenatchee River basin, 2014.

Sampling location	Target sample size		
	Wild spring Chinook	Wild steelhead	Wild Sockeye
Chiwawa Trap	2,500-8,000	500-2,000	NA
Nason Creek Trap	2,500-8,000	500-2,000	NA
Lower Wenatchee Trap	500-1,000	50-250	3,000-5,000
Chiwawa Remote Sampling	3,000	NA	NA
Nason Remote Sampling	3,000	NA	NA

Survival rates for various juvenile life-stages were calculated based on estimates of seeding levels (total egg deposition), parr abundance, numbers of emigrants, and smolt abundance. Total egg deposition was estimated as the product of the number of redds counted in the basin times the mean fecundity of female spawners. Fecundity was estimated from females collected for broodstock using an electronic egg counter. Numbers of emigrants and smolts were estimated at trapping sites and numbers of parr were estimated using snorkel observations only in the Chiwawa River basin. Survival estimates could not be calculated for some stocks (e.g., summer Chinook) because specific life-stage abundance estimates were lacking.

## 2.4 Spawning/Carcass Surveys

Methods for conducting carcass and spawning ground surveys are detailed in Hillman et al. (2013). Information collected during spawning surveys included spawn time, redd distribution, and redd abundance. Data collected during carcass surveys included sex, size (fork length and postorbital-to-hypural length), scales for aging<sup>2</sup>, degree of egg voidance, DNA samples, and identification of marks or tags. The sampling goal for carcasses was 20% of the spawning population. Crews also conducted snorkel surveys to assess the incidence of precociously maturing fish spawning naturally in streams.

Steelhead surveys were conducted in major and minor spawning areas in the mainstem Wenatchee River and downstream from PIT-tag interrogation systems on the Chiwawa River,

<sup>2</sup> In this report we use two methods of describing age. One is termed the “European Method.” This method has two digits, separated by a period. The first digit represents the number of winters the fish spent in freshwater before migrating to the sea. The second digit indicates the number of winters the fish spent in the ocean. For example, a fish designated as 1.2 spent one winter in freshwater and two in the ocean. A fish designated as 0.3 migrated to the ocean in its first year and spent three winters in the ocean. The other method describes the total age of the fish (egg-to-spawning adult, i.e., gravel-to-gravel), so fish demarcated as 0.3 or 1.2 are considered 4-year-olds, from the same brood.

Nason Creek, and Peshastin Creek. These surveys were conducted during March through June in reaches and index areas described in Table 2.7. Total redd counts in these reaches were estimated by expanding counts within non-index areas by expansion factors developed within index areas.

**Table 2.7.** Description of reaches and index areas surveyed for steelhead redds in the Wenatchee River basin.

Stream	Code	Reach*	Index/reference area
Wenatchee River	W2	Sleepy Hollow Br to L. Cashmere Br	Sleepy Hollow Br to Cashmere Boat Rmp
	W6	Leavenworth Br to Icicle Rd Br	Leavenworth Boat Ramp to Icicle Ck
	W8	Tumwater Dam to Tumwater Br	Island below Swiftwater to Swiftwater CG
	W9	Tumwater Br to Chiwawa R	Tumwater Br to Plain
	W10	Chiwawa R to Lk Wenatchee	Chiwawa Pump St. to Lk Wenatchee
Peshastin Creek	P1	Mouth to PIT Detection Site	Mouth to PIT Detection Site
Chiwawa River	C1	Mouth to Rd 62 Br RM 6.4	Mouth to PIT Detection Site
Nason Creek	N1	Mouth to PIT Detection Site	Mouth to PIT Detection Site

\* Reaches 2, 6, 8, 9, and 10 (major spawning areas) are surveyed weekly, while Reaches 1, 3, and 5 (minor survey areas) are surveyed during peak spawning.

Beginning in 2014, adult steelhead escapement estimates in the majority of tributaries in the Wenatchee River basin were generated using mark-recapture techniques based on steelhead PIT tagged at Priest Rapids Dam. Mark-recapture estimates in the tributaries were then added to the estimates based on redd surveys to generate a total spawning escapement to the Wenatchee River basin.

Spring Chinook redd and carcass surveys were conducted during August through September in the Chiwawa River (including Rock and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek (including Ingalls Creek, but not in 2014 because of wildfires), upper Wenatchee River, Little Wenatchee River, and the White River (including the Napeequa River and Panther Creek). Survey reaches for spring Chinook are described in Table 2.8.

**Table 2.8.** Description of reaches surveyed for spring Chinook redds and carcasses in the Wenatchee River basin.

Stream	Code	Reach	River mile (RM)
Chiwawa River	C1	Mouth to Grouse Creek	0.0-11.7
	C2	Grouse Creek to Rock Creek	11.7-19.3
	C3	Rock Creek to Schaefer Creek	19.3-22.4
	C4	Schaefer Creek to Atkinson Flats	22.4-25.6
	C5	Atkinson Flats to Maple Creek	25.6-27.0
	C6	Maple Creek to Phelps Creek	27.0-30.3
	C7	Phelps Creek to Buck Creek	30.3-31.4
Rock Creek	R1	Mouth to Chiwawa River Road Bridge	0.0-0.5
Chikamin Creek	K1	Mouth to Chiwawa River Road Bridge	0.0-0.5
Nason Creek	N1	Mouth to Kahler Creek Bridge	0.0-3.9
	N2	Kahler Creek Bridge to Hwy 2 Bridge	3.9-8.3
	N3	Hwy 2 Bridge to Lower RR Bridge	8.3-13.2

Stream	Code	Reach	River mile (RM)
	N4	Lower RR Bridge to Whitepine Creek	13.2-15.4
Little Wenatchee River	L2	Old Fish Weir to Lost Creek	2.7-5.2
	L3	Lost Creek to Rainy Creek	5.2-9.2
	L4	Rainy Creek to Falls	9.2-Falls
White River	H2	Sears Creek Bridge to Napeequa River	6.4-11.0
	H3	Napeequa River to Grasshopper Meadows	11.0-12.9
	H4	Grasshopper Meadows to Falls	12.9-16.1
Napeequa River	Q1	Mouth to Take Out	0.0-1.0
Panther Creek	T1	Mouth to Boulder Field	0.0-1.0
Wenatchee River	W8	Tumwater Dam to Tumwater Bridge	30.9-35.6
	W9	Tumwater Bridge to Chiwawa River	35.6-48.4
	W10	Chiwawa River to Lake Wenatchee	48.4-54.2
Chiwaukum Creek	U1	Mouth to Metal Bridge	0.0-1.0
Icicle Creek	I1	Mouth to Hatchery	0.0-2.8
	I2	Hatchery to Sleeping Lady	2.8-3.3
	I3	Sleeping Lady to Snow Creek	3.3-3.8
Peshastin Creek	P1	Mouth to Camas Creek	0.0-5.9
	P2	Camas Creek to Mouth of Scotty Creek	5.9-16.3
Ingalls Creek	D1	Mouth to Trailhead	0.0-1.0

The sockeye salmon hatchery program ended after the 2011 brood year. As a result, monitoring activities that focused on evaluating the effects of the supplementation program on the natural population switched to monitoring the abundance and productivity of the natural population (McElhaney et al. 2000). Thus, estimation of spawn time and carcass surveys were discontinued in 2014. Nevertheless, this report retains the results of carcass sampling during the period 1993-2013. Survey reaches in which carcasses and live fish (for area-under-the-curve estimates) were conducted are identified in Table 2.9.

From 2009-2013, mark-recapture methods were used to estimate spawning escapement within the White River, while area-under-the-curve (AUC) methods were used to estimate spawning escapement within the Little Wenatchee River. Beginning in 2014, mark-recapture methods were used to estimate the spawning escapement of sockeye in both the White River and Little Wenatchee watersheds.

**Table 2.9.** Description of reaches surveyed for sockeye salmon carcasses and live fish in the Wenatchee River basin during survey years 1993-2013.

Stream	Code	Reach	River mile (RM)
Little Wenatchee River	L1	Mouth to Old Fish Weir	0.0-2.7
	L2	Old Fish Weir to Lost Creek	2.7-5.2
	L3	Lost Creek to Rainy Creek	5.2-9.2
White River	H1	Mouth to Sears Creek Bridge	0.0-6.4
	H2	Sears Creek Bridge to Napeequa River	6.4-11.0

Stream	Code	Reach	River mile (RM)
	H3	Napeequa River to Grasshopper Meadows	11.0-12.9
Napeequa River	Q1	Mouth to End	0.0-1.0

Wenatchee summer Chinook redd and carcass surveys were conducted from September through November throughout the entire mainstem Wenatchee River, which was divided into ten reaches (Table 2.10). Surveys were conducted weekly in all reaches. All redds were enumerated during weekly census counts.

**Table 2.10.** Description of reaches and index areas surveyed for summer Chinook redds in the Wenatchee River basin.

Code	Reach	River mile	Index/reference area (RM)
W1	Mouth to Sleepy Hollow Br	0.0-3.3	River Bend to Sleepy Hollow Br (1.7-3.3)
W2	Sleepy Hollow Br to L. Cashmere Br	3.3-9.5	L. Cashmere Br to Old Monitor Br (7.1-9.5)
W3	L. Cashmere Br to Dryden Dam	9.5-17.8	Williams Canyon to Dryden Dam (15.5-17.8)
W4	Dryden Dam to Peshastin Br	17.8-20.0	Dryden Dam to Peshastin Br (17.8-20.0)
W5	Peshastin Br to Leavenworth Br	20.0-23.9	Irrigation Flume to Leavenworth Br (22.8-23.9)
W6	Leavenworth Br to Icicle Rd Br	23.9-26.4	Icicle to Boat Takeout (24.5-25.6)
W7	Icicle Rd Br to Tumwater Dam	26.4-30.9	Icicle Br to Penstock Br (26.4-28.7)
W8	Tumwater Dam to Tumwater Br	30.9-35.6	Swiftwater Campgd to Tumwater Br (33.5-35.6)
W9	Tumwater Br to Chiwawa River	35.6-47.9	Swing Pool to Railroad Tunnel (36.7-39.3)
W10	Chiwawa River to Lake Wenatchee	47.9-54.2	Swamp to Bridge (52.7-53.6)

Summer Chinook redd and carcass surveys were also conducted in the Methow and Chelan rivers from September through November. Total (map) redd counts were conducted in these rivers. Table 2.11 describes the survey reaches on the Methow River. The Colville Tribes conducted summer Chinook redd and carcass surveys in the Okanogan River basin. Those results are reported in a separate report (annual report to BPA).

**Table 2.11.** Description of reaches surveyed for summer Chinook redds and carcasses on the Methow, Okanogan, and Similkameen rivers.

Stream	Code	Reach	River mile (RM)
Methow River	M1	Mouth to Methow Bridge	0.0-14.8
	M2	Methow Bridge to Carlton Bridge	14.8-27.2
	M3	Carlton Bridge to Twisp Bridge	27.2-39.6
	M4	Twisp Bridge to MVID	39.6-44.9
	M5	MVID to Winthrop Bridge	44.9-49.8
	M6	Winthrop Bridge to Hatchery Dam	49.8-51.6
Okanogan River	O1	Mouth to Mallot Bridge	0.0-16.9
	O2	Mallot Bridge to Okanogan Bridge	16.9-26.1
	O3	Okanogan Bridge to Omak Bridge	26.1-30.7
	O4	Omak Bridge to Riverside Bridge	30.7-40.7

Stream	Code	Reach	River mile (RM)
	O5	Riverside Bridge to Tonasket Bridge	40.7-56.8
	O6	Tonasket Bridge to Zosel Dam	56.8-77.4
Similkameen River	S1	Driscoll Channel to Oroville Bridge	0.0-1.8
	S2	Oroville Bridge to Enloe Dam	1.8-5.7

For summer and spring Chinook, total spawning escapements for each population were estimated as the product of total number of redds times the ratio of fish per redd for a specific stock. Fish per redd ratios were estimated as the ratio of males to females sampled at broodstock collection sites and monitoring sites. For steelhead, spawning escapement was estimated with a combination of PIT-tag-based tributary and redd-based mainstem Wenatchee River estimates. Total spawning escapement for sockeye salmon in the Little Wenatchee and White River watersheds was estimated using mark-recapture methods. Adult sockeye were PIT tagged at Tumwater Dam and Bonneville Dam<sup>3</sup> and detected in the Little Wenatchee and White rivers with stationary PIT-tag interrogation systems.

Derived metrics calculated from carcass surveys, broodstock sampling, stock assessments, and harvest records included proportion of hatchery spawners, stray rates, age-at-maturity, length-at-age, smolt-to-adult survival (SAR), hatchery replacement rates (HRR), harvest rates, and natural replacement rates (NRR). The expected SARs and HRRs (from Peven and Murdoch 2005) for different stocks raised in the PUD hatchery programs are provided in Table 2.12. Methods for calculating these variables are described in Hillman et al. (2013) and in “White Papers” developed by the Hatchery Evaluation Technical Team (HETT) (see Appendices in Hillman et al. 2012).

**Table 2.12.** Expected smolt-to-adult (SAR) and hatchery replacement rates (HRR) for stocks raised in the PUD Hatchery Programs.

Program	Number of broodstock	Smolts released	SAR	Adult equivalents	Number of smolts/adult	HRR
Chiwawa Spring Chinook	74	144,026	0.003	432	333	5.8
Nason Creek Spring Chinook	66	125,000	0.003	375	333	5.7
Wenatchee Summer Chinook	278	500,001	0.003	1,500	333	5.4
Methow Summer Chinook	100	200,000	0.003	600	333	6.0
Wenatchee Steelhead	130	247,300	0.010	2,473	100	19.0

Derived data that rely on CWTs (e.g., HRR, SAR, stray rates, etc.) are five or more years behind release information because of the lag time for returning adult fish to enter the fishery and spawning grounds, and the processing of tags. Consequently, complete information on rates and ratios based on CWTs is generally only available for brood years before 2008.

<sup>3</sup>Adult sockeye that were tagged at Bonneville Dam and detected at Tumwater Dam were included in the mark-recapture analyses.



## SECTION 3: WENATCHEE STEELHEAD

The goal of summer steelhead supplementation in the Wenatchee Basin is to use artificial production to replace adult production lost because of mortality at Rock Island and Rocky Reach dams, as well as inundation compensation for Rocky Reach Dam, while not reducing the natural production or long-term fitness of steelhead in the basin. The Rock Island Fish Hatchery Complex began operation in 1989 under funding from Chelan PUD. The Complex operated originally through the Rock Island Settlement Agreement, but since 2004 has operated under the Rock Island and Rocky Reach Anadromous Fish Agreement and Habitat Conservation Plans.

Prior to 1998, steelhead eggs were received from Wells Hatchery (adult broodstock were collected at Wells Dam); fish were reared at Eastbank Fish Hatchery and then released into the Wenatchee River. Beginning in 1998, the program changed to collecting broodstock within the Wenatchee Basin. Currently, HxH adult steelhead are collected from the run-at-large at the right and left-bank traps at Dryden Dam, and at Tumwater Dam if the weekly quotas cannot be achieved at Dryden Dam. Wild by wild (WxW) adult steelhead are collected from the run-at-large at Tumwater and Dryden dams if the weekly quotas cannot be achieved at Dryden Dam.

Before 2012, the goal was to collect up to 208 adult steelhead (50% natural-origin fish and 50% hatchery-origin fish) for the Wenatchee steelhead program. In 2011, the Hatchery Committees reevaluated the amount of hatchery compensation needed to achieve NNI. Based on that evaluation, the goal of the program was revised. The current goal (beginning in 2012) is to collect 130 adult steelhead (64 natural-origin and 66 hatchery-origin fish) for a 247,300 smolt program, but the number of broodstock collected cannot exceed 33% of the natural Wenatchee steelhead population. Broodstock collection occurs from about 1 July through 15 November at Dryden Dam and 1 September through 15 November at Tumwater Dam, with trapping occurring up to 24 hours per day, five days a week, at Dryden Dam left and right-bank traps and at Tumwater Dam. The intent of the current program is to target adults necessary to meet a 50% natural-origin, conservation-oriented program and a 50% hatchery-origin safety-net program.

Prior to the 2012 brood year, adult steelhead were held and spawned at Wells Fish Hatchery because of unsuitable adult holding temperatures at Eastbank Fish Hatchery. Beginning with the 2012 brood year, spawning has occurred at Eastbank Fish Hatchery. Before 2012, juvenile steelhead were reared at a combination of facilities including Eastbank, Chelan, Turtle Rock, Rocky Reach Annex, and Chiwawa facilities. Juvenile steelhead reared in these facilities were trucked to release locations on the Wenatchee River, Chiwawa River, and Nason Creek. A percentage of the fish have also been released voluntarily from Blackbird Pond and Roling Pond. Beginning in the fall of 2012, the entire Wenatchee steelhead program overwinters at the Chiwawa Acclimation Facility. Some of these fish are transferred to short-term remote acclimation sites (e.g., Blackbird Pond and Roling Pond), while others are planted from trucks throughout the Wenatchee, Nason, and Chiwawa basins.

Before 2012, the production goal for the Wenatchee steelhead supplementation program was to release 400,000 yearling smolts into the Wenatchee Basin at six fish per pound. Since 2012, the revised production goal is to release 247,300 smolts (123,650 for conservation and 123,650 for safety net). Targets for fork length and weight are 191 mm (CV = 9.0) and 75.6 g, respectively; the target size at release is six fish per pound. Over 96% of these fish receive CWTs. In addition,

since 2006, juvenile steelhead from different parental-cross groups (e.g., WxW, HxW, and HxH) have been PIT tagged annually.

### 3.1 Broodstock Sampling

This section focuses on results from sampling 2013 and 2014 brood years of Wenatchee steelhead, which were collected at Dryden and Tumwater dams. The 2013 brood begins the tracking of the life cycle of steelhead released in 2014. The 2014 brood is included because juveniles from this brood are still maintained within the hatchery.

#### Origin of Broodstock

A total of 147 Wenatchee steelhead from the 2012 return (2013 brood) were collected at Dryden and Tumwater dams (Table 3.1). About 43% of these were natural-origin (adipose fin present, no CWT, and no elastomer tags) fish and the remaining 57% were hatchery-origin (elastomer tagged and/or adipose fin absent) adults. Origin was determined by analyzing scales and/or otoliths. The total number of steelhead spawned from the 2013 brood was 117 adults (42% natural-origin and 58% hatchery-origin).

A total of 135 steelhead were collected from the 2013 return (2014 brood) at Dryden and Tumwater dams; 65 (48%) natural-origin (adipose fin present, no CWT, and no elastomer tags) and 70 (52%) hatchery-origin (elastomer tagged and adipose present or CWT and adipose fin present) adults. A total of 132 steelhead were spawned; 48.5% were natural-origin fish and 51.5% were hatchery fish (Table 3.1). Origin was confirmed by sampling scales and/or otoliths.

**Table 3.1.** Numbers of wild and hatchery steelhead collected for broodstock, numbers that died before spawning, and numbers of steelhead spawned, 1998-2014. Unknown origin fish (i.e., undetermined by scale analysis, no elastomer, CWT, or fin clips, and no additional hatchery marks) were considered naturally produced. Mortality includes fish that died of natural causes, fish killed at spawning, and surplus broodstock.

Brood year	Wild steelhead					Hatchery steelhead					Total number spawned
	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	
1998	35	0	0	35	0	43	4	2	37	0	72
1999	58	5	1	52	0	67	1	2	64	0	116
2000	39	2	1	36	0	101	9	12	60	20	96
2001	64	5	8	51	0	114	5	6	103	0	154
2002	99	0	1	96	2	113	1	0	64	48	160
2003	63	10	4	49	0	92	2	0	90	0	139
2004	85	3	0	75	7	132	1	0	61	70	136
2005	95	8	0	87	0	114	7	1	104	2	191
2006	101	5	0	93	3	98	0	0	69	29	162
2007	79	0	2	76	1	97	0	14	58	25	134
2008	104	0	3	77	22	107	0	28	54	25	131
2009	101	2	0	86	13	107	1	4	73	29	159
2010	106	1	1	96	8	105	2	23	75	5	171
2011	104	8	1	91	4	104	13	2	70	0	161
<i>Average<sup>b</sup></i>	<i>81</i>	<i>4</i>	<i>2</i>	<i>71</i>	<i>4</i>	<i>100</i>	<i>3</i>	<i>7</i>	<i>70</i>	<i>18</i>	<i>142</i>
2012	63	3	0	59	1	66	0	1	65	0	124
2013	63	8	1	49	5	84	9	7	68	0	117

Brood year	Wild steelhead					Hatchery steelhead					Total number spawned
	Number collected	Prespawm loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawm loss <sup>a</sup>	Mortality	Number spawned	Number released	
2014	65	0	1	64	0	70	68	2	68	0	132
Average <sup>c</sup>	64	4	1	57	2	73	26	3	67	0	124

<sup>a</sup> Pre-spawn loss represents the number of fish that died during the holding period before spawning. Mortality is the number of fish that were surplus following spawning.

<sup>b</sup> This average represents the program before recalculation in 2011.

<sup>c</sup> This average represents the current program, which began in 2012.

### Age/Length Data

Broodstock ages were determined from examination of scales and/or otoliths. For the 2013 brood year, both natural-origin and hatchery steelhead consisted primarily of 2-salt adults (Table 3.2). For the 2014 brood year, both hatchery and natural-origin steelhead consisted primarily of 2-salt adults (Table 3.2).

**Table 3.2.** Percent of hatchery and wild steelhead of different ages (saltwater ages) collected from broodstock, 1998-2014.

Brood year	Origin	Saltwater age		
		1	2	3
1998	Wild	39.4	60.6	0.0
	Hatchery	20.9	79.1	0.0
1999	Wild	50.0	48.3	1.7
	Hatchery	81.8	18.2	0.0
2000	Wild	56.4	43.6	0.0
	Hatchery	67.9	32.1	0.0
2001	Wild	51.7	48.3	0.0
	Hatchery	14.9	85.1	0.0
2002	Wild	55.6	44.4	0.0
	Hatchery	94.6	5.4	0.0
2003	Wild	13.1	85.3	1.6
	Hatchery	29.4	70.6	0.0
2004	Wild	94.8	5.2	0.0
	Hatchery	95.2	4.8	0.0
2005	Wild	22.1	77.9	0.0
	Hatchery	20.5	79.5	0.0
2006	Wild	28.7	71.3	0.0
	Hatchery	60.3	39.7	0.0
2007	Wild	40.3	59.3	0.0
	Hatchery	62.1	37.9	0.0
2008	Wild	65.4	33.7	0.9
	Hatchery	88.8	11.2	0.0
2009	Wild	39.8	57.8	2.4
	Hatchery	23.4	76.6	0.0

Brood year	Origin	Saltwater age		
		1	2	3
2010	Wild	65.2	33.7	1.1
	Hatchery	76.5	23.5	0.0
2011	Wild	27.5	72.5	0.0
	Hatchery	36.0	64.0	0.0
2012	Wild	42.4	52.5	5.1
	Hatchery	40.9	59.1	0.0
2013	Wild	40.7	57.4	1.9
	Hatchery	45.5	54.5	0.0
2014	Wild	47.5	50.8	1.6
	Hatchery	29.4	70.6	0.0
Average	Wild	45.9	53.1	1.0
	Hatchery	52.2	47.8	0.0

There was little difference between mean lengths of hatchery and natural-origin steelhead in the 2013 and 2014 brood years (Table 3.3). Natural-origin fish were on average 1 to 3 cm larger than hatchery-origin fish of the same age.

**Table 3.3.** Mean fork length (cm) at age (saltwater ages) of hatchery and wild steelhead collected from broodstock, 1998-2014; N = sample size and SD = 1 standard deviation.

Brood year	Origin	Steelhead fork length (cm)								
		1-Salt			2-Salt			3-Salt		
		Mean	N	SD	Mean	N	SD	Mean	N	SD
1998	Wild	63	15	4	79	20	5	-	0	-
	Hatchery	61	9	4	73	34	4	-	0	-
1999	Wild	65	29	5	74	28	5	77	1	-
	Hatchery	62	54	4	73	12	4	-	0	-
2000	Wild	64	22	3	74	17	5	-	0	-
	Hatchery	60	57	3	71	27	4	-	0	-
2001	Wild	61	33	6	77	31	5	-	0	-
	Hatchery	62	17	4	72	97	4	-	0	-
2002	Wild	64	55	4	77	44	4	-	0	-
	Hatchery	63	106	4	73	6	4	-	0	-
2003	Wild	69	8	6	77	52	5	91	1	-
	Hatchery	66	27	4	75	65	4	-	0	-
2004	Wild	63	73	6	78	4	2	-	0	-
	Hatchery	61	59	3	73	3	1	-	0	-
2005	Wild	59	21	4	74	74	5	-	0	-
	Hatchery	59	23	4	72	89	4	-	0	-
2006	Wild	63	27	5	75	67	6	-	0	-

Brood year	Origin	Steelhead fork length (cm)								
		1-Salt			2-Salt			3-Salt		
		Mean	N	SD	Mean	N	SD	Mean	N	SD
	Hatchery	61	41	4	72	27	5	-	0	-
2007	Wild	64	31	6	76	46	5	-	0	-
	Hatchery	60	60	4	71	36	5	-	0	-
2008	Wild	64	68	4	77	35	4	80	1	-
	Hatchery	60	95	4	72	12	2	-	0	-
2009	Wild	65	33	5	76	48	6	81	2	0
	Hatchery	63	18	4	75	59	5	-	-	-
2010	Wild	64	60	5	74	31	5	76	1	-
	Hatchery	61	53	5	73	23	5	-	-	-
2011	Wild	62	28	5	76	74	5	-	0	-
	Hatchery	60	36	4	74	64	4	-	0	-
2012	Wild	63	25	3	74	31	5	74	3	2
	Hatchery	59	27	3	74	39	4	-	0	-
2013	Wild	61	22	5	77	31	5	74	1	-
	Hatchery	60	35	3	74	42	4	-	0	-
2014	Wild	61	29	4	75	31	4	61	1	-
	Hatchery	60	20	3	72	48	4	-	0	-
Average	Wild	63	34	5	76	39	5	77	1	1
	Hatchery	61	43	4	73	40	4	-	0	-

### Sex Ratios

Male steelhead in the 2013 brood year made up about 48% of the adults collected, resulting in an overall male to female ratio of 0.93:1.00 (Table 3.4). For the 2014 brood year, males made up about 49% of the adults collected, resulting in an overall male to female ratio of 0.96:1.00. On average (1998-2014), the sex ratio is slightly less than the 1:1 ratio assumed in the broodstock protocol (Table 3.4).

**Table 3.4.** Numbers of male and female wild and hatchery steelhead collected for broodstock, 1998-2014. Ratios of males to females are also provided.

Brood year	Number of wild steelhead			Number of hatchery steelhead			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
1998	13	22	0.59:1.00	15	28	0.54:1.00	0.56:1.00
1999	22	36	0.61:1.00	35	32	1.09:1.00	0.84:1.00
2000	18	21	0.86:1.00	60	41	1.46:1.00	1.26:1.00
2001	38	26	1.46:1.00	40	74	0.54:1.00	0.78:1.00
2002	32	67	0.48:1.00	81	32	2.53:1.00	1.14:1.00
2003	19	44	0.43:1.00	44	48	0.92:1.00	0.68:1.0
2004	43	42	1.02:1.00	90	42	2.14:1.00	1.58:1.00

Brood year	Number of wild steelhead			Number of hatchery steelhead			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
2005	36	59	0.61:1.00	46	68	0.68:1.00	0.65:1.00
2006	38	63	0.60:1.00	47	51	0.92:1.00	0.75:1.00
2007	36	43	0.84:1.00	49	48	1.02:1.00	0.93:1.00
2008	61	43	1.42:1.00	68	39	1.74:1.00	1.57:1.00
2009	44	57	0.77:1.00	54	53	1.02:1.00	0.89:1.00
2010	49	57	0.86:1.00	62	43	1.44:1.00	1.11:1.00
2011	44	60	0.73:1.00	50	54	0.93:1.00	0.82:1.00
2012	30	33	0.91:1.00	31	35	0.89:1.00	0.90:1.00
2013	33	30	1.10:1.00	38	46	0.83:1.00	0.93:1.00
2014	30	33	0.91:1.00	36	36	1.00:1.00	0.96:1.00
<b>Total</b>	<b>586</b>	<b>736</b>	<b>0.80:1.00</b>	<b>846</b>	<b>770</b>	<b>1.10:1.00</b>	<b>0.95:1.00</b>

### Fecundity

Fecundities for Wenatchee steelhead in brood years 2013 and 2014 averaged 5,762 and 5,839 eggs per female, respectively (Table 3.5). Mean fecundities for the 2013 and 2014 brood years were also greater than the 5,678 eggs per female assumed in the broodstock protocol.

**Table 3.5.** Mean fecundity of wild, hatchery, and all female steelhead collected for broodstock, 1998-2014.

Brood year	Mean fecundity		
	Wild	Hatchery	Total
1998	6,202	5,558	5,924
1999	5,691	5,186	5,424
2000	5,858	5,729	5,781
2001	5,951	6,359	6,270
2002	5,776	5,262	5,626
2003	6,561	6,666	6,621
2004	5,118	5,353	5,238
2005	5,545	6,061	5,832
2006	5,688	5,251	5,492
2007	5,840	5,485	5,660
2008	5,693	5,153	5,433
2009	6,199	6,586	6,408
2010	5,458	5,423	5,442
2011	6,276	6,100	6,203
2012	5,309	6,388	5,891
2013	5,749	5,770	5,762
2014	5,831	5,847	5,839
<b>Average</b>	<b>5,809</b>	<b>5,775</b>	<b>5,814</b>

## 3.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

From 1998-2011, a total of 493,827 eggs were required to meet the program release goal of 400,000 smolts. This was based on the unfertilized egg-to-release survival standard of 81%. In 2012, the egg take target was reduced to 305,309, which is needed to meet the revised release target of 247,300 smolts. Between 1998 and 2011, the egg take goal was reached 57% of the time (Table 3.6). Since 2011, the target has been reached or exceeded 100% of the time (Table 3.6).

**Table 3.6.** Numbers of eggs taken from steelhead broodstock, 1998-2014.

Brood year	Number of eggs taken
1998	224,315
1999	303,083
2000	280,872
2001	549,464
2002	503,030
2003	532,708
2004	408,538
2005	672,667
2006	546,382
2007	462,662
2008	439,980
2009	633,229
2010	499,499
2011	522,049
<i>Average (1998-2011)</i>	<i>488,782</i>
2012	371,151
2013	339,949
2014	395,453
<i>Average (2012-present)</i>	<i>368,851</i>

#### *Number of acclimation days*

Juvenile WxW steelhead were transferred from Chelan Fish Hatchery to the Chiwawa Acclimation Facility in October 2013 and HxH steelhead were transferred from Eastbank Fish Hatchery to Chiwawa Acclimation Facility in November 2013. In April 2014, about 25,000 steelhead were transferred to Blackbird Pond near Leavenworth for acclimation on Wenatchee River water. Fish were acclimated for 7 d before a volitional release was initiated on 22 April.

The remainder stayed at the Chiwawa Acclimation Facility until they were volitionally and forced released from the facility during late April to mid-May.

Juvenile Wenatchee steelhead at the Chiwawa Acclimation Facility were acclimated and reared on Wenatchee and Chiwawa River water. Before 2012, Wenatchee steelhead were reared on Columbia River water from January through May before being trucked and released into the Wenatchee River basin (Table 3.7).

**Table 3.7.** Water source and mean acclimation period for Wenatchee steelhead, brood years 1998-2013.

Brood year	Release year	Parental origin	Water source	Number of Days
1998	1999	H x H	Wenatchee/Chiwawa	36
		H x W	Wenatchee/Chiwawa	36
		W x W	Wenatchee/Chiwawa	36
1999	2000	H x H	Wenatchee/Chiwawa	138
		H x W	Wenatchee/Chiwawa	138
		W x W	Wenatchee/Chiwawa	138
		H x W	Eastbank	0
		W x W	Eastbank	0
2000	2001	H x H	Wenatchee/Chiwawa	122
		H x W	Wenatchee/Chiwawa	122
		H x W	Wenatchee/Chiwawa	122
		W x W	Wenatchee/Chiwawa	122
2001	2002	H x H	Columbia	92
		H x H	Wenatchee/Chiwawa	63
		H x W	Columbia	92
		H x W	Wenatchee/Chiwawa	63
		W x W	Columbia	153
2002	2003	H x H	Columbia	98
		H x W	Columbia	98
		W x W	Columbia	117
2003	2004	H x H	Columbia	88
		H x W	Wenatchee/Chiwawa	84
		W x W	Columbia	148
2004	2005	H x H	Columbia	160
		H x W	Columbia	160
		W x W	Columbia	160
2005	2006	H x H	Columbia	116
		H x W	Columbia	113
		W x W	Columbia	141
2006	2007	Early H x W	Columbia	111



Brood year	Release year	Parental origin	Water source	Number of Days
		Late H x W	Columbia	112
		W x W	Columbia	148
2007	2008	Early H x W	Columbia	94-95
		Late H x W	Columbia	91-93
		W x W	Columbia	138
2008	2009	Early H x W	Columbia	120-121
		Early H x W	Columbia/Wenatchee	120-121/28-95
		Late H x W	Columbia	114-115
		W x W	Columbia	152-153
2009	2010	Early H x W	Columbia	93-94
		Early H x W	Columbia/Wenatchee	99-111
		Early H x W	Wenatchee	31-129
		Late H x W	Columbia	84-87
		W x W	Columbia/Nason	118-120/28
2010	2011	H x H	Wenatchee	188-192
		H x H	Wenatchee	37-87
		H x H	Columbia	181
		W x W	Columbia	148-149
		W x W	Columbia/Nason	113-114/42-101
		W x W	Columbia	148-149
2011	2012	W x W	Wenatchee	160-201
		W x W	Wenatchee	179-188
		W x W	Wenatchee	21-72
		W x W	Nason	56-107
2012	2013	H x H	Wenatchee	168-189
		H x H	Wenatchee	168-225
		W x W	Wenatchee	168-225
		W x W	Wenatchee	168-189
		W x W	Chiwawa	187
2013	2014	H x H	Wenatchee	7-67
		H x H	Wenatchee	168-169
		W x W	Wenatchee	176-197
		W x W	Wenatchee	179-204

## Release Information

### Numbers released

In 2011, the HCP Hatchery Committee agreed to reduce the Wenatchee summer steelhead program from 400,000 smolts to 247,300 smolts. Based on this new goal and the number of WxW steelhead present, all HxH steelhead were transferred to the Ringold Fish Hatchery to be included in their production program.

The release of 2013 brood Wenatchee steelhead achieved 93% of the 247,300 target goal with about 229,836 smolts released into the Wenatchee and Chiwawa rivers and Nason Creek (Table 3.8). Distribution of juvenile steelhead released in each of the three subbasins was determined by the mean proportion of steelhead redds in each basin. About 38.9% and 10.2% of the steelhead were released in Nason Creek and the Chiwawa River, respectively. The balance of the program was split between the Wenatchee River downstream from Tumwater Dam (11.0%) and the Wenatchee River upstream from the dam (39.9%).

**Table 3.8.** Numbers of steelhead smolts released from the hatchery, brood years 1998-2013. Before brood year 2011, the release target for steelhead was 400,000 smolts. Beginning with brood year 2011, the release target is 247,300 smolts.

Brood year	Release year	Number of smolts
1998	1999	172,078
1999	2000	175,701
2000	2001	184,639
2001	2002	335,933
2002	2003	302,060
2003	2004	374,867
2004	2005	294,114
2005	2006	452,184
2006	2007	299,937
2007	2008	306,690
2008	2009	327,143
2009	2010	484,772
2010	2011	354,314
<i>Average (1998-2010)</i>		<b>312,649</b>
2011	2012	206,397
2012	2013	249,004
2013	2014	229,836
<i>Average (2011-present)</i>		<b>228,412</b>

### Numbers marked

Wenatchee hatchery steelhead from the 2013 brood were marked with coded wire tags (CWT) in the snout. About 58.0% of the juveniles released were also adipose fin clipped (Table 9).

**Table 3.9.** Release location and marking scheme for the 1998-2013 brood Wenatchee steelhead.

Brood year	Release location	Parental origin	Proportion Ad-clip	CWT or VIE color/side	Tag rate	Number released
1998	Chiwawa River	H x H	0.000	Red Left	0.994	52,765
	Chiwawa River	H x W	0.000	Green Left	0.990	37,013
	Chiwawa River	W x W	0.000	Orange Left	0.827	82,300
1999	Wenatchee River	H x H	0.000	Green Left	0.911	45,347
	Wenatchee River	H x W	0.000	Orange Left	0.927	30,713
	Chiwawa River	H x H	0.000	Red Right	0.936	25,622
	Chiwawa River	H x W	0.000	Green Right	0.936	43,379
	Chiwawa River	W x W	0.000	Orange Right	0.936	30,600
2000	Chiwawa River	H x H	0.000	Red Left	0.963	33,417
	Chiwawa River	H x W	0.000	Green Left	0.963	57,716
	Chiwawa River	H x W	0.000	Green Right	0.949	48,029
	Chiwawa River	W x W	0.000	Orange Right	0.949	45,477
2001	Nason Creek	H x W	0.000	Green Right	0.934	75,276
	Nason Creek	W x W	0.000	Orange Right	0.934	48,115
	Chiwawa River	H x W	0.000	Green Left	0.895	92,487
	Chiwawa River	H x H	0.000	Red Left	0.895	120,055
2002	Chiwawa River	H x H	0.000	Red Left	0.920	156,145
	Chiwawa River	H x W	0.000	Green Left	0.928	33,528
	Nason Creek	W x W	0.000	Orange Right	0.928	112,387
2003	Wenatchee River	H x H	0.000	Red Left	0.968	117,663
	Chiwawa River	H x W	0.000	Green Left	0.927	191,796
	Nason Creek	W x W	0.000	Orange Right	0.962	65,408
2004	Wenatchee River	H x H	0.500	Red Left	0.804	39,636
	Chiwawa River	H x W	0.000	Green Left	0.977	153,959
	Nason Creek	W x W	0.000	Pink Right	0.940	100,519
2005	Wenatchee River	H x H	1.000	Red Left	0.983	104,552
	Wenatchee River	H x W	0.616	Green Left	0.979	190,319
	Chiwawa River	H x W	0.616	Green Left	0.979	18,634
	Chiwawa River	W x W	0.000	Pink Right	0.969	14,124
	Nason Creek	W x W	0.000	Pink Right	0.969	124,555
2006	Wenatchee River	H x W (early)	1.000	Green Right	0.918	66,022
	Wenatchee River	H x W (late)	0.671	Green Left	0.935	92,176

Brood year	Release location	Parental origin	Proportion Ad-clip	CWT or VIE color/side	Tag rate	Number released
	Chiwawa River	H x W (late)	0.671	Green Left	0.935	41,240
	Chiwawa River	W x W	0.000	Pink Right	0.945	7,500
	Nason Creek	W x W	0.000	Pink Right	0.945	92,999
2007	Wenatchee River	H x W (early)	0.967	Green Right	0.950	64,310
	Wenatchee River	H x W (late)	0.586	Green Left	0.951	97,549
	Chiwawa River	H x W (late)	0.586	Green Left	0.951	43,011
	Chiwawa River	W x W	0.000	Pink Right	0.952	7,026
	Nason Creek	W x W	0.000	Pink Right	0.952	94,794
2008	Blackbird Pond	HxW (early)	0.917	Green Right	0.910	49,878
	Wenatchee River	H x W (early)	0.917	Green Right	0.910	48,624
	Wenatchee River	H x W (late)	0.595	Green Left	0.908	74,848
	Chiwawa River	H x W (late)	0.595	Green Left	0.908	25,835
	Chiwawa River	W x W	0.000	Pink Right	0.904	25,778
	Nason Creek	W x W	0.000	Pink Right	0.904	102,170
2009	Blackbird Pond	H x W (early)	0.969	Green Right	0.934	50,248
	Wenatchee River	H x W (early)	0.969	Green Right	0.934	105,239
	Wenatchee River	H x W (late)	0.973	Green Left	0.975	27,612
	Wenatchee River	H x W (late)	0.000	Green Left	0.975	45,435
	Chiwawa River	H x W (early)	0.969	Green Right	0.934	23,835
	Chiwawa River	H x W (late)	0.973	Green Left	0.975	33,047
	Chiwawa River	H x W (late)	0.000	Green Left	0.975	54,381
	Nason Creek	W x W	0.000	Pink Right	0.979	145,029
2010	Wenatchee River	H x H	0.994	-	0.984	24,838
	Wenatchee River	H x H	0.994	-	0.984	45,000
	Wenatchee River	H x H	0.994	-	0.984	92,113
	Chiwawa River	W x W	0.000	Pink Right	0.917	81,174
	Nason Creek	W x W	0.000	Pink R/Pink L	0.884	20,000
	Nason Creek	W x W	0.000	Pink Right	0.917	91,189
2011	Wenatchee River	W x W	0.985	CWT	0.953	70,885
	Wenatchee River	W x W	0.985	CWT	0.953	24,992
	Wenatchee River	W x W	0.000	CWT	0.987	25,569
	Chiwawa River	W x W	0.985	CWT	0.953	31,050
	Nason Creek	W x W	0.000	CWT	0.989	18,254

Brood year	Release location	Parental origin	Proportion Ad-clip	CWT or VIE color/side	Tag rate	Number released
	Nason Creek	W x W	0.985	CWT	0.953	36,225
2012	Wenatchee River	W x W	0.000	CWT	0.965	14,824
	Wenatchee River	H x H	1.000	AD/CWT	0.920	9,841
	Wenatchee River	W x W	0.000	CWT	0.965	28,362
	Wenatchee River	H x H	1.000	AD/CWT	0.920	76,695
	Chiwawa River	W x W	0.000	CWT	0.965	12,760
	Chiwawa River	H x H	1.000	AD/CWT	0.920	34,503
	Nason Creek	W x W	0.000	CWT	0.965	43,854
	Nason Creek	W x W	0.000	CWT	0.965	28,165
2013	Wenatchee River	W x W	0.000	CWT	0.963	36,736
	Wenatchee River	H x H	0.998	AD/CWT	0.990	55,055
	Wenatchee River	H x H	0.998	AD/CWT	0.990	25,316
	Chiwawa River	W x W	0.000	CWT	0.963	9,360
	Chiwawa River	H x H	0.998	AD/CWT	0.990	14,040
	Nason Creek	W x W	0.000	CWT	0.963	50,503
	Nason Creek	H x H	0.998	AD/CWT	0.990	38,826

**Numbers PIT tagged**

Table 3.10 summarizes the number of hatchery steelhead of different parental origins that have been PIT-tagged and released into the Wenatchee River basin.

**Table 3.10.** Summary of PIT-tagging activities for Wenatchee hatchery steelhead, brood years 2006-2013.

Brood year	Release location	Parental origin	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2006	Wenatchee River	H x W (early)	10,036	479	24	9,533
	Wenatchee/Chiwawa rivers	H x W (late)	10,031	922	20	9,089
	Chiwawa River/Nason	W x W	10,019	152	352	9,515
2007	Wenatchee River	H x W (early)	9,852	22	10	9,820
	Wenatchee/Chiwawa rivers	H x W (late)	10,063	73	78	9,912
	Chiwawa River/Nason	W x W	10,038	55	1	9,982
2008	Wenatchee River	H x W (early)	10,101	59	15	10,027
	Wenatchee/Chiwawa rivers	H x W (late)	10,104	106	17	9,981
	Chiwawa River/Nason	W x W	10,101	159	80	9,862

Brood year	Release location	Parental origin	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2009	Wenatchee/Chiwawa rivers	H x W (early)	10,114	574	11	9,529
	Wenatchee (Blackbird)	H x W (early)	8,100	0	0	8,100
	Wenatchee/Chiwawa rivers	H x W (late)	10,115	271	11	9,833
	Chiwawa pilot	H x W (early)	10,107	532	103	9,472
	Chiwawa River/Nason	W x W	10,101	38	3	10,060
2010	Wenatchee River	HxH	10,100	624	21	9,455
	Chiwawa River/Nason	WxW	10,100	206	0	9,894
	Wenatchee (Blackbird)	HxH	10,101	235	8	9,858
	Wenatchee River	HxH	10,100	46	28	10,026
2011	Wenatchee/Chiwawa/Nason	WxW (circular)	10,101	139	30	9,932
	Wenatchee/Chiwawa/Nason	WxW (raceway)	20,220	121	35	20,064
2012	Wenatchee/Chiwawa/Nason	WxW (circular)	15,244	176	4	15,064
	Wenatchee/Chiwawa/Nason	HxH (raceway)	10,223	140	13	10,070
2013	Wenatchee/Chiwawa/Nason	WxW	5,100	95	1	5,004
	Wenatchee/Chiwawa/Nason	HxH	10,201	84	12	10,105

**2014 Brood Wenatchee HxH Summer Steelhead**—A total of 10,129 Wenatchee HxH summer steelhead were tagged at Eastbank Hatchery on 15-18 September 2014. These fish were tagged in raceway #4. Fish were not fed during tagging or for two days before and after tagging. Fish averaged 74 mm in length and 5.9 g at time of tagging.

**2014 Brood Wenatchee WxW Summer Steelhead**—A total of 5,100 Wenatchee WxW summer steelhead were tagged at Chelan Hatchery on 2-4 September 2014. These fish were tagged in raceway #2. Fish were not fed during tagging or for two days before and after tagging. Fish averaged 97 mm in length and 11.0 g at time of tagging.

#### **Fish size and condition at release**

With the exception of the Blackbird Pond release, all 2013 brood steelhead were trucked and released as yearling smolts in April and May 2014. The Blackbird Pond group was released volitionally beginning on 22 April. The WxW fish did not meet the length or weight target, but exceeded the target for coefficient of variation (CV) for fork length (Table 3.11). The HxH group was combined with the WxW group in Pond 2 once they were transferred to Chiwawa Acclimation Facility. The HxH and the mixed WxW and HxH groups did not meet the length or weight targets and these fish were smaller than the WxW fish.

**Table 3.11.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of steelhead smolts released from the hatchery, brood years 1998-2013. Size targets are provided in the last row of the table.

Brood year	Release year	Parental origin	Fork length (mm)		Mean weight	
			Mean	CV	Grams (g)	Fish/pound
1998	1999	H x H	201	11.1	92.3	5
		H x W	190	12.8	76.9	6
		W x W	173	12.0	55.3	8
1999	2000	H x H	181	8.9	70.6	6
		H x W	187	7.2	75.3	6
		W x W	184	11.3	71.5	6
2000	2001	H x H	218	15.2	122.4	4
		H x W	209	10.6	107.5	4
		W x W	205	10.7	100.9	5
2001	2002	H x H	179	17.4	67.0	7
		H x W	192	15.6	82.8	6
		W x W	206	11.6	102.6	4
2002	2003	H x H	194	13.1	83.0	6
		H x W	191	13.0	77.4	6
		W x W	180	19.1	70.3	7
2003	2004	H x H	191	14.4	73.1	6
		H x W	199	12.9	83.9	5
		W x W	200	11.1	90.1	5
2004	2005	H x H	204	11.3	87.2	6
		H x W	202	13.5	71.9	5
		W x W	198	12.4	76.6	6
2005	2006	H x H	215	12.6	116.6	4
		H x W	198	11.8	86.3	5
		W x W	189	15.4	55.3	6
2006	2007	H x H (early)	213	12.1	109.6	4
		H x W (late)	186	11.8	68.3	7
		W x W	178	11.1	58.6	8
2007	2008	H x W (early)	192	17.4	77.1	6
		H x W (late)	179	19.3	63.8	7
		W x W	183	12.3	62.8	7
2008	2009	H x W (early)	184	11.6	68.0	7
		H x W (late)	186	11.6	73.5	6
		W x W	181	13.0	59.7	8

Brood year	Release year	Parental origin	Fork length (mm)		Mean weight	
			Mean	CV	Grams (g)	Fish/pound
2009	2010	H x W (early)	197	11.3	84.2	5
		H x W (late)	192	11.1	72.7	6
		W x W	190	9.6	70.5	6
2010	2011	H x H	183	14.1	68.9	4
		W x W	188	10.5	68.1	7
2011	2012	H x H	NA	NA	NA	NA
		W x W	156	17.1	45.2	10
2012	2013	H x H / W x W	150	16.1	40.8	11
		H x H / W x W	157	16.4	45.0	10
		W x W	156	18.7	49.0	9
2013	2014	H x H / W x W	157	14.5	49.4	9
		H x H	127	16.2	26.8	17
		W x W	162	20.4	55.8	8
<i>Targets</i>			<b>191</b>	<b>9.0</b>	<b>75.6</b>	<b>6</b>

### Survival Estimates

Overall survival of Wenatchee steelhead (WxW and HxH) from green (unfertilized) egg to release was below the standard set for the program. This is in large part because of poor unfertilized egg to eyed egg survival (Table 3.12).

The Wenatchee steelhead program, from its inception, has experienced highly variable fertilization rates. It is unknown at this time what mechanisms may be influencing stock performance at these stages.

**Table 3.12.** Hatchery life-stage survival rates (%) for steelhead, brood years 1998-2013. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
1998	92.0	100.0	85.5	91.7	99.2	98.8	97.8	99.9	76.7
1999	91.2	100.0	66.9	93.0	95.9	94.9	93.1	99.7	58.0
2000	83.9	96.2	77.6	86.7	99.3	98.9	97.7	99.5	65.7
2001	90.0	100.0	73.0	91.8	99.1	97.8	91.3	99.7	61.1
2002	99.0	100.0	69.2	93.1	95.9	94.4	89.6	89.6	60.0
2003	87.0	96.8	86.3	83.8	97.2	94.8	97.6	85.3	70.4
2004	97.6	98.5	83.4	93.7	97.8	94.1	92.2	99.9	72.0
2005	91.3	95.1	81.3	92.1	95.6	91.8	89.7	99.6	67.2
2006	99.1	95.3	73.2	85.4	95.4	94.6	87.8	98.5	54.9



Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
2007	100.0	100.0	80.3	92.0	95.7	92.7	89.8	99.1	66.3
2008	100.0	100.0	87.1	88.4	99.0	97.4	96.6	99.5	74.4
2009	97.3	100.0	89.0	97.2	96.0	95.2	88.6	96.6	76.6
2010	96.7	100.0	93.8	93.9	91.0	86.2	80.6	96.0	70.9
2011 <sup>a</sup>	96.3	94.4	74.2	97.7	96.6	89.5	86.4	98.4	62.7
2012	95.2	98.4	74.7	99.7	97.8	94.0	90.1	98.9	67.1
2013	80.8	97.0	75.0	96.5	97.8	96.6	93.4	99.2	67.6
<b>Average</b>	<b>93.6</b>	<b>98.2</b>	<b>79.4</b>	<b>92.3</b>	<b>96.8</b>	<b>94.5</b>	<b>91.4</b>	<b>97.5</b>	<b>67.0</b>
<b>Standard</b>	<b>90.0</b>	<b>85.0</b>	<b>92.0</b>	<b>98.0</b>	<b>97.0</b>	<b>93.0</b>	<b>90.0</b>	<b>95.0</b>	<b>81.0</b>

<sup>a</sup> Survival estimates are only for WxW steelhead.

### 3.3 Disease Monitoring

Rearing of the 2013 brood Wenatchee summer steelhead was similar to previous years with fish being held on Chelan spring water, Eastbank well water, and Chelan well water before being transferred for overwinter acclimation at the Chiwawa Acclimation Facility. Volitional and non-migratory released fish were released into Nason Creek, Chiwawa River, and the Wenatchee River. There were no major fish health concerns for brood year 2013.

### 3.4 Natural Juvenile Productivity

During 2014, juvenile steelhead were sampled at the Lower Wenatchee, Chiwawa, and Nason Creek traps and counted during snorkel surveys within the Chiwawa River basin. Because the snorkel surveys targeted juvenile Chinook salmon, the entire distribution of juvenile steelhead in the Chiwawa River basin was not surveyed. Therefore, the parr numbers presented below represent a minimum estimate.

#### Parr Estimates

A total of 16,083 ( $\pm 10.0\%$ ) age-0 (<100 mm) and 5,084 ( $\pm 12.0\%$ ) age-1+ (100-200 mm)<sup>4</sup> steelhead/rainbow were estimated in the Chiwawa River basin in August 2014 (Table 3.13 and 3.14). During the survey period 1992-2014, numbers of age-0 and 1+ steelhead/rainbow have ranged from 1,410 to 45,727 and 2,533 to 22,130, respectively, in the Chiwawa River basin (Table 3.13 and 3.14; Figure 3.1). Numbers of all fish counted in the Chiwawa River basin are reported in Appendix A.

Juvenile steelhead/rainbow were distributed primarily throughout the lower seven reaches of the Chiwawa River (downstream from Rock Creek). Their densities were highest in the lower portions of the river and in tributaries. Age-0 steelhead/rainbow most often used riffle and multiple channel habitats in the Chiwawa River, although they also associated with woody debris in pool and glide habitat. In tributaries they were generally most abundant in small pools. Those that were observed in riffles selected stations in quiet water behind small and large boulders, or

<sup>4</sup> A steelhead/rainbow trout larger than 200 mm (8 in) was considered a resident trout.

occupied stations in quiet water along the stream margin. In pool and multiple-channel habitats, age-0 steelhead/rainbow used the same kinds of habitat as age-0 Chinook salmon.

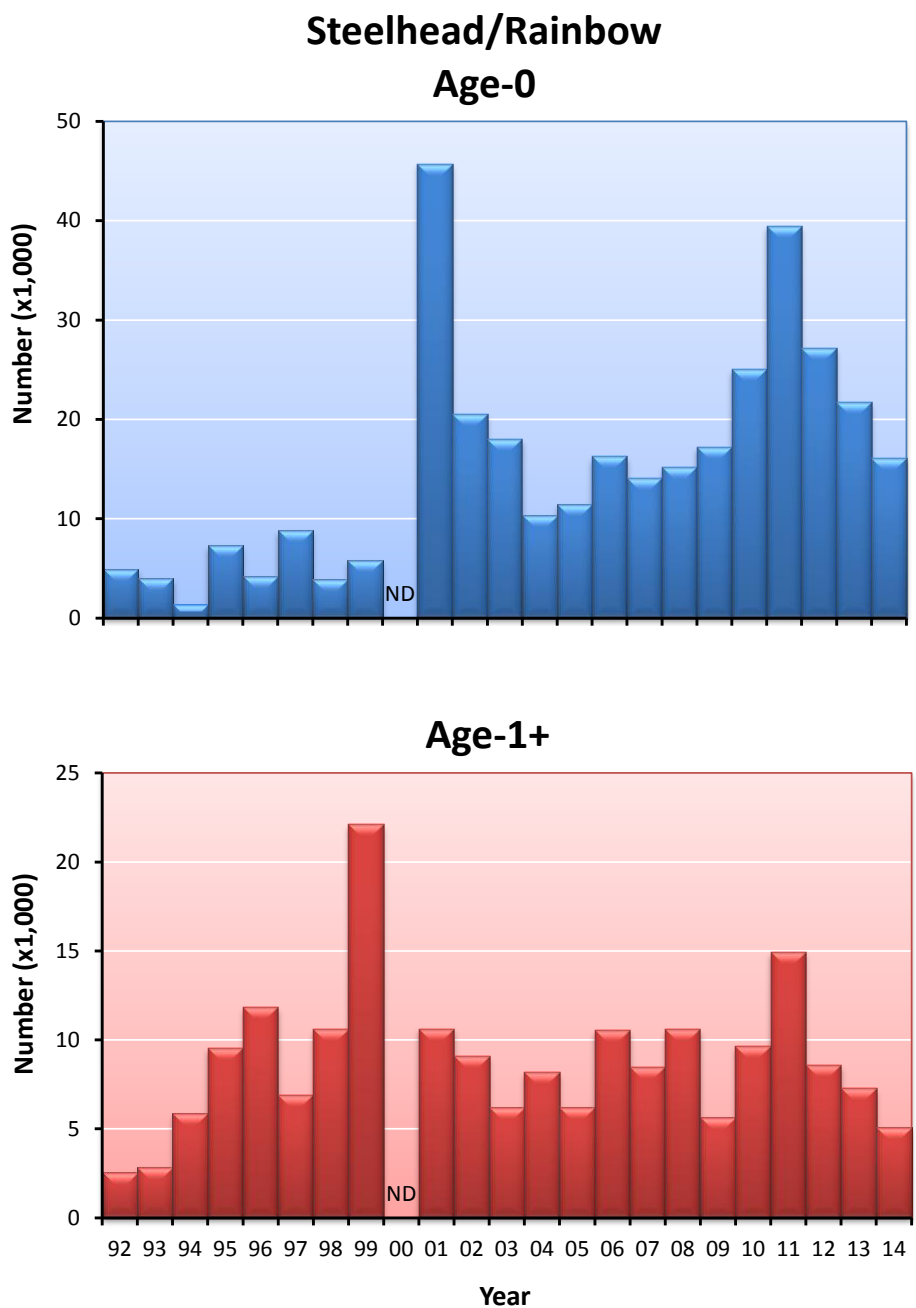
Age-1+ steelhead/rainbow most often used pool, riffle, and multiple-channel habitats. Those that used pools were usually in deeper water than subyearling steelhead/rainbow and Chinook salmon. Like age-0 steelhead/rainbow, age-1+ steelhead/rainbow selected stations in quiet water behind boulders in riffles, but the two age groups rarely occurred together. Age-1+ steelhead/rainbow used deeper and faster water than did subyearling steelhead/rainbow.

**Table 3.13.** Total numbers of age-0 steelhead/rainbow trout estimated in different streams in the Chiwawa River basin during snorkel surveys in August 1992-2014; NS = not sampled.

Sample Year	Chiwawa River	Phelps Creek	Chikamin Creek	Rock Creek	Unnamed Creek	Big Meadow Creek	Alder Creek	Brush Creek	Clear Creek	Total
1992	4,927	NS	NS	NS	NS	NS	NS	NS	NS	4,927
1993	3,463	0	356	185	NS	NS	NS	NS	NS	4,004
1994	953	0	256	24	0	177	0	0	0	1,410
1995	6,005	0	744	90	0	371	40	107	0	7,357
1996	3,244	0	71	40	0	763	127	0	0	4,245
1997	6,959	224	84	324	0	1,124	58	50	0	8,823
1998	2,972	22	280	96	113	397	18	22	0	3,921
1999	5,060	20	253	189	0	255	34	27	0	5,838
2000	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
2001	35,759	192	1,449	1,826	0	6,345	156	0	0	45,727
2002	12,137	0	2,252	889	0	4,948	277	18	0	20,521
2003	9,911	296	996	1,166	96	5,366	73	116	0	18,020
2004	8,464	110	583	113	40	957	35	78	0	10,380
2005	4,852	120	2,931	477	45	2,973	65	0	0	11,463
2006	10,669	21	858	872	34	3,647	73	71	0	16,245
2007	8,442	53	2,137	348	11	2,955	65	28	34	14,073
2008	9,863	0	2,260	859	0	1,987	57	168	36	15,230
2009	13,231	0	1,183	449	0	2,062	170	67	17	17,179
2010	17,572	0	2,870	1,478	5	2,843	182	35	33	25,018
2011	35,825	0	1,503	804	0	1,066	56	152	40	39,446
2012	21,537	0	1,817	1,501	0	2,164	42	54	19	27,134
2013	17,889	0	602	816	0	2,189	44	99	43	21,682
2014	12,256	21	1,617	1,039	0	1,005	32	56	57	16,083
<i>Average</i>	<i>11,454</i>	<i>51</i>	<i>1,195</i>	<i>647</i>	<i>17</i>	<i>2,180</i>	<i>80</i>	<i>57</i>	<i>14</i>	<i>15,397</i>

**Table 3.14.** Total numbers of age-1+ steelhead/rainbow trout estimated in different steams in the Chiwawa River basin during snorkel surveys in August 1992-2014; NS = not sampled.

Sample Year	Chiwawa River	Phelps Creek	Chikamin Creek	Rock Creek	Unnamed Creek	Big Meadow Creek	Alder Creek	Brush Creek	Clear Creek	Total
1992	2,533	NS	NS	NS	NS	NS	NS	NS	NS	2,533
1993	2,530	0	228	102	NS	NS	NS	NS	NS	2,860
1994	4,972	0	476	296	5	107	0	0	0	5,856
1995	8,769	0	494	71	0	183	0	0	0	9,517
1996	11,381	0	6	27	0	435	0	0	0	11,849
1997	6,574	160	0	105	0	66	0	0	0	6,905
1998	10,403	0	133	49	0	0	0	0	0	10,585
1999	21,779	0	68	201	0	82	0	0	0	22,130
2000	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
2001	9,368	16	186	407	0	646	0	0	0	10,623
2002	7,200	0	199	165	0	1,526	0	0	0	9,090
2003	4,745	362	426	599	0	47	0	0	0	6,179
2004	7,700	107	209	0	0	174	0	0	0	8,190
2005	4,624	63	957	257	0	287	0	0	0	6,188
2006	7,538	76	748	1,186	0	985	0	0	0	10,533
2007	6,976	0	945	96	0	431	0	0	0	8,448
2008	8,317	0	1,168	298	0	793	0	0	0	10,576
2009	4,998	16	320	102	0	167	21	0	5	5,629
2010	8,324	32	366	393	0	780	21	0	0	9,916
2011	13,329	0	415	470	0	689	0	0	0	14,903
2012	7,671	0	285	410	0	210	0	0	0	8,576
2013	6,439	0	0	48	0	766	0	0	0	7,253
2014	4,568	13	96	211	0	165	0	0	31	5,084
<i>Average</i>	<i>7,761</i>	<i>40</i>	<i>368</i>	<i>262</i>	<i>0</i>	<i>427</i>	<i>2</i>	<i>0</i>	<i>2</i>	<i>8,792</i>



**Figure 3.1.** Numbers of subyearling and yearling steelhead/rainbow trout within the Chiwawa River basin in August 1992-2014; ND = no data.

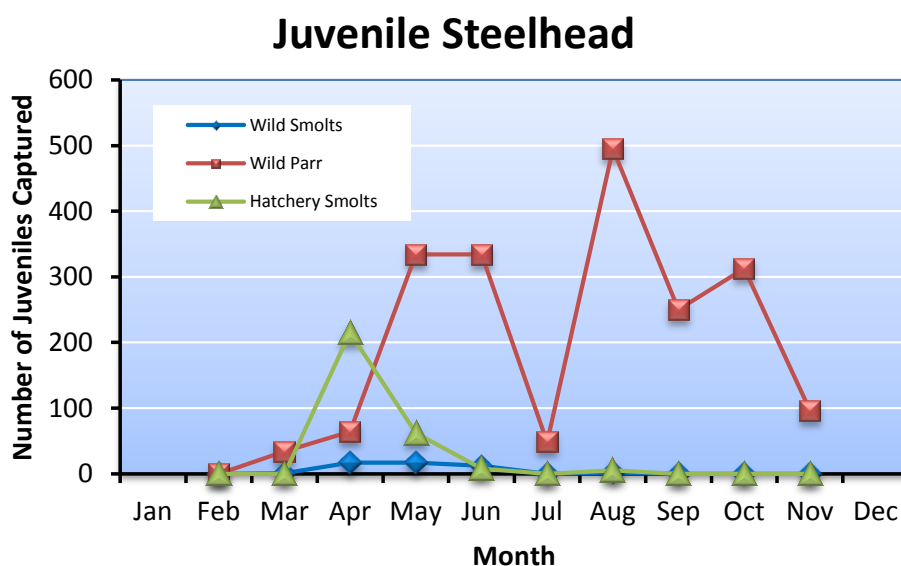
### Emigrant and Smolt Estimates

Numbers of steelhead smolts and emigrants were estimated at the Chiwawa, Nason, and Lower Wenatchee traps in 2014.

### Chiwawa Trap

The Chiwawa Trap operated between 18 March and 13 November 2014. During the trapping period, the trap was inoperable for 21 days because of high river discharge, debris, snow/ice, or mechanical failure. The trap operated in two different positions depending on stream flow; lower position at flows greater than 12 m<sup>3</sup>/s and an upper position at flows less than 12 m<sup>3</sup>/s. Monthly captures of all fish collected at the Chiwawa Trap are reported in Appendix B.

A total of 49 wild steelhead/rainbow smolts, 290 hatchery smolts, and 1,889 wild parr were captured at the Chiwawa Trap. Most (74%) of the hatchery steelhead smolts were collected in April, while most (94%) of the wild steelhead smolts were captured from April through June (Figure 3.2). Although steelhead/rainbow parr emigrated throughout the sampling period, peaks in emigration were observed in May and June, and August through October (Figure 3.2). Of the total number of wild steelhead captured (smolt and parr) 97% were classified as parr. Because of low numbers of steelhead/rainbow captured (a capture of at least 250 fish is needed for a statistically valid trial), no mark-recapture efficiency trials could be conducted with steelhead/rainbow at the Chiwawa Trap to estimate total population abundance.



**Figure 3.2.** Monthly captures of wild smolts, wild parr, and hatchery smolt steelhead/rainbow at the Chiwawa Trap, 2014.

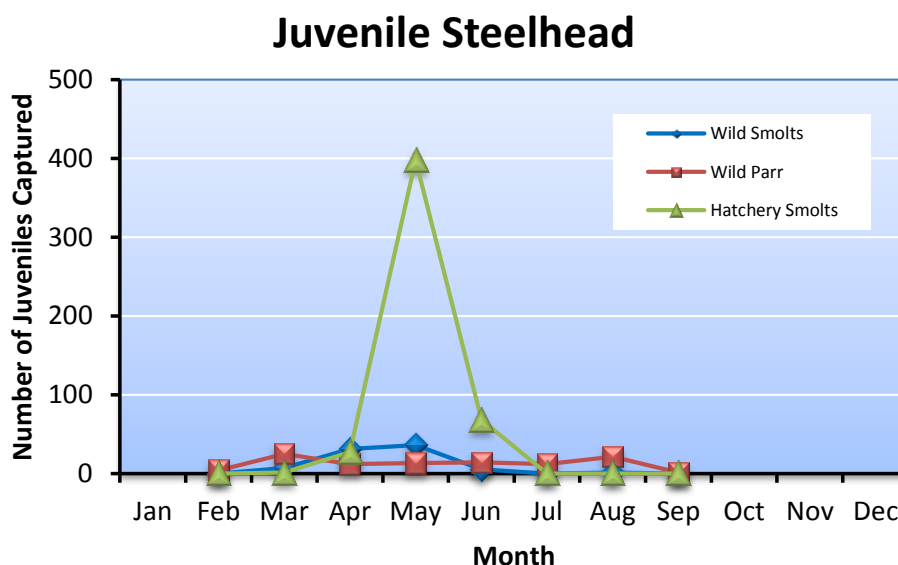
### Nason Creek Trap

The Nason Creek Trap operated between 1 March and 30 November 2014. During the nine-month sampling period the trap was inoperable for 48 days because of low discharge and ice accumulation. The trap captured a total of 18 wild steelhead smolts, 1,571 hatchery smolts, 991 wild parr, and 258 wild fry.

### Lower Wenatchee Trap

The Lower Wenatchee Trap operated between 12 February and 7 September 2014. During that time period the trap was inoperable for 12 days because of high river discharge, debris, snow/ice, or major hatchery releases. During the seven-month sampling period, a total of 102 wild

steelhead parr, 80 wild steelhead smolts, and 494 hatchery steelhead were captured at the trap. Because of the low numbers of steelhead encountered daily at the trap, it was not possible to carry out mark-recapture trials using steelhead. To gain insight into capture efficiency, 473 WxW hatchery steelhead (“surrogates” for natural fish) were marked and transported from Chiwawa Acclimation Facility and released at Dryden Dam. Two separate trials were conducted, resulting in a pooled efficiency estimate (95% C.I.) of 6,149 ( $\pm$  32,095) parr and smolt emigrants. Figure 3.3 shows the monthly captures of steelhead collected at the Lower Wenatchee Trap. All fish captured in the trap are reported in Appendix B.



**Figure 3.3.** Monthly captures of wild smolts, wild parr, and hatchery smolt steelhead/rainbow at the Lower Wenatchee Trap, 2014.

### PIT Tagging Activities

As part of the Comparative Survival Study (CSS), a total of 1,349 juvenile steelhead/rainbow trout (1,342 wild and seven hatchery) were PIT tagged and released in 2014 in the Wenatchee River basin (Table 3.15a). Most of these were tagged at the Chiwawa Trap. See Appendix C for a complete list of all fish captured, tagged, lost, and released.

**Table 3.15a.** Numbers of wild and hatchery steelhead/rainbow trout that were captured, tagged, and released at different locations within the Wenatchee River basin, 2014. Numbers of fish that died or shed tags are also given.

Sampling Location	Species and Life Stage	Number captured	Number of recaptures	Number tagged	Number died	Shed Tags	Total released	Percent mortality
Chiwawa Trap	Wild Steelhead/Rainbow	1,288	4	1,195	9	0	1,186	0.70
	Hatchery Steelhead/Rainbow	11	3	3	0	0	3	0.00
	<b>Total</b>	<b>1,299</b>	<b>7</b>	<b>1,198</b>	<b>9</b>	<b>0</b>	<b>1,189</b>	<b>0.69</b>
Chiwawa River (Electrofishing)	Wild Steelhead/Rainbow	94	0	23	0	0	23	0.00
	Hatchery Steelhead/Rainbow	0	0	0	0	0	0	--

Sampling Location	Species and Life Stage	Number captured	Number of recaptures	Number tagged	Number died	Shed Tags	Total released	Percent mortality
	<b>Total</b>	<b>94</b>	<b>0</b>	<b>23</b>	<b>0</b>	<b>0</b>	<b>23</b>	<b>0.00</b>
Lower Wenatchee Trap	Wild Steelhead/Rainbow	143	5	133	0	0	133	0.00
	Hatchery Steelhead/Rainbow	8	4	4	0	0	4	0.00
	<b>Total</b>	<b>151</b>	<b>9</b>	<b>137</b>	<b>0</b>	<b>0</b>	<b>137</b>	<b>0.00</b>
<b>Total:</b>	<b>Wild Steelhead/Rainbow</b>	<b>1,525</b>	<b>9</b>	<b>1,351</b>	<b>9</b>	<b>0</b>	<b>1,342</b>	<b>0.59</b>
	<b>Hatchery Steelhead/Rainbow</b>	<b>19</b>	<b>7</b>	<b>7</b>	<b>0</b>	<b>0</b>	<b>7</b>	<b>0.00</b>
<b>Grand Total:</b>		<b>1,544</b>	<b>16</b>	<b>1,358</b>	<b>9</b>	<b>0</b>	<b>1,349</b>	<b>0.58</b>

Numbers of steelhead/rainbow PIT-tagged and released as part of CSS during the period 2006-2014 are shown in Table 3.15b.

**Table 3.15b.** Summary of the numbers of wild and hatchery steelhead/rainbow trout that were tagged and released at different locations within the Wenatchee River basin, 2006-2014.

Sampling Location	Species and Life Stage	Numbers of PIT-tagged steelhead/rainbow released								
		2006	2007	2008	2009	2010	2011	2012	2013	2014
Chiwawa Trap	Wild Steelhead/Rainbow	1,366	832	1,431	1,127	930	1,012	1,011	1,228	1,186
	Hatchery Steelhead/Rainbow	0	3	2	1	2	1	2	0	3
	<b>Total</b>	<b>1,366</b>	<b>835</b>	<b>1,433</b>	<b>1,128</b>	<b>932</b>	<b>1,013</b>	<b>1,013</b>	<b>1,228</b>	<b>1,189</b>
Chiwawa River (Angling or Electrofishing)	Wild Steelhead/Rainbow	33	167	94	35	99	0	0	0	23
	Hatchery Steelhead/Rainbow	1	47	35	43	64	0	0	0	0
	<b>Total</b>	<b>34</b>	<b>214</b>	<b>129</b>	<b>78</b>	<b>163</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>23</b>
Upper Wenatchee Trap	Wild Steelhead/Rainbow	21	37	24	46	69	82	70	43	0
	Hatchery Steelhead/Rainbow	0	0	0	0	0	0	0	0	0
	<b>Total</b>	<b>21</b>	<b>37</b>	<b>24</b>	<b>46</b>	<b>69</b>	<b>82</b>	<b>70</b>	<b>43</b>	<b>0</b>
Nason Creek (Angling or Electrofishing)	Wild Steelhead/Rainbow	174	452	255	459	318	0	0	0	0
	Hatchery Steelhead/Rainbow	26	75	87	197	32	0	0	0	0
	<b>Total</b>	<b>200</b>	<b>527</b>	<b>342</b>	<b>656</b>	<b>350</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Upper Wenatchee (Angling or Electrofishing)	Wild Steelhead/Rainbow	413	1,001	21	7	30	0	0	0	0
	Hatchery Steelhead/Rainbow	2	64	26	23	9	0	0	0	0
	<b>Total</b>	<b>415</b>	<b>1,065</b>	<b>47</b>	<b>30</b>	<b>39</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Middle Wenatchee (Angling or Electrofishing)	Wild Steelhead/Rainbow	0	0	981	867	1,517	0	0	850	0
	Hatchery Steelhead/Rainbow	0	0	11	5	57	0	0	2	0
	<b>Total</b>	<b>0</b>	<b>0</b>	<b>992</b>	<b>872</b>	<b>1,574</b>	<b>0</b>	<b>0</b>	<b>852</b>	<b>0</b>
Lower Wenatchee (Angling or Electrofishing)	Wild Steelhead/Rainbow	0	0	102	69	0	0	0	0	0
	Hatchery Steelhead/Rainbow	0	0	10	9	0	0	0	0	0
	<b>Total</b>	<b>0</b>	<b>0</b>	<b>112</b>	<b>78</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Peshastin Creek (Angling or Electrofishing)	Wild Steelhead/Rainbow	0	0	0	92	307	0	0	0	0
	Hatchery Steelhead/Rainbow	0	0	0	0	0	0	0	0	0

Sampling Location	Species and Life Stage	Numbers of PIT-tagged steelhead/rainbow released								
		2006	2007	2008	2009	2010	2011	2012	2013	2014
Electrofishing)	<b>Total</b>	0	0	0	92	307	0	0	0	0
Lower Wenatchee Trap	Wild Steelhead/Rainbow	131	461	285	227	465	0	0	613	133
	Hatchery Steelhead/Rainbow	0	0	0	1	0	0	0	0	4
	<b>Total</b>	131	461	285	228	465	0	0	613	137
<b>Total:</b>	Wild Steelhead/Rainbow	2,138	2,950	3,193	2,929	3,735	1,094	1,081	2,734	1,342
	Hatchery Steelhead/Rainbow	29	189	171	279	164	1	2	2	7
<b>Grand Total:</b>		2,167	3,139	3,364	3,208	3,899	1,095	1,083	2,736	1,349

### 3.5 Spawning Surveys

Surveys for steelhead redds were conducted during March through early June, 2014, in the mainstem Wenatchee River and portions of select tributaries (Chiwawa River, Nason Creek, and Peshastin Creek). Beginning in 2014, adult steelhead escapement estimates in the majority of tributaries in the Wenatchee River basin were generated using mark-recapture techniques based on steelhead PIT tagged at Priest Rapids Dam (see Appendix D and Truscott et al. 2015 for details).

#### Redd Counts

A total of 109 steelhead redds were counted in the Wenatchee River and the lower portions of select tributaries in 2014 (Table 3.16). Because steelhead escapement estimates in tributaries are based on mark-recapture techniques, there are no or limited redd counts in tributaries beginning in 2014. Additionally, mainstem redd counts in 2014 were expanded based on estimates of observer efficiency (see Appendix D). Thus, evaluation of trends in redd counts is appropriate only before 2014.

**Table 3.16.** Numbers of steelhead redds estimated within different streams/watersheds within the Wenatchee River basin, 2001-2014; NS = not surveyed. Redd counts from 2004-2013 have been conducted within the same areas and with the same methods. Beginning in 2014, complete redd counts were conducted only within the mainstem Wenatchee River. Therefore, trends in redd counts are only appropriate for the mainstem Wenatchee River from 2004 through 2013.

Survey year	Number of steelhead redds							Total
	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River <sup>a</sup>	Icicle	Peshastin	
2001	25	27	NS	NS	116	19	NS	<b>187</b>
2002	80	80	1	0	315	27	NS	<b>503</b>
2003	64	121	5	3	248	16	15	<b>472</b>
2004	62	127	0	0	151	23	34	<b>397</b>
2005	162	412	0	2	459	8	97	<b>1,140</b>
2006	19	77	NS	0	191	41	67	<b>395</b>
2007	11	78	0	1	46	6	17	<b>159</b>
2008	11	88	NS	1	100	37	49	<b>286</b>



Survey year	Number of steelhead redds							
	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River <sup>a</sup>	Icicle	Peshastin	Total
2009	75	126	0	0	327	102	32	<b>662</b>
2010	74	270	4	3	380	120	118	<b>969</b>
2011	77	235	2	0	323	180	115	<b>932</b>
2012	8	158	0	0	137	47	65	<b>415</b>
2013	27	135	NS	NS	200	48	62	<b>472</b>
2014	5	0	NS	NS	195 <sup>b</sup>	NS	5	<b>205</b>

<sup>a</sup> Includes redds in Beaver and Chiwaukum creeks.

<sup>b</sup> Steelhead redd counts in the mainstem Wenatchee River were expanded based on estimated observer efficiency (see Appendix D).

### Redd Distribution

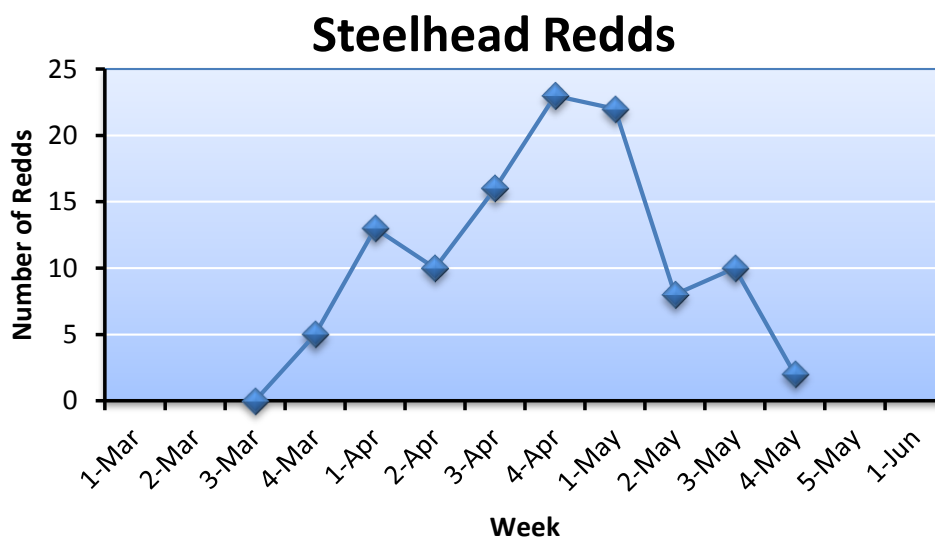
Steelhead redds were not evenly distributed among reaches within survey reaches on the Wenatchee River in 2014 (Table 3.17). About 63.8% of the spawning in the Wenatchee River occurred upstream from Tumwater Dam (Table 3.17).

**Table 3.17.** Numbers and percentages of steelhead redds counted within different reaches on the Wenatchee River during March through early June, 2014; SE = standard error.

Reach	Reach type	Number of redds counted	Expanded redd counts		Percent of redds within stream/watershed
			Estimated	SE	
Wenatchee 1 (W1)	Non-index	0	0.1	0.0	0.1
Wenatchee 2 (W2)	Index	4	4.9	4.7	2.5
Wenatchee 3 (W3)	Non-index	2	2.2	0.8	1.1
Wenatchee 4 (W4)	Non-index	0	0.2	0.1	0.1
Wenatchee 5 (W5)	Non-index	0	0.2	0.1	0.1
Wenatchee 6 (W6)	Index	25	63.1	39.1	32.4
Wenatchee 7 (W7)	NS	NS	NS	NS	--
Wenatchee 8 (W8)	Index	4	16.1	7.5	8.3
Wenatchee 9 (W9)	Index	46	77.9	70.2	40.1
Wenatchee 10 (W10)	Index	16	29.9	21.7	15.4
<b>Total</b>		<b>97</b>	<b>194.5</b>	<b>83.8</b>	<b>100.0</b>

### Spawn Timing

Steelhead began spawning during the third week of March in the Wenatchee River. Spawning activity appeared to begin once the mean daily stream temperature reached about 4.8°C and was observed in water temperatures ranging from 2.0 - 7.0°C. Steelhead spawning peaked during the fourth week of April in the Wenatchee River (Figure 3.4).



**Figure 3.4.** Numbers of steelhead redds counted during different weeks in different index areas within the Wenatchee River basin, March through early June 2014.

### Spawning Escapement

Before 2014, steelhead spawning escapement upstream from Tumwater Dam was calculated as the number of redds (in the Wenatchee River and tributaries upstream from the dam) times the fish per redd ratio (based on sex ratios estimated at Tumwater Dam using video surveillance). Beginning in 2014, escapement in tributaries was estimated using PIT-tag mark-recapture techniques (Truscott et al. 2015; Table 3.18), while observer efficiency expanded redd counts were used to estimate escapement in the mainstem Wenatchee River (Appendix D). Total redd counts were also used to estimate escapement in the lower portions of the main tributaries (downstream from the PIT interrogation sites).

**Table 3.18.** Spawning escapement estimates for natural-origin and hatchery-origin steelhead within tributaries of the Wenatchee River, brood year 2014. Escapement estimates were based on PIT-tag mark-recapture techniques (Truscott et al. 2015). SE = standard error and CV = coefficient of variation.

Tributary	Natural-origin steelhead			Hatchery-origin steelhead		
	Estimate	SE	CV	Estimate	SE	CV
Mission Creek	94	24	0.259	31	16	0.406
Peshastin Creek	226	39	0.174	6	10	0.733
Chumstick Creek	78	23	0.286	7	10	0.701
Icicle Creek	76	24	0.275	45	19	0.357
Chiwaukum Creek	37	17	0.372	9	9	0.683
Chiwawa River	142	31	0.207	103	26	0.238
Nason Creek	190	34	0.180	148	31	0.210

The estimated fish per redd ratio for steelhead in 2014 was 1.70 (Table 3.19). Multiplying this ratio by the total number of redds estimated in the Wenatchee River upstream from Tumwater

Dam resulted in a spawning escapement of 210 steelhead (Table 3.19). Adding this estimate to the mark-recapture estimates of tributary escapement (260 hatchery + 369 wild = 629 total) indicates that 839 escaped to spawning areas upstream from Tumwater Dam in 2014. This assumes that all steelhead that escaped into tributaries (based on mark-recapture) spawned. Therefore, of the 863 steelhead counted at Tumwater, about 97% were estimated to have spawned upstream from the dam.

**Table 3.19.** Numbers of steelhead counted at Tumwater Dam, fish/redd estimates (based on male-to-female ratios estimated at Tumwater Dam), numbers of steelhead redds counted upstream from Tumwater Dam, total spawning escapement upstream from Tumwater Dam (estimated as the total number of redds times the fish/redd ratio), and the proportion of the Tumwater Dam count that made up the spawning escapement. Beginning in 2014, escapements include estimates from redd counts in the Wenatchee River and mark-recapture techniques in tributaries.

Survey year	Total count at Tumwater Dam	Fish/redd	Number of redds			Spawning escapement <sup>a</sup>	Proportion of Tumwater count that spawned
			Index area	Non-index area	Total redds		
2001	820	2.08	118	19	137	285	0.35
2002	1,720	2.68	296	179	475	1,273	0.74
2003	1,810	1.60	353	88	441	706	0.39
2004	1,869	2.21	277	92	369	815	0.44
2005	2,650	1.61	828	136	964	1,552	0.59
2006	1,053	2.05	192	34	226	463	0.44
2007	657	1.94	105	29	134	260	0.40
2008	1,328	2.81	124	35	159	447	0.34
2009	1,781	1.83	284	107	391	716	0.40
2010	2,270	2.33	546	95	641	1,494	0.66
2011	1,130	1.79	427	33	460	823	0.73
2012	1,055	2.00	273	22	295	590	0.56
2013	1,087	1.65	276	9	285	470	0.43
<i>Average<sup>b</sup></i>	<i>1,488</i>	<i>2.02</i>	<i>333</i>	<i>59</i>	<i>392</i>	<i>763</i>	<i>0.50</i>
2014	863	1.70	124	0	124	839	0.97
<i>Average<sup>c</sup></i>	<i>863</i>	<i>1.70</i>	<i>124</i>	<i>0</i>	<i>124</i>	<i>839</i>	<i>0.97</i>

<sup>a</sup> Escapement estimates before 2014 were based on expanded redd counts in the Wenatchee River and tributaries; escapement estimates beginning in 2014 were based on expanded redd counts within the Wenatchee River and mark-recapture techniques in tributaries.

<sup>b</sup> The average is based on estimates from 2004 to 2013.

<sup>c</sup> The average is based on estimates from 2014 to present.

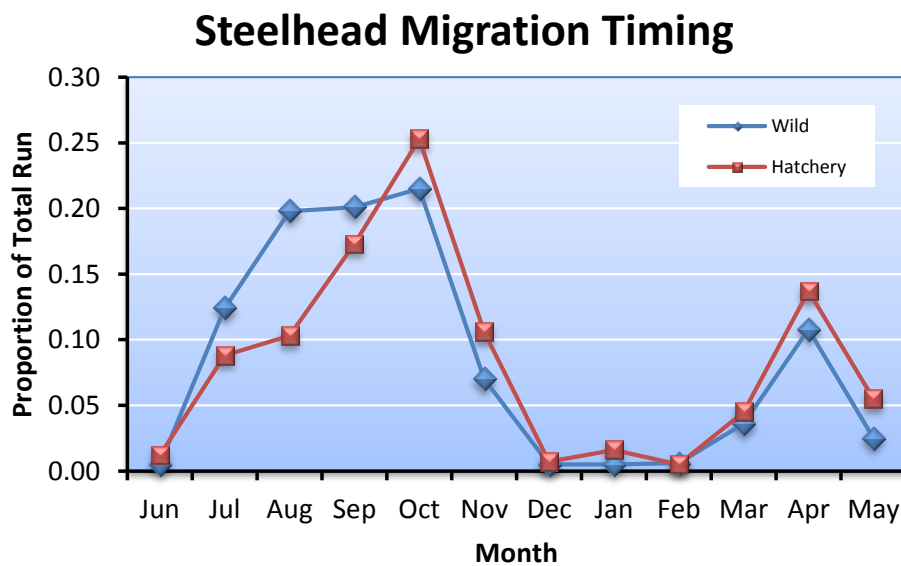
### 3.6 Life History Monitoring

Life history characteristics of steelhead were assessed by examining fish collected at broodstock collection sites, examining videotape at Tumwater Dam, and by reviewing tagging data and fisheries statistics. Prior to brood year 2011, some statistics could not be calculated because few steelhead were tagged with CWTs. Since brood year 2011, all steelhead released from the hatchery program are tagged with CWTs. In addition, about 15,109 of the 2013 brood were PIT

tagged. With the placement of remote PIT tag detectors in spawning streams in 2007 and 2008, statistics such as origin on spawning grounds, stray rates, and SARs can be estimated more accurately.

### Migration Timing

Sampling at Tumwater Dam indicates that steelhead migrate throughout the year; however, the migration distribution is bimodal, indicating that steelhead migrate past Tumwater Dam in two pulses: one pulse during summer-autumn the year before spawning and another during winter-spring the year of spawning (Figure 3.5). Most steelhead passed Tumwater Dam during July through October and April. The highest proportion of both wild and hatchery fish migrated during October.



**Figure 3.5.** Proportion of wild and hatchery steelhead sampled at Tumwater Dam for the combined brood years of 1999-2014.

Because the migration of steelhead is bimodal, we estimated migration statistics separately for each migration pulse (i.e., summer-autumn migration and winter-spring migration). That is, we compared migration statistics for wild and hatchery steelhead passing Tumwater Dam during the summer-autumn period independent of those for the winter-spring migration period. We estimated the week and month that 10%, 50% (median), and 90% of the wild and hatchery steelhead passed Tumwater Dam during the two migration periods. We also estimated the mean weekly and monthly migration timing for wild and hatchery steelhead.

Overall, there was little difference in migration timing of wild and hatchery fish enumerated at Tumwater Dam (Table 3.20a and b; Figure 3.5). For both the summer-autumn and winter-spring migration periods, wild and hatchery steelhead arrived at the dam during the same week and month. The mean and median migration timing for wild and hatchery steelhead were also similar. However, during the summer-autumn migration period, on average, wild steelhead appeared to end their migration about one week earlier than hatchery steelhead.

**Table 3.20a.** The week that 10%, 50% (median), and 90% of the wild and hatchery steelhead passed Tumwater Dam during their summer-autumn migration (June through December) and during their winter-spring migration (January through May), 1999-2014. The average week is also provided for both migration periods. Migration timing is based on video sampling at Tumwater. The presence of eroded fins and/or missing adipose fins was used to distinguish hatchery fish from wild fish during video monitoring at Tumwater Dam. Estimates also include steelhead collected for broodstock.

Spawn year	Origin	Steelhead Migration Time (week)									
		Summer-Autumn Migration (Jun-Dec)					Winter-Spring Migration (Jan-May)				
		10%	50%	90%	Mean	Sample size	10%	50%	90%	Mean	Sample size
1999	Wild	27	32	47	35	81	12	16	17	15	29
	Hatchery	25	31	47	34	47	12	16	18	15	27
2000	Wild	31	36	41	36	238	11	14	18	14	40
	Hatchery	31	34	41	36	194	12	14	16	14	69
2001	Wild	29	34	41	35	391	13	15	17	15	84
	Hatchery	30	38	41	36	227	12	16	17	15	156
2002	Wild	29	39	46	38	810	13	14	17	14	181
	Hatchery	35	42	46	41	610	12	15	18	15	124
2003	Wild	30	33	40	35	731	3	9	16	9	193
	Hatchery	30	35	51	37	372	3	9	15	9	538
2004	Wild	30	40	45	39	644	13	16	18	16	222
	Hatchery	29	40	44	38	677	11	17	19	16	361
2005	Wild	30	39	43	38	986	10	15	17	15	206
	Hatchery	27	38	42	36	1112	12	16	18	15	377
2006	Wild	29	40	43	39	428	12	15	17	15	191
	Hatchery	29	41	43	39	334	4	13	16	12	181
2007	Wild	30	36	41	35	277	11	17	17	15	108
	Hatchery	29	38	43	36	90	11	17	18	16	214
2008	Wild	30	38	43	38	397	13	15	18	16	123
	Hatchery	33	41	45	40	554	14	18	19	17	311
2009	Wild	30	37	46	37	338	13	15	19	15	87
	Hatchery	29	35	46	36	1133	13	16	19	16	229
2010	Wild	31	37	45	38	648	11	15	18	15	171
	Hatchery	31	40	45	40	1207	12	16	19	16	309
2011	Wild	29	36	44	36	797	13	17	19	17	118
	Hatchery	31	39	45	39	991	15	18	19	18	240
2012	Wild	31	34	41	35	642	15	20	20	17	83
	Hatchery	32	39	43	38	715	15	19	19	17	223
2013	Wild	31	36	43	37	755	13	16	18	15	55
	Hatchery	31	42	45	40	1431	16	17	18	16	210
2014	Wild	29	35	41	35	549	14	18	19	17	57

Spawn year	Origin	Steelhead Migration Time (week)									
		Summer-Autumn Migration (Jun-Dec)					Winter-Spring Migration (Jan-May)				
		10%	50%	90%	Mean	Sample size	10%	50%	90%	Mean	Sample size
	Hatchery	32	40	42	38	511	15	17	19	17	78
Average	Wild	30	36	43	37	545	12	15	18	15	122
	Hatchery	30	38	44	38	638	12	16	18	15	228

**Table 3.20b.** The month that 10%, 50% (median), and 90% of the wild and hatchery steelhead passed Tumwater Dam during their summer-autumn migration (June through December) and during their winter-spring migration (January through May), 1999-2014. The average month is also provided for both migration periods. Migration timing is based on video sampling at Tumwater. The presence of eroded fins and/or missing adipose fins was used to distinguish hatchery fish from wild fish during video monitoring at Tumwater Dam. Estimates also include steelhead collected for broodstock.

Spawn year	Origin	Steelhead Migration Time (month)									
		Summer-Autumn Migration (Jun-Dec)					Winter-Spring Migration (Jan-May)				
		10%	50%	90%	Mean	Sample size	10%	50%	90%	Mean	Sample size
1999	Wild	7	8	11	8	81	3	4	4	4	29
	Hatchery	6	8	11	8	47	3	4	4	4	27
2000	Wild	8	9	10	9	238	3	4	5	4	40
	Hatchery	8	8	10	9	194	3	4	4	4	69
2001	Wild	7	8	10	8	391	3	4	4	4	84
	Hatchery	7	9	10	9	227	3	4	4	4	156
2002	Wild	7	9	11	9	810	3	4	4	4	181
	Hatchery	9	10	11	10	610	3	4	5	4	124
2003	Wild	7	8	10	8	731	1	3	4	3	193
	Hatchery	7	8	12	9	372	1	3	4	2	538
2004	Wild	7	10	11	9	644	3	4	4	4	222
	Hatchery	7	10	10	9	677	3	4	5	4	361
2005	Wild	7	9	10	9	986	3	4	4	4	206
	Hatchery	7	9	10	9	1112	3	4	5	4	377
2006	Wild	7	10	10	10	428	3	4	4	4	191
	Hatchery	7	10	10	9	334	1	3	4	3	181
2007	Wild	7	9	10	9	277	3	4	4	4	108
	Hatchery	7	9	10	9	90	3	4	5	4	214
2008	Wild	7	9	10	9	397	3	4	5	4	123
	Hatchery	8	10	11	10	554	4	4	5	4	311
2009	Wild	7	9	11	9	338	3	4	5	4	87
	Hatchery	7	8	11	9	1133	3	4	5	4	229
2010	Wild	8	9	11	9	648	3	4	5	4	171
	Hatchery	8	10	11	10	1207	3	4	5	4	309

Spawn year	Origin	Steelhead Migration Time (month)									
		Summer-Autumn Migration (Jun-Dec)					Winter-Spring Migration (Jan-May)				
		10%	50%	90%	Mean	Sample size	10%	50%	90%	Mean	Sample size
2011	Wild	7	9	11	9	797	4	4	5	4	118
	Hatchery	8	9	11	9	991	4	5	5	5	240
2012	Wild	8	8	10	9	642	4	4	5	4	83
	Hatchery	8	9	10	9	715	4	4	5	4	223
2013	Wild	8	9	10	9	755	4	4	5	4	55
	Hatchery	8	10	11	10	1431	4	4	5	4	210
2014	Wild	7	9	10	9	549	4	4	5	4	57
	Hatchery	8	10	10	9	511	4	4	5	4	78
Average	Wild	7	9	10	9	545	3	4	5	4	122
	Hatchery	7	9	11	9	638	3	4	5	4	228

### Age at Maturity

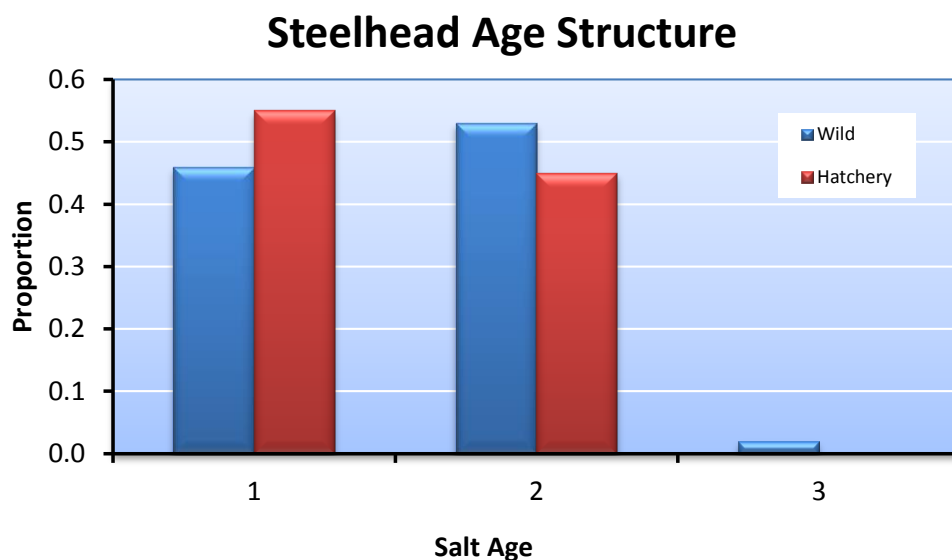
Nearly all steelhead broodstock collected at Tumwater and Dryden dams lived in saltwater 1 to 2 years (saltwater age) (Table 3.21). Very few saltwater age-3 fish returned and those that did were wild fish. On average, there was a difference between the saltwater age at return of wild and hatchery fish. A greater proportion of hatchery fish returned as saltwater age-1 fish than did wild fish. In contrast, a greater number of wild fish returned as saltwater-2 fish than did hatchery fish (Figure 3.6).

**Table 3.21.** Proportions of wild and hatchery steelhead broodstock of different ages collected at Tumwater and Dryden dams, 1998-2014. Age represents the number of years the fish lived in salt water.

Sample year	Origin	Saltwater age			Sample size
		1	2	3	
1998	Wild	0.39	0.61	0.00	35
	Hatchery	0.21	0.79	0.00	43
1999	Wild	0.50	0.48	0.02	58
	Hatchery	0.82	0.18	0.00	67
2000	Wild	0.56	0.44	0.00	39
	Hatchery	0.68	0.32	0.00	101
2001	Wild	0.52	0.48	0.00	64
	Hatchery	0.15	0.85	0.00	114
2002	Wild	0.56	0.44	0.00	99
	Hatchery	0.95	0.05	0.00	113
2003	Wild	0.13	0.85	0.02	63
	Hatchery	0.29	0.71	0.00	92
2004	Wild	0.95	0.05	0.00	85
	Hatchery	0.95	0.05	0.00	132

Sample year	Origin	Saltwater age			Sample size
		1	2	3	
2005	Wild	0.22	0.78	0.00	95
	Hatchery	0.21	0.79	0.00	114
2006	Wild	0.29	0.71	0.00	101
	Hatchery	0.60	0.40	0.00	98
2007	Wild	0.40	0.59	0.00	79
	Hatchery	0.62	0.38	0.00	97
2008	Wild	0.65	0.34	0.01	104
	Hatchery	0.89	0.11	0.00	107
2009	Wild	0.40	0.58	0.20	83
	Hatchery	0.23	0.77	0.0	77
2010	Wild	0.65	0.34	0.01	92
	Hatchery	0.77	0.23	0.00	98
2011	Wild	0.28	0.73	0.00	102
	Hatchery	0.36	0.64	0.00	100
2012	Wild	0.42	0.53	0.05	59
	Hatchery	0.41	0.59	0.00	66
2013	Wild	0.41	0.57	0.02	54
	Hatchery	0.46	0.55	0.00	77
2014	Wild	0.48	0.51	0.02	61
	Hatchery	0.29	0.71	0.00	68
<i>Average</i>	<i>Wild</i>	<i>0.46</i>	<i>0.53</i>	<i>0.02</i>	<i>75</i>
	<i>Hatchery</i>	<i>0.55</i>	<i>0.45</i>	<i>0.00</i>	<i>92</i>





**Figure 3.6.** Proportions of wild and hatchery steelhead of different saltwater ages sampled at Tumwater Dam for the combined years 1998-2014.

### Size at Maturity

On average, hatchery steelhead collected at Tumwater and Dryden dams were about 3 cm smaller than wild steelhead (Table 3.22). This may be related to the fact that more wild steelhead return as saltwater age-2 fish than hatchery steelhead.

**Table 3.22.** Mean fork length (cm) at age (saltwater ages) of hatchery and wild steelhead collected from broodstock, 1998-2014; N = sample size and SD = 1 standard deviation.

Return year	Origin	Steelhead fork length (cm)								
		1-Salt			2-Salt			3-Salt		
		Mean	N	SD	Mean	N	SD	Mean	N	SD
1998	Wild	63	15	4	79	20	5	-	0	-
	Hatchery	61	9	4	73	34	4	-	0	-
1999	Wild	65	29	5	74	28	5	77	1	-
	Hatchery	62	54	4	73	12	4	-	0	-
2000	Wild	64	22	3	74	17	5	-	0	-
	Hatchery	60	57	3	71	27	4	-	0	-
2001	Wild	61	33	6	77	31	5	-	0	-
	Hatchery	62	17	4	72	97	4	-	0	-
2002	Wild	64	55	4	77	44	4	-	0	-
	Hatchery	63	106	4	73	6	4	-	0	-
2003	Wild	69	8	6	77	52	5	91	1	-
	Hatchery	66	27	4	75	65	4	-	0	-
2004	Wild	63	73	6	78	4	2	-	0	-
	Hatchery	61	59	3	73	3	1	-	0	-

Return year	Origin	Steelhead fork length (cm)								
		1-Salt			2-Salt			3-Salt		
		Mean	N	SD	Mean	N	SD	Mean	N	SD
2005	Wild	59	21	4	74	74	5	-	0	-
	Hatchery	59	23	4	72	89	4	-	0	-
2006	Wild	63	27	5	75	67	6	-	0	-
	Hatchery	61	41	4	72	27	5	-	0	-
2007	Wild	64	31	6	76	46	5	-	0	-
	Hatchery	60	60	4	71	36	5	-	0	-
2008	Wild	64	68	4	77	35	4	80	2	-
	Hatchery	60	95	4	72	12	2	-	0	-
2009	Wild	65	33	5	76	48	6	81	2	0
	Hatchery	63	18	4	75	59	5	-	0	-
2010	Wild	64	60	5	74	31	5	76	1	-
	Hatchery	61	53	5	73	23	5	-	0	-
2011	Wild	62	28	5	76	74	5	-	0	-
	Hatchery	60	36	4	74	64	4	-	0	-
2012	Wild	63	25	3	74	31	5	74	3	2
	Hatchery	59	27	3	74	39	4	-	0	-
2013	Wild	61	22	5	77	31	5	74	1	-
	Hatchery	60	35	3	74	42	4	-	0	-
2014	Wild	61	29	4	75	31	4	61	1	-
	Hatchery	60	20	3	72	48	4	-	0	-
<i>Average</i>	<i>Wild</i>	<i>63</i>	<i>34</i>	<i>5</i>	<i>76</i>	<i>39</i>	<i>5</i>	<i>77</i>	<i>1</i>	<i>1</i>
	<i>Hatchery</i>	<i>61</i>	<i>43</i>	<i>4</i>	<i>73</i>	<i>40</i>	<i>4</i>	<i>-</i>	<i>0</i>	<i>-</i>

### Contribution to Fisheries

Nearly all harvest on Wenatchee steelhead occurs within the Columbia basin. Harvest rates on steelhead in the Lower Columbia River fisheries (both tribal and non-tribal) are generally less than 5-10% (NMFS 2004). WDFW regulates steelhead harvest in the Upper Columbia. Under certain conditions, WDFW may allow a harvest on hatchery steelhead (adipose fin clipped fish). The intent is to reduce the number of hatchery steelhead that exceed habitat seeding levels in spawning areas and to increase the proportion of wild steelhead in spawning populations.

### Origin on Spawning Grounds

With the implementation of PIT-tag mark-recapture techniques in 2014, we can estimate the contribution of natural-origin and hatchery-origin fish on the spawning grounds (Table 3.23). Based on mark-recapture estimates, naturally produced steelhead made up about 64.2% of the escapement. Importantly, the abundance of hatchery fish in the upper Wenatchee Basin was regulated at Tumwater Dam. A total of 219 hatchery fish were surplus at the dam resulting in

the passage of 863 steelhead over the dam. Natural-origin steelhead comprised 62.3% (N = 538) of the fish that passed the dam.

**Table 3.23.** Spawning escapement estimates for natural-origin and hatchery-origin steelhead within the Wenatchee River, brood year 2014. Escapement estimates were based on PIT-tag mark-recapture techniques (Truscott et al. 2015). SE = standard error and CV = coefficient of variation.

Tributary	Natural-origin steelhead			Hatchery-origin steelhead		
	Estimate	SE	CV	Estimate	SE	CV
Mission Creek	94	24	0.259	31	16	0.406
Peshastin Creek	226	39	0.174	6	10	0.733
Chumstick Creek	78	23	0.286	7	10	0.701
Icicle Creek	76	24	0.275	45	19	0.357
Chiwaukum Creek	37	17	0.372	9	9	0.683
Chiwawa River	142	31	0.207	103	26	0.238
Nason Creek	190	34	0.180	148	31	0.210
Wenatchee River	340	48	0.141	251	60	0.239
<i>Total</i>	<i>978</i>	<i>96</i>	<i>0.098</i>	<i>545</i>	<i>97</i>	<i>0.178</i>

### Straying

Stray rates of Wenatchee steelhead can be estimated by examining the locations where PIT-tagged hatchery steelhead were last detected. PIT tagging of steelhead began with brood year 2005, which allows estimation of stray rates by brood return. These data only provide estimates for brood years 2005 through 2010, because later brood years are still rearing in the ocean. The target for brood year stray rates should be less than 5%.

Based on PIT-tag analyses, on average, about 25% of the hatchery steelhead returns were last detected in streams outside the Wenatchee River basin (Table 3.24). The numbers in Table 3.24 should be considered rough estimates because they are not based on confirmed spawning (only last detections) and the numbers have not been adjusted for detection efficiencies, which currently do not exist for most PIT-tag detection arrays in tributaries. What these data do indicate is that large numbers of hatchery steelhead from the Wenatchee program have wandered or strayed into the Entiat and Methow rivers, and also into the Deschutes and Tucannon rivers. About 31% of the fish were last detected at Wells Dam.

**Table 3.24.** Number and percent of hatchery-origin Wenatchee steelhead that homed to target spawning areas and the target hatchery program, and number and percent that strayed to non-target spawning areas and hatchery programs for brood years 2005 to 2010. Estimates were based on last detections of PIT-tagged hatchery steelhead. Percent strays should be less than 5%.

Brood Year	Homing				Straying			
	Target streams		Target hatchery*		Non-target stream		Non-target hatchery	
	Number	%	Number	%	Number	%	Number	%
2005	76	75.5	0	0.0	27	24.5	0	0.0
2006	72	61.7	1	0.9	43	37.4	0	0.0

Brood Year	Homing				Straying			
	Target streams		Target hatchery*		Non-target stream		Non-target hatchery	
	Number	%	Number	%	Number	%	Number	%
2007	171	60.6	0	0.0	110	39.4	0	0.0
2008	79	88.8	0	0.0	10	11.2	0	0.0
2009	185	84.3	0	0.0	35	15.7	0	0.0
2010	79	81.4	0	0.0	18	18.6	0	0.0
<b>Average</b>	<b>110</b>	<b>75.2</b>	<b>0</b>	<b>0.1</b>	<b>41</b>	<b>24.5</b>	<b>0</b>	<b>0.0</b>

\* Homing to the target hatchery includes Wenatchee hatchery steelhead that are captured and included as broodstock in the Wenatchee Hatchery program. These hatchery fish are typically collected at Dryden and Tumwater dams.

At this time, we cannot estimate among population stray rates by return year, because we have no estimates of detection efficiencies for PIT-tag interrogation sites within different populations. These data are needed to estimate the total number of Wenatchee steelhead that stray into areas outside the Wenatchee River basin. Finally, for the same reason, we cannot evaluate within-population stray rates.

## Genetics

Genetic studies were conducted to determine the potential effects of the Wenatchee Supplementation Program on natural-origin summer steelhead in the Wenatchee River basin (Seamons et al. 2012; the entire report is appended as Appendix E). Temporal collections of tissue samples from Wenatchee hatchery-produced and natural-origin adults sampled at Dryden and Tumwater dams and from natural-origin juveniles from three Wenatchee River tributaries and the Entiat River were surveyed for genetic variation with 132 genetic (single nucleotide polymorphism loci; SNPs) markers. Peshastin Creek and the Entiat River served as no-hatchery-outplant controls. Genetic data were interrogated for the presence or absence of spatial and temporal trends in allele frequencies, genetic distances, and effective population size.

**Allele Frequencies**—Changes to the summer steelhead hatchery supplementation program had no detectable effect on genetic diversity of wild populations. On average, hatchery-origin adults had higher minor allele frequencies (MAF) than natural-origin adults, which may simply reflect the mixed ancestry of hatchery adults. Both hatchery and natural-origin adults had MAF similar to juveniles collected in spawning tributaries and in the Entiat River. There was no temporal trend in allele frequencies or observed heterozygosity in adult or juvenile collections and allele frequencies in control populations were no different than those still receiving hatchery outplants. This suggests that the hatchery program has had little effect on allele frequencies since broodstock sources changed in 1998 from mixed-ancestry broodstock collected in the Columbia River to using broodstock collected in the Wenatchee River.

**Genetic Distances**—As intended, interbreeding of Wenatchee River hatchery and natural-origin adults reduced the genetic differences between Wells Hatchery adults and Wenatchee River natural-origin adults observed in the first few years after changing the broodstock collection protocol. Although there were detectable genetic differences between hatchery and natural-origin adults, the magnitude of that difference declined over time. Hatchery adults were genetically different from natural-origin adults and juveniles based on pair-wise  $F_{ST}$  and principal components analysis, most likely because of the smaller effective population size ( $N_b$ ) in the

hatchery population (see below). Pair-wise  $F_{ST}$  estimates and genetic distances between hatchery and natural-origin adults collected the same year declined over time suggesting that the interbreeding of hatchery and natural-origin adults in the hatchery (and presumably in the wild) is slowly homogenizing Wenatchee River summer steelhead. Analyses using brood year were inconclusive because of limitations in the data.

**Effective Population Size**—Although the effective population size of the Wenatchee River hatchery steelhead program was consistently small, it does not appear to have caused a reduction in the effective population size of wild populations. On average, estimates of  $N_b$  were much lower and varied less for hatchery adults than for natural-origin adults and juveniles. Estimates of  $N_b$  for hatchery adults declined from the earliest brood years to a stable new low value after broodstock practices were changed in 1998. There was no indication that this had any effect on  $N_b$  in natural-origin adults and juveniles;  $N_b$  estimates for natural-origin adults and juveniles were, on average, higher and varied considerably over the 1998-2010 time period and showed no temporal trend.

### Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For brood years 2001-2014, the approximate PNI Index was less than 0.67 (Table 3.25), suggesting that the hatchery environment has a greater influence on adaptation of Wenatchee steelhead than does the natural environment.

**Table 3.25.** Approximate Proportionate Natural Influence (PNI) Index of the Wenatchee steelhead supplementation program for brood years 2001-2014. PNI was calculated as the proportion of naturally produced steelhead in the hatchery broodstock (pNOB) divided by the proportion of hatchery steelhead on the spawning grounds (pHOS) plus pNOB. NOS = number of natural-origin steelhead on the spawning grounds; HOS = number of hatchery-origin steelhead on the spawning grounds; NOB = number of natural-origin steelhead collected for broodstock; and HOB = number of hatchery-origin steelhead included in hatchery broodstock.

Brood year	Spawners <sup>a</sup>			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
2001	158	127	0.45	51	103	0.33	0.43
2002	731	542	0.43	96	64	0.60	0.59
2003	355	350	0.50	49	90	0.35	0.42
2004	371	445	0.55	75	61	0.55	0.50
2005	690	862	0.56	87	104	0.46	0.45
2006	253	210	0.45	93	69	0.57	0.56
2007	145	115	0.44	76	58	0.57	0.56

Brood year	Spawners <sup>a</sup>			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
2008	168	279	0.62	77	54	0.59	0.48
2009	171	545	0.76	86	73	0.54	0.42
2010	524	970	0.65	96	75	0.56	0.46
2011	351	472	0.57	91	70	0.57	0.50
2012	381	209	0.35	59	65	0.48	0.57
2013	322	148	0.31	49	68	0.42	0.57
2014	476	363	0.43	64	68	0.48	0.53
<b>Average</b>	<b>364</b>	<b>403</b>	<b>0.51</b>	<b>75</b>	<b>73</b>	<b>0.50</b>	<b>0.50</b>

<sup>a</sup> The presence of eroded fins or missing adipose fins was used to distinguish hatchery fish from wild fish during video monitoring at Tumwater Dam. The PNI estimates are appropriate for steelhead spawning upstream from Tumwater Dam. They may not represent PNI for steelhead spawning downstream from Tumwater Dam.

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel time (arithmetic mean days) of hatchery steelhead from release sites (e.g., Chiwawa River, Nason Creek, and Wenatchee River) to McNary Dam, and smolt to adult ratios (SARs) from release to detection at Bonneville Dam (Table 3.26).<sup>5</sup> Over the nine brood years for which PIT-tagged hatchery fish are available, survival rates from the release sites to McNary Dam ranged from 0.173 to 0.785 (note that survival rates of 0.000 were associated with very small sample sizes); SARs from release to detection at Bonneville Dam ranged from 0.006 to 0.038. Average travel time from the release sites to McNary Dam ranged from 15 to 80 days.

Some of the variation in survival rates and travel time was related to release location, type of release, and rearing scenario. For example, on average, steelhead released in the Chiwawa River appeared to have higher survival rates to McNary Dam than did steelhead released in the Wenatchee River or Nason Creek. Within the Chiwawa River, steelhead released volitionally from the circulars appeared to have higher survival rates to McNary Dam than those released volitionally from raceways. In contrast, for steelhead released into Nason Creek and the Wenatchee River, there did not appear to be a difference in survival rates to McNary Dam for fish reared in circulars or within raceways. Based on the available data, SARs varied little among the release locations or rearing scenarios.

Travel time from release to McNary Dam varied among release locations and rearing scenario. In general, steelhead released into the Chiwawa River and Nason Creek appeared to travel more quickly to McNary Dam than did steelhead released into the Wenatchee River. Of those released into the Chiwawa River, steelhead released volitionally from raceways appeared to travel to McNary Dam more quickly than those released from circulars; although there are few replicates and differences in travel times are small. The same also appeared to be true for releases into the Wenatchee River. In contrast, there appeared to be little differences in travel times for steelhead reared in raceways or circulars that were released into Nason Creek.

<sup>5</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

**Table 3.26.** Total number of Wenatchee hatchery summer steelhead released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2005-2012. Standard errors are shown in parentheses. NA = not available (i.e., for SARs, not all the adults from the release groups have returned to the Columbia River).

Brood year	Release location <sup>a</sup>	Crosses <sup>b</sup>	Type of release	Rearing scenario <sup>c</sup>	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2004	Chiwawa	HxW	NA	Turtle Rock	2,439	0.480 (0.037)	26.9 (59.5)	0.011 (0.002)
	Chiwawa	WxW	NA	Turtle Rock	853	0.485 (0.054)	21.1 (8.8)	0.008 (0.003)
	Nason	WxW	NA	Turtle Rock	8,826	0.412 (0.017)	26.7 (56.1)	0.010 (0.001)
	Wenatchee	HxH	NA	Turtle Rock	6,662	0.645 (0.029)	15.8 (6.1)	0.032 (0.002)
	Wenatchee	HxW	NA	Turtle Rock	10,422	0.592 (0.022)	18.1 (7.4)	0.019 (0.001)
2005	Chiwawa	HxW	NA	Turtle Rock	3,448	0.540 (0.065)	22.6 (27.2)	0.017 (0.002)
	Wenatchee	HxH	NA	Turtle Rock	8,610	0.656 (0.057)	20.1 (35.8)	0.017 (0.001)
	Wenatchee	HxW	NA	Turtle Rock	5,021	0.649 (0.074)	20.2 (9.0)	0.014 (0.002)
2006	NA	NA	NA	NA	NA	NA	NA	NA
2007	Chiwawa	HxW	NA	Turtle Rock	2,882	0.520 (0.057)	22.3 (7.9)	0.020 (0.003)
	Chiwawa	WxW	NA	Turtle Rock	785	0.467 (0.069)	18.7 (9.0)	0.038 (0.007)
	Nason	WxW	NA	Turtle Rock	8,060	0.505 (0.030)	22.3 (24.1)	0.030 (0.002)
	Wenatchee	HxW	NA	Turtle Rock	9,047	0.631 (0.041)	18.2 (17.2)	0.038 (0.002)
2008	Chiwawa	HxW L	NA	Turtle Rock	2,008	0.574 (0.080)	20.3 (7.0)	0.006 (0.002)
	Chiwawa	WxW	NA	Turtle Rock	1,457	0.546 (0.090)	31.6 (108.5)	0.010 (0.003)
	Nason	WxW	NA	Turtle Rock	7,951	0.500 (0.037)	21.4 (17.5)	0.014 (0.001)
	Wenatchee	HxW E	NA	Turtle Rock	4,517	0.511 (0.044)	19.5 (7.7)	0.008 (0.001)
	Wenatchee	HxW L	NA	Turtle Rock	6,710	0.545 (0.038)	19.3 (6.8)	0.010 (0.001)
2009	Chiwawa	HxW E	Forced	Turtle Rock	4,874	0.576 (0.076)	24.3 (8.3)	0.012 (0.002)
	Chiwawa	HxW E	Volitional	Chiwawa Circ	8,653	0.785 (0.100)	19.4 (26.0)	0.007 (0.001)
	Nason	WxW	Forced	Turtle Rock	8,918	0.504 (0.042)	27.2 (26.6)	0.017 (0.001)
	Wenatchee	HxW E	Forced	Turtle Rock	11,300	0.543 (0.041)	25.8 (54.8)	0.014 (0.001)
	Wenatchee	HxW E	Volitional	Blackbird	2,184	0.317 (0.054)	80.4 (11.7)	0.010 (0.002)
	Wenatchee	WxW	Volitional	Rohlfing	566	0.443 (0.187)	78.1 (8.6)	0.014 (0.005)
2010	Chiwawa	WxW	Forced	Turtle Rock	4,226	0.586 (0.057)	24.4 (60.1)	NA
	Nason	WxW	Forced	Turtle Rock	5,256	0.548 (0.044)	23.5 (53.3)	NA
	Wenatchee	HxH	Forced	Turtle Rock	8,506	0.583 (0.053)	30.2 (50.1)	NA
	Wenatchee	HxH	Volitional	Blackbird	6,493	0.659 (0.062)	18.2 (20.8)	NA
	Wenatchee	HxH	Volitional	Chiwawa Circ	10,031	0.413 (0.043)	21.6 (66.1)	NA
2011	Chiwawa	WxW	Volitional	RCY	3,603	0.407 (0.056)	15.1 (8.3)	NA

Brood year	Release location <sup>a</sup>	Crosses <sup>b</sup>	Type of release	Rearing scenario <sup>c</sup>	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
	Nason	WxW	Volitional	RCY	4,065	0.334 (0.042)	20.9 (60.9)	NA
	Wenatchee	WxW	Non-movers	Circular	1,122	0.354 (0.228)	40.6 (89.1)	NA
	Wenatchee	WxW	Non-movers	RCY	2,395	0.368 (0.084)	22.7 (57.0)	NA
	Wenatchee	WxW	Volitional	Blackbird	2,099	0.660 (0.016)	48.2 (90.0)	NA
	Wenatchee	WxW	Volitional	Circular	7,206	0.277 (0.042)	31.6 (74.3)	NA
	Wenatchee	WxW	Volitional	RCY	4,422	0.327 (0.032)	15.2 (25.6)	NA
	All	WxW	NA	Circ or RCY	5,107	0.213 (0.023)	24.4 (59.5)	NA
2012	Chiwawa	HxH	Volitional	RCY	2,891	0.407 (0.057)	15.2 (7.2)	NA
	Nason	WxW	Forced	Circular	4,271	0.378 (0.065)	25.0 (33.1)	NA
	Nason	WxW	Volitional	Circular	5,404	0.364 (0.048)	24.9 (31.6)	NA
	L. Wenatchee	HxH	Volitional	RCY	2,224	0.573 (0.138)	18.7 (8.4)	NA
	U. Wenatchee	HxH	Forced	RCY	2,556	0.490 (0.102)	27.8 (54.7)	NA
	Wenatchee	HxH	Volitional	Blackbird	1,658	0.428 (0.092)	50.0 (7.6)	NA
	All	HxH	NA	RCY	769	0.455 (0.291)	23.5 (11.8)	NA
2013	All	WxW	NA	Circular	5,397	0.327 (0.049)	25.4 (45.0)	NA
	Chiwawa	Mixed	Volitional	RCY	1,567	0.352 (0.063)	15.2 (7.0)	NA
	Nason	Mixed	Volitional	RCY	3,796	0.447 (0.115)	20.2 (9.4)	NA
	Nason	WxW	Non-movers	Circular	74	0.000 (-)	0.0 (-)	NA
	Nason	WxW	Volitional	Circular	1,594	0.173 (0.043)	18.0 (5.4)	NA
	L. Wenatchee	Mixed	Non-movers	RCY	3,275	0.359 (0.167)	21.3 (10.2)	NA
	U. Wenatchee	Mixed	Volitional	RCY	2,862	0.452 (0.080)	16.3 (9.7)	NA
	Wenatchee	HxH	Volitional	Blackbird	819	0.337 (0.128)	33.5 (11.9)	NA
	All	HxH	NA	RCY	907	0.000 (-)	0.0 (-)	NA
All	WxW	NA	Circ or RCY	232	0.000 (-)	0.0 (-)	NA	

<sup>a</sup> All = Chiwawa River, Nason Creek, and the Wenatchee River.

<sup>b</sup> HxH = hatchery by hatchery cross; WxW = wild by wild cross; Mixed = both HxH and WxW crosses; E = early; and L = late.

<sup>c</sup> Circ = circulars; RCY = raceway.

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). For brood years 1998-2007, NRR for summer steelhead in the Wenatchee



River basin averaged 0.74 (range, 0.13-3.10) if harvested fish were included in the estimate (Table 3.27).

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and were calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. These rates should be greater than the NRRs and greater than or equal to 19.2 (the calculated target value in Hillman et al. 2013). In nearly all years, HRRs were greater than NRRs (Table 3.27). HRRs exceeded the estimated target value of 19.2 in one of the ten years.

**Table 3.27.** Broodstock collected, spawning escapements, natural and hatchery-origin recruits (NOR and HOR), and natural and hatchery replacement rates (NRR and HRR with harvest) for summer steelhead in the Wenatchee River basin, brood years 1998-2007.

Brood year	Broodstock Collected	Spawning Escapement	Harvest included			
			HOR	NOR	HRR	NRR
1998	78	602	148	1,867	1.89	3.10
1999	125	343	1,944	334	15.55	0.97
2000	120	1,030	312	878	2.60	0.85
2001	178	1,655	10,335	1,050	58.06	0.66
2002	162	5,000	1,905	515	11.76	0.13
2003	155	2,598	956	504	6.17	0.27
2004	217	2,949	2,538	728	11.70	0.25
2005	209	3,609	3,106	904	14.86	0.25
2006	199	2,219	1,454	1,007	7.31	0.45
2007	176	880	535	430	3.04	0.49
<i>Average</i>	<i>162</i>	<i>2,089</i>	<i>2,323</i>	<i>822</i>	<i>13.29</i>	<i>0.74</i>

### Smolt-to-Adult Survivals

Smolt-to-adult ratios (SARs) are calculated as the number of returning hatchery adults divided by the number of tagged hatchery smolts released. SARs are generally based on CWT returns. However, prior to brood year 2011, Wenatchee steelhead were not extensively tagged with CWTs. Therefore, elastomer-tagged fish were used to estimate SARs from release to capture at Priest Rapids Dam. With the return of brood year 2011, SARs will be based on CWT returns.

SARs (not adjusted for tag loss) for Wenatchee steelhead ranged from 0.0009 to 0.0315 (mean = 0.0097) for brood years 1996-2007 (Table 3.28).

**Table 3.28.** Smolt-to-adult ratios (SARs) for Wenatchee hatchery steelhead, 1996-2007. Estimates were based on elastomer tags recaptured at Priest Rapids Dam. SARs were not adjusted for tag loss after release.

Brood year	Number of tagged smolts released	SAR
1996	348,693	0.0034
1997	429,422	0.0041
1998	172,078	0.0009

Brood year	Number of tagged smolts released	SAR
1999	175,661	0.0111
2000	184,639	0.0017
2001	335,933	0.0308
2002	302,060	0.0063
2003	374,867	0.0025
2004	294,114	0.0038
2005	452,184	0.0107
2006	258,697	0.0100
2007	306,690	0.0315
<i>Average</i>	<i>302,920</i>	<i>0.0097</i>

## 3.7 ESA/HCP Compliance

### Broodstock Collection

Collection of brood year 2013 broodstock for Wenatchee summer steelhead at Dryden and Tumwater dams began on 1 July and ended on 4 October 2013 at Dryden Dam and 17 November 2013 at Tumwater Dam consistent with the collection period identified in the 2013 broodstock collection protocol. The broodstock collection achieved a total collection of 147 steelhead, including 63 natural-origin steelhead identified in the annual broodstock collection protocols.

About 2,117 steelhead were handled and released (or surplused) at Tumwater and Dryden dams during brood year 2013 Wenatchee steelhead broodstock collection. Most were hatchery-origin fish handled at Tumwater Dam and ultimately surplused to meet the pHOS objective upstream from Tumwater Dam. Fish released at Dryden Dam were released because the weekly quota for hatchery or wild steelhead had been attained, but not for both hatchery and wild fish, or because they were non-target fish (red/green VIE tagged), or they were unidentifiable hatchery-origin steelhead. All steelhead released were allowed to fully recover from the anesthesia and released immediately upstream from the trap sites.

In addition to steelhead encountered at Dryden Dam during steelhead broodstock collection, an estimated 65 spring Chinook salmon were captured and released unharmed immediately upstream from the trap facility. Consistent with ESA Section 10 Permit 1395 impact minimization measures, all ESA species handled were subject of water-to-water transfers.

### Hatchery Rearing and Release

The 2013 brood Wenatchee steelhead reared throughout all life stages without significant mortality (defined as >10% population mortality associated with a single event). However, the 2013 brood had both poor survival of females to spawn combined and poor fertilization to eyed-egg survival resulting in an unfertilized-to-release survival of 67.6%, which was less than the program target of 81% (see Section 3.2).

Juvenile rearing occurred at three separate facilities including Eastbank Fish Hatchery, Chelan Fish Hatchery, and the Chiwawa Acclimation Facility. Multiple facilities were used to take

advantage of variable water temperatures to manipulate growth of juveniles from different parental crosses. Typically, wild steelhead spawn later than their hatchery cohort and are therefore reared at Chelan Fish Hatchery on warmer water to accelerate their growth so they achieve a size-at-release similar to HxH parental cross progeny reared on cooler water at Eastbank Fish Hatchery. All parental cross groups received final rearing and over-winter acclimation at the Chiwawa Acclimation Facility on Wenatchee River and Chiwawa River surface water before direct release (scatter planting) in the Wenatchee River basin.

The 2013 brood steelhead smolt release in the Wenatchee River basin totaled 229,836 smolts, representing about 92.9% of the program target of 247,300 smolts identified in the Rocky Reach and Rock Island Dam HCPs and in ESA Section 10 Permit 1395. As specified in ESA Section 10 Permit 1395, all steelhead smolts released were externally marked or internally tagged and a representative number were PIT tagged (see Section 3.2).

### Hatchery Effluent Monitoring

Per ESA Permits 1196, 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January 2014 through 31 December 2014. NPDES monitoring and reporting for Chelan PUD Hatchery Programs during 2014 are provided in Appendix F.

### Smolt and Emigrant Trapping

Per ESA Section 10 Permit No. 1395, the permit holders are authorized a direct take of up to 20% of the emigrating steelhead population and a lethal take not to exceed 2% of the fish captured (NMFS 2003). Based on the estimated wild steelhead population (smolt trap expansion) and hatchery juvenile steelhead population estimate (hatchery release data) for the Wenatchee River basin, the reported steelhead encounters during the 2014 emigration complied with take provisions in the Section 10 permit and are detailed in Table 3.29. Additionally, juvenile fish captured at the trap locations were handled consistent with provisions in ESA Section 10 Permit 1395 Section B.

**Table 3.29.** Estimated take of Upper Columbia River steelhead resulting from juvenile emigration monitoring in the Wenatchee River basin, 2014. NA = not available.

Trap location	Population estimate				Number trapped				Total	Take allowed by Permit
	Wild	Hatchery <sup>b</sup>	Parr	Fry	Wild	Hatchery	Parr	Fry		
<b>Chiwawa Trap</b>										
Population	NA	23,400	NA	NA	49	239	1,889	529	2,706	
Encounter rate	NA	NA	NA	NA	NA	0.0102	NA	NA	NA	0.20
Mortality <sup>b</sup>	NA	NA	NA	NA	0	0	20	4	24	
Mortality rate	NA	NA	NA	NA	0.0000	0.0042	0.0106	0.0076	0.0089	0.02
<b>Lower Wenatchee Trap</b>										
Population	NA	229,836	NA	NA	80	494	102	117	793	
Encounter rate	NA	NA	NA	NA	NA	0.0022	NA	NA	NA	0.20
Mortality <sup>b</sup>	NA	NA	NA	NA	1	12	1	0	14	

Trap location	Population estimate				Number trapped				Total	Take allowed by Permit
	Wild	Hatchery <sup>a</sup>	Parr	Fry	Wild	Hatchery	Parr	Fry		
Mortality rate	NA	NA	NA	NA	0.0125	0.0243	0.0098	0.0000	0.0177	0.02
<b>Wenatchee River Basin Total</b>										
Population	NA	229,836	NA	NA	129	733	1,991	646	3,499	
Encounter rate	NA	NA	NA	NA	NA	0.0032	NA	NA	NA	0.20
Mortality <sup>b</sup>	NA	NA	NA	NA	1	12	21	4	38	
Mortality rate	NA	NA	NA	NA	0.0076	0.0164	0.0105	0.0062	0.0109	0.02

<sup>a</sup> 2014 smolt release data for the Wenatchee River basin.

<sup>b</sup> Mortality includes trapping and PIT-tag mortalities.

### Spawning Surveys

Steelhead spawning ground surveys were conducted in the Wenatchee River basin during 2014, as authorized by ESA Section 10 Permit No. 1395. Because of the difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical, and extreme caution was used to avoid established redds when wading was required.

### Stock Assessment at Priest Rapids Dam

Upper Columbia River steelhead stock assessment sampling at Priest Rapids Dam (PRD) is authorized through ESA Section 10 Permit No. 1395 (NMFS 2003). Permit authorizations include interception and biological sampling of up to 15% of the Upper Columbia River steelhead passing PRD to determine upriver adult population size, estimate hatchery to wild ratios, determine age-class contribution, and evaluate the need for managing hatchery steelhead consistent with ESA recovery objectives, which include fully seeding spawning habitat with naturally produced Upper Columbia River steelhead supplemented with artificially propagated steelhead (NMFS 2003). The 2012-2013 run-cycle report (BY 2013) for stock assessment sampling at Priest Rapids Dam was compiled under provisions of ESA Section 10 Permit 1395. Data and reporting information are included in Appendix G.

## SECTION 4: WENATCHEE SOCKEYE SALMON

The goal of sockeye salmon supplementation in the Wenatchee Basin was to use artificial production to replace adult production lost because of mortality at Rock Island Dam, while not reducing the natural production or long-term fitness of sockeye in the basin. The Rock Island Fish Hatchery Complex began operation in 1989 under funding from Chelan PUD. The Complex operated originally through the Rock Island Settlement Agreement, but since 2004 has operated under the Anadromous Fish Agreement and Habitat Conservation Plans.

Adult sockeye were collected for broodstock from the run-at-large at Tumwater Dam. Beginning in 2011, because of passage delays at Tumwater Dam during trapping operations, sockeye broodstock were collected at Dryden Dam. The goal was to collect up to 260 natural-origin adult sockeye for the program. Broodstock collection occurred from about 7 July through 28 August with trapping occurring no more than 16 hours per day, three days a week at Tumwater Dam and up to seven days per week at the Dryden Dam left and right-bank facilities.

Adult sockeye were held and spawned at Eastbank Fish Hatchery. The fertilized eggs were also incubated at the hatchery. For brood years 1989 through 1998, unfed fry were transferred from the hatchery to Lake Wenatchee net pens. From 1998 to 2011, juvenile sockeye were reared at Eastbank Fish Hatchery until July when they were transferred to the net pens. The initial rearing at Eastbank was to increase growth rates. During most years up through 2005, juvenile sockeye were released from net pens at two different times, August and November. Since 2006, all juvenile sockeye were released in late October.

The production goal for the Wenatchee sockeye supplementation program was to release 200,000 subyearlings into Lake Wenatchee at 20 fish per pound. Targets for fork length and weight were 133 mm (CV = 9.0) and 22.7 g, respectively. Over 90% of these fish were marked with CWTs. In addition, from 2006-2011, about 15,000 juvenile sockeye were PIT tagged annually. Following an evaluation of the supplementation program in 2011, the Hatchery Committees decided to convert the Wenatchee sockeye hatchery program to summer steelhead in 2012. Monitoring occurs annually to track the status of the natural population.

### 4.1 Broodstock Sampling

As noted above, the Wenatchee sockeye program was terminated in 2012. Thus, no broodstock have been collected since 2011 and the release of juvenile sockeye into Lake Wenatchee in 2012 (2011 brood) was the last. Therefore, this section presents the history of the program and tracks the juveniles from the 2011 brood that were released as parr into Lake Wenatchee in 2012. Some of these fish began their smolt migrations in 2013.

#### Origin of Broodstock

Wenatchee sockeye broodstock have not been collected since 2011. Table 4.1 shows the history of the number of broodstock that were collected during the period 1989 to 2011.

**Table 4.1.** Numbers of wild and hatchery sockeye salmon collected for broodstock, numbers that died before spawning, and numbers of sockeye spawned, 1989-2011. Unknown origin fish (i.e., undetermined by scale analysis, no CWT or fin clips, and no additional hatchery marks) were considered naturally produced. Mortality includes sockeye that died of natural causes typically near the end of spawning and were not needed for the program, surplus sockeye killed at spawning, sockeye that died but were not recovered from the net pens, and sockeye that may have jumped out of the net pens.

Brood year	Wild sockeye					Hatchery sockeye					Total number spawned
	Number collected	Prespawm loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawm loss <sup>a</sup>	Mortality	Number spawned	Number released	
1989	299	93	47	115	44	0	0	0	0	0	115
1990	333	7	7	302	17	0	0	0	0	0	302
1991	357	18	16	199	124	0	0	0	0	0	199
1992	362	18	5	320	19	0	0	0	0	0	320
1993	307	79	21	207	0	0	0	0	0	0	207
1994	329	15	9	236	69	5	0	0	5	0	241
1995	218	5	7	194	12	3	0	0	3	0	197
1996	291	2	0	225	64	20	0	0	0	20	225
1997	283	12	3	192	76	19	0	0	19	0	211
1998	225	37	25	122	41	6	0	0	6	0	128
1999	90	7	1	79	3	60	0	0	60	0	139
2000	256	19	1	170	66	5	0	0	5	0	175
2001	252	27	10	200	15	8	1	0	7	0	207
2002	257	0	1	256	0	0	0	0	0	0	256
2003	261	12	9	198	42	0	0	0	0	0	198
2004	211	13	12	177	9	0	0	0	0	0	177
2005	243	29	12	166	36	0	0	0	0	0	166
2006	260	2	4	214	40	0	0	0	0	0	214
2007	248	15	3	210	20	0	0	0	0	0	210
2008	258	4	11	243	0	2	0	0	2	0	245
2009	258	5	14	239	0	3	0	3	0	0	239
2010	256	3	0	198	55	0	0	0	0	0	256
2011	204	0	8	196	0	0	0	0	0	0	196
<b>Average</b>	<b>263</b>	<b>18</b>	<b>10</b>	<b>203</b>	<b>33</b>	<b>6</b>	<b>0</b>	<b>0</b>	<b>5</b>	<b>1</b>	<b>210</b>

<sup>a</sup> Pre-spawn loss represents the number of fish that died during the holding period before spawning. Mortality is the number of fish that were surplus following spawning.

### Age/Length Data

Ages of sockeye were determined from scales and otoliths collected from broodstock and are shown in Table 4.2.

**Table 4.2.** Percent of hatchery and wild sockeye salmon of different ages (total age) collected from broodstock, 1994-2011.

Return year	Origin	Total age		
		4	5	6
1994	Wild	57.3	41.7	1.0
	Hatchery	40.0	60.0	0.0

Return year	Origin	Total age		
		4	5	6
1995	Wild	77.3	20.7	2.0
	Hatchery	66.7	33.3	0.0
1996	Wild	65.8	34.2	0.0
	Hatchery	0.0	0.0	0.0
1997	Wild	86.5	13.5	0.0
	Hatchery	57.9	42.1	0.0
1998	Wild	9.9	88.6	1.5
	Hatchery	66.7	33.3	0.0
1999	Wild	21.8	74.7	3.5
	Hatchery	90.0	8.3	1.7
2000	Wild	97.7	2.3	0.0
	Hatchery	100.0	0.0	0.0
2001	Wild	69.9	29.6	0.5
	Hatchery	71.4	28.6	0.0
2002	Wild	31.6	67.6	0.8
	Hatchery	0.0	0.0	0.0
2003	Wild	2.6	90.5	6.9
	Hatchery	0.0	0.0	0.0
2004	Wild	97.5	2.0	0.5
	Hatchery	0.0	0.0	0.0
2005	Wild	74.2	25.8	0.0
	Hatchery	0.0	0.0	0.0
2006	Wild	34.0	65.5	0.5
	Hatchery	0.0	0.0	0.0
2007	Wild	1.9	88.4	9.7
	Hatchery	0.0	0.0	0.0
2008	Wild	95.0	4.0	1.0
	Hatchery	100.0	0.0	0.0
2009	Wild	78.5	21.5	0.0
	Hatchery	100.0	0.0	0.0
2010	Wild	67.4	32.6	0.0
	Hatchery	0.0	0.0	0.0
2011	Wild	53.7	44.3	2.0
	Hatchery	0.0	0.0	0.0
<b>Average</b>	<b>Wild</b>	<b>56.8</b>	<b>41.5</b>	<b>1.7</b>
	<b>Hatchery</b>	<b>38.5</b>	<b>11.4</b>	<b>0.1</b>

Lengths and ages of sockeye sampled during the life of the program are provided in Table 4.3.

**Table 4.3.** Mean fork length (cm) at age (total age) of hatchery and wild sockeye salmon collected for broodstock, 1994-2011; SD = 1 standard deviation.

Return year	Origin	Sockeye fork length (cm)								
		Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD
1994	Wild	56	125	3	55	91	3	54	2	3
	Hatchery	57	2	1	56	3	1	-	0	-
1995	Wild	51	153	2	55	41	4	54	4	5
	Hatchery	53	2	4	59	1	-	-	0	-
1996	Wild	52	146	4	53	76	3	-	0	-
	Hatchery	-	0	-	-	0	-	-	0	-
1997	Wild	50	166	3	53	26	5	-	0	-
	Hatchery	54	11	4	59	8	2	-	0	-
1998	Wild	51	13	4	55	117	3	53	2	3
	Hatchery	52	4	2	55	2	8	-	0	-
1999	Wild	52	19	4	50	65	4	56	3	1
	Hatchery	50	54	3	56	5	4	56	1	-
2000	Wild	52	167	2	54	4	3	-	0	-
	Hatchery	54	5	1	-	0	-	-	0	-
2001	Wild	54	151	3	56	65	4	58	1	-
	Hatchery	51	5	5	55	2	4	-	0	-
2002	Wild	54	77	2	56	165	4	57	2	0
	Hatchery	-	0	-	-	0	-	-	0	-
2003	Wild	54	5	4	60	172	2	60	13	4
	Hatchery	-	0	-	-	0	-	-	0	-
2004	Wild	53	192	3	56	4	3	63	1	-
	Hatchery	-	0	-	-	0	-	-	0	-
2005	Wild	51	132	3	57	46	4	-	0	-
	Hatchery	-	0	-	-	0	-	-	0	-
2006	Wild	52	70	3	56	135	4	54	2	3
	Hatchery	-	0	-	-	0	-	-	0	-
2007	Wild	57	4	2	58	182	5	58	20	5
	Hatchery	-	0	-	-	0	-	-	0	-
2008	Wild	52	245	3	52	11	3	62	2	6
	Hatchery	53	2	3	-	-	-	-	-	-
2009	Wild	54	197	3	59	54	4	-	-	-
	Hatchery	54	2	1	-	-	-	-	-	-
2010	Wild	56	130	2	57	63	4	-	-	-
	Hatchery	-	-	-	-	-	-	-	-	-
2011	Wild	55	109	2	59	90	3	61	4	3



Return year	Origin	Sockeye fork length (cm)								
		Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD
	Hatchery	-	-	-	-	-	-	-	-	-
Average	Wild	53	116	3	55	78	4	57	3	3
	Hatchery	53	5	3	57	2	4	56	1	-

### Sex Ratios

Sex ratios of wild and hatchery sockeye collected during the life of the sockeye hatchery program are presented in Table 4.4.

**Table 4.4.** Numbers of male and female wild and hatchery sockeye collected for broodstock, 1989-2011. Ratios of males to females are also provided.

Return year	Number of wild sockeye			Number of hatchery sockeye			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
1989	162	137	1.18:1.00	0	0	-	1.18:1.00
1990	177	156	1.13:1.00	0	0	-	1.13:1.00
1991	260	97	2.68:1.00	0	0	-	2.68:1.00
1992	180	182	0.99:1.00	0	0	-	0.99:1.00
1993	130	177	0.73:1.00	0	0	-	0.73:1.00
1994	162	167	0.97:1.00	1	4	0.25:1.00	0.95:1.00
1995	102	116	0.88:1.00	1	2	0.50:1.00	0.87:1.00
1996	150	161	0.93:1.00	0	0	-	0.93:1.00
1997	139	144	0.97:1.00	10	9	1.11:1.00	0.97:1.00
1998	115	110	1.05:1.00	2	4	0.50:1.00	1.03:1.00
1999	22	68	0.32:1.00	37	23	1.61:1.00	0.65:1.00
2000	155	101	1.53:1.00	3	2	1.50:1.00	1.53:1.00
2001	114	138	0.83:1.00	4	4	1.00:1.00	0.83:1.00
2002	128	129	0.99:1.00	0	0	-	0.99:1.00
2003	161	100	1.61:1.00	0	0	-	1.61:1.00
2004	108	103	1.05:1.00	0	0	-	1.05:1.00
2005	130	113	1.15:1.00	0	0	-	1.15:1.00
2006	130	130	1.00:1.00	0	0	-	1.00:1.00
2007	127	121	1.05:1.00	0	0	-	1.05:1.00
2008	127	131	0.97:1.00	1	1	1.00:1.00	0.97:1.00
2009	133	125	1.06:1.00	0	3	0.00:1.00	1.04:1.00
2010	127	129	0.98:1.00	0	0	-	0.98:1.00
2011	106	98	1.08:1.00	0	0	-	1.08:1.00
<b>Total</b>	<b>2,074</b>	<b>2,017</b>	<b>1.03:1.00</b>	<b>58</b>	<b>48</b>	<b>1.21</b>	<b>1.03:1.00</b>

## Fecundity

Fecundities of sockeye collected during the life of the hatchery program are presented in Table 4.5.

**Table 4.5.** Mean fecundity of female sockeye salmon collected for broodstock, 1989-2011. Fecundities were determined from pooled egg lots and were not identified for individual females.

Return year	Mean fecundity
1989	2,344
1990	2,225
1991	2,598
1992	2,341
1993	2,340
1994	2,798
1995	2,295
1996	2,664
1997	2,447
1998	2,813
1999	2,319
2000	2,673
2001	2,960
2002	2,856
2003	3,511
2004	2,505
2005	2,718
2006	2,656
2007	3,115
2008	2,555
2009	2,459
2010	2,782
2011	2,960
<i>Average</i>	<i>2,649</i>

## 4.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Numbers of eggs taken from sockeye broodstock during the life of the sockeye hatchery program are shown in Table 4.6.

**Table 4.6.** Numbers of eggs taken from sockeye broodstock, 1989-2011.

Return year	Number of eggs taken
1989	133,600
1990	326,267
1991	231,254
1992	381,561
1993	231,700
1994	338,562
1995	247,900
1996	314,390
1997	254,459
1998	163,278
1999	190,732
2000	227,234
2001	301,925
2002	356,982
2003	319,470
2004	225,499
2005	211,985
2006	292,136
2007	302,363
2008	316,476
2009	304,963
2010	278,171
2011	290,046
<i>Average</i>	<i>271,389</i>

*Number of acclimation days*

During the life of the program, Wenatchee sockeye were only acclimated on Lake Wenatchee water in net pens. Acclimation days are presented in Table 4.7.

**Table 4.7.** Water source and mean acclimation period for Wenatchee sockeye, brood years 1989-2011.

Brood year	Release year	Transfer date	Release date	Number of Days	Water source
1989	1990	5-Apr	24-Oct	202	Lake Wenatchee
1990	1991	10-Apr	19-Oct	192	Lake Wenatchee
1991	1992	1-Apr	20-Oct	202	Lake Wenatchee
1992	1993	5-Apr	7-Sep	155	Lake Wenatchee
		5-Apr	26-Oct	204	Lake Wenatchee
1993	1994	5-Apr	1-Sep	149	Lake Wenatchee

Brood year	Release year	Transfer date	Release date	Number of Days	Water source
		5-Apr	17-Oct	195	Lake Wenatchee
1994	1995	4-Apr	15-Sep	164	Lake Wenatchee
		4-Apr	23-Oct	202	Lake Wenatchee
1995	1996	4-Apr	25-Oct	204	Lake Wenatchee
1996	1997	4-Apr	22-Oct	201	Lake Wenatchee
1997	1998	1-Apr	9-Nov	222	Lake Wenatchee
1998	1999	1-Apr	29-Oct	211	Lake Wenatchee
1999	2000	25-Jul	28-Aug	34	Lake Wenatchee
		26-Jul	1-Nov	98	Lake Wenatchee
2000	2001	2-Jul	27-Aug	56	Lake Wenatchee
		3-Jul	27-Sep	86	Lake Wenatchee
2001	2002	15-Jul	28-Aug	44	Lake Wenatchee
		16-Jul	22-Sep	68	Lake Wenatchee
2002	2003	30-Jun	25-Aug	56	Lake Wenatchee
		1-Jul	22-Oct	113	Lake Wenatchee
2003	2004	6-Jul	25-Aug	50	Lake Wenatchee
		7-Jul	3-Nov	119	Lake Wenatchee
2004	2005	5-Jul	29-Aug	55	Lake Wenatchee
		6-Jul	2-Nov	120	Lake Wenatchee
2005	2006	11-Jul	30-Oct	111	Lake Wenatchee
2006	2007	9-10 Jul	31-Oct	113-114	Lake Wenatchee
2007	2008	7-8 Jul	29-Oct	113-114	Lake Wenatchee
2008	2009	21-Jul	28-Oct	100	Lake Wenatchee
2009	2010	19-20, 23-Jul	27-Oct	97-101	Lake Wenatchee
2010	2011	6, 11-12-Jul	26-Oct	107-113	Lake Wenatchee
2011	2012	9-10-Jul	29-Oct	112-113	Lake Wenatchee

## Release Information

### *Numbers released*

Numbers of juvenile sockeye released into Lake Wenatchee during the life of the program are shown in Table 4.8a. Coded wire tag marking rates and numbers of PIT-tagged juvenile sockeye released are also shown in Table 4.8a.

**Table 4.8.** Total number of sockeye parr released and numbers of released fish with CWTs and PIT tags for brood years 1989-2011. The release target for sockeye was 200,000 fish.

Brood year	Release year	CWT mark rate	Number of released fish with PIT tags	Number released
1989	1990	Not marked	0	108,400
1990	1991	0.9308	0	270,802
1991	1992	0.8940	0	167,523
1992	1993	0.9240	0	340,597
1993	1994	0.7278	0	190,443
1994	1995	0.8869	0	252,859
1995 <sup>a</sup>	1996	1.0000	0	150,808
1996 <sup>a</sup>	1997	0.9680	0	284,630
1997 <sup>a</sup>	1998	0.9642	0	197,195
1998 <sup>a</sup>	1999	0.8713	0	121,344
1999	2000	0.9527	0	167,955
2000	2001	0.9558	0	190,174
2001	2002	0.9911	0	200,938
2002	2003	0.9306	0	315,783
2003	2004	0.9291	0	240,459
2004	2005	0.8995	14,859	172,923
2005	2006	0.9811	14,764	140,542
2006	2007	0.9735	14,947	225,670
2007	2008	0.9863	14,858	252,133
2008	2009	0.9576	14,486	154,772
2009	2010	0.9847	5,039	227,743
2010	2011	0.9564	5,074	243,260
2011	2012	0.9690	0	241,918
<i>Average</i>		<i>0.9379</i>	<i>11,994<sup>b</sup></i>	<i>211,255</i>

<sup>a</sup> These groups were only adipose fin clipped.

<sup>b</sup> Average is based on brood years 2004 to 2010.

### *Fish size and condition at release*

The size and condition of the juvenile sockeye released into Lake Wenatchee during the life of the program are presented in Table 4.9.

**Table 4.9.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of sockeye released, brood years 1989-2011. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1989	1990	128	-	18.2	25
1990	1991	131	-	18.9	24
1991	1992	117	3.0	20.6	22
1992	1993	73	6.8	4.2	44
1993	1994	103	-	13.6	40
1994	1995	75	6.1	4.5	38
1995	1996	137	8.2	14.7	30
1996	1997	107	5.6	15.1	30
1997	1998	122	6.1	21.3	21
1998	1999	112	5.4	17.0	27
1999	2000	94	9.5	9.5	48
		134	11.5	31.3	15
2000	2001	123	6.5	22.3	20
		146	8.4	26.0	12
2001	2002	118	7.4	20.7	22
		135	7.3	30.5	15
2002	2003	73	5.6	4.4	104
		118	7.7	13.7	23
		145	9.4	38.6	13
2003	2004	79	4.6	4.8	96
		118	5.9	17.0	26
		158	8.1	44.3	10
2004	2005	116	4.5	17.2	18
		151	7.0	39.3	12
2005	2006	149	7.5	43.7	10
2006	2007	138	10.6	32.4	14
2007	2008	137	9.3	33.0	14
2008	2009	138	9.6	34.6	13
2009	2010	143	8.9	35.5	13
2010	2011	132	14.3	30.7	15
2011	2012	142	9.6	35.3	13

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
<i>Targets</i>		<i>133</i>	<i>9.0</i>	<i>22.7</i>	<i>20</i>

### Survival Estimates

Life-stage survival estimates for juvenile sockeye during the life of the hatchery program are shown in Table 4.10.

**Table 4.10.** Hatchery life-stage survival rates (%) for sockeye salmon, brood years 1989-2011. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
1989	41.6	100.0	88.1	63.9	99.2	98.9	98.1	65.2	83.0
1990	96.2	99.4	90.8	96.3	99.9	99.2	98.4	98.4	81.1
1991	91.8	94.1	79.2	94.8	99.8	99.3	96.4	96.4	72.4
1992	91.1	98.8	92.3	98.0	99.9	99.8	98.6	98.8	89.2
1993	57.1	99.2	89.2	98.3	99.6	99.1	93.7	93.8	82.2
1994	89.8	99.2	79.2	96.0	99.5	98.6	98.3	98.2	74.7
1995	97.5	99.1	87.5	95.0	99.0	93.3	73.2	73.2	60.8
1996	99.2	100.0	95.1	98.7	99.7	99.3	96.4	96.5	90.5
1997	92.8	99.3	84.8	97.9	97.9	97.6	95.5	94.9	77.5
1998	75.4	95.5	77.7	98.4	98.6	98.2	97.1	97.2	74.3
1999	92.3	100.0	92.2	97.3	99.6	99.3	98.2	99.7	88.1
2000	84.5	98.1	93.8	97.7	96.7	96.1	91.4	96.8	83.7
2001	75.4	99.2	78.5	97.6	98.0	97.6	86.9	95.1	66.6
2002	100.0	100.0	95.7	97.8	99.6	99.2	94.6	99.8	88.5
2003	91.0	98.1	87.2	96.9	99.0	98.2	94.8	95.5	74.6
2004	88.7	92.6	88.0	93.1	97.9	97.4	93.7	96.1	76.7
2005	98.5	98.5	85.3	94.9	97.8	96.6	95.5	99.2	66.3
2006	95.3	99.1	73.2	85.4	95.4	94.6	87.8	98.5	54.9
2007	88.4	99.2	89.1	98.6	97.0	95.9	94.9	99.0	83.4
2008	97.0	100.0	59.0	88.3	99.1	97.2	93.8	97.4	48.9
2009	95.8	98.3	89.1	94.8	96.9	96.2	88.4	92.3	74.7
2010	99.0	98.0	92.6	98.2	97.5	96.5	95.6	99.6	87.0
2011	100.0	100.0	92.6	100.0	96.8	96.0	95.4	99.7	88.3
<i>Average</i>	<i>88.6</i>	<i>98.5</i>	<i>86.1</i>	<i>94.7</i>	<i>98.5</i>	<i>97.6</i>	<i>93.8</i>	<i>94.8</i>	<i>76.8</i>
<i>Standard</i>	<i>90.0</i>	<i>85.0</i>	<i>92.0</i>	<i>98.0</i>	<i>97.0</i>	<i>93.0</i>	<i>90.0</i>	<i>95.0</i>	<i>81.0</i>

### 4.3 Disease Monitoring

Because the sockeye hatchery program was terminated in 2012, there are no disease-monitoring results.

### 4.4 Natural Juvenile Productivity

Sockeye smolt abundance was estimated at a trap located near the mouth of Lake Wenatchee during the period 1997 to 2011. Because the efficiency of the trap was difficult to assess, the operation was terminated in 2011. In 2012, the trap was relocated downstream near the mouth of the Chiwawa River and operated there for two years. Again, because few marked sockeye smolts were recaptured, the operation was terminated in 2013. Beginning in 2013, smolt abundance has been estimated at the Lower Wenatchee Trap.

#### Emigrant and Smolt Estimates

The Lower Wenatchee Trap operated between 12 February and 7 October 2014. During that time period the trap was inoperable for 12 days because of high river flows, debris, snow/ice, or major hatchery releases. During the eight-month sampling period, a total of 7,678 wild juvenile sockeye and 72 hatchery juvenile sockeye were captured at the Lower Wenatchee Trap. An emigrant estimate was calculated for juvenile sockeye salmon for the 2014 run year (Table 4.11). The same model used in calculation of this estimate was also used to calculate emigration of wild sockeye salmon for the 2013 run year. Figure 4.1 shows the monthly captures of sockeye collected at the Lower Wenatchee Trap in 2014. All fish captured in the Lower Wenatchee trap are reported in Appendix B.

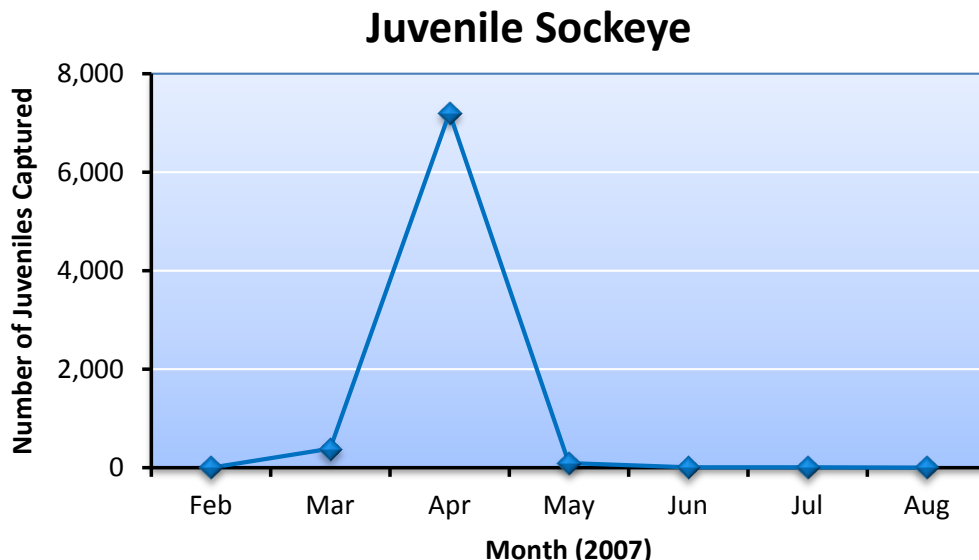
**Table 4.11.** Estimated numbers of wild and hatchery sockeye smolts that emigrated from Lake Wenatchee during run years 1997-2011; ND = no data. Estimates for the run years 1997-2011 were based on sampling at the Upper Wenatchee smolt trap; estimates beginning in 2013 were based on sampling at the Lower Wenatchee smolt trap.

Run year	Numbers of sockeye smolts	
	Wild smolts	Hatchery smolts
1997	55,359	28,828
1998	1,447,259	55,985
1999	1,944,966	112,524
2000	985,490	24,684
2001	39,353	94,046
2002	729,716	121,511
2003	5,439,032	140,322
2004	5,771,187	216,023
2005	723,413	122,399
2006	1,266,971	159,500
2007	2,797,313	140,542
2008 <sup>a</sup>	549,682	121,843
2009 <sup>a</sup>	355,549	119,908
2010 <sup>a</sup>	3,958,888	126,326
2011	1,500,730	159,089



Run year	Numbers of sockeye smolts	
	Wild smolts	Hatchery smolts
2012	ND	ND
2013	873,096	--
2014	1,275,027	--
<i>Average</i>	<i>1,837,661</i>	<i>116,235</i>

<sup>a</sup> Estimates refined based on PIT tag survival to McNary Dam.



**Figure 4.1.** Monthly captures of wild sockeye salmon smolts at the Lower Wenatchee Trap, 2014.

Age classes of wild sockeye smolts were determined from a length frequency analysis based on scales collected randomly each year since 1997 (Table 4.12). For the available run years, most wild sockeye smolts migrated as age 1+ fish. Only in two years (1997 and 2005) did more smolts migrate as age 2+ fish. Relatively few smolts migrated at age 3+.

**Table 4.12.** Age structure and estimated number of wild sockeye smolts that emigrated from Lake Wenatchee, 1997-2014; ND = no data. Estimates for the run years 1997-2011 were based on sampling at the Upper Wenatchee smolt trap; estimates beginning in 2013 were based on sampling at the Lower Wenatchee smolt trap.

Run year	Proportion of wild smolts			Total wild emigrants
	Age 1+	Age 2+	Age 3+	
1997	0.075	0.906	0.019	55,359
1998	0.955	0.037	0.008	1,447,259
1999	0.619	0.381	0.000	1,944,966
2000	0.599	0.400	0.001	985,490
2001	0.943	0.051	0.006	39,353
2002	0.961	0.039	0.000	729,716

Run year	Proportion of wild smolts			Total wild emigrants
	Age 1+	Age 2+	Age 3+	
2003	0.740	0.026	0.000	5,439,032
2004	0.929	0.071	0.000	5,771,187
2005	0.230	0.748	0.022	723,413
2006	0.994	0.006	0.000	1,266,971
2007	0.996	0.004	0.000	2,797,313
2008	0.804	0.195	0.001	549,682
2009	0.927	0.073	0.000	355,549
2010	0.963	0.036	0.001	3,958,888
2011	0.786	0.214	0.000	1,500,730
2012	ND	ND	ND	ND
2013	0.933	0.067	0.000	873,096
2014	0.953	0.047	0.000	1,275,027
<i>Average</i>	<i>0.789</i>	<i>0.194</i>	<i>0.003</i>	<i>1,747,825</i>

### Freshwater Productivity

Egg-smolt survival estimates for wild sockeye salmon are provided in Table 4.13. Estimates of egg deposition were calculated based on the spawner escapement at Tumwater Dam and the sex ratio and fecundity of the broodstock. For the 2012 brood year (a year where brood was not collected), a linear relationship with post-orbital to hypural length as the independent variable was used to calculate average fecundity of sockeye sampled at Tumwater Dam ( $r^2 = 0.40$ ,  $P < 0.01$ ). Smolts for brood years 1995-2009 were based on captures at the Upper Wenatchee Trap. No smolt estimates are available for brood year 2010. Smolt estimates for brood years since 2012 are based on captures made at the Lower Wenatchee Trap. Egg-smolt survival rates for brood years 1995-2012 have ranged from 0.012 to 0.212 (mean = 0.091).

**Table 4.13.** Estimated egg deposition (estimated as mean fecundity times estimated number of females), numbers of smolts, and survival rates for wild Wenatchee sockeye salmon, 1995-2012; NA = not available.

Brood year	Number of females	Mean fecundity	Total eggs	Numbers of wild smolts				Egg-smolt survival
				Age 1+	Age 2+	Age 3+	Total	
1995	2,136	2,295	4,902,120	4,174	53,549	0	57,723	0.012
1996	3,767	2,664	10,035,288	1,382,133	741,032	985	2,124,150	0.212
1997	5,404	2,447	13,223,588	1,203,934	394,196	236	1,598,366	0.121
1998	2,024	2,813	5,693,512	590,309	2,007	0	592,316	0.104
1999	513	2,319	1,189,647	37,110	28,459	0	65,569	0.055
2000	11,413	2,673	30,506,949	701,257	1,414,148	0	2,115,405	0.069
2001	21,685	2,960	64,187,600	4,024,884	409,754	15,915	4,450,553	0.069
2002	17,226	2,856	49,197,456	5,361,433	541,113	0	5,902,546	0.120
2003	2,158	3,511	7,576,738	166,385	7,602	0	173,987	0.023
2004	15,469	2,505	38,749,845	1,259,369	11,189	275	1,270,833	0.033

Brood year	Number of females	Mean fecundity	Total eggs	Numbers of wild smolts				Egg-smolt survival
				Age 1+	Age 2+	Age 3+	Total	
2005	5,867	2,718	15,946,506	2,786,123	107,243	0	2,893,366	0.181
2006	2,747	2,656	7,296,032	442,164	25,919	1,507	469,590	0.064
2007	2,001	3,115	6,232,804	329,629	142,916	594	473,139	0.076
2008	11,775	2,555	30,084,691	3,814,226	320,567	0	4,134,794	0.137
2009	3,939	2,459	9,684,965	1,179,569	NA	0	NA	NA
2010 <sup>a</sup>	11,918	2,785	33,190,467	NA	58,136	0	NA	NA
2011 <sup>b</sup>	9,722	2,970	28,873,491	814,960	60,382	NA	NA	NA
2012 <sup>b</sup>	14,753	2,745	40,496,573	1,214,645	NA	NA	NA	NA
<b>Average</b>	<b>8,029</b>	<b>2,725</b>	<b>22,059,348</b>	<b>1,488,959</b>	<b>269,888</b>	<b>1,220</b>	<b>1,880,167</b>	<b>0.091</b>

<sup>a</sup> There is no emigrant estimate for trapping during 2012 or 2013.

<sup>b</sup> Emigrant estimates are based on captures at the Lower Wenatchee Trap.

Juvenile survival rates for hatchery sockeye salmon are provided in Table 4.14. Release-smolt survival rates for brood years 1995-2009 have ranged from 0.000 to 1.000 (mean = 0.570). Egg-smolt survival rates for the same brood years ranged from 0.000 to 0.710 (mean = 0.294). On average, egg-smolt survival of hatchery sockeye is about three times greater than egg-smolt survival of wild sockeye.

**Table 4.14.** Juvenile survival rates for hatchery Wenatchee sockeye, brood years 1995-2009.

Brood year	Number of eggs	Number of parr released	Date of release	Estimated number of smolts	Egg-smolt survival	Release-smolt survival
1995	247,900	150,808	10/25/96	28,828	0.116	0.191
1996	314,390	284,630	10/22/97	55,985	0.178	0.197
1997	254,459	197,195	11/9/98	112,524	0.442	0.571
1998	163,278	121,344	10/27/99	24,684	0.151	0.203
1999	190,732	84,466	8/28/00	30,326	0.159	0.359
		83,489	11/1/00	63,720	0.334	0.763
2000	227,234	92,055	8/27/01	30,918	0.136	0.336
		98,119	9/27/01	90,593	0.399	0.923
2001	301,925	96,486	8/28/02	36,484	0.121	0.378
		104,452	9/23/02	103,838	0.344	0.994
2002	356,982	98,509	6/16/03	5,192	0.015	0.053
		104,855	8/25/03	98,412	0.276	0.939
		112,419	10/22/03	112,419	0.315	1.000
2003	319,470	32,755	6/15/04	0	0.000	0.000
		104,879	8/25/04	19,574	0.061	0.187
		102,825	11/3/04	102,825	0.322	1.000
2004	225,499	81,428	8/29/05	159,500	0.707	0.922
		91,495	11/2/05			
2005	211,985	70,386	10/30/06	140,542	0.663	1.000

Brood year	Number of eggs	Number of parr released	Date of release	Estimated number of smolts	Egg-smolt survival	Release-smolt survival
		70,156	10/30/06			
2006	292,136	225,670	10/31/07	121,843	0.412	0.540
2007	302,363	252,133	10/29/08	119,908	0.397	0.476
2008	316,476	154,772	10/28/09	126,326	0.399	0.813
2009	304,963	227,743	10/27/10	159,089	0.522	0.699

<sup>a</sup> There is no emigrant estimate for the 2010 or 2011 brood years.

### PIT Tagging Activities

A total of 4,821 wild juvenile sockeye salmon were PIT tagged and released in 2014 at the Lower Wenatchee Trap. Numbers of wild sockeye salmon PIT-tagged and released as part of the Comparative Survival Study during the period 2006-2014 are shown in Table 4.15. See Appendix C for a complete list of all fish captured, tagged, lost, and released.

**Table 4.15.** Summary of the numbers of wild sockeye salmon that were tagged and released at the Upper and Lower Wenatchee Traps within the Wenatchee River basin, 2006-2014.

Sampling Location	Numbers of PIT-tagged sockeye salmon released								
	2006	2007	2008	2009	2010	2011	2012	2013	2014
Upper Wenatchee Trap	0	0	3,165	3,683	10,006	0	0	0	0
Lower Wenatchee Trap	0	0	0	0	0	0	0	0	4,821

## 4.5 Spawning Escapement

The sockeye salmon hatchery program ended after the 2011 brood year. As a result, monitoring activities that focused on evaluating the effects of the supplementation program on the natural population switched to monitoring the abundance and productivity of the natural population. Thus, estimation of spawn time and carcass surveys were discontinued.

From 2009-2013, mark-recapture methods were used to estimate spawning escapement within the White River, while area-under-the-curve (AUC) methods were used to estimate spawning escapement within the Little Wenatchee River. Beginning in 2014, mark-recapture methods were used to estimate the spawning escapement of sockeye in the White River and Little Wenatchee watersheds (see Appendix H for more details).

### Mark-Recapture Estimates

As noted above, spawning escapement of sockeye salmon in 2014 was estimated using mark-recapture methods. This method relied on PIT tags to estimate sockeye spawning escapement (see Appendix H for more details).

Using mark-recapture methods, the estimated total escapement of sockeye in the Upper Wenatchee River basin in 2014 was 53,412 (Table 4.16). About 92% of the escapement entered the White River watershed (including the Napeequa River).

**Table 4.16.** Estimated escapement of adult sockeye into the Little Wenatchee and White River watersheds for return years 2009-2014. Escapement was based on recapture of PIT-tagged fish.

Return year	Tumwater Dam count	Recreational harvest	Little Wenatchee escapement	White River escapement	Total spawning escapement
2009	16,034	2,285	576	13,876	14,452
2010	35,821	4,129	2,062	19,542	21,604
2011 <sup>a</sup>	18,634	0	2,431	14,582	17,013
2012	66,520	12,107	4,607	23,866	28,473
2013 <sup>a</sup>	29,015	6,262	2,426	14,294	16,720
2014	99,898	16,255	4,391	49,021	53,412
<b>Average</b>	<b>44,320</b>	<b>6,840</b>	<b>2,749</b>	<b>22,530</b>	<b>25,279</b>

<sup>a</sup> Spawning escapements in 2011 and 2012 were calculated using AUC counts and a regression model (Keller and Murauskas 2012).

The spawning escapement of 53,412 Wenatchee sockeye was greater than the overall average of 17,282 (Table 4.17).

**Table 4.17.** Spawning escapements for sockeye salmon in the Wenatchee River basin for return years 1989-2014; NA = not available and AUC = area under the curve.

Return year	Escapement estimation method	Spawning escapement		
		Little Wenatchee	White	Total
1989	Counts at Tumwater Dam	NA	NA	<b>21,802</b>
1990	Counts at Tumwater Dam	NA	NA	<b>27,325</b>
1991	Counts at Tumwater Dam	NA	NA	<b>26,689</b>
1992	Counts at Tumwater Dam	NA	NA	<b>16,461</b>
1993	Counts at Tumwater Dam	NA	NA	<b>27,726</b>
1994	Counts at Tumwater Dam	NA	NA	<b>7,330</b>
1995	Counts at Tumwater Dam	NA	NA	<b>3,448</b>
1996	Counts at Tumwater Dam	NA	NA	<b>6,573</b>
1997	Counts at Tumwater Dam	NA	NA	<b>9,693</b>
1998	Counts at Tumwater Dam	NA	NA	<b>4,014</b>
1999	Counts at Tumwater Dam	NA	NA	<b>1,025</b>
2000	Counts at Tumwater Dam	NA	NA	<b>20,735</b>
2001	Counts at Tumwater Dam	NA	NA	<b>29,103</b>
2002	Counts at Tumwater Dam	NA	NA	<b>27,565</b>
2003	Counts at Tumwater Dam	NA	NA	<b>4,855</b>
2004	Counts at Tumwater Dam	NA	NA	<b>27,556</b>
2005	Counts at Tumwater Dam	NA	NA	<b>14,011</b>
2006	AUC	574	5,634	<b>6,208</b>
2007	AUC	150	1,720	<b>1,870</b>
2008	AUC	3,491	16,757	<b>20,248</b>
2009	AUC and Mark-Recapture	763	7,004	<b>7,767</b>

Return year	Escapement estimation method	Spawning escapement		
		Little Wenatchee	White	Total
2010	AUC and Mark-Recapture	2,543	19,157	<b>21,700</b>
2011	AUC and Mark-Recapture	2,431	14,582	<b>17,013</b>
2012	AUC and Mark-Recapture	4,607	23,866	<b>28,473</b>
2013	AUC and Mark-Recapture	2,426	14,294	<b>16,720</b>
2014	Mark-Recapture	4,391	49,021	<b>53,412</b>
<i>Average</i>		<i>2,375</i>	<i>16,893</i>	<i>17,282</i>

## 4.6 Carcass Surveys

As described earlier, carcass surveys were not conducted in 2014. The information contained in this section represents carcass data collected before 2014.

### Number sampled

Table 4.18 shows the number of carcasses sampled within different survey streams during the period 1993-2013.

**Table 4.18.** Numbers of sockeye carcasses sampled within different streams/watersheds within the Wenatchee River basin, 1989-2013.

Survey year	Numbers of sockeye carcasses			
	Little Wenatchee	White	Napeequa	Total
1993	90	195	0	<b>285</b>
1994	121	165	0	<b>286</b>
1995	0	56	0	<b>56</b>
1996	43	1,387	3	<b>1,433</b>
1997	69	1,425	41	<b>1,535</b>
1998	61	524	4	<b>589</b>
1999	40	186	0	<b>226</b>
2000	821	5,494	0	<b>6,315</b>
2001	650	3,127	0	<b>3,777</b>
2002	506	7,258	55	<b>7,819</b>
2003	86	1,002	14	<b>1,102</b>
2004	625	6,960	138	<b>7,723</b>
2005	1	7	0	<b>8</b>
2006	101	2,158	38	<b>2,297</b>
2007	17	363	3	<b>383</b>
2008	476	5,132	125	<b>5,733</b>
2009	84	3,103	103	<b>3,290</b>
2010	217	7,832	70	<b>8,119</b>
2011	372	3,322	48	<b>3,742</b>
2012	1,309	7,479	31	<b>8,819</b>

Survey year	Numbers of sockeye carcasses			
	Little Wenatchee	White	Napeequa	Total
2013	179	2,996	27	3,202
<i>Average</i>	<i>279</i>	<i>2,865</i>	<i>33</i>	<i>3,178</i>

### Carcass Distribution and Origin

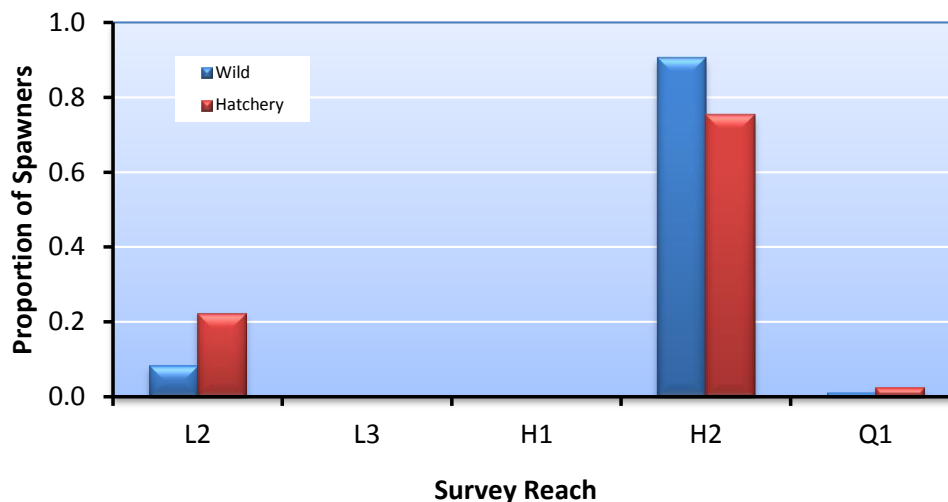
Based on the available data (1993-2013), the largest percentage of both wild and hatchery sockeye spawned in Reach 2 on the White River (Table 4.19 and Figure 4.2). However, a greater percentage of wild fish was found in Reach 2 than hatchery fish.

**Table 4.19.** Numbers of wild and hatchery sockeye carcasses sampled within different reaches in the Wenatchee River basin, 1993-2013. Reach codes are described in Table 2.9.

Survey year	Origin	Numbers of sockeye carcasses					Total
		Little Wenatchee		White River			
		L2	L3	H1	H2	Q1	
1993	Wild	86	0	0	183	0	269
	Hatchery	4	0	0	12	0	16
1994	Wild	112	0	0	155	0	267
	Hatchery	9	0	0	9	0	18
1995	Wild	0	0	0	55	0	55
	Hatchery	0	0	0	1	0	1
1996	Wild	41	0	0	1,299	3	1,343
	Hatchery	2	0	0	88	0	90
1997	Wild	65	0	0	1,411	40	1,516
	Hatchery	4	0	0	11	1	16
1998	Wild	61	0	0	515	4	580
	Hatchery	0	0	0	9	0	9
1999	Wild	30	0	0	164	0	194
	Hatchery	10	0	0	22	0	32
2000	Wild	694	0	3	5,239	0	5,936
	Hatchery	127	0	0	252	0	379
2001	Wild	625	0	0	3,063	0	3,688
	Hatchery	25	0	0	64	0	89
2002	Wild	504	0	0	7,207	55	7,766
	Hatchery	2	0	0	51	0	53
2003	Wild	81	0	0	993	14	1,088
	Hatchery	5	0	0	9	0	14
2004	Wild	606	0	0	6,755	166	7,527
	Hatchery	19	0	0	205	22	246
2005	Wild	201	0	5	2,966	21	3,193
	Hatchery	1	0	0	8	0	9
2006	Wild	80	0	0	2,112	36	2,228
	Hatchery	21	0	0	46	2	69

Survey year	Origin	Numbers of sockeye carcasses					Total
		Little Wenatchee		White River			
		L2	L3	H1	H2	Q1	
2007	Wild	17	0	0	346	3	366
	Hatchery	0	0	0	17	0	17
2008	Wild	472	0	0	5,118	124	5,714
	Hatchery	4	0	0	14	1	19
2009	Wild	80	0	0	3,084	103	3,267
	Hatchery	4	0	0	19	0	23
2010	Wild	210	0	0	7,711	69	7,990
	Hatchery	7	0	0	121	1	129
2011	Wild	266	0	0	3,079	43	3,388
	Hatchery	106	0	0	243	5	354
2012	Wild	1,270	0	21	7,368	30	8,689
	Hatchery	39	0	3	87	1	130
2013	Wild	174	0	1	2,936	26	3,137
	Hatchery	3	0	0	56	1	60
Average	Wild	270	0	1	2,941	35	3,248
	Hatchery	18	0	0	61	2	81

### Wenatchee Sockeye Salmon



**Figure 4.2.** Distribution of wild and hatchery produced carcasses in different reaches in the Wenatchee River basin, pooled data from 1993-2013. Reach codes are described in Table 2.9; L = Little Wenatchee, H = White River, and Q = Napeequa River.

### 4.7 Life History Monitoring

Life history characteristics of Wenatchee sockeye were assessed by examining carcasses on spawning grounds and fish sampled at broodstock collection sites or during stock assessment, and by reviewing tagging data and fisheries statistics.



## Migration Timing

There was little difference in migration timing of hatchery and wild sockeye past Tumwater Dam (Table 4.20a and b; Figure 4.3). On average, early in the run, hatchery and wild sockeye arrived at the dam at about the same time. Toward the end of the migration period, hatchery sockeye tended to arrive at the dam slightly later than did wild sockeye. Most hatchery and wild sockeye migrated upstream past Tumwater Dam during July through early August. The peak migration time for both hatchery and wild sockeye was the last two weeks of July (Figure 4.3).

**Table 4.20a.** The Julian day and date that 10%, 50% (median), and 90% of the wild and hatchery sockeye salmon passed Tumwater Dam, 1998-2014. The average Julian day and date are also provided. Migration timing is based on video sampling at Tumwater. Data for 1998 through 2003 were based on videotapes and broodstock trapping and may not reflect the actual number of hatchery sockeye salmon. All sockeye were visually examined during trapping from 2004 to present.

Survey year	Origin	Sockeye Migration Time (days)								Sample size
		10 Percentile		50 Percentile		90 Percentile		Mean		
		Julian	Date	Julian	Date	Julian	Date	Julian	Date	
1998	Wild	195	14-Jul	201	20-Jul	208	27-Jul	202	21-Jul	4,173
	Hatchery	196	15-Jul	204	23-Jul	220	8-Aug	206	25-Jul	31
1999	Wild	226	14-Aug	233	21-Aug	241	29-Aug	234	22-Aug	908
	Hatchery	228	16-Aug	234	22-Aug	242	30-Aug	235	23-Aug	264
2000	Wild	200	18-Jul	206	24-Jul	213	31-Jul	207	25-Jul	18,390
	Hatchery	199	17-Jul	206	24-Jul	213	31-Jul	206	24-Jul	2,589
2001	Wild	189	8-Jul	194	13-Jul	214	2-Aug	198	17-Jul	32,554
	Hatchery	199	18-Jul	212	31-Jul	240	28-Aug	214	2-Aug	79
2002	Wild	204	23-Jul	208	27-Jul	219	7-Aug	210	29-Jul	27,241
	Hatchery	204	23-Jul	209	28-Jul	222	10-Aug	211	30-Jul	580
2003	Wild	194	13-Jul	200	19-Jul	208	27-Jul	201	20-Jul	4,699
	Hatchery	194	13-Jul	201	20-Jul	211	30-Jul	203	22-Jul	375
2004	Wild	191	9-Jul	196	14-Jul	207	25-Jul	198	16-Jul	31,408
	Hatchery	189	7-Jul	194	12-Jul	203	21-Jul	196	14-Jul	1,758
2005	Wild	192	11-Jul	199	18-Jul	227	15-Aug	204	23-Jul	14,176
	Hatchery	187	6-Jul	200	19-Jul	251	8-Sep	212	31-Jul	42
2006	Wild	201	20-Jul	204	23-Jul	214	2-Aug	206	25-Jul	9,151
	Hatchery	202	21-Jul	219	7-Aug	228	16-Aug	215	3-Aug	507
2007	Wild	201	20-Jul	210	29-Jul	227	15-Aug	213	1-Aug	2,542
	Hatchery	205	24-Jul	213	1-Aug	231	19-Aug	216	4-Aug	65
2008	Wild	200	18-Jul	207	25-Jul	219	6-Aug	208	26-Jul	29,229
	Hatchery	201	19-Jul	206	24-Jul	215	2-Aug	208	26-Jul	103
2009	Wild	198	17-Jul	204	23-Jul	213	1-Aug	206	25-Jul	15,552
	Hatchery	199	18-Jul	205	24-Jul	215	3-Aug	207	26-Jul	534
2010	Wild	199	18-Jul	205	24-Jul	220	8-Aug	208	27-Jul	34,519
	Hatchery	200	19-Jul	215	3-Aug	244	1-Sep	218	6-Aug	1,302

Survey year	Origin	Sockeye Migration Time (days)								Sample size
		10 Percentile		50 Percentile		90 Percentile		Mean		
		Julian	Date	Julian	Date	Julian	Date	Julian	Date	
2011	Wild	213	1-Aug	216	4-Aug	224	12-Aug	217	5-Aug	17,680
	Hatchery	213	1-Aug	213	1-Aug	231	19-Aug	216	4-Aug	954
2012 <sup>a</sup>	Wild	207	25-Jul	212	30-Jul	216	3-Aug	212	30-Jul	21,246
	Hatchery	207	25-Jul	207	25-Jul	228	15-Aug	213	31-Jul	348
2013	Wild	196	15-Jul	200	19-Jul	207	26-Jul	201	20-Jul	28,245
	Hatchery	197	16-Jul	201	20-Jul	211	30-Jul	203	22-Jul	770
2014	Wild	194	13-Jul	199	18-Jul	210	29-Jul	201	20-Jul	97,670
	Hatchery	196	15-Jul	201	20-Jul	211	30-Jul	203	22-Jul	2,229
<i>Average</i>	Wild	<i>200</i>		<i>206</i>		<i>217</i>		<i>207</i>		<i>22,905</i>
	Hatchery	<i>201</i>		<i>208</i>		<i>224</i>		<i>211</i>		<i>737</i>

<sup>a</sup> The origin of sockeye passing Tumwater Dam during 8 through 11 August 2012 was not assessed. The total number of sockeye passing Tumwater Dam in 2012 was 30,617 adults. Thus, about 9,023 adults of unknown origin passed Tumwater Dam in 2012.

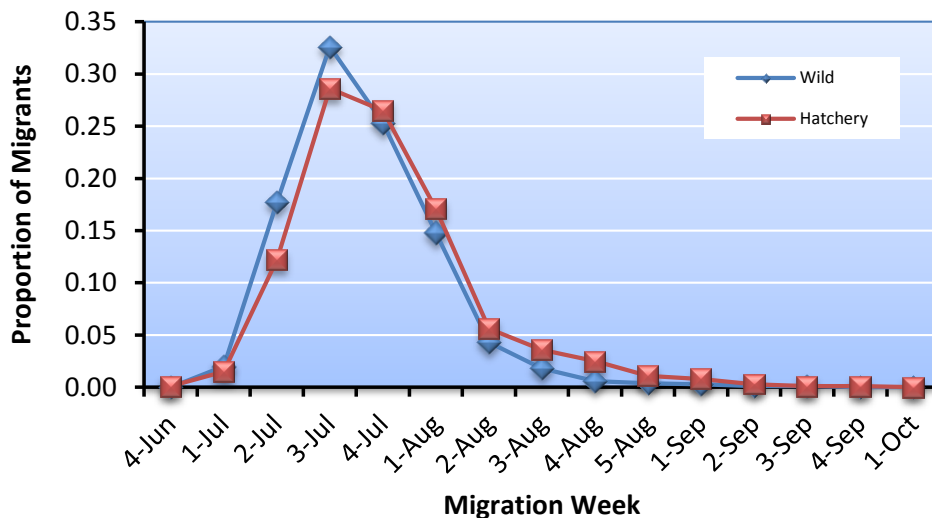
**Table 4.20b.** The week that 10%, 50% (median), and 90% of the wild and hatchery sockeye salmon passed Tumwater Dam, 1998-2014. The average week is also provided. Migration timing is based on video sampling at Tumwater. Data for 1998 through 2003 were based on videotapes and broodstock trapping and may not reflect the actual number of hatchery sockeye salmon. All sockeye were visually examined during trapping from 2004 to present.

Survey year	Origin	Sockeye Migration Time (week)				Sample size
		10 Percentile	50 Percentile	90 Percentile	Mean	
1998	Wild	28	29	30	29	4,173
	Hatchery	28	30	32	30	31
1999	Wild	33	34	35	34	908
	Hatchery	33	34	35	34	264
2000	Wild	29	30	31	30	18,390
	Hatchery	29	30	31	30	2,589
2001	Wild	27	28	31	29	32,554
	Hatchery	29	31	35	31	79
2002	Wild	30	30	32	30	27,241
	Hatchery	30	30	32	31	580
2003	Wild	28	29	30	29	4,699
	Hatchery	28	29	31	29	375
2004	Wild	28	28	28	29	31,408
	Hatchery	27	28	29	28	1,758
2005	Wild	28	29	33	30	14,176
	Hatchery	27	29	36	31	42
2006	Wild	29	29	31	30	9,151
	Hatchery	29	32	33	31	507

Survey year	Origin	Sockeye Migration Time (week)				Sample size
		10 Percentile	50 Percentile	90 Percentile	Mean	
2007	Wild	29	30	33	31	2,542
	Hatchery	30	31	33	31	65
2008	Wild	29	30	32	30	29,229
	Hatchery	29	30	31	30	103
2009	Wild	29	30	31	30	15,552
	Hatchery	29	29	31	30	534
2010	Wild	29	30	32	30	34,519
	Hatchery	29	31	35	32	1,302
2011	Wild	31	31	32	31	17,680
	Hatchery	31	31	33	31	954
2012 <sup>a</sup>	Wild	30	31	31	31	21,246
	Hatchery	30	30	33	31	348
2013	Wild	28	29	30	29	28,245
	Hatchery	29	29	31	29	770
2014	Wild	28	29	30	29	97,670
	Hatchery	28	29	29	29	2,229
<i>Average</i>	Wild	<i>29</i>	<i>30</i>	<i>31</i>	<i>30</i>	<i>22,905</i>
	Hatchery	<i>29</i>	<i>30</i>	<i>32</i>	<i>30</i>	<i>737</i>

<sup>a</sup> The origin of sockeye passing Tumwater Dam during 8 through 11 August 2012 was not assessed. The total number of sockeye passing Tumwater Dam in 2012 was 30,617 adults. Thus, about 9,023 adults of unknown origin passed Tumwater Dam in 2012.

### Sockeye Migration Timing



**Figure 4.3.** Proportion of wild and hatchery sockeye observed (using video) passing Tumwater Dam each week during their migration period late-June through early-October; data were pooled over survey years 1998-2014.

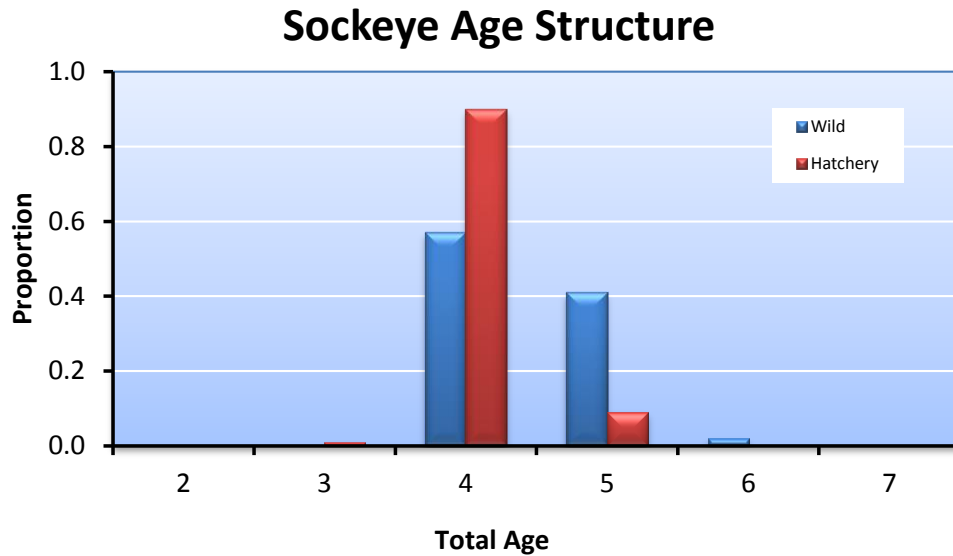
### Age at Maturity

Although sample sizes are small, it appears that most hatchery sockeye returned as age-4 fish, while most wild sockeye returned as age-4 and 5 fish (Table 4.21; Figure 4.4). Only wild fish have returned at age-6.

**Table 4.21.** Proportions of wild and hatchery sockeye of different ages (total age) sampled in broodstock and on spawning grounds, 1994-2013. Since 2012, only wild and hatchery sockeye sampled on spawning grounds were used to establish proportions.

Survey year	Origin	Total age						Sample size
		2	3	4	5	6	7	
1994	Wild	-	-	-	-	-	-	0
	Hatchery	0.00	0.00	0.88	0.13	0.00	0.00	16
1995	Wild	-	-	-	-	-	-	0
	Hatchery	0.00	0.00	0.00	1.00	0.00	0.00	1
1996	Wild	-	-	-	-	-	-	0
	Hatchery	0.00	0.00	1.00	0.00	0.00	0.00	82
1997	Wild	-	-	-	-	-	-	0
	Hatchery	0.00	0.00	0.77	0.23	0.00	0.00	13
1998	Wild	0.00	0.08	0.85	0.08	0.00	0.00	26
	Hatchery	0.00	0.00	0.64	0.36	0.00	0.00	11
1999	Wild	0.00	0.00	0.18	0.73	0.10	0.00	113
	Hatchery	0.00	0.00	0.65	0.35	0.00	0.00	31
2000	Wild	0.00	0.00	0.00	1.00	0.00	0.00	1
	Hatchery	0.00	0.00	0.98	0.02	0.00	0.00	359
2001	Wild	0.00	0.00	0.76	0.24	0.00	0.00	29
	Hatchery	0.00	0.00	0.75	0.25	0.00	0.00	171
2002	Wild	0.00	0.00	0.20	0.80	0.00	0.00	5
	Hatchery	0.00	0.00	0.29	0.71	0.00	0.00	63
2003	Wild	0.00	0.00	0.00	1.00	0.00	0.00	5
	Hatchery	0.00	0.33	0.67	0.00	0.00	0.00	6
2004	Wild	-	-	-	-	-	-	0
	Hatchery	0.00	0.02	0.93	0.05	0.00	0.00	244
2005	Wild	-	-	-	-	-	-	0
	Hatchery	0.00	0.13	0.75	0.13	0.00	0.00	8
2006	Wild	0.00	0.00	0.34	0.65	0.01	0.00	207
	Hatchery	0.00	0.00	1.00	0.00	0.00	0.00	65
2007	Wild	0.00	0.00	0.02	0.88	0.10	0.00	206
	Hatchery	0.00	0.00	0.35	0.65	0.00	0.00	17
2008	Wild	0.00	0.00	0.95	0.04	0.01	0.00	258
	Hatchery	0.00	0.08	0.92	0.00	0.00	0.00	12
2009	Wild	0.00	0.00	0.79	0.21	0.00	0.00	251

Survey year	Origin	Total age						Sample size
		2	3	4	5	6	7	
	Hatchery	0.00	0.00	1.00	0.00	0.00	0.00	2
2010	Wild	0.00	0.00	0.67	0.33	0.00	0.00	193
	Hatchery	0.00	0.00	0.98	0.02	0.00	0.00	130
2011	Wild	0.00	0.00	0.63	0.36	0.01	0.00	270
	Hatchery	0.00	0.02	0.96	0.02	0.00	0.00	274
2012	Wild	0.00	0.00	0.92	0.08	0.00	0.00	13
	Hatchery	0.00	0.00	0.96	0.03	0.01	0.00	128
2013	Wild	0.00	0.00	0.25	0.75	0.00	0.00	4
	Hatchery	0.00	0.00	0.89	0.11	0.00	0.00	44
Average	Wild	0.00	0.00	0.57	0.41	0.02	0.00	79
	Hatchery	0.00	0.01	0.90	0.09	0.00	0.00	86



**Figure 4.4.** Proportions of wild and hatchery sockeye salmon of different total ages sampled at Tumwater Dam and on spawning grounds in the Wenatchee River basin for the combined years 1994-2013.

### Size at Maturity

Although sample sizes are small, wild and hatchery sockeye were similar in size in 2013 (Table 4.22). In addition, the pooled data indicate that there is little difference in mean sizes of hatchery and wild sockeye salmon sampled in the Wenatchee River basin (Table 4.22). Analyses for the five-year reports will compare sizes of hatchery and wild fish of the same age groups and sex.

**Table 4.22.** Mean lengths (POH; cm) and variability statistics for wild and hatchery sockeye salmon sampled at Dryden Dam (broodstock) and on spawning grounds in the Wenatchee River basin, 1994-2013; SD = 1 standard deviation.

Survey year	Origin	Sample size	Sockeye length (POH; cm)			
			Mean	SD	Minimum	Maximum
1994	Wild	0	-	-	-	-
	Hatchery	14	42	3	37	47
1995	Wild	0	-	-	-	-
	Hatchery	1	53	-	53	53
1996	Wild	0	-	-	-	-
	Hatchery	5	51	3	49	55
1997	Wild	6	40	3	38	45
	Hatchery	17	41	3	37	50
1998	Wild	585	43	3	34	50
	Hatchery	20	43	3	40	51
1999	Wild	99	42	3	36	50
	Hatchery	31	41	3	36	47
2000	Wild	1	48	-	48	48
	Hatchery	377	40	2	30	49
2001	Wild	29	42	2	38	47
	Hatchery	184	43	3	35	51
2002	Wild	5	42	1	40	43
	Hatchery	52	44	3	37	49
2003	Wild	5	44	4	38	47
	Hatchery	13	42	5	30	48
2004	Wild	0	-	-	-	-
	Hatchery	230	40	3	33	49
2005	Wild	0	-	-	-	-
	Hatchery	8	43	9	35	64
2006	Wild	248	45	4	34	52
	Hatchery	17	41	5	31	48
2007	Wild	248	45	3	32	52
	Hatchery	16	41	5	31	48
2008	Wild	261	52	3	44	66
	Hatchery	20	39	3	30	41
2009	Wild	260	43	3	33	53
	Hatchery	22	41	2	36	46
2010	Wild	200	56	3	48	66
	Hatchery	131	41	2	35	45
2011	Wild	277	43	3	35	51

Survey year	Origin	Sample size	Sockeye length (POH; cm)			
			Mean	SD	Minimum	Maximum
	Hatchery	282	40	3	32	49
2012	Wild	15	40	4	34	48
	Hatchery	130	40	3	31	48
2013	Wild	2	49	3	47	51
	Hatchery	64	50	4	43	65
<i>Pooled</i>	<i>Wild</i>	<i>2,241</i>	<i>43</i>	<i>3</i>	<i>32</i>	<i>66</i>
	<i>Hatchery</i>	<i>1,634</i>	<i>45</i>	<i>4</i>	<i>30</i>	<i>65</i>

### Contribution to Fisheries

The total number of hatchery and wild sockeye captured in different fisheries is provided in Tables 4.23 and 4.24. Harvest on hatchery-origin sockeye has been less than the harvest on wild sockeye.

**Table 4.23.** Estimated number and percent (in parentheses) of hatchery-origin Wenatchee sockeye captured in different fisheries, 1989-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational <sup>a</sup> (sport)	
1989	0 (0)	279 (30)	4 (0)	639 (69)	922
1990	0 (0)	23 (100)	0 (0)	0 (0)	23
1991	0 (0)	6 (100)	0 (0)	0 (0)	6
1992	0 (0)	38 (97)	1 (3)	0 (0)	39
1993	0 (0)	4 (100)	0 (0)	0 (0)	4
1994	0 (0)	3 (100)	0 (0)	0 (0)	3
1995	0 (0)	10 (100)	0 (0)	0 (0)	10
1996	0 (0)	61 (81)	9 (12)	5 (7)	75
1997	0 (0)	69 (73)	11 (12)	15 (16)	95
1998	0 (0)	7 (100)	0 (0)	0 (0)	7
1999	0 (0)	3 (20)	0 (0)	12 (80)	15
2000	0 (0)	59 (12)	9 (2)	414 (86)	482
2001	0 (0)	0 (0)	0 (0)	3 (100)	3
2002	0 (0)	16 (100)	0 (0)	0 (0)	16
2003	0 (0)	3 (100)	0 (0)	0 (0)	3
2004	0 (0)	7 (4)	0 (0)	192 (96)	199
2005	0 (0)	61 (41)	7 (5)	79 (54)	147
2006	0 (0)	124 (23)	1 (0)	409 (77)	534
2007	0 (0)	95 (82)	12 (10)	9 (8)	116
2008	0 (0)	83 (20)	10 (2)	322 (78)	415
<i>Average</i>	<i>0 (0)</i>	<i>48 (64)</i>	<i>3 (2)</i>	<i>105 (34)</i>	<i>156</i>

<sup>a</sup> Includes the Lake Wenatchee fishery.

**Table 4.24.** Estimated number and percent (in parentheses) of wild Wenatchee sockeye captured in different fisheries, 1989-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational <sup>a</sup> (sport)	
1989	0 (0)	2,192 (31)	26 (0)	4,838 (69)	7,056
1990	0 (0)	191 (100)	0 (0)	0 (0)	191
1991	0 (0)	293 (99)	2 (1)	0 (0)	295
1992	0 (0)	345 (99)	5 (1)	0 (0)	350
1993	0 (0)	661 (99)	4 (1)	0 (0)	665
1994	0 (0)	146 (100)	0 (0)	0 (0)	146
1995	0 (0)	63 (86)	3 (4)	7 (10)	73
1996	0 (0)	1,554 (56)	247 (9)	993 (36)	2,794
1997	0 (0)	3,061 (54)	370 (6)	2,266 (40)	5,697
1998	0 (0)	938 (99)	4 (0)	10 (1)	952
1999	0 (0)	22 (19)	3 (3)	90 (78)	115
2000	0 (0)	1,189 (19)	162 (3)	4,881 (78)	6,232
2001	0 (0)	827 (100)	0 (0)	0 (0)	827
2002	0 (0)	379 (83)	2 (0)	73 (16)	454
2003	0 (0)	129 (25)	11 (2)	383 (73)	523
2004	0 (0)	1,559 (24)	147 (2)	4,825 (74)	6,531
2005	0 (0)	2,497 (44)	177 (3)	2,996 (53)	5,670
2006	0 (0)	2,844 (52)	107 (2)	2,505 (46)	5,456
2007	0 (0)	1,533 (57)	202 (8)	944 (35)	2,679
2008	0 (0)	5,446 (25)	648 (3)	15,414 (72)	21,508
<b>Average</b>	<b>0 (0)</b>	<b>1,293 (64)</b>	<b>106 (2)</b>	<b>2,011 (34)</b>	<b>3,411</b>

<sup>a</sup> Includes the Lake Wenatchee fishery.

## Straying

Stray rates were determined by examining CWTs recovered on spawning grounds within and outside the Wenatchee River basin. In addition, PIT tagging of hatchery sockeye, which began with brood year 2005, allows estimation of stray rates by brood return. Targets for strays based on return year (recovery year) outside the Wenatchee River basin should be less than 5%. The target for brood year strays should also be less than 5%.

Based on CWTs and brood year analysis, virtually no hatchery-origin Wenatchee sockeye strayed into non-target spawning areas or hatchery programs before brood year 2006 (Table 4.25). However, sockeye from brood years 2006 and 2007 strayed into the Entiat River and a few into the Methow River (non-target streams) and a non-target hatchery (Umpqua Trap) (Table 4.25). Stray rates of Wenatchee sockeye from brood year 2006 exceeded the target of 5%.



**Table 4.25.** Number and percent of hatchery-origin Wenatchee sockeye that homed to target spawning areas and the target hatchery program, and number and percent that strayed to non-target spawning areas and hatchery programs, by brood years 1990-2008. Hatchery-origin sockeye from brood years 1995-1998 were not tagged because of columnaris disease. Percent stays should be less than 5%.

Brood year	Homing				Straying			
	Target streams		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1990	402	99.5	2	0.5	0	0.0	0	0.0
1991	1	100.0	0	0.0	0	0.0	0	0.0
1992	92	98.9	0	0.0	0	0.0	1	1.1
1993	29	96.7	1	3.3	0	0.0	0	0.0
1994	66	94.3	4	5.7	0	0.0	0	0.0
1995	-	-	-	-	-	-	-	-
1996	-	-	-	-	-	-	-	-
1997	-	-	-	-	-	-	-	-
1998	-	-	-	-	-	-	-	-
1999	65	100.0	0	0.0	0	0.0	0	0.0
2000	571	100.0	0	0.0	0	0.0	0	0.0
2001	17	100.0	0	0.0	0	0.0	0	0.0
2002	251	100.0	0	0.0	0	0.0	0	0.0
2003	11	100.0	0	0.0	0	0.0	0	0.0
2004	56	100.0	0	0.0	0	0.0	0	0.0
2005	67	97.1	2	2.9	0	0.0	0	0.0
2006	117	41.9	0	0.0	160	57.3	2	0.7
2007	260	97.4	1	0.4	6	2.2	0	0.0
2008	85	90.4	0	0.0	9	9.6	0	0.0
<b>Average</b>	<b>139</b>	<b>94.4</b>	<b>1</b>	<b>0.9</b>	<b>12</b>	<b>4.6</b>	<b>0</b>	<b>0.1</b>

\* Homing to the target hatchery includes Wenatchee hatchery sockeye that are captured and included as broodstock in the Wenatchee Hatchery program. These hatchery fish were collected at Tumwater Dam.

Based on PIT-tag analyses, on average, about 9% of the hatchery sockeye returns were last detected in streams outside the Wenatchee River basin (Table 4.26). The numbers in Table 4.26 should be considered rough estimates because they are not based on confirmed spawning (only last detections) and the numbers have not been adjusted for detection efficiencies, which currently do not exist for PIT-tag detection arrays in tributaries. What these data do indicate is that some hatchery sockeye from the Wenatchee program have wandered or strayed into the Entiat and Methow rivers and possibly into the Okanogan system (based on sockeye detected at Wells Dam but not in the Methow River).

**Table 4.26.** Number and percent of hatchery-origin Wenatchee sockeye that homed to target spawning areas and the target hatchery program, and number and percent that strayed to non-target spawning areas and hatchery programs for brood years 2005-2009. Estimates were based on last detections of PIT-tagged hatchery sockeye. Percent strays should be less than 5%.

Brood Year	Homing				Straying			
	Target streams		Target hatchery*		Non-target stream		Non-target hatchery	
	Number	%	Number	%	Number	%	Number	%
2005	166	92.2	0	0	14	7.8	0	0
2006	440	94.6	0	0	25	5.4	0	0
2007	192	95.0	0	0	10	5.0	0	0
2008	127	89.4	0	0	15	10.6	0	0
2009	41	82.0	0	0	9	18.0	0	0
<b>Average</b>	<b>193</b>	<b>90.6</b>	<b>0</b>	<b>0</b>	<b>15</b>	<b>9.4</b>	<b>0</b>	<b>0</b>

\* Homing to the target hatchery includes Wenatchee hatchery sockeye that are captured and included as broodstock in the Wenatchee Hatchery program. These hatchery fish were collected at Tumwater Dam.

## Genetics

Genetic studies were conducted to determine the potential effects of the Wenatchee sockeye supplementation program on natural-origin sockeye in the upper Wenatchee River basin (Blankenship et al. 2008; the entire report is appended as Appendix I). Specifically, the objective of the study was to determine if the genetic composition of the Lake Wenatchee sockeye population had been altered by the supplementation program, which was based on the artificial propagation of a small subset of the Wenatchee population. Microsatellite DNA allele frequencies were used to differentiate between temporally replicated collections of natural and hatchery-origin sockeye in the Wenatchee River basin. A total of 13 collections of Wenatchee sockeye were analyzed; eight temporally replicated collections of natural-origin sockeye and five temporally replicated collections of hatchery-origin sockeye. Paired natural-hatchery collections were available from return years 2000, 2001, 2004, 2006, and 2007.

Overall, the study showed that allele frequency distributions were consistent over time, regardless of origin, resulting in small, insignificant measures of genetic differentiation among collections. This indicates that there was no year-to-year differences in allele frequencies between natural and hatchery-origin sockeye. In addition, the analyses found no differences between pre- and post-supplementation collections. Thus, it was concluded that the allele frequencies of the broodstock collections equaled the allele frequency of the natural collections.

## Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate

selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

The approximate PNI Index values for the life of the program (brood years 1989-2011) are shown in Table 4.27. Throughout the program, PNI was consistently greater than 0.67. The hatchery program was terminated in 2012.

**Table 4.27.** Approximate Proportionate Natural Influence (PNI) Index of the Wenatchee sockeye supplementation program for brood years 1989-2011. PNI was calculated as the proportion of naturally produced sockeye in the hatchery broodstock (pNOB) divided by the proportion of hatchery sockeye counted at Tumwater Dam (pHOS) plus pNOB. NOS = number of natural-origin sockeye counted at Tumwater Dam; HOS = number of hatchery-origin sockeye counted at Tumwater Dam; NOB = number of natural-origin sockeye collected for broodstock; and HOB = number of hatchery-origin sockeye included in hatchery broodstock.

Brood year	Spawners <sup>a</sup>			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
1989	21,802	0	0.00	115	0	1.00	1.00
1990	27,325	0	0.00	302	0	1.00	1.00
1991	26,689	0	0.00	199	0	1.00	1.00
1992	16,461	0	0.00	320	0	1.00	1.00
1993	25,064	2,662	0.10	207	0	1.00	0.91
1994	6,934	396	0.05	236	5	0.98	0.95
1995	3,262	186	0.05	194	3	0.98	0.95
1996	6,027	546	0.08	225	0	1.00	0.93
1997	8,376	68	0.01	192	19	0.91	0.99
1998	3,982	32	0.01	122	6	0.95	0.99
1999	961	64	0.06	79	60	0.57	0.90
2000	19,574	1,161	0.06	170	5	0.97	0.94
2001	28,288	815	0.03	200	7	0.97	0.97
2002	27,372	193	0.01	256	0	1.00	0.99
2003	4,797	58	0.01	198	0	1.00	0.99
2004	26,095	1,460	0.05	177	0	1.00	0.95
2005	13,983	28	0.00	166	0	1.00	1.00
2006	9,183	255	0.03	214	0	1.00	0.97
2007	2,320	59	0.02	210	0	1.00	0.98
2008	22,931	92	0.00	243	2	0.99	1.00
2009	13,093	447	0.03	239	0	1.00	0.97
2010	30,357	1,134	0.04	198	0	1.00	0.96
2011	17,490	940	0.05	196	0	1.00	0.95
2012	18,214	296	0.02	--	--	--	--
2013	22,118	614	0.03	--	--	--	--
<b>Average</b>	<b>16,108</b>	<b>460</b>	<b>0.03</b>	<b>203</b>	<b>5</b>	<b>0.97</b>	<b>0.97</b>

<sup>a</sup> Proportions of natural-origin and hatchery-origin spawners were determined from video tape at Tumwater Dam.

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel time (arithmetic mean days) of hatchery sockeye salmon from Lake Wenatchee to McNary Dam, and smolt to adult ratios (SARs) from release to detection at Bonneville Dam (Table 4.28).<sup>6</sup> Over the seven brood years for which PIT-tagged hatchery fish were released, survival rates from Lake Wenatchee to McNary Dam ranged from 0.211 to 0.370; SARs from release to detection at Bonneville Dam ranged from 0.005 to 0.044. Average travel time from Lake Wenatchee to McNary Dam ranged from 176 to 202 days.

**Table 4.28.** Total number of hatchery sockeye parr released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2004-2010. Standard errors are shown in parentheses. NA = not available (i.e., not all the adults from the release groups have returned to the Columbia River).

Brood year	Number of sockeye released with PIT tags	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2004	14,859	0.334 (0.013)	176.4 (61.9)	0.020 (0.001)
2005	14,764	0.370 (0.030)	202.0 (9.1)	0.044 (0.002)
2006	14,947	0.312 (0.013)	199.9 (8.6)	0.024 (0.001)
2007	14,858	0.307 (0.020)	192.9 (35.7)	0.015 (0.001)
2008	14,486	0.211 (0.015)	194.2 (29.1)	0.005 (0.001)
2009	5,039	0.302 (0.048)	191.7 (26.6)	0.014 (0.002)
2010	5,074	0.315 (0.038)	196.7 (7.3)	NA

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population. Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). We calculated NORs with and without harvest. NORs without harvest include all returning fish that either returned to the basin or were collected as wild broodstock. NORs with harvest include all fish harvested and are based on a brood year harvest rates from the hatchery program. For brood years 1989-2008, NRR in the Wenatchee averaged 1.51 (range, 0.13-5.30) if harvested fish were not included in the estimate and 1.79 (range, 0.14-6.42) if harvested fish were included in the estimate (Table 4.29).

Hatchery replacement rates (HRR) were estimated as hatchery adult-to-adult returns. These rates should be greater than the NRRs and greater than or equal to 5.40 (the calculated target value in Hillman et al. 2013). HRRs exceeded NRRs in 12 or 13 of the 20 years of data depending on if harvest was or was not included in the estimates (Table 4.29). Hatchery replacement rates for

<sup>6</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

Wenatchee sockeye have equaled or exceeded the estimated target value of 5.40 in four or five of the 20 years depending on if harvest was or was not included in the estimate (Table 4.29).

**Table 4.29.** Broodstock collected, spawning escapements, natural and hatchery-origin recruits (NOR and HOR), and natural and hatchery replacement rates (NRR and HRR; with and without harvest) for sockeye salmon in the Wenatchee River basin, 1989-2008.

Brood year	Broodstock Collected	Spawning Escapement	Harvest not included				Harvest included			
			HOR	NOR	HRR	NRR	HOR	NOR	HRR	NRR
1989	255	21,802	2757	23,616	10.81	1.08	3680	30,672	14.43	1.41
1990	316	27,325	401	3,509	1.27	0.13	423	3,701	1.34	0.14
1991	233	26,689	95	4,820	0.41	0.18	101	5,116	0.43	0.19
1992	343	16,461	576	5,336	1.68	0.32	615	5,685	1.79	0.35
1993	307	27,726	71	11,151	0.23	0.40	75	11,815	0.24	0.43
1994	265	7,330	47	1,191	0.18	0.16	50	1,337	0.19	0.18
1995	209	3,448	121	838	0.58	0.24	131	912	0.63	0.26
1996	227	6,573	1,348	28,049	5.94	4.27	1423	30,840	6.27	4.69
1997	226	8,444	739	36,097	3.27	4.27	834	41,794	3.69	4.95
1998	190	4,014	104	16,166	0.55	4.03	111	17,118	0.58	4.26
1999	147	1,025	68	566	0.46	0.55	83	682	0.56	0.67
2000	195	20,735	1,425	29,082	7.31	1.40	1907	35,314	9.78	1.70
2001	245	29,103	24	17,242	0.10	0.59	28	18,069	0.11	0.62
2002	257	27,565	281	5,752	1.09	0.21	297	6,206	1.16	0.23
2003	219	4,855	32	2,054	0.15	0.42	35	2,588	0.16	0.53
2004	202	27,555	94	23,599	0.47	0.86	293	30,130	1.45	1.09
2005	207	14,011	462	20,833	2.23	1.49	608	26,504	2.94	1.89
2006	220	9,438	1,145	26,966	5.20	2.86	1679	32,421	7.63	3.44
2007	228	2,379	911	12,604	4.00	5.30	1029	15,283	4.51	6.42
2008	260	23,023	11,072	33,346	42.58	1.45	14187	54,853	54.57	2.38
<i>Average</i>	<i>238</i>	<i>15,475</i>	<i>1,089</i>	<i>15,141</i>	<i>4.43</i>	<i>1.51</i>	<i>1379</i>	<i>18,552</i>	<i>5.62</i>	<i>1.79</i>

### Juvenile-to-Adult Survivals

When possible, both parr-to-adult ratios (PAR) and smolt-to-adult ratios (SAR) were calculated for hatchery sockeye salmon. Ratios were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery parr released or the estimated number of smolts emigrating from Lake Wenatchee. Here, survival ratios were based on CWT returns, when available, or on the estimated number of hatchery adults recovered on the spawning grounds, in broodstock, and harvested. For the available brood years, PARs have ranged from 0.0001 to 0.0339 for hatchery sockeye salmon and SARs have ranged from 0.0002 to 0.0254 (Table 4.30).

**Table 4.30.** Parr-to-adult ratios (PAR) and smolt-to-adult ratios (SAR) for Wenatchee hatchery sockeye salmon, brood years 1990-2007; NA = not available.

Brood year	Number of parr released	Number of smolts	Estimated adult recaptures	PAR	SAR
1989	108,400	NA	3,680	0.0339	NA
1990	270,802	NA	423	0.0016	NA
1991	167,523	NA	101	0.0006	NA
1992	340,597	NA	615	0.0018	NA
1993	190,443	NA	75	0.0004	NA
1994	252,859	NA	50	0.0002	NA
1995	150,808	28,828	131	0.0009	0.0045
1996	284,630	55,985	1,423	0.0050	0.0254
1997	197,195	112,524	834	0.0042	0.0074
1998	121,344	24,684	111	0.0009	0.0045
1999	167,955	94,046	83	0.0005	0.0009
2000	190,174	121,511	1,907	0.0100	0.0157
2001	200,938	140,322	28	0.0001	0.0002
2002	315,783	216,023	297	0.0009	0.0014
2003	240,459	122,399	35	0.0001	0.0003
2004	172,923	159,500	293	0.0017	0.0018
2005	140,542	140,542	608	0.0043	0.0043
2006	225,670	121,843	1,679	0.0074	0.0138
2007	252,133	119,908	1,029	0.0041	0.0086
<i>Average</i>	<i>210,062</i>	<i>112,163</i>	<i>705</i>	<i>0.0041</i>	<i>0.0068</i>

## 4.8 ESA/HCP Compliance

### Smolt and Emigrant Trapping

ESA-listed spring Chinook and steelhead were encountered during operation of the Lower Wenatchee trap. ESA takes are reported in the steelhead (Section 3.8) and spring Chinook (Section 5.8) sections and will not be repeated here.

### Spawning Surveys

Sockeye spawning ground surveys conducted in the Wenatchee River basin during 2014 were consistent with ESA Section 10 Permit No. 1347. Because of the difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical and extreme caution was used to avoid established redds when wading was required.

## SECTION 5: WENATCHEE (CHIWAWA) SPRING CHINOOK

The goal of Chiwawa spring Chinook salmon supplementation is to achieve “No Net Impact” to the productivity of spring Chinook caused by the operation of the Rock Island Hydroelectric Project. The Rock Island Fish Hatchery Complex began operation in 1989 under funding from Chelan PUD. The Complex operated originally through the Rock Island Settlement Agreement, but since 2004 has operated under the Rock Island and Rocky Reach Anadromous Fish Agreement and Habitat Conservation Plans.

Adult spring Chinook are collected for broodstock at the Chiwawa Weir and Tumwater Dam. From 2011 through 2013, all spring Chinook broodstock were collected at the Chiwawa Weir in order to reduce passage delays caused by trapping at Tumwater Dam. Prior to 2012, the goal was to collect up to 379 adult spring Chinook for the program with natural-origin fish making up not less than 33% of the broodstock. In 2011, the Hatchery Committees reevaluated the amount of hatchery compensation needed to achieve NNI. Based on that evaluation, the goal of the program was revised. The current goal (beginning in 2012) is to collect 74 natural-origin spring Chinook. The number collected cannot exceed 33% of the natural-origin spring Chinook returns to Tumwater. Beginning in 2014, previously PIT-tagged hatchery-origin Chiwawa spring Chinook are collected at Tumwater Dam, while the Chiwawa Weir is used to collect natural-origin brood for the Chiwawa spring Chinook program. Broodstock collection occurs from May through July at Tumwater with trapping occurring up to 24 hours per day, seven days a week and at the Chiwawa Weir with trapping occurring from 15 June to 1 August (not to exceed 15 cumulative trapping days) on a 24-hour-up/24-hour-down schedule consistent with annual broodstock collection protocols.

Adult spring Chinook are spawned and reared at Eastbank Fish Hatchery. Juvenile spring Chinook are transferred from the hatchery to the Chiwawa Acclimation Facility in late September or early October. They are released voluntarily from the Chiwawa Acclimation Facility during April the following year.

Before 2012, the production goal for the Chiwawa spring Chinook supplementation program was to release 672,000 yearling smolts into the Chiwawa River at 12 fish per pound. Beginning with the 2014 brood, the revised production goal is to release 144,026 smolts as part of a conservation program at 18 fish per pound. The Wenatchee spring Chinook safety-net program is now part of the Nason Creek spring Chinook program. Targets for fork length and weight are 155 mm (CV = 9.0) and 37.8 g, respectively. Over 90% of these fish are marked with CWTs. In addition, since 2006, juvenile spring Chinook have been PIT tagged annually.

Although this section of the report focuses on results from monitoring the Chiwawa spring Chinook program, information on spring Chinook collected throughout the Wenatchee River basin is also provided. Information specific to the Nason Creek spring Chinook conservation program is presented in Section 6 and the White River Captive Broodstock Program is presented in Section 7.

### 5.1 Broodstock Sampling

This section focuses on results from sampling 2012-2014 Chiwawa spring Chinook broodstock, which were collected at the Chiwawa Weir and at Tumwater Dam, consistent with methods in

the broodstock collections protocols (Hillman et al. 2013). Some information for the 2014 return is not available at this time (e.g., age structure and final origin determination). This information will be provided in the 2015 annual report.

### Origin of Broodstock

Hatchery-origin adults made up between 68% of the Chiwawa spring Chinook broodstock for return years 2012-2014 (Table 5.1). Natural and hatchery-origin adults were collected at Tumwater Dam and the Chiwawa Weir for return year 2014. Broodstock were trapped at Tumwater Dam from mid-June through mid-July of 2014, and at the Chiwawa Weir from mid-July through mid-August. Hatchery-origin broodstock were collected at Tumwater Dam in 2014 to meet the Nason Creek Safety Net requirements. Additional hatchery-origin broodstock were collected to ensure production obligations were achieved in the event that insufficient natural-origin collections could be made. A total of 67 hatchery-origin fish collected in 2014 were surplus at Eastbank Fish Hatchery.

**Table 5.1.** Numbers of wild and hatchery Chiwawa spring Chinook collected for broodstock, numbers that died before spawning, and numbers of Chinook spawned, 1989-2014. Unknown origin fish (i.e., undetermined by scale analysis, no CWT or fin clips, and no additional hatchery marks) were considered naturally produced. Mortality includes fish that died of natural causes typically near the end of spawning and were not needed for the program or were surplus fish killed at spawning.

Brood year	Wild spring Chinook					Hatchery spring Chinook					Total number spawned
	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	
1989	28	0	0	28	0	0	0	0	0	0	28
1990	19	1	0	18	0	0	0	0	0	0	18
1991	32	0	5	27	0	0	0	0	0	0	27
1992	113	0	0	78	35	0	0	0	0	0	78
1993	100	3	3	94	0	0	0	0	0	0	94
1994	9	0	1	8	0	4	0	0	4	0	12
1995	No Program										
1996	8	0	0	8	0	10	0	0	10	0	18
1997	37	0	5	32	0	83	1	3	79	0	111
1998	13	0	0	13	0	35	1	0	34	0	47
1999	No Program										
2000	10	0	1	9	0	38	1	16	21	0	30
2001	115	2	0	113	0	267	8	0	259	0	372
2002	21	0	1	20	0	63	1	11	51	0	71
2003	44	1	2	41	0	75	2	20	53	0	94
2004	100	1	16	83	0	196	30	34	132	0	215
2005	98	1	6	91	0	185	3	1	181	0	279
2006	95	0	4	91	0	303	0	29	224	50	315
2007	45	1	1	43	0	124	2	18	104	0	147
2008	88	2	3	83	0	241	5	16	220	0	303
2009	113	6	11	96	0	151	3	37	111	0	207
2010	83	0	6	77	0	103	0	5	98	0	175
2011	80	0	0	80	0	101	2	6	93	0	173
<i>Average<sup>b</sup></i>	<i>60</i>	<i>1</i>	<i>3</i>	<i>54</i>	<i>2</i>	<i>94</i>	<i>3</i>	<i>9</i>	<i>80</i>	<i>2</i>	<i>134</i>
2012	75	1	1	73	0	41	3	38	0	0	111



Brood year	Wild spring Chinook					Hatchery spring Chinook					Total number spawned
	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	
2013	170	5	0	70	95	52	1	50	0	1	70
2014	61	0	0	61	0	203	1	68	134	0	195
<i>Average<sup>c</sup></i>	<i>102</i>	<i>2</i>	<i>0</i>	<i>68</i>	<i>32</i>	<i>99</i>	<i>2</i>	<i>52</i>	<i>45</i>	<i>0</i>	<i>125</i>

<sup>a</sup> Pre-spawn loss represents the number of fish that died during the holding period before spawning. Mortality is the number of fish that were surplus following spawning.

<sup>b</sup> This average represents the program before recalculation in 2011.

<sup>c</sup> This average represents the current program, which began in 2012. Origin determinations should be considered preliminary pending scale analyses.

### Age/Length Data

Ages were determined from scales and/or coded wire tags (CWT) collected from broodstock. For both the 2012 and 2013 returns, most adults, regardless of origin, were age-4 Chinook (Table 5.2). A larger percentage of the age-5 Chinook were natural-origin fish, whereas a larger percentage of the age-3 fish were hatchery-origin fish.

**Table 5.2.** Percent of hatchery and wild spring Chinook of different ages (total age) collected from broodstock, 1991-2013.

Return year	Origin	Total age			
		2	3	4	5
1991	Wild	0.0	0.0	22.0	78.0
	Hatchery	0.0	0.0	0.0	0.0
1992	Wild	0.0	0.0	28.6	71.4
	Hatchery	0.0	0.0	50.0	50.0
1993	Wild	0.0	0.0	22.0	78.0
	Hatchery	0.0	0.0	0.0	0.0
1994	Wild	0.0	0.0	28.6	71.4
	Hatchery	0.0	0.0	50.0	50.0
1995	Wild	No program			
	Hatchery				
1996	Wild	0.0	28.6	71.4	0.0
	Hatchery	0.0	50.0	50.0	0.0
1997	Wild	0.0	0.0	87.5	12.5
	Hatchery	0.0	1.2	98.8	0.0
1998	Wild	0.0	0.0	63.6	36.4
	Hatchery	0.0	0.0	62.9	37.1
1999	Wild	No program			
	Hatchery				
2000	Wild	0.0	20.0	70.0	10.0
	Hatchery	0.0	59.1	40.9	0.0
2001	Wild	0.0	2.8	94.4	2.8
	Hatchery	0.0	1.5	98.5	0.0

Return year	Origin	Total age			
		2	3	4	5
2002	Wild	0.0	0.0	66.7	33.3
	Hatchery	0.0	0.0	93.4	6.6
2003	Wild	0.0	27.0	2.7	70.3
	Hatchery	0.0	21.3	5.3	73.3
2004	Wild	1.0	6.1	88.8	4.1
	Hatchery	0.0	40.4	59.6	0.0
2005	Wild	0.0	1.0	85.0	14.0
	Hatchery	0.0	4.4	95.6	0.0
2006	Wild	0.0	2.0	70.4	27.6
	Hatchery	0.0	1.3	81.2	17.4
2007	Wild	0.0	15.6	53.3	31.1
	Hatchery	0.0	27.4	60.5	12.1
2008	Wild	0.0	6.3	78.8	15.0
	Hatchery	0.0	8.2	86.8	4.9
2009	Wild	0.0	8.6	79.0	12.4
	Hatchery	0.0	18.5	79.5	2.0
2010	Wild	0.0	5.3	94.7	0.0
	Hatchery	0.0	0.0	99.0	1.0
2011	Wild	0.0	2.7	52.7	44.6
	Hatchery	0.0	20.4	60.2	19.4
2012	Wild	0.0	0.0	79.0	21.0
	Hatchery	0.0	4.3	95.7	0.0
2013	Wild	0.0	0.0	65.7	34.3
	Hatchery	0.0	2.2	86.7	11.1
Average	Wild	0.0	6.0	62.1	31.8
	Hatchery	0.0	12.4	64.5	13.6

There was little difference in mean lengths between hatchery and natural-origin broodstock of age-4 and 5 Chinook in 2012 and 2013 (Table 5.3).

**Table 5.3.** Mean fork length (cm) at age (total age) of hatchery and wild spring Chinook collected from broodstock, 1991-2013; N = sample size and SD = 1 standard deviation.

Return year	Origin	Spring Chinook fork length (cm)											
		Age-2			Age-3			Age-4			Age-5		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
1991	Wild	-	0	-	-	5	-	-	19	-	-	8	-
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-
1992	Wild	-	0	-	-	0	-	-	0	-	-	0	-
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-

Return year	Origin	Spring Chinook fork length (cm)											
		Age-2			Age-3			Age-4			Age-5		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
1993	Wild	-	0	-	-	0	-	79	4	3	92	8	4
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-
1994	Wild	-	0	-	-	0	-	79	2	3	96	5	6
	Hatchery	-	0	-	-	0	-	82	2	11	92	2	2
1995	Wild	No program											
	Hatchery												
1996	Wild	-	0	-	51	2	1	79	5	7	-	0	-
	Hatchery	-	0	-	56	5	4	74	5	6	-	0	-
1997	Wild	-	0	-	-	0	-	80	28	5	99	4	8
	Hatchery	-	0	-	56	1	-	82	82	4	-	0	-
1998	Wild	-	0	-	-	0	-	78	7	13	83	4	18
	Hatchery	-	0	-	-	0	-	77	22	8	93	13	7
1999	Wild	No program											
	Hatchery												
2000	Wild	-	0	-	51	2	3	82	7	4	98	1	-
	Hatchery	-	0	-	59	13	4	79	9	8	-	0	-
2001	Wild	-	0	-	49	3	6	82	101	6	95	3	3
	Hatchery	-	0	-	56	4	7	83	261	5	-	0	-
2002	Wild	-	0	-	-	0	-	79	12	4	96	6	10
	Hatchery	-	0	-	-	0	-	81	57	6	94	4	9
2003	Wild	-	0	-	55	10	5	83	1	-	99	26	6
	Hatchery	-	0	-	59	16	5	86	4	18	96	55	6
2004	Wild	47	1	-	60	6	6	80	87	5	99	4	3
	Hatchery	-	0	-	51	80	7	80	118	5	-	0	-
2005	Wild	-	0	-	49	1	-	80	85	6	96	14	8
	Hatchery	-	0	-	56	8	5	82	175	6	-	0	-
2006	Wild	-	0	-	50	2	2	79	69	7	97	27	5
	Hatchery	-	0	-	46	1	-	80	205	6	95	43	7
2007	Wild	-	0	-	54	7	3	79	24	6	93	14	7
	Hatchery	-	0	-	59	34	8	81	75	5	93	15	7
2008	Wild	-	0	-	54	5	9	83	63	5	93	12	6
	Hatchery	-	0	-	56	20	10	82	211	6	96	12	7
2009	Wild	-	0	-	52	9	6	81	83	5	94	13	6
	Hatchery	-	0	-	56	28	6	82	120	5	87	3	11
2010	Wild	-	0	-	58	4	9	80	72	6	-	0	-
	Hatchery	-	0	-	-	0	-	82	102	6	101	1	-
2011	Wild	-	0	-	56	2	3	79	39	5	95	33	7

Return year	Origin	Spring Chinook fork length (cm)											
		Age-2			Age-3			Age-4			Age-5		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
	Hatchery	-	0	-	63	21	7	80	62	6	95	20	6
2012	Wild	-	0	-	-	0	-	81	49	6	97	13	8
	Hatchery	-	0	-	51	2	0	80	41	5	-	0	-
2013	Wild	-	0	-	-	1	-	74	44	6	92	23	8
	Hatchery	-	0	-	60	1	-	78	39	6	88	5	7
Average	Wild	47	0	-	53	3	5	80	38	6	95	10	7
	Hatchery	-	0	-	56	11	5	81	76	7	94	8	7

### Sex Ratios

Male spring Chinook in the 2012-2014 return years made up 49.5%, 49.1%, and 49.2%, respectively, of the adults collected. This resulted in overall male to female ratios of 0.90:1.00, 0.96:1.00, and 0.97:1.00, respectively (Table 5.4). For the 2014 return year, natural-origin and hatchery-origin fish both consisted of a slightly higher proportion of females than males (Table 5.4).

**Table 5.4.** Numbers of male and female wild and hatchery spring Chinook collected for broodstock, 1989-2014. Ratios of males to females are also provided.

Return year	Number of wild spring Chinook			Number of hatchery spring Chinook			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
1989	11	17	0.65:1.00	-	-	-	0.65:1.00
1990	7	12	0.58:1.00	-	-	-	0.58:1.00
1991	13	19	0.68:1.00	-	-	-	0.68:1.00
1992	39	39	1.00:1.00	-	-	-	1.00:1.00
1993	50	50	1.00:1.00	-	-	-	1.00:1.00
1994	5	4	1.25:1.00	2	2	1.00:1.00	1.17:1.00
1995	No program						
1996	6	2	3.00:1.00	8	2	4.00:1.00	3.50:1.00
1997	14	23	0.61:1.00	34	49	0.69:1.00	0.67:1.00
1998	9	4	2.25:1.00	18	17	1.06:1.00	1.29:1.00
1999	No program						
2000	5	5	1.00:1.00	32	6	5.33:1.00	3.36:1.00
2001	45	70	0.64:1.00	90	177	0.51:1.00	0.55:1.00
2002	9	12	0.75:1.00	30	33	0.91:1.00	0.87:1.00
2003	28	16	1.75:1.00	42	33	1.27:1.00	1.43:1.00
2004	58	42	1.38:1.00	102	94	1.09:1.00	1.18:1.00
2005	58	40	1.45:1.00	89	96	0.93:1.00	1.08:1.00
2006	49	46	1.07:1.00	123	179	0.69:1.00	0.77:1.00
2007	20	25	0.80:1.00	66	58	1.14:1.00	1.04:1.00

Return year	Number of wild spring Chinook			Number of hatchery spring Chinook			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
2008	41	47	0.87:1.00	109	132	0.83:1.00	0.84:1.00
2009	53	60	0.88:1.00	79	72	1.10:1.00	1.00:1.00
2010	41	42	0.98:1.00	53	50	1.06:1.00	1.02:1.00
2011	38	42	0.90:1.00	53	48	1.10:1.00	1.01:1.00
2012	35	40	0.87:1.00	20	21	0.95:1.00	0.90:1.00
2013	83	87	0.95:1.00	26	26	1.00:1.00	0.96:1.00
2014	29	32	0.91:1.00	101	102	0.99:1.00	0.97:1.00
<b>Total</b>	<b>746</b>	<b>776</b>	<b>0.96:1.00</b>	<b>1,077</b>	<b>1,197</b>	<b>0.91:1.00</b>	<b>0.92:1.00</b>

### Fecundity

Mean fecundities for the 2012-2014 returns of spring Chinook ranged from 4,045-4,716 eggs per female (Table 5.5). These fecundities were generally less than the overall average of 4,654 eggs per female, but were close to the expected fecundity of 4,400 eggs per female assumed in the broodstock protocol. For the 2014 return year, natural-origin Chinook produced more eggs per female than did hatchery-origin fish. This could be attributed to differences in size and age of hatchery and natural-origin fish described above (Tables 5.2 and 5.3).

**Table 5.5.** Mean fecundity of wild, hatchery, and all female spring Chinook collected for broodstock, 1989-2014; NA = not available.

Return year	Mean fecundity		
	Wild	Hatchery	Total
1989*	NA	NA	2,832
1990*	NA	NA	5,024
1991*	NA	NA	4,600
1992*	NA	NA	5,199 <sup>a</sup>
1993*	NA	NA	5,249
1994*	NA	NA	5,923
1995	No program		
1996*	NA	NA	4,645
1997	4,752	4,479	4,570
1998	5,157	5,376	5,325
1999	No program		
2000	5,028	5,019	5,023
2001	4,530	4,663	4,624
2002	5,024	4,506	4,654
2003	6,191	5,651	5,844
2004	4,846	4,775	4,799
2005	4,365	4,312	4,327
2006	4,773	4,151	4,324

Return year	Mean fecundity		
	Wild	Hatchery	Total
2007	4,656	4,351	4,441
2008	4,691	4,560	4,592
2009	4,691	4,487	4,573
2010	4,548	4,114	4,314
2011	4,969	3,884	4,385
2012	4,522	3,682	4,223
2013	4,716	0	4,716
2014	4,467	3,834	4,045
<i>Average</i>	<i>4,819</i>	<i>4,226</i>	<i>4,654</i>

\* Individual fecundities were not tracked with females until 1997.

<sup>a</sup> Estimated as the mean of fecundities two years before and two years after 1992.

## 5.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Based on the unfertilized egg-to-release survival standard of 81%, a total of 829,630 eggs were required to meet the program release goal of 672,000 smolts for brood years 1989-2010. For the 2011 and 2012 brood years, a total of 367,536 and 252,410 eggs were required to meet the release goals of 298,000 and 204,452 smolts, respectively. Since 2013, 169,442 eggs have been required to achieve a release goal of 144,026 smolts for the Chiwawa spring Chinook Program. Between 1989 and 2014, the egg take goal was reached only in 2001 (Table 5.6). The green egg takes for 2012-2014 brood years were 99.3%, 97.4%, and 99.7% of program goals, respectively.

ESA Permit 18121 sets limits on the percentage of the total run and natural-origin fish in the broodstock to meet the conservation program. Applying these criteria to the low total abundance of spring Chinook salmon to the Chiwawa River basin and the low abundance of natural-origin fish returning to the basin has resulted in the program not meeting production goals.

**Table 5.6.** Numbers of eggs taken from spring Chinook broodstock, 1989-2014; NP = no program. Egg take for the Nason Creek Safety Net Program began in 2014.

Return year	Number of eggs taken	
	Chiwawa Program	Nason Creek Safety Net Program
1989	45,311	NP
1990	60,287	NP
1991	73,601	NP
1992	111,624	NP
1993	257,208	NP
1994	35,539	NP
1995	NP	NP

Return year	Number of eggs taken	
	Chiwawa Program	Nason Creek Safety Net Program
1996	18,579	NP
1997	312,182	NP
1998	90,521	NP
1999	NP	NP
2000	55,256	NP
2001	1,099,630	NP
2002	196,186	NP
2003	247,501	NP
2004	538,176	NP
2005	536,490	NP
2006	744,344	NP
2007	359,739	NP
2008	761,821	NP
2009	564,912	NP
2010	383,944	NP
2011	366,244	NP
<b>Average (1989-2011)</b>	<b>326,624</b>	<b>NP</b>
2012	250,695	NP
2013	165,047	NP
2014	169,007	217,290
<b>Average (2012-present)</b>	<b>227,843</b>	<b>217,290</b>

### **Number of acclimation days**

Early rearing of the 2012 brood Chiwawa spring Chinook was similar to previous years with fish being held on well water before being transferred to Chiwawa Acclimation Facility for final acclimation. Beginning in 2006 (2005 brood acclimation), modifications were made to the Chiwawa Acclimation Facility intakes so that Wenatchee River water could be applied to the Chiwawa River intakes during severe cold periods to prevent the formation of frazzle ice. During acclimation of the 2012 brood, fish were acclimated for 204 to 211 days on Chiwawa River water, with 107 of those days containing a small percentage of Wenatchee River water to prevent freezing of hatchery intakes (Table 5.7).

**Table 5.7.** Number of days spring Chinook broods were acclimated and water source, brood years 1989-2012; NA = not available.

Brood year	Release year	Transfer date	Release date	Number of days and water source		
				Total	Chiwawa	Wenatchee
1989	1991	19-Oct	11-May	204	NA	NA
1990	1992	13-Sep	27-Apr	227	NA	NA
1991	1993	24-Sep	24-Apr	212	NA	NA
1992	1994	30-Sep	20-Apr	202	NA	NA
1993	1995	28-Sep	20-Apr	204	NA	NA
1994	1996	1-Oct	25-Apr	207	NA	NA
1995	1997	No Program				
1996	1998	25-Sep	29-Apr	216	NA	NA
1997	1999	28-Sep	22-Apr	206	NA	NA
1998	2000	27-Sep	24-Apr	210	NA	NA
1999	2001	No Program				
2000	2002	26-Sep	25-Apr	211	NA	NA
2001	2003	22-Oct	1-May	191	NA	NA
2002	2004	25-Sep	2-May	220	NA	NA
2003	2005	30-Sep	3-May	215	NA	NA
		30-Sep	18-Apr-18-May	200	NA	NA
2004	2006	3-Sep	1-May	240	88-104	124
		3-Sep	17-Apr-17-May	226	NA	NA
2005	2007	25-Sep	1-May	217	217	98 <sup>a</sup>
		26-Sep	16-Apr-15-May	202-232	202-232	98 <sup>a</sup>
2006	2008	24-27-Sep	14-Apr-13-May	231	231	95 <sup>a</sup>
2007	2009	1-Oct	15-Apr-13-May	223	223	103 <sup>a</sup>
2008	2010	14-15-Sep	14-Apr-12-May	212-241	212-241	129
2009	2011	14-15-Sep	26-Apr-19-May	225-249	225-249	88
2010	2012	3, 5-6-Oct	17-Apr-1-May	195-212	195-212	132
2011	2013	24-26-Sep	16-22-Apr	202-210	202-210	40
2012	2014	23-25 Sep	14-21 Apr	204-211	204-211	107 <sup>a</sup>

<sup>a</sup> Represents the number of days Wenatchee River water was applied to the Chiwawa River intake screen to prevent the formation of frazzle ice.



## Release Information

### Numbers released

The 2012 brood Chiwawa spring Chinook program achieved 108.8% of the 204,452 target goal with about 222,504 smolts being released volitionally into the Chiwawa River in 2014 (Table 5.8).

**Table 5.8.** Numbers of spring Chinook smolts tagged and released from the hatchery, brood years 1989-2012. The release target for Chiwawa spring Chinook is 298,000 smolts.

Brood year	Release year	Type of release	CWT mark rate	Number released that were PIT tagged	Number of smolts released	Total number of smolts released	
1989	1991	Volitional	0.9932	0	43,000	43,000	
1990	1992	Volitional	0.9931	0	53,170	53,170	
1991	1993	Volitional	0.9831	0	62,138	62,138	
1992	1994	Volitional	0.9747	0	85,113	85,113	
1993	1995	Volitional	0.9892	0	223,610	223,610	
1994	1996	Volitional	0.9967	0	27,226	27,226	
1995	1997	No program					
1996	1998	Forced	0.8413	0	15,176	15,176	
1997	1999	Volitional	0.9753	0	266,148	266,148	
1998	2000	Volitional	0.9429	0	75,906	75,906	
1999	2001	No program					
2000	2002	Volitional	0.9920	0	47,104	47,104	
2001	2003	Forced	0.9961	0	192,490 <sup>a</sup>	377,544	
		Volitional	0.9856	0	185,054 <sup>a</sup>		
2002	2004	Volitional	0.9693	0	149,668	149,668	
2003	2005	Forced	0.9783	0	69,907	222,131	
		Volitional	0.9743	0	152,224		
2004	2006	Forced	0.9533	0	243,505	494,517	
		Volitional	0.9493	0	251,012		
2005	2007	Forced	0.9882	4,993	245,406	494,012	
		Volitional	0.9864	4,988	248,606		
2006	2007	Direct	0.0000	0	12,977 <sup>b</sup>	612,482	
	2008	Volitional	0.9795	9,894	612,482		
2007	2008	Direct	0.0000	0	9,494	305,542	
	2009	Volitional	0.9948	10,035	296,048		
2008	2010	Volitional	0.9835	10,006	609,789	609,789	
2009	2011	Forced	0.9874	0	241,181	438,561	
		Volitional	0.9874	9,412	197,380		

Brood year	Release year	Type of release	CWT mark rate	Number released that were PIT tagged	Number of smolts released	Total number of smolts released
2010 <sup>c</sup>	2012	Volitional	0.9904	5,020	346,248	346,248
2011	2013	Volitional	0.9902	9,945	281,821	281,821
2012	2014	Volitional	0.9841	5,061	222,504	222,504

<sup>a</sup> This does not include the 226,456 eyed eggs that were planted in the Chiwawa River.

<sup>b</sup> This high ELISA group was only adipose fin clipped and directly planted into Big Meadow Creek in May.

<sup>c</sup> This does not include 18,480 eyed eggs that were culled because of high ELISA.

### Numbers tagged

The 2012 brood Chiwawa spring Chinook were 98% CWT and adipose fin clipped (Table 5.8).

In 2014, a total of 10,114 spring Chinook from the 2013 brood were PIT tagged at Eastbank Hatchery on 4 to 8 August. These fish were tagged in raceway #12. Fish were not fed the day before tagging, during tagging, or for two days after tagging. Fish averaged 98 mm in length and 12.0 g at time of tagging. These fish were transferred to the Chiwawa Raceway in September 2014. These fish will be released in the Chiwawa River during spring 2015.

Table 5.9 summarizes the number of hatchery spring Chinook that have been PIT-tagged and released into the Chiwawa River.

**Table 5.9.** Summary of PIT-tagging activities for Chiwawa hatchery spring Chinook, brood years 2005-2012.

Brood year	Release year	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2005	2007	10,063	74	8	9,981 <sup>a</sup>
2006	2008	10,055	134	27	9,894
2007	2009	10,112	61	16	10,035
2008	2010	10,101	81	14	10,006
2009	2011	10,101	655	34	9,412
2010	2012	5,102	82	0	5,020
2011	2013	10,200	254	1	9,945
2012	2014	5,100	37	2	5,061

<sup>a</sup> This release consisted of 4,988 tagged Chinook that were released volitionally and 4,993 that were forced released.

### Fish size and condition at release

Spring Chinook from the 2012 brood were released as yearling smolts between 14 and 21 April 2014. Size at release (16 fpp) was larger than the target of 18 fpp established for the program. The CV for fork length was 26% short of the target (Table 5.10).

**Table 5.10.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of spring Chinook smolts released from the hatchery, brood years 1989-2012. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1989	1991	147	4.4	37.8	12
1990	1992	137	5.0	32.4	14
1991	1993	135	4.2	30.3	15
1992	1994	133	5.0	28.4	16
1993	1995	136	4.5	30.2	15
1994	1996	139	7.1	34.4	13
1995	1997	No Program			
1996	1998	157	5.3	52.1	9
1997	1999	146	7.2	38.7	12
1998	2000	143	9.1	39.5	12
1999	2001	No Program			
2000	2002	150	6.8	46.7	10
2001	2003	142	7.1	37.6	12
2002	2004	146	8.5	40.3	11
2003	2005	167 <sup>a</sup>	5.9	59.4	8
		151 <sup>b</sup>	7.4	44.2	10
2004	2006	146 <sup>a</sup>	6.4	39.1	12
		139 <sup>b</sup>	5.7	34.3	13
2005	2007	136 <sup>a</sup>	4.6	30.8	15
		129 <sup>b</sup>	5.8	26.6	17
2006	2008	124	8.8	23.5	19
2007	2008	70 <sup>a</sup>	4.0	3.7	122
	2009	140 <sup>b</sup>	11.0	33.6	14
2008	2010	141	10.7	36.0	13
2009	2011	167	12.9	56.8	8
2010	2012	129	8.1	25.8	18
2011	2013	134	6.4	29.5	15
2012	2014	130	6.7	28.5	16
<b>Average</b>		<b>139</b>	<b>6.9</b>	<b>35.4</b>	<b>17</b>
<b>Targets</b>		<b>155</b>	<b>9.0</b>	<b>37.8</b>	<b>18</b>

<sup>a</sup> Forced release group.

<sup>b</sup> Volitional release group.

### Survival Estimates

Overall survival of Chiwawa spring Chinook from green (unfertilized) egg to release was above the standard set for the program (Table 5.11). There was higher than expected survivals

throughout all stages contributing to increased program performance. Pre-spawn survival of adults was also above the standard set for the program.

**Table 5.11.** Hatchery life-stage survival rates (%) for spring Chinook, brood years 1989-2012. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
1989	100.0	100.0	98.0	99.1	99.1	99.0	96.4	99.3	94.8
1990	100.0	85.7	91.8	98.1	99.5	98.9	97.9	99.2	88.2
1991	100.0	100.0	94.4	96.1	99.6	97.9	93.2	95.0	84.4
1992	100.0	100.0	98.4	96.7	99.9	99.9	80.0	80.6	76.2
1993	96.0	98.0	89.7	98.0	99.7	99.3	98.9	99.7	86.9
1994	100.0	100.0	98.6	100.0	99.8	99.4	77.0	78.9	76.6
1995	No program								
1996	100.0	100.0	88.3	100.0	93.8	93.0	89.9	97.7	81.7
1997	98.6	100.0	93.2	95.7	98.3	99.6	95.6	99.3	85.3
1998	95.2	100.0	94.5	99.0	98.5	98.3	89.6	99.1	83.9
1999	No program								
2000	100.0	100.0	91.0	98.1	97.2	96.6	95.4	99.3	85.2
2001	97.6	97.0	88.9	98.1	99.7	99.6	51.3	51.8	34.3
2002	97.8	100.0	82.1	98.0	97.4	96.7	94.8	99.1	76.3
2003	93.9	100.0	93.2	97.7	99.5	99.3	98.5	98.1	89.7
2004	97.8	82.5	93.3	98.4	98.8	94.3	93.9	97.2	91.9
2005	97.1	100.0	95.9	98.0	99.2	99.0	97.9	99.1	92.1
2006	100.0	100.0	90.1	98.1	99.2	99.0	95.3	97.7	84.2
2007	98.8	97.7	92.9	97.2	99.4	99.0	98.0	99.4	88.5
2008	96.6	99.3	90.8	93.2	97.4	97.1	95.6	97.6	80.0
2009	94.4	97.6	92.5	88.3	97.6	97.4	89.2	92.8	77.6
2010 <sup>a</sup>	98.9	100.0	99.2	100.0	97.9	97.5	95.6	98.2	94.8
2011	98.9	98.9	93.2	88.4	96.8	96.4	93.4	97.1	76.9
2012	98.3	100.0	94.6	98.3	99.7	99.3	98.5	99.4	91.6
<b>Average</b>	<b>98.2</b>	<b>98.0</b>	<b>92.9</b>	<b>97.0</b>	<b>98.5</b>	<b>98.0</b>	<b>91.6</b>	<b>94.3</b>	<b>82.8</b>
<b>Standard</b>	<b>90.0</b>	<b>85.0</b>	<b>92.0</b>	<b>98.0</b>	<b>97.0</b>	<b>93.0</b>	<b>90.0</b>	<b>95.0</b>	<b>81.0</b>

<sup>a</sup> Survival estimates do not include the 18,840 eyed eggs that were culled because of high ELISA levels.

### 5.3 Disease Monitoring

Results of 2014 adult broodstock bacterial kidney disease (BKD) monitoring indicated that all females had ELISA values less than 0.199. About 88.9% of females had ELISA values less than 0.120, which would have required about 11.1% of the progeny to be reared at densities not to exceed 0.06 fish per pound (Table 5.12).

For the 2012 brood, mortalities resulting from external fungal infections began increasing shortly after transfer to the Chiwawa Acclimation Facility. A formalin drip treatments was used to control the infection. No significant health issues were encountered for the remainder of juvenile rearing.

**Table 5.12.** Proportion of bacterial kidney disease (BKD) titer groups for the Chiwawa spring Chinook broodstock, brood years 1996-2014. Also included are the proportions to be reared at either 0.125 fish per pound or 0.060 fish per pound.

Brood year <sup>a</sup>	Optical density values by titer group				Proportion at rearing densities (fish per pound, fpp)	
	Very Low (≤ 0.099)	Low (0.1-0.199)	Moderate (0.2-0.449)	High (≥ 0.450)	≤ 0.125 fpp (<0.119)	≤ 0.060 fpp (>0.120)
1996	0.0000	0.2500	0.2500	0.5000	0.0000	1.0000
1997	0.1176	0.7353	0.0588	0.0882	0.3529	0.6471
1998	0.1176	0.8235	0.0588	0.0000	0.4706	0.5294
1999	No Program					
2000	0.0000	0.9091	0.0909	0.0000	0.1818	0.8182
2001	0.4066	0.5436	0.0373	0.0124	0.6515	0.3485
2002	0.2195	0.6585	0.0732	0.0488	0.5610	0.4390
2003	0.6957	0.1087	0.0652	0.1304	0.7174	0.2826
2004	0.8182	0.1515	0.0227	0.0076	0.8939	0.1061
2005	0.9084	0.0916	0.0000	0.0000	0.9695	0.0305
2006	0.7222	0.2556	0.0000	0.0222	0.8444	0.1556
2007	0.5854	0.3415	0.0244	0.0488	0.7073	0.2927
2008	0.8304	0.1520	0.0058	0.0117	0.9357	0.0643
2009	0.7600	0.1840	0.0080	0.0480	0.8480	0.1520
2010	0.8791	0.0769	0.0000	0.0439	0.9451	0.0549
2011	0.7640	0.2022	0.0000	0.0337	0.8764	0.1236
2012	0.8333	0.1333	0.0167	0.0167	0.9170	0.0830
2013	0.0829	0.1429	0.0286	0.0000	0.8857	0.1143
2014	0.8282	0.1720	0.0000	0.0000	0.8889	0.1111
<b>Average</b>	<b>0.5316</b>	<b>0.3296</b>	<b>0.0411</b>	<b>0.0562</b>	<b>0.7026</b>	<b>0.2974</b>

<sup>a</sup> Individual ELISA samples were not collected before the 1996 brood.

## 5.4 Natural Juvenile Productivity

During 2014, juvenile spring Chinook were sampled at the Lower Wenatchee, Nason Creek, and Chiwawa traps and counted during snorkel surveys within the Chiwawa River basin. Results from sampling at the Nason Creek Trap are provided in Section 6.

### Parr Estimates

Based on snorkel surveys, a total of 121,240 (±11%) subyearling and 939 (±28%) yearling spring Chinook were estimated in the Chiwawa River basin in August 2014 (Table 5.13 and 5.14).

During the survey period 1992-2014, numbers of subyearling and yearling Chinook have ranged from 5,815 to 149,563 and 5 to 967, respectively, in the Chiwawa River basin (Table 5.13 and 5.14; Figure 5.1). Numbers of all fish counted in the Chiwawa River basin are reported in Appendix A.

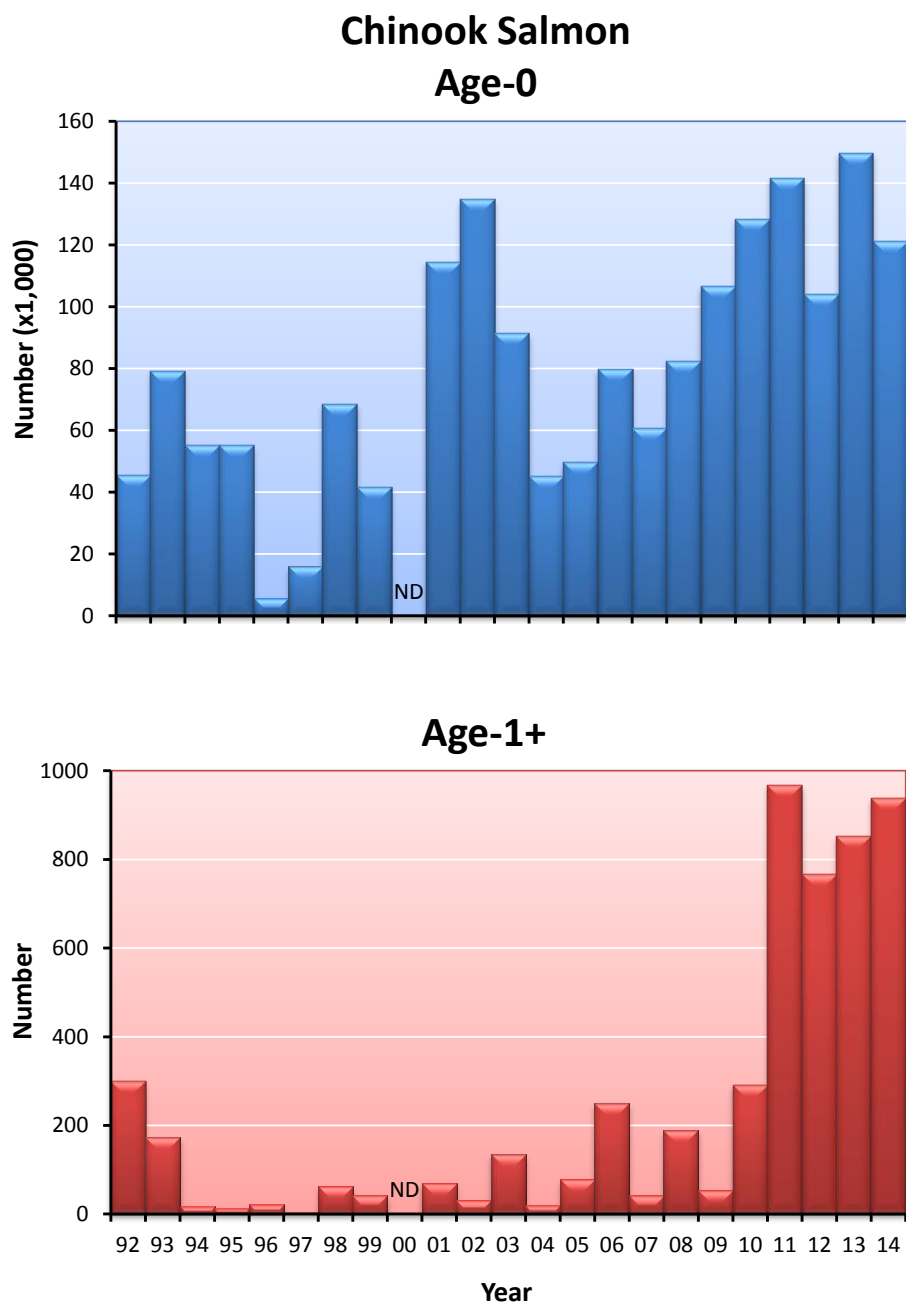
**Table 5.13.** Total numbers of subyearling spring Chinook estimated in different streams in the Chiwawa River basin during snorkel surveys in August 1992-2014; NS = not sampled.

Sample Year	Number of subyearling spring Chinook									
	Chiwawa River	Phelps Creek	Chikamin Creek	Rock Creek	Unnamed Creek	Big Meadow Creek	Alder Creek	Brush Creek	Clear Creek	Total
1992	45,483	NS	NS	NS	NS	NS	NS	NS	NS	45,483
1993	77,269	0	1,258	586	NS	NS	NS	NS	NS	79,113
1994	53,492	0	398	474	68	624	0	0	0	55,056
1995	52,775	0	1,346	210	0	683	67	160	0	55,241
1996	5,500	0	29	10	0	248	28	0	0	5,815
1997	15,438	0	56	92	0	480	0	0	0	16,066
1998	65,875	0	1,468	496	57	506	0	13	0	68,415
1999	40,051	0	366	592	0	598	22	0	0	41,629
2000	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
2001	106,753	168	2,077	2,855	354	2,332	78	0	0	114,617
2002	117,230	75	8,233	2,953	636	5,021	429	0	297	134,874
2003	80,250	4,508	1,570	3,255	118	1,510	22	45	0	91,278
2004	43,360	102	717	215	54	637	21	71	0	45,177
2005	45,999	71	2,092	660	17	792	0	0	0	49,631
2006	73,478	113	2,500	1,681	51	1,890	62	127	0	79,902
2007	53,863	125	5,235	870	51	538	20	28	22	60,752
2008	72,431	214	3,287	4,730	163	1,221	28	255	22	82,351
2009	101,085	125	2,486	1,849	14	1,082	29	18	17	106,705
2010	117,499	526	4,571	4,052	0	1,449	56	42	25	128,220
2011	136,424	64	2,762	1,330	53	581	42	214	40	141,510
2012	96,036	78	4,125	2,227	49	1,322	35	31	37	103,940
2013	140,485	120	3,301	3,214	0	2,345	31	21	46	149,563
2014	113,869	361	2,384	3,124	28	1,367	11	28	68	121,240
<i>Average</i>	<i>75,211</i>	<i>317</i>	<i>2,393</i>	<i>1,689</i>	<i>86</i>	<i>1,261</i>	<i>49</i>	<i>53</i>	<i>29</i>	<i>80,754</i>

**Table 5.14.** Total numbers of yearling spring Chinook estimated in different streams in the Chiwawa River basin during snorkel surveys in August 1992-2014; NS = not sampled.

Sample Year	Number of yearling spring Chinook									
	Chiwawa River	Phelps Creek	Chikamin Creek	Rock Creek	Unnamed Creek	Big Meadow Creek	Alder Creek	Brush Creek	Y Creek	Total
1992	563	NS	NS	NS	NS	NS	NS	NS	NS	563
1993	174	0	0	0	NS	NS	NS	NS	NS	174
1994	14	0	0	4	0	0	0	0	0	18

Sample Year	Number of yearling spring Chinook									
	Chiwawa River	Phelps Creek	Chikamin Creek	Rock Creek	Unnamed Creek	Big Meadow Creek	Alder Creek	Brush Creek	Y Creek	Total
1995	13	0	0	0	0	0	0	0	0	13
1996	22	0	0	0	0	0	0	0	0	22
1997	5	0	0	0	0	0	0	0	0	5
1998	63	0	0	0	0	0	0	0	0	63
1999	41	0	0	0	0	0	0	0	0	41
2000	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
2001	66	0	3	0	0	0	0	0	0	69
2002	32	0	0	0	0	0	0	0	0	32
2003	134	0	0	0	0	0	0	0	0	134
2004	14	0	0	0	0	7	0	0	0	21
2005	62	0	17	0	0	0	0	0	0	79
2006	345	0	0	43	0	0	0	0	0	388
2007	41	0	0	0	0	0	0	0	0	41
2008	144	0	45	0	0	0	0	0	0	189
2009	49	0	0	5	0	0	0	0	0	54
2010	207	27	19	38	0	0	0	0	0	291
2011	645	0	71	194	0	57	0	0	0	967
2012	748	0	0	19	0	0	0	0	0	767
2013	836	0	0	8	0	8	0	0	0	852
2014	867	28	4	38	0	2	0	0	0	939
<b>Average</b>	<b>231</b>	<b>3</b>	<b>8</b>	<b>17</b>	<b>0</b>	<b>4</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>260</b>



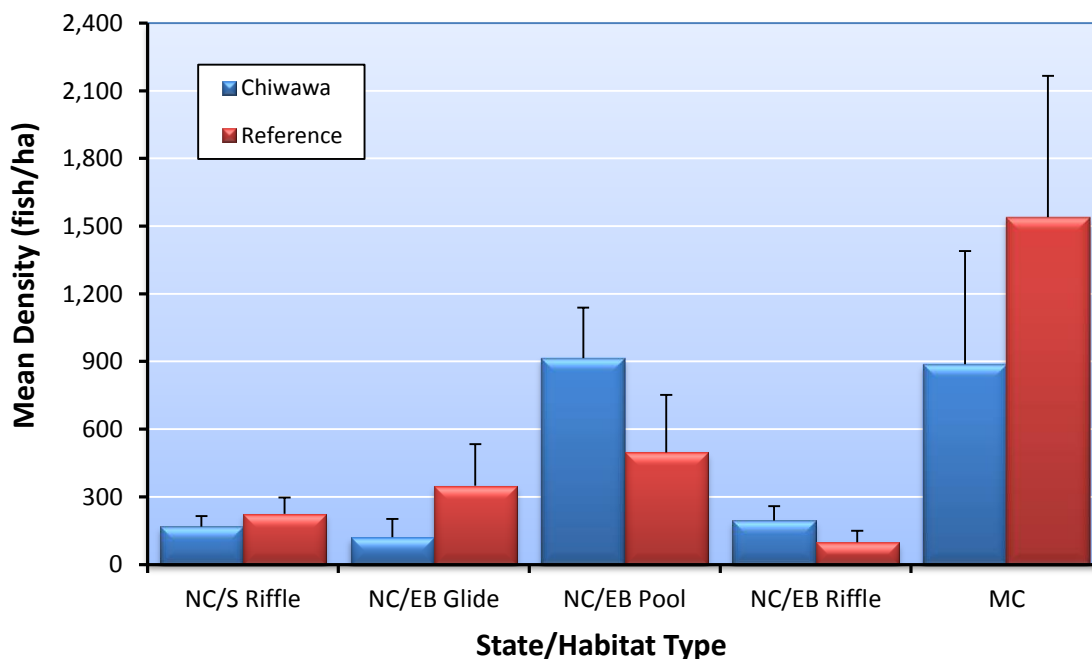
**Figure 5.1.** Numbers of subyearling and yearling Chinook salmon within the Chiwawa River Basin in August 1992-2014; ND = no data.

Juvenile Chinook were distributed contagiously among reaches in the Chiwawa River. Their densities were highest in the upper portions of the basin, with the highest densities within tributaries. Juvenile Chinook were most abundant in multiple channels and least abundant in glides and riffles. Most Chinook associated closely with woody debris in multiple channels. These sites (multiple channels) made up 17% of the total area of the Chiwawa River basin, but they provided habitat for 52% of all subyearling Chinook in the basin in 2014. In contrast, riffles



made up 54% of the total area, but provided habitat for only 11% of all juvenile Chinook in the Chiwawa River basin. Pools made up 22% of the total area and provided habitat for 37% of all juvenile Chinook in the basin. Virtually no Chinook used glides that lacked woody debris.

Mean densities of juvenile Chinook in two reaches of the Chiwawa River were generally less than those in corresponding reference areas (Figure 5.2). Within both the Chiwawa River and its reference areas, pools and multiple channels consistently had the highest densities of juvenile Chinook.



**Figure 5.2.** Comparison of the 21-year means of subyearling spring Chinook densities within state/habitat types in reaches 3 and 8 of the Chiwawa River and their matched reference areas on Nason Creek and the Little Wenatchee River. NC = natural channel; S = straight channel; EB = eroded banks; MC = multiple channel. There was no sampling in 2000 and no sampling within reference areas in 1992.

### Smolt and Emigrant Estimates

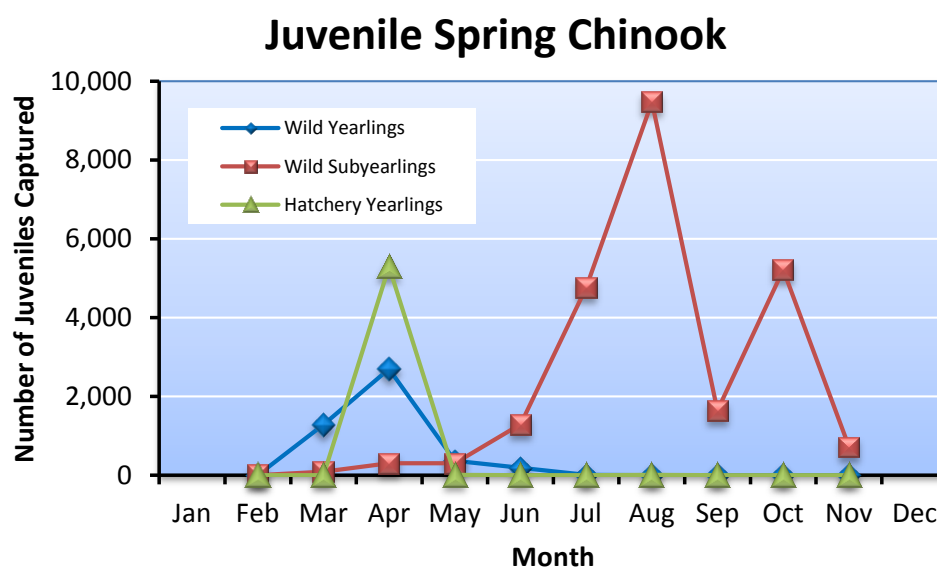
Numbers of spring Chinook smolts and emigrants were estimated at the Chiwawa and Lower Wenatchee traps in 2014.

#### Chiwawa Trap

The Chiwawa Trap operated between 18 March and 13 November 2014. During that time period the trap was inoperable for 21 days because of high river flows, debris, snow/ice, or mechanical failure. The trap operated in two different positions depending on stream flow; lower position at flows greater than 12 m<sup>3</sup>/s and an upper position at flows less than 12 m<sup>3</sup>/s. Daily trap efficiencies were estimated from two regression models depending on trap position and age class of fish (e.g., subyearling and yearling). The daily number of fish captured was expanded by the

estimated trap efficiency to estimate daily total emigration. Monthly captures of all fish and results of mark-recapture efficiency tests at the Chiwawa Trap are reported in Appendix B.

Wild yearling spring Chinook (2012 brood year) were primarily captured from March through June 2014 (Figure 5.3). Based on capture efficiencies estimated from the flow model, the total number of wild yearling Chinook emigrating from the Chiwawa River was 34,334 ( $\pm 6,488$ ). Combining the total number of subyearling spring Chinook (49,774) that emigrated during the fall of 2013 with the total number of yearling Chinook (34,334) that emigrated during 2014 and the number of estimated Chinook that were not trapped (25,305) resulted in a total emigrant estimate of 109,413 ( $\pm 11,723$ ) spring Chinook for the 2012 brood year (Table 5.15). The method for estimating emigration during the non-trapping period is explained in Appendix B.



**Figure 5.3.** Monthly captures of wild subyearling, wild yearling, and hatchery yearling spring Chinook at the Chiwawa Trap, 2014.

**Table 5.15.** Numbers of redds and juvenile spring Chinook at different life stages in the Chiwawa River basin for brood years 1991-2014; NS = not sampled.

Brood year	Number of redds	Egg deposition	Number of parr	Number of smolts produced within Chiwawa River basin <sup>a</sup>	Total number of smolts <sup>b</sup>	Number of emigrants
1991	104	478,400	45,483 <sup>c</sup>	42,525	42,525	NS
1992	302	1,570,098	79,113	39,723	56,763	65,541
1993	106	556,394	55,056	8,662	17,926	22,698
1994	82	485,686	55,240	16,472	22,145	25,067
1995	13	66,248	5,815	3,830	5,230	5,951
1996	23	106,835	16,066	15,475	17,922	19,183
1997	82	374,740	68,415	28,334	39,044	44,562

Brood year	Number of redds	Egg deposition	Number of parr	Number of smolts produced within Chiwawa River basin <sup>a</sup>	Total number of smolts <sup>b</sup>	Number of emigrants
1998	41	218,325	41,629	23,068	24,953	25,923
1999	34	166,090	NS	10,661	13,953	15,649
2000	128	642,944	114,617	40,831	50,634	55,685
2001	1,078	4,984,672	134,874	86,482	389,940	546,266
2002	345	1,605,630	91,278	90,948	152,547	184,279
2003	111	648,684	45,177	16,755	27,897	33,637
2004	241	1,156,559	49,631	72,080	101,172	116,158
2005	332	1,436,564	79,902	69,064	140,737	177,659
2006	297	1,284,228	60,752	45,050	86,579	107,972
2007	283	1,256,803	82,351	25,809	65,539	86,006
2008	689	3,163,888	106,705	35,023	91,229	120,184
2009	421	1,925,233	128,220	30,959	51,417	61,955
2010	502	2,165,628	141,510	47,511	82,911	101,130
2011	492	2,157,420	103,940	37,185	82,053	108,832
2012	808	3,412,184	149,563	34,334	92,490	109,413
2013	714	3,367,224	121,240	-	-	-
<b>Average</b>	<b>314</b>	<b>1,444,803</b>	<b>80,754</b>	<b>37,308</b>	<b>75,260</b>	<b>96,845</b>

<sup>a</sup> The estimated number of smolts (yearlings) that are produced entirely within the Chiwawa River basin. Smolt estimates for brood years 1992-1996 were calculated with a mark-recapture model; brood years 1997-present were calculated with a flow model.

<sup>b</sup> These numbers represent Chiwawa smolts produced within the entire Wenatchee River basin. This assumes that 66% of the subyearling migrants from the Chiwawa River basin survive to smolt in the Wenatchee River basin, regardless of the number of subyearling migrants (i.e., no density dependence). Smolt estimates for brood years 1992-1996 were calculated with a mark-recapture model; brood years 1997-present were calculated with a flow model.

<sup>c</sup> Estimate only includes numbers of Chinook in the Chiwawa River. Tributaries were not sampled at that time.

Wild subyearling spring Chinook (2013 brood year) were captured between March and November 2014. Based on capture efficiencies estimated from the flow model for both the upper trap position and lower position, the total number of wild subyearling (fry and parr) Chinook from the Chiwawa River basin was 114,049 ( $\pm 10,839$ ). Removing fry from the estimate, a total of 73,695 ( $\pm 8,464$ ) parr emigrated from the Chiwawa River basin in 2014. Although subyearling parr migrated during most months of sampling, the majority (82%) migrated during July, August, and October (Figure 5.3).

Yearling spring Chinook sampled in 2014 averaged 89 mm in length, 7.7 g in weight, and had a mean condition of 1.05 (Table 5.16). These size estimates were less than the overall mean of yearling spring Chinook sampled in previous years (overall means: 93 mm, 9.1 g, and condition of 1.08). Subyearling spring Chinook sampled in 2014 at the Chiwawa Trap averaged 71 mm in length, averaged 3.7 g, and had a mean condition of 1.08 (Table 5.16). These sizes were less than the overall mean of subyearling spring Chinook sampled in previous years (overall means, 76 mm, 5.3 g, and condition of 1.09).

**Table 5.16.** Mean fork length (mm), weight (g), and condition factor of subyearling and yearling spring Chinook collected in the Chiwawa Trap, 1996-2014. Numbers in parentheses indicate 1 standard deviation.

Sample year	Life stage	Sample size <sup>a</sup>	Mean size		
			Length (mm)	Weight (g)	Condition (K)
1996	Subyearling	514	78 (25)	6.9 (4.2)	1.11 (0.11)
	Yearling	1,589	94 (9)	9.5 (3.0)	1.11 (0.08)
1997	Subyearling	840	86 (8)	7.5 (2.1)	1.16 (0.08)
	Yearling	1,114	100 (7)	10.2 (2.6)	1.02 (0.10)
1998	Subyearling	3,743	82 (11)	6.2 (2.2)	1.08 (0.09)
	Yearling	2,663	97 (7)	10.3 (2.8)	1.12 (0.23)
1999	Subyearling	569	89 (9)	8.5 (2.4)	1.15 (0.07)
	Yearling	3,664	95 (8)	9.6 (3.4)	1.09 (0.19)
2000	Subyearling	1,810	85 (10)	7.4 (2.4)	1.15 (0.10)
	Yearling	1,891	97 (8)	10.5 (5.2)	1.13 (0.07)
2001	Subyearling	4,657	82 (11)	6.6 (3.4)	1.14 (0.09)
	Yearling	2,935	97 (7)	10.5 (2.4)	1.15 (0.08)
2002	Subyearling	6,130	64 (12)	3.0 (1.6)	1.06 (0.10)
	Yearling	1,735	94 (8)	9.0 (2.3)	1.09 (0.08)
2003	Subyearling	3,679	64 (12)	3.2 (1.7)	1.08 (0.10)
	Yearling	2,657	87 (9)	7.2 (3.5)	1.07 (0.10)
2004	Subyearling	2,278	75 (16)	4.3 (2.1)	0.92 (0.16)
	Yearling	1,032	91 (9)	8.5 (2.7)	1.09 (0.10)
2005	Subyearling	2,702	73 (12)	4.6 (2.2)	1.08 (0.09)
	Yearling	803	96 (9)	9.9 (2.8)	1.08 (0.08)
2006	Subyearling	3,462	76 (11)	5.1 (2.0)	1.12 (0.21)
	Yearling	4,645	95 (7)	9.4 (2.3)	1.10 (0.13)
2007	Subyearling	1,718	72 (12)	4.5 (2.1)	1.13 (0.16)
	Yearling	2,245	91 (8)	8.6 (2.5)	1.10 (0.09)
2008	Subyearling	10,443	79 (12)	5.9 (2.3)	1.15 (0.15)
	Yearling	8,792	93 (7)	8.8 (2.1)	1.08 (0.10)
2009	Subyearling	10,536	75 (10)	5.0 (2.2)	0.91 (0.11)
	Yearling	3,630	92 (7)	8.8 (2.1)	0.89 (0.07)
2010	Subyearling	3,888	77 (12)	5.4 (2.3)	1.11 (0.16)
	Yearling	5,799	91 (8)	8.9 (2.2)	1.15 (0.14)
2011	Subyearling	6,870	73 (11)	4.8 (2.2)	1.15 (0.16)
	Yearling	4,734	94 (8)	8.7 (2.2)	1.04 (0.10)
2012	Subyearling	8,756	75 (10)	4.8 (2.2)	1.13 (0.28)
	Yearling	7,290	90 (7)	8.0 (2.6)	1.06 (0.24)
2013	Subyearling	10,181	71 (10)	4.1 (1.7)	1.09 (0.39)

Sample year	Life stage	Sample size <sup>a</sup>	Mean size		
			Length (mm)	Weight (g)	Condition (K)
	Yearling	3,135	88 (9)	7.7 (2.8)	1.09 (0.20)
2014	Subyearling	7,122	71 (10)	3.7 (1.6)	1.08 (0.10)
	Yearling	3,956	89 (8)	7.7 (2.2)	1.05 (0.08)
Average	Subyearling	4,731	76 (7)	5.3 (1.5)	1.09 (0.07)
	Yearling	3,385	93 (3)	9.0 (1.0)	1.08 (0.06)

<sup>a</sup> Sample size represents the number of fish that were measured for both length and weight.

### Lower Wenatchee Trap

The lower Wenatchee Trap operated in a new location beginning in 2013. Hence, historic flow-discharge relationships are invalid and new models to estimate trap efficiency must be developed for all species. Until new models are developed (2-3 years) all estimates of juvenile abundance should be considered preliminary.

The Lower Wenatchee Trap operated between 12 February and 7 September 2014. During that time period the trap was inoperable for 12 days because of high river flows, debris, snow/ice, or major hatchery releases. During the seven-month sampling period, a total of 1,700 wild yearling Chinook, 81,455 wild subyearling Chinook (mostly summer Chinook), and 31,290 hatchery yearling Chinook were captured at the Lower Wenatchee Trap. Based on capture efficiencies estimated from the pooled model, the total number of wild yearling Chinook that emigrated past the Lower Wenatchee Trap was 67,973 ( $\pm 431,135$ ). Monthly captures of all fish collected at the Lower Wenatchee Trap are reported in Appendix B.

### PIT Tagging Activities

As part of the Comparative Survival Study (CSS), a total of 17,002 wild juvenile Chinook (12,103 subyearling and 4,899 yearlings) were PIT tagged and released in 2013 in the Wenatchee River basin (Table 5.17a). Most of these (71.6%) were tagged at the Chiwawa trap. See Appendix C for a complete list of all fish captured, tagged, lost, and released.

**Table 5.17a.** Numbers of wild Chinook that were captured, tagged, and released at different locations within the Wenatchee River basin, 2014. Numbers of fish that died or shed tags are also given.

Sampling Location	Species and Life Stage	Number captured	Number of recaptures	Number tagged	Number died	Shed Tags	Total released	Percent mortality
Chiwawa Trap	Wild Subyearling Chinook	11,803	3,782	11,375	14	3	11,358	0.12
	Wild Yearling Chinook	4,476	75	4,399	16	0	4,383	0.36
	<b>Total</b>	<b>16,279</b>	<b>3,857</b>	<b>15,774</b>	<b>30</b>	<b>3</b>	<b>15,741</b>	<b>0.48</b>
Chiwawa River (Electrofishing)	Wild Subyearling Chinook	1,082	2	1,034	0	2	1,032	0.00
	Wild Yearling Chinook	0	0	0	0	0	0	--
	<b>Total</b>	<b>1,082</b>	<b>2</b>	<b>1,034</b>	<b>0</b>	<b>2</b>	<b>1,032</b>	<b>0.00</b>
Nason Creek (Electrofishing)	Wild Subyearling Chinook	1,908	28	1,821	4	1	1,816	0.21
	Wild Yearling Chinook	0	0	0	0	0	0	--
	<b>Total</b>	<b>1,908</b>	<b>28</b>	<b>1,821</b>	<b>4</b>	<b>1</b>	<b>1,816</b>	<b>0.21</b>
Lower Wenatchee Trap	Wild Subyearling Chinook	36	0	36	0	0	36	0.00

Sampling Location	Species and Life Stage	Number captured	Number of recaptures	Number tagged	Number died	Shed Tags	Total released	Percent mortality
	Wild Yearling Chinook	1,631	107	1,521	15	0	1,506	0.92
	<b>Total</b>	<b>1,667</b>	<b>107</b>	<b>1,557</b>	<b>15</b>	<b>0</b>	<b>1,542</b>	<b>0.92</b>
<b>Total:</b>	<b>Wild Subyearling Chinook</b>	<b>14,829</b>	<b>3,812</b>	<b>14,266</b>	<b>18</b>	<b>6</b>	<b>14,242</b>	<b>0.12</b>
	<b>Wild Yearling Chinook</b>	<b>6,107</b>	<b>182</b>	<b>5,920</b>	<b>31</b>	<b>0</b>	<b>5,889</b>	<b>0.51</b>
<b>Grand Total:</b>		<b>20,936</b>	<b>3,994</b>	<b>20,186</b>	<b>49</b>	<b>6</b>	<b>20,131</b>	<b>0.63</b>

Numbers of wild Chinook salmon PIT-tagged and released as part of CSS during the period 2006-2014 are shown in Table 5.17b.

**Table 5.17b.** Summary of the numbers of wild Chinook that were tagged and released at different locations within the Wenatchee River basin, 2006-2014.

Sampling Location	Species and Life Stage	Numbers of PIT-tagged Chinook salmon released								
		2006	2007	2008	2009	2010	2011	2012	2013	2014
Chiwawa Trap	Wild Subyearling Chinook	5,130	6,137	8,755	8,765	3,324	6,030	7,644	9,086	11,358
	Wild Yearling Chinook	2,793	4,659	8,397	3,694	6,281	4,318	7,980	3,093	4,383
	<b>Total</b>	<b>7,923</b>	<b>10,796</b>	<b>17,152</b>	<b>12,459</b>	<b>9,605</b>	<b>10,348</b>	<b>15,624</b>	<b>12,179</b>	<b>15,741</b>
Chiwawa River (Angling or Electrofishing)	Wild Subyearling Chinook	111	20	43	128	531	0	3,181	3,017	1,032
	Wild Yearling Chinook	0	0	0	3	4	0	0	0	0
	<b>Total</b>	<b>111</b>	<b>20</b>	<b>43</b>	<b>131</b>	<b>535</b>	<b>0</b>	<b>3,181</b>	<b>3,017</b>	<b>1,032</b>
Upper Wenatchee Trap	Wild Subyearling Chinook	0	15	0	37	3	1	1	0	0
	Wild Yearling Chinook	81	1,434	159	296	486	714	75	94	0
	<b>Total</b>	<b>81</b>	<b>1,449</b>	<b>159</b>	<b>333</b>	<b>489</b>	<b>715</b>	<b>76</b>	<b>94</b>	<b>0</b>
Nason Creek (Angling or Electrofishing)	Wild Subyearling Chinook	68	6	4	701	595	0	0	0	1,816
	Wild Yearling Chinook	1	7	0	13	3	0	0	0	0
	<b>Total</b>	<b>69</b>	<b>13</b>	<b>4</b>	<b>714</b>	<b>598</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>1,816</b>
Upper Wenatchee (Angling or Electrofishing)	Wild Subyearling Chinook	0	61	1	0	2	0	0	0	0
	Wild Yearling Chinook	27	0	0	0	0	0	0	0	0
	<b>Total</b>	<b>27</b>	<b>61</b>	<b>1</b>	<b>0</b>	<b>2</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Middle Wenatchee (Angling or Electrofishing)	Wild Subyearling Chinook	0	0	65	284	233	0	0	0	0
	Wild Yearling Chinook	0	0	0	0	0	0	0	0	0
	<b>Total</b>	<b>0</b>	<b>0</b>	<b>65</b>	<b>284</b>	<b>233</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Lower Wenatchee (Angling or Electrofishing)	Wild Subyearling Chinook	0	0	0	0	0	0	0	0	0
	Wild Yearling Chinook	0	0	0	0	0	0	0	0	0
	<b>Total</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Peshastin Creek (Angling or Electrofishing)	Wild Subyearling Chinook	0	0	0	0	1	0	0	0	0
	Wild Yearling Chinook	0	0	0	0	0	0	0	0	0
	<b>Total</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>1</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>
Lower Wenatchee	Wild Subyearling Chinook	0	0	2	0	0	0	0	0	36

Sampling Location	Species and Life Stage	Numbers of PIT-tagged Chinook salmon released								
		2006	2007	2008	2009	2010	2011	2012	2013	2014
Trap	Wild Yearling Chinook	522	1,641	506	468	917	0	0	1,712	1,506
	<b>Total</b>	<b>522</b>	<b>1,641</b>	<b>508</b>	<b>468</b>	<b>917</b>	<b>0</b>	<b>0</b>	<b>1,712</b>	<b>1,542</b>
<b>Total:</b>	<b>Wild Subyearling Chinook</b>	<b>5,309</b>	<b>6,239</b>	<b>8,870</b>	<b>9,915</b>	<b>4,689</b>	<b>6,031</b>	<b>10,826</b>	<b>12,103</b>	<b>14,242</b>
	<b>Wild Yearling Chinook</b>	<b>3,424</b>	<b>7,741</b>	<b>9,062</b>	<b>4,474</b>	<b>7,691</b>	<b>5,032</b>	<b>8,055</b>	<b>4,899</b>	<b>5,889</b>
<b>Grand Total:</b>		<b>8,733</b>	<b>13,980</b>	<b>17,932</b>	<b>14,389</b>	<b>12,380</b>	<b>11,063</b>	<b>18,881</b>	<b>17,002</b>	<b>20,131</b>

### Freshwater Productivity

Both productivity and survival estimates for different life stages of spring Chinook in the Chiwawa River basin are provided in Table 5.18. Estimates for brood year 2012 generally fall within the ranges estimated over the period of brood years 1991-2011. During that period, freshwater productivities ranged from 125-1,015 parr/redd, 114-779 smolts/redd, and 135-834 emigrants/redd. Survivals during the same period ranged from 2.7-19.1% for egg-parr, 2.7-16.8% for egg-smolt, and 3.2-18.0% for egg-emigrants. Overwinter survival rates for juvenile spring Chinook within the Chiwawa River basin have ranged from 15.7-100.0%.

**Table 5.18.** Productivity (fish/redd) and survival (%) estimates for different juvenile life stages of spring Chinook in the Chiwawa River basin for brood years 1991-2013; ND = no data. These estimates were derived from data in Table 5.15.

Brood year	Parr/Redd	Smolts/Redd <sup>a</sup>	Emigrants/Redd	Egg-Parr (%)	Parr-Smolt <sup>b</sup> (%)	Egg-Smolt <sup>a</sup> (%)	Egg-Emigrant (%)
1991	437	409	ND	9.5	93.5	8.9	ND
1992	262	188	217	5.0	50.2	3.6	4.2
1993	519	169	214	9.9	15.7	3.2	4.1
1994	674	270	306	11.4	29.8	4.6	5.2
1995	447	402	458	8.8	65.9	7.9	9.0
1996	699	779	834	15.0	96.3	16.8	18.0
1997	834	476	543	18.3	41.4	10.4	11.9
1998	1,015	609	632	19.1	55.4	11.4	11.9
1999	ND	410	460	ND	ND	8.4	9.4
2000	895	396	435	17.8	35.6	7.9	8.7
2001	125	362	507	2.7	64.1	7.8	11.0
2002	265	442	534	5.7	99.6	9.5	11.5
2003	407	251	303	7.0	37.1	4.3	5.2
2004	206	420	482	4.3	100.0	8.7	10.0
2005	241	424	535	5.6	86.4	9.8	12.4
2006	205	292	364	4.7	74.2	6.7	8.4
2007	291	232	304	6.6	31.3	5.2	6.8
2008	155	132	174	3.4	32.8	2.9	3.8
2009	305	122	147	6.7	24.1	2.7	3.2

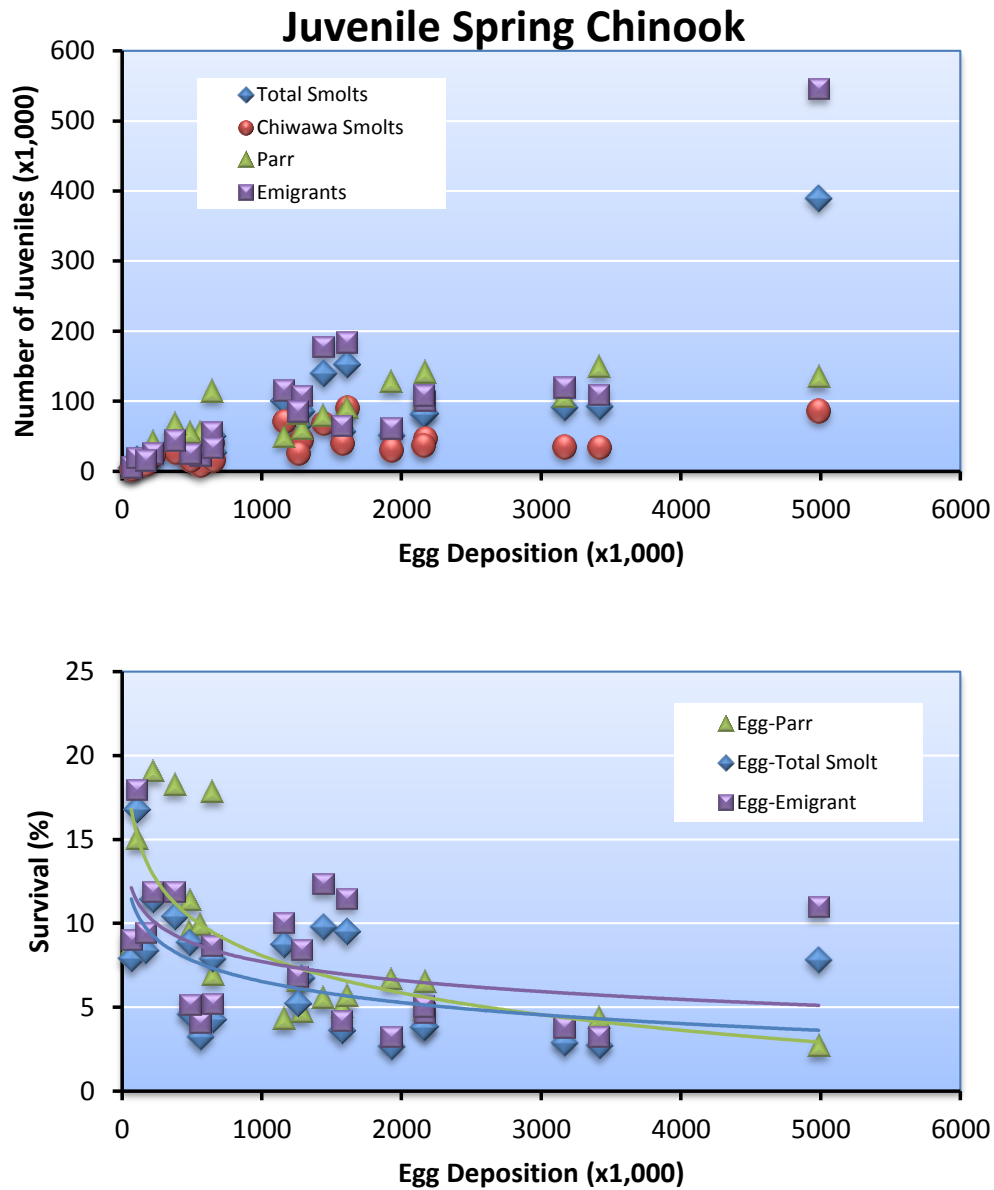
Brood year	Parr/Redd	Smolts/Redd <sup>a</sup>	Emigrants/ Redd	Egg-Parr (%)	Parr-Smolt <sup>b</sup> (%)	Egg-Smolt <sup>a</sup> (%)	Egg- Emigrant (%)
2010	282	165	201	6.5	33.6	3.8	4.7
2011	211	172	221	4.8	35.8	3.9	5.0
2012	185	114	135	4.4	23.0	2.7	3.2
2013	170	-	-	3.6	-	-	-
<b>Average</b>	<b>401</b>	<b>329</b>	<b>381</b>	<b>8.2</b>	<b>53.6</b>	<b>6.9</b>	<b>8.0</b>

<sup>a</sup> These estimates include Chiwawa smolts produced within the Wenatchee River basin. This assumes that 66% of the subyearling migrants survive to smolt, regardless of the number of subyearling migrants (i.e., no density dependence). The assumed 66% survival estimate is being revised; however, an additional year of data is needed to generate a more precise estimate. Smolt estimates for brood years 1992-1996 were calculated with a mark-recapture model; brood years 1997-present were calculated with a flow model.

<sup>b</sup> These estimates represent overwinter survival within the Chiwawa River basin. It does not include Chiwawa smolts produced outside the Chiwawa River basin. As noted in footnote *a*, smolts/redd and egg-smolt survival include Chiwawa smolts produced in the Wenatchee River basin.

Seeding level (egg deposition) explained most of the variability in productivity and survival of juvenile spring Chinook in the Chiwawa River basin. That is, for estimates based on “within-Chiwawa-Basin” life stages (e.g., parr and smolts), survival and productivity decreased as seeding levels increased (Figure 5.4). This suggests that density dependence regulates juvenile productivity and survival within the Chiwawa River basin. This form of population regulation is less apparent with total smolts (i.e., Chiwawa smolts produced within the Wenatchee River basin) and total emigrants. However, one would expect the number of emigrants to increase as seeding levels exceed the rearing capacity of the Chiwawa River basin.





**Figure 5.4.** Relationships between seeding levels (egg deposition) and juvenile life-stage survivals and productivities for Chiwawa spring Chinook, brood years 1991-2012. Total smolts are Chiwawa smolts produced within and outside the Chiwawa River basin (assumes a 66% survival on subyearling emigrants; the survival estimate will be modified after next year). Chiwawa smolts are smolts produced only in the Chiwawa River basin.

### 5.5 Spawning Surveys

Surveys for spring Chinook redds were conducted during August through September, 2014, in the Chiwawa River (including Rock, Phelps, and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek, Upper Wenatchee River, Little Wenatchee River, and the White River (including the Napeequa River and Panther Creek). Ingalls Creek (tributary to Peshastin Creek) and Chiwaukum Creek (tributary to the upper Wenatchee) were not surveyed in 2014 because wildfires prevented access.

Spawning escapement for spring Chinook was calculated as the number of redds times the male-to-female ratio (i.e., fish per redd expansion factor) estimated from broodstock and fish sampled at adult trapping sites. In the future, spawning escapement will be estimated using the area-under-the-curve (AUC) method (Millar et al. 2012). WDFW is currently developing an observer efficiency model that will be used to estimate variance.

### Redd Counts

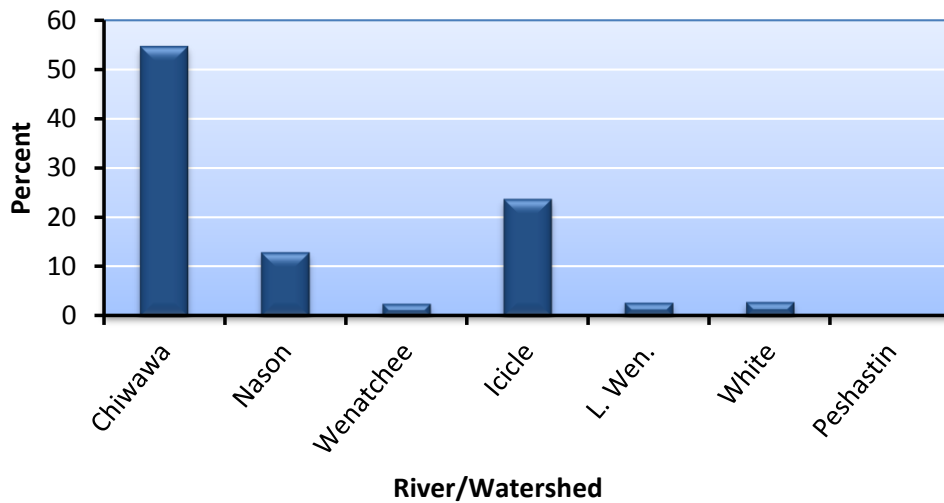
A total of 885 spring Chinook redds were counted in the Wenatchee River basin in 2014 (Table 5.19). This is higher than the average of 665 redds counted during the period 1989-2014 in the Wenatchee River basin. Most spawning occurred in the Chiwawa River (54.8% or 485 redds) (Table 5.19; Figure 5.5). Nason Creek contained 13.0% (115 redds), Icicle Creek contained 23.8% (211 redds), White River contained 2.9% (26 redds), Little Wenatchee contained 2.8% (25 redds), the Upper Wenatchee River 2.6% (23 redds), and Peshastin Creek contained 0.0% (0 redds).

**Table 5.19.** Numbers of spring Chinook redds counted within different streams/watersheds within the Wenatchee River basin, 1989-2014. Redd counts in Peshastin Creek in 2001 and 2002 (\*) were elevated because the U.S. Fish and Wildlife Service planted 487 and 350 spring Chinook adults, respectively, into the stream. These counts were not included in the total or average calculations. WDFW began full implementation of adult management in 2014.

Sample year	Number of spring Chinook redds							Total
	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River	Icicle	Peshastin	
1989	314	98	45	64	94	24	NS	<b>639</b>
1990	255	103	30	22	36	50	4	<b>500</b>
1991	104	67	18	21	41	40	1	<b>292</b>
1992	302	81	35	35	38	37	0	<b>528</b>
1993	106	223	61	66	86	53	5	<b>600</b>
1994	82	27	7	3	6	15	0	<b>140</b>
1995	13	7	0	2	1	9	0	<b>32</b>
1996	23	33	3	12	1	12	1	<b>85</b>
1997	82	55	8	15	15	33	1	<b>209</b>
1998	41	29	8	5	0	11	0	<b>94</b>
1999	34	8	3	1	2	6	0	<b>54</b>
2000	128	100	9	8	37	68	0	<b>350</b>
2001	1,078	374	74	104	218	88	173*	<b>2,109</b>
2002	345	294	42	42	64	245	107*	<b>1,139</b>
2003	111	83	12	15	24	18	60	<b>323</b>
2004	239	169	13	22	46	30	55	<b>574</b>
2005	333	193	64	86	143	8	3	<b>830</b>
2006	297	152	21	31	27	50	10	<b>588</b>
2007	283	101	22	20	12	17	11	<b>466</b>
2008	689	336	38	31	180	116	21	<b>1,411</b>
2009	421	167	39	54	5	32	15	<b>733</b>

Sample year	Number of spring Chinook redds							Total
	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River	Icicle	Peshastin	
2010	502	188	38	33	47	155	5	968
2011	492	170	30	20	12	122	26	872
2012	880	413	43	86	73	199	10	1,704
2013	714	212	51	54	17	107	4	1,159
2014	485	115	25	26	23	211	0	885
<i>Average</i>	<i>321</i>	<i>146</i>	<i>28</i>	<i>34</i>	<i>48</i>	<i>68</i>	<i>10</i>	<i>665</i>

### Spring Chinook Redds



**Figure 5.5.** Percent of the total number of spring Chinook redds counted in different streams/watersheds within the Wenatchee River basin during August through September, 2014.

#### Redd Distribution

Spring Chinook redds were not evenly distributed among reaches within survey streams in 2014 (Table 5.20). Most of the spawning in the Chiwawa River basin occurred in Reaches 1 through 6. About 60% of the spawning in the Chiwawa River basin occurred in the lower two reaches (RM 0.0-19.3; from the mouth to Rock Creek). Relatively few fish spawned in Rock and Chikamin creeks. The spatial distribution of redds in Nason Creek was weighted towards Reach 1, having 37% of the Nason Creek redds. In the Little Wenatchee River, 100% of all spawning occurred in Reach 3 (RM 5.2-9.2; Lost Creek to Rainy Creek). On the White River, 85% of the spawning occurred in Reach 3 (RM 11.0-12.9; Napeequa River to Grasshopper Meadows). About 75.7% of all the spawning in the Wenatchee River occurred upstream from the mouth of the Chiwawa River.

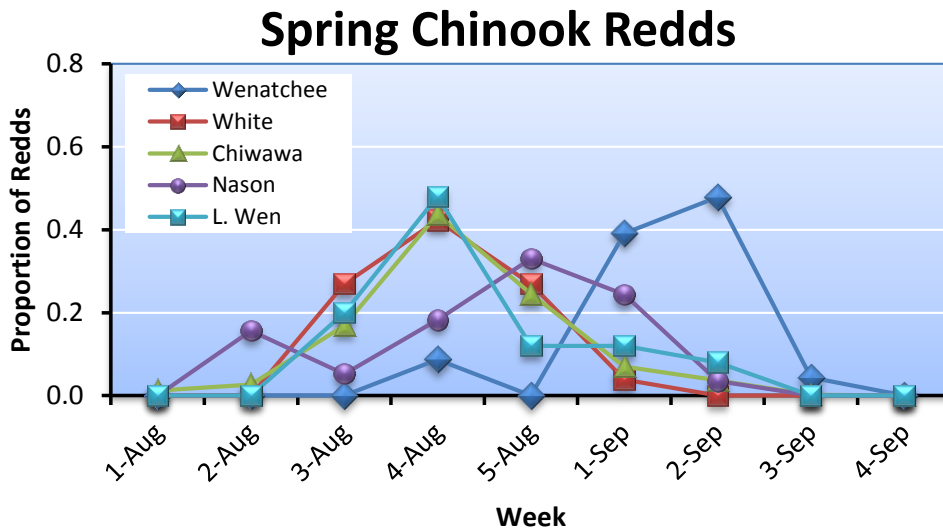
**Table 5.20.** Numbers and proportions of spring Chinook redds counted within different streams/watersheds within the Wenatchee River basin during August through September, 2014. NS = not surveyed. See Table 2.8 for description of survey reaches.

Stream/watershed	Reach	Number of redds	Proportion of redds within stream/watershed
Chiwawa	Chiwawa 1 (C1)	116	0.24
	Chiwawa 2 (C2)	175	0.36
	Chiwawa 3 (C3)	20	0.04
	Chiwawa 4 (C4)	51	0.11
	Chiwawa 5 (C5)	43	0.09
	Chiwawa 6 (C6)	62	0.13
	Chiwawa 7 (C7)	7	0.01
	Phelps 1 (S1)	0	0.00
	Rock 1 (R1)	5	0.01
	Chikamin 1 (K1)	6	0.01
	<b>Total</b>	<b>485</b>	<b>1.00</b>
Nason	Nason 1 (N1)	42	0.37
	Nason 2 (N2)	11	0.10
	Nason 3 (N3)	36	0.31
	Nason 4 (N4)	26	0.23
		<b>Total</b>	<b>115</b>
Little Wenatchee	Little Wen 2 (L2)	0	0.00
	Little Wen 3 (L3)	25	1.00
		<b>Total</b>	<b>25</b>
White	White 2 (H2)	1	0.04
	White 3 (H3)	22	0.85
	White 4 (H4)	0	0.00
	Napeequa 1 (Q1)	2	0.08
	Panther 1 (T1)	1	0.04
		<b>Total</b>	<b>26</b>
Wenatchee River	Wen 9 (W9)	4	0.17
	Wen 10 (W10)	19	0.83
	Chiwaukum (U1)	NS	--
		<b>Total</b>	<b>23</b>
Icicle	Icicle 1 (I1)	59	0.30
	Icicle 2 (I2)	127	0.60
	Icicle 3 (I3)	25	0.10
		<b>Total</b>	<b>211</b>
Peshastin	Peshastin 1 (P1)	0	0.00
	Peshastin 2 (P2)	0	0.00
	Ingalls (D1)	NS	--

Stream/watershed	Reach	Number of redds	Proportion of redds within stream/watershed
	Total	0	1.00
<i>Grand Total</i>		885	1.00

### Spawn Timing

Spring Chinook began spawning during the first week of August in the Chiwawa River, the second week of August in Nason Creek, and the end of August in the White River, Little Wenatchee River, and the Wenatchee River (Figure 5.6). Spawning peaked the fourth week of August in the Chiwawa River, White River and the Little Wenatchee, and the fifth week of August in Nason Creek. Spawning in the Wenatchee River peaked the second week of September. All spawning was completed by the end of September.



**Figure 5.6.** Proportion of spring Chinook redds counted during different weeks in different sampling streams within the Wenatchee River basin, August through September 2014.

### Spawning Escapement

Spawning escapement for spring Chinook was calculated as the number of redds times the male-to-female ratio (i.e., fish per redd expansion factor) estimated from broodstock and fish sampled at adult trapping sites. The estimated fish per redd ratio for spring Chinook upstream from Tumwater in 2014 was 2.06 (based on sex ratios estimated at Tumwater Dam). The estimated fish per redd ratio for spring Chinook downstream from Tumwater (Icicle and Peshastin creeks) was 2.01 (derived from broodstock collected at the Leavenworth National Fish Hatchery). Multiplying these ratios by the number of redds counted in the Wenatchee River basin resulted in a total spawning escapement of 1,813 spring Chinook (Table 5.21). The Chiwawa River basin had the highest spawning escapement (999 Chinook), while Peshastin Creek had the lowest.

**Table 5.21.** Number of redds, fish per redd ratios, and total spawning escapement for spring Chinook in the Wenatchee River basin, 2014. Spawning escapement was estimated as the product of redds times fish per redd.

Sampling area	Total number of redds	Fish/redd	Total spawning escapement*
Chiwawa	485	2.06	999
Nason	115	2.06	237
Upper Wenatchee River	23	2.06	47
Icicle	211	2.01	424
Little Wenatchee	25	2.06	52
White	26	2.06	54
Peshastin	0	2.01	0
<b>Total</b>	<b>885</b>	<b>--</b>	<b>1,813</b>

\* Spawning escapement estimate is based on total number of redds by stream. If escapement is calculated at the reach scale, then the total escapement may vary from what is shown here because of rounding errors.

The estimated total spawning escapement of 1,813 spring Chinook in 2014 was greater than the overall average of 1,469 spring Chinook (Table 5.22). The escapement in the Chiwawa River basin in 2014 was 2.4 times the escapement in Icicle Creek, the second most abundant escapement in the Wenatchee River basin (Table 5.22).

**Table 5.22.** Spawning escapements for spring Chinook in the Wenatchee River basin for return years 1989-2014; NA = not available.

Return year	Upper basin spawning escapement						Lower basin spawning escapement			Total
	Fish/redd	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River	Fish/redd	Icicle	Peshastin	
1989	2.27	713	222	102	145	213	2.27	54	NA	<b>1,449</b>
1990	2.24	571	231	67	49	81	2.24	112	9	<b>1,120</b>
1991	2.33	242	156	42	49	96	2.33	93	2	<b>680</b>
1992	2.24	676	181	78	78	85	2.24	83	0	<b>1,181</b>
1993	2.20	233	491	134	145	189	2.20	117	11	<b>1,320</b>
1994	2.24	184	60	16	7	13	2.24	34	0	<b>314</b>
1995	2.51	33	18	0	5	3	2.51	23	0	<b>82</b>
1996	2.53	58	83	8	30	3	2.53	30	3	<b>215</b>
1997	2.22	182	122	18	33	33	2.22	73	2	<b>463</b>
1998	2.21	91	64	18	11	0	2.21	24	0	<b>208</b>
1999	2.77	94	22	8	3	6	2.77	17	0	<b>150</b>
2000	2.70	346	270	24	22	100	2.70	184	0	<b>946</b>
2001	1.60	1,725	598	118	166	349	1.60	141	277	<b>3,374</b>
2002	2.05	707	603	86	86	131	2.05	502	219	<b>2,334</b>
2003	2.43	270	202	29	36	58	2.43	44	146	<b>785</b>
2004 <sup>a</sup>	3.56/3.00	851	507	39	66	138	1.79	54	98	<b>1,753</b>
2005	1.80	599	347	115	155	257	1.75	14	5	<b>1,492</b>
2006	1.78	529	271	37	55	48	1.80	90	18	<b>1,048</b>
2007	4.58	1,296	463	101	92	55	1.86	32	20	<b>2,059</b>
2008	1.68	1,158	565	64	52	302	1.77	205	37	<b>2,383</b>

Return year	Upper basin spawning escapement						Lower basin spawning escapement			Total
	Fish/redd	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River	Fish/redd	Icicle	Peshastin	
2009	3.20	1,347	534	125	173	16	2.72	87	41	2,323
2010	2.18	1,094	410	83	72	102	2.72	422	14	2,197
2011	4.13	2,032	702	124	83	50	2.66	325	69	3,385
2012	1.68	1,478	694	72	144	123	1.90	378	19	2,908
2013	1.93	1,378	409	98	104	33	1.75	187	7	2,216
2014	2.06	999	237	52	54	47	2.01	424	0	1,813
<i>Average</i>	--	726	325	64	74	97	--	144	40	1,469

<sup>a</sup> In 2004 the fish/redd expansion estimate of 3.56 was applied to the Chiwawa River only and 3.00 fish/redd for the rest of the upper basin.

### 5.6 Carcass Surveys

Surveys for spring Chinook carcasses were conducted during August through September, 2014, in the Chiwawa River (including Rock, Phelps, and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek, Upper Wenatchee River, Little Wenatchee River, and White River (including the Napeequa River and Panther Creek). Ingalls Creek (tributary to Peshastin Creek) and Chiwaukum Creek (tributary to the upper Wenatchee) were not surveyed in 2014 because wildfires prevented access.

#### Number sampled

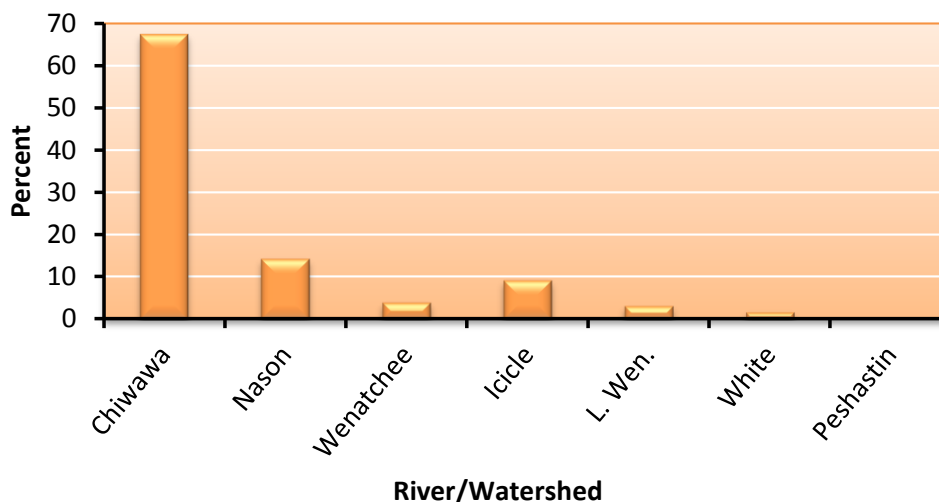
A total of 474 spring Chinook carcasses were sampled during August through September in the Wenatchee River basin (Table 5.23). Most were sampled in the Chiwawa River basin (68% or 320 carcasses) and Nason Creek (14% or 68 carcasses) (Figure 5.7). A total of 44 carcasses were sampled in Icicle Creek, 19 in the upper Wenatchee River, 15 in the Little Wenatchee, 8 in the White River, and none in Peshastin Creek.

**Table 5.23.** Numbers of spring Chinook carcasses sampled within different streams/watersheds within the Wenatchee River basin, 1996-2014.

Survey year	Number of spring Chinook carcasses							
	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River	Icicle	Peshastin	Total
1996	22	3	0	2	0	1	0	28
1997	17	42	3	8	1	28	1	100
1998	24	25	3	2	1	6	0	61
1999	15	5	0	0	2	1	0	23
2000	122	110	8	1	37	52	0	330
2001	763	388	68	81	213	163	63	1,739
2002	210	292	30	25	34	91	65	747
2003	70	100	8	8	11	37	64	298
2004	178	186	1	13	29	16	40	463
2005	391	217	48	52	120	2	0	830
2006	241	190	13	25	15	7	0	491

Survey year	Number of spring Chinook carcasses							Total
	Chiwawa	Nason	Little Wenatchee	White	Wenatchee River	Icicle	Peshastin	
2007	250	201	16	13	24	15	6	525
2008	386	243	15	13	94	67	5	823
2009	240	128	20	20	1	67	2	478
2010	192	141	7	11	29	39	2	421
2011	177	98	7	4	3	40	3	332
2012	390	332	24	21	23	61	3	854
2013	396	142	20	22	8	28	1	671
2014	320	68	15	8	19	44	0	474
<b>Average</b>	<b>232</b>	<b>153</b>	<b>16</b>	<b>17</b>	<b>35</b>	<b>40</b>	<b>13</b>	<b>510</b>

### Spring Chinook Carcasses



**Figure 5.7.** Percent of the total number of spring Chinook carcasses sampled in different streams/watersheds within the Wenatchee River basin during August through September, 2014.

#### Carcass Distribution and Origin

Spring Chinook carcasses were not evenly distributed among reaches within survey streams in 2014 (Table 5.24). Most of the carcasses in the Chiwawa River basin occurred in Reaches 1 and 2 (downstream from Rock Creek). In Nason Creek, most carcasses (65%) were collected in Reach 1 and the fewest (3%) in Reach 4. Most of the carcasses in the Little Wenatchee River were sampled in Reach 3 (Lost Creek to Rainy Creek). On the White River, all occurred in Reach 3 (Napeequa River to Grasshopper Meadows). On the Wenatchee River, 79% of the carcasses were found upstream from the confluence of the Chiwawa River and 21% were found below the confluence.



**Table 5.24.** Numbers and proportions of carcasses sampled within different streams/watersheds within the Wenatchee River basin during August through September, 2014. See Table 2.8 for description of survey reaches.

Stream/watershed	Reach	Number of carcasses	Proportion of redds within stream/watershed
Chiwawa	Chiwawa 1 (C1)	82	0.26
	Chiwawa 2 (C2)	128	0.40
	Chiwawa 3 (C3)	12	0.04
	Chiwawa 4 (C4)	38	0.12
	Chiwawa 5 (C5)	25	0.08
	Chiwawa 6 (C6)	30	0.09
	Chiwawa 7 (C7)	1	0.00
	Phelps 1 (S1)	0	0.00
	Rock 1 (R1)	2	0.01
	Chikamin 1 (K1)	2	0.01
	<b>Total</b>	<b>320</b>	<b>1.00</b>
Nason	Nason 1 (N1)	44	0.65
	Nason 2 (N2)	6	0.09
	Nason 3 (N3)	16	0.23
	Nason 4 (N4)	2	0.03
	<b>Total</b>	<b>68</b>	<b>1.00</b>
Little Wenatchee	Little Wen 2 (L2)	3	0.20
	Little Wen 3 (L3)	12	0.80
	<b>Total</b>	<b>15</b>	<b>1.00</b>
White	White 2 (H2)	0	0.00
	White 3 (H3)	8	1.00
	White 4 (H4)	0	0.00
	Napeequa 1 (Q1)	0	0.00
	Panther 1 (T1)	0	0.00
	<b>Total</b>	<b>8</b>	<b>1.00</b>
Wenatchee River	Wen 9 (W9)	4	0.21
	Wen 10 (W10)	15	0.79
	Chiwaukum 1	NS	--
	<b>Total</b>	<b>19</b>	<b>1.00</b>
Icicle	Icicle 1 (I1)	43	0.98
	Icicle 2 (I2)	0	0.00
	Icicle 3 (I3)	1	0.02
	<b>Total</b>	<b>44</b>	<b>1.00</b>
Peshastin	Peshastin 1 (P1)	0	0
	Peshastin 2 (P2)	0	0
	Ingalls (D1)	NS	--

Stream/watershed	Reach	Number of carcasses	Proportion of redds within stream/watershed
<b>Grand Total</b>		<b>474</b>	<b>1.00</b>

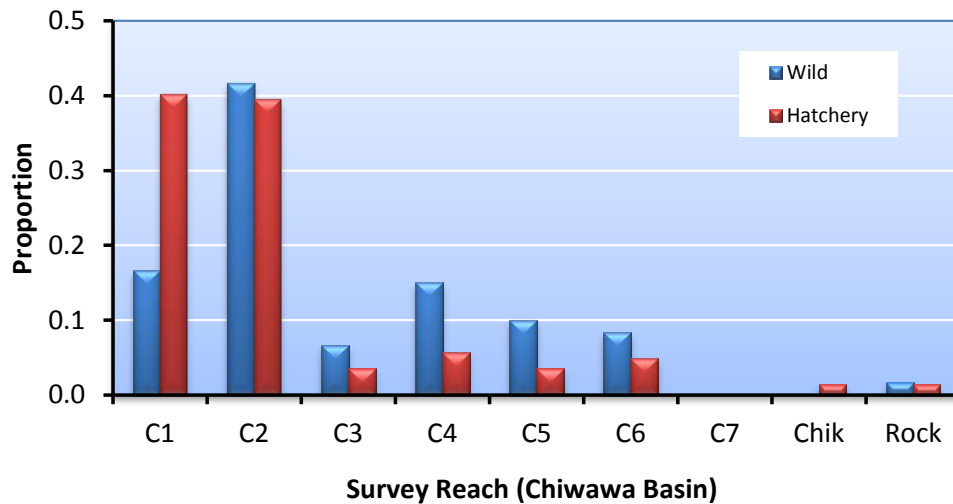
Of the 320 carcasses sampled in the Chiwawa River basin in 2014, 47% were hatchery fish (Table 5.25; these numbers may change after analysis of CWTs). In the Chiwawa River basin, the spatial distribution of hatchery and wild fish was not equal (Table 5.25). A larger percentage of hatchery fish were found in the lower reach (C1; i.e., Mouth to Grouse Creek) than were wild fish. This general trend was also apparent in the pooled data (Figure 5.8).

**Table 5.25.** Numbers of wild and hatchery spring Chinook carcasses sampled within different reaches in the Chiwawa River basin, 1993-2014. See Table 2.8 for description of survey reaches.

Survey year	Origin	Survey Reach									Total
		C-1	C-2	C-3	C-4	C-5	C-6	C-7	Chikamin	Rock	
1993	Wild	0	0	0	0	0	0	--	0	0	<b>0</b>
	Hatchery	1	0	0	0	0	0	--	0	0	<b>1</b>
1994	Wild	0	6	0	2	0	2	--	0	0	<b>10</b>
	Hatchery	1	1	0	2	0	0	--	0	0	<b>4</b>
1995	Wild	0	0	0	0	0	0	--	0	0	<b>0</b>
	Hatchery	2	3	0	1	0	0	--	0	0	<b>6</b>
1996	Wild	13	1	1	1	0	0	--	0	0	<b>16</b>
	Hatchery	6	0	0	0	0	0	--	0	0	<b>6</b>
1997	Wild	5	2	0	1	0	0	--	0	0	<b>8</b>
	Hatchery	3	1	0	0	0	1	--	1	3	<b>9</b>
1998	Wild	0	3	6	1	2	4	--	0	0	<b>16</b>
	Hatchery	1	3	2	0	1	1	--	0	0	<b>8</b>
1999	Wild	1	8	0	5	0	0	--	0	0	<b>14</b>
	Hatchery	0	0	0	0	1	0	--	0	0	<b>1</b>
2000	Wild	29	29	1	1	1	1	--	0	0	<b>62</b>
	Hatchery	42	12	0	0	0	2	--	0	0	<b>56</b>
2001	Wild	27	60	15	43	16	21	--	1	3	<b>186</b>
	Hatchery	164	284	19	58	14	21	--	8	0	<b>568</b>
2002	Wild	22	15	10	6	9	7	--	1	0	<b>70</b>
	Hatchery	46	41	12	5	1	15	--	15	4	<b>139</b>
2003	Wild	7	13	0	12	4	2	--	0	0	<b>38</b>
	Hatchery	14	14	0	3	1	0	--	0	0	<b>32</b>
2004	Wild	25	50	2	12	7	2	--	0	1	<b>99</b>
	Hatchery	48	21	1	1	1	4	--	0	2	<b>78</b>
2005	Wild	18	36	3	5	3	2	--	0	0	<b>67</b>
	Hatchery	170	132	7	7	4	3	--	0	1	<b>324</b>
2006	Wild	10	17	2	8	4	3	--	1	0	<b>45</b>
	Hatchery	84	75	5	7	6	13	--	3	3	<b>196</b>
2007	Wild	3	15	3	4	2	2	--	0	0	<b>29</b>
	Hatchery	42	118	15	14	18	12	--	2	0	<b>221</b>

Survey year	Origin	Survey Reach									Total
		C-1	C-2	C-3	C-4	C-5	C-6	C-7	Chikamin	Rock	
2008	Wild	4	23	0	4	4	8	--	0	0	43
	Hatchery	174	122	2	9	15	15	--	4	1	342
2009	Wild	3	21	4	8	4	1	--	0	3	44
	Hatchery	89	70	6	14	7	5	--	0	5	196
2010	Wild	4	30	7	8	10	3	--	0	0	62
	Hatchery	64	35	2	10	7	5	--	0	5	128
2011	Wild	8	26	10	6	8	6	--	0	1	65
	Hatchery	43	40	4	5	5	10	--	1	4	112
2012	Wild	11	74	6	21	13	18	0	0	3	146
	Hatchery	94	91	9	13	16	16	0	0	6	245
2013	Wild	8	38	7	21	16	14	1	0	3	105
	Hatchery	101	112	19	23	13	15	0	5	3	291
2014	Wild	18	78	8	28	19	21	0	0	0	172
	Hatchery	64	50	4	10	6	9	1	2	2	148
Average	Wild	10	25	4	9	6	5	0	0	1	60
	Hatchery	57	56	5	8	5	7	0	2	2	142

### Spring Chinook Carcass Distribution



**Figure 5.8.** Distribution of wild and hatchery produced carcasses in different reaches in the Chiwawa River basin, 1993-2014; Chik = Chikamin Creek and Rock = Rock Creek. Reach codes are described in Table 2.8.

### Sampling Rate

Overall, 26% of the estimated total spawning escapement of spring Chinook in the Wenatchee River basin was sampled in 2014 (Table 5.26). Sampling rates among streams/watershed varied from 10 to 40%.

**Table 5.26.** Number of redds and carcasses, total spawning escapement, and sampling rates for spring Chinook salmon in the Wenatchee River basin, 2014.

Sampling area	Total number of redds	Total number of carcasses	Total spawning escapement	Sampling rate
Chiwawa	485	320	999	0.32
Nason	115	68	237	0.29
Upper Wenatchee	23	19	47	0.40
Icicle	211	44	424	0.10
Little Wenatchee	25	15	52	0.29
White	26	8	54	0.15
Peshastin	0	0	0	--
<b>Total</b>	<b>885</b>	<b>474</b>	<b>1,813</b>	<b>0.26</b>

### Length Data

Mean lengths (POH, cm) of male and female spring Chinook carcasses sampled during surveys in the Wenatchee River basin in 2014 are provided in Table 5.27. The average sizes of males and females sampled in the Wenatchee River basin were 60 and 62 cm, respectively.

**Table 5.27.** Mean lengths (postorbital-to-hypural length; cm) and standard deviations (in parentheses) of male and female spring Chinook carcasses sampled in different streams/watersheds in the Wenatchee River basin, 2014.

Stream/watershed	Mean lengths (cm)	
	Male	Female
Chiwawa	62 (9.9)	62 (4.7)
Nason	52 (9.7)	62 (4.5)
Upper Wenatchee	65 (5.8)	63 (4.9)
Icicle	65 (8.8)	59 (3.9)
Little Wenatchee	58 (10.6)	67 (5.8)
White	54 (0)	62 (6.4)
Peshastin	--	--
<b>Total</b>	<b>60 (10.6)</b>	<b>62 (4.8)</b>

## 5.7 Life History Monitoring

Life history characteristics of spring Chinook were assessed by examining carcasses on spawning grounds and fish collected at broodstock collection sites, and by reviewing tagging data and fisheries statistics.

### Migration Timing

In 2014, there was a difference in migration timing of hatchery and wild spring Chinook past Tumwater Dam (Table 5.28a and b; Figure 5.9). Hatchery fish arrived at the dam earlier than did wild fish. On average, however, early in the migration, wild Chinook arrived at Tumwater Dam

slightly earlier than hatchery fish, but by the end of the migration, both were arriving at about the same time. Most hatchery and wild spring Chinook migrated upstream past Tumwater Dam during June and July (Figure 5.9).

**Table 5.28a.** The Julian day and date that 10%, 50% (median), and 90% of the wild and hatchery spring Chinook salmon passed Tumwater Dam, 1998-2014. The average Julian day and date are also provided. Migration timing is based on video sampling at Tumwater. Data for 1998 through 2003 were based on videotapes and broodstock trapping and may not reflect the actual number of hatchery spring Chinook. All spring Chinook were visually examined during trapping from 2004 to present.

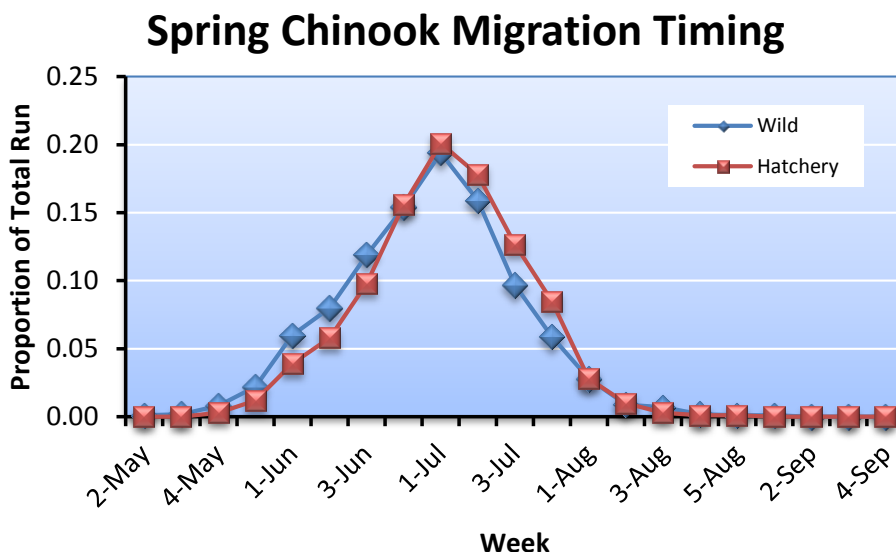
Survey year	Origin	Spring Chinook Migration Time (days)								Sample size
		10 Percentile		50 Percentile		90 Percentile		Mean		
		Julian	Date	Julian	Date	Julian	Date	Julian	Date	
1998	Wild	156	5-Jun	156	5-Jun	156	5-Jun	156	5-Jun	49
	Hatchery	156	5-Jun	156	5-Jun	156	5-Jun	156	5-Jun	25
1999	Wild	192	11-Jul	207	26-Jul	224	12-Aug	207	26-Jul	173
	Hatchery	200	19-Jul	211	30-Jul	229	17-Aug	213	1-Aug	25
2000	Wild	171	19-Jun	186	4-Jul	194	12-Jul	184	2-Jul	651
	Hatchery	179	27-Jun	189	7-Jul	201	19-Jul	190	8-Jul	357
2001	Wild	154	3-Jun	166	15-Jun	185	4-Jul	167	16-Jun	2,073
	Hatchery	157	6-Jun	169	18-Jun	185	4-Jul	170	19-Jun	4,244
2002	Wild	174	23-Jun	189	8-Jul	204	23-Jul	189	8-Jul	1,033
	Hatchery	178	27-Jun	189	8-Jul	199	18-Jul	189	8-Jul	1,363
2003	Wild	162	11-Jun	181	30-Jun	200	19-Jul	181	30-Jun	919
	Hatchery	157	6-Jun	179	28-Jun	192	11-Jul	178	27-Jun	423
2004	Wild	156	4-Jun	172	20-Jun	189	7-Jul	172	20-Jun	969
	Hatchery	161	9-Jun	177	25-Jun	189	7-Jul	177	25-Jun	1,295
2005	Wild	153	2-Jun	172	21-Jun	193	12-Jul	173	22-Jun	1,038
	Hatchery	153	2-Jun	173	22-Jun	187	6-Jul	172	21-Jun	2,808
2006	Wild	177	26-Jun	184	3-Jul	193	12-Jul	185	4-Jul	577
	Hatchery	178	27-Jun	185	4-Jul	194	13-Jul	186	5-Jul	1601
2007	Wild	169	18-Jun	185	4-Jul	203	22-Jul	185	4-Jul	351
	Hatchery	174	23-Jun	192	11-Jul	209	28-Jul	192	11-Jul	3,232
2008	Wild	173	21-Jun	188	6-Jul	209	27-Jul	189	7-Jul	634
	Hatchery	177	25-Jun	193	11-Jul	210	28-Jul	193	11-Jul	5,368
2009	Wild	174	23-Jun	186	5-Jul	201	20-Jul	187	6-Jul	1,008
	Hatchery	175	24-Jun	187	6-Jul	202	21-Jul	188	7-Jul	4,106
2010	Wild	173	22-Jun	190	9-Jul	214	2-Aug	191	10-Jul	977
	Hatchery	180	29-Jun	194	13-Jul	213	1-Aug	195	14-Jul	4,450
2011	Wild	183	2-Jul	198	17-Jul	213	1-Aug	198	17-Jul	1,433
	Hatchery	187	6-Jul	200	19-Jul	210	29-Jul	199	18-Jul	4,707
2012	Wild	180	28-Jun	191	9-Jul	205	23-Jul	192	10-Jul	1,482
	Hatchery	182	30-Jun	194	12-Jul	206	24-Jul	194	12-Jul	4,449

Survey year	Origin	Spring Chinook Migration Time (days)								Sample size
		10 Percentile		50 Percentile		90 Percentile		Mean		
		Julian	Date	Julian	Date	Julian	Date	Julian	Date	
2013	Wild	163	12-Jun	182	1-Jul	199	18-Jul	183	2-Jul	1,106
	Hatchery	164	13-Jun	181	30-Jun	195	14-Jul	181	30-Jun	3,681
2014	Wild	171	20-Jun	188	7-Jul	202	21-Jul	187	6-Jul	1,329
	Hatchery	167	16-Jun	182	1-Jul	195	14-Jul	181	30-Jun	2,510
Average	Wild	<b>169</b>	-	<b>184</b>	-	<b>199</b>	-	<b>184</b>	-	<b>930</b>
	Hatchery	<b>172</b>	-	<b>185</b>	-	<b>198</b>	-	<b>186</b>	-	<b>2,626</b>

**Table 5.28b.** The week that 10%, 50% (median), and 90% of the wild and hatchery spring Chinook salmon passed Tumwater Dam, 1998-2014. The average week is also provided. Migration timing is based on video sampling at Tumwater. Data for 1998 through 2003 were based on videotapes and broodstock trapping and may not reflect the actual number of hatchery spring Chinook. All spring Chinook were visually examined during trapping from 2004 to present.

Survey year	Origin	Spring Chinook Migration Time (week)				Sample size
		10 Percentile	50 Percentile	90 Percentile	Mean	
1998	Wild	23	23	23	23	49
	Hatchery	23	23	23	23	25
1999	Wild	28	30	32	30	173
	Hatchery	29	31	34	31	25
2000	Wild	24	27	27	27	651
	Hatchery	26	27	29	28	357
2001	Wild	22	24	27	24	2,073
	Hatchery	23	25	27	25	4,244
2002	Wild	25	27	30	27	1,033
	Hatchery	26	27	29	27	1,363
2003	Wild	24	26	29	26	919
	Hatchery	23	26	28	26	423
2004	Wild	23	25	27	25	969
	Hatchery	23	26	27	26	1,295
2005	Wild	22	25	28	25	1,038
	Hatchery	22	25	27	25	2,808
2006	Wild	26	27	28	27	577
	Hatchery	26	27	28	27	1,601
2007	Wild	25	27	29	27	351
	Hatchery	25	28	30	28	3,232
2008	Wild	25	27	30	27	634
	Hatchery	26	28	30	28	5,368
2009	Wild	25	27	29	27	1,008

Survey year	Origin	Spring Chinook Migration Time (week)				Sample size
		10 Percentile	50 Percentile	90 Percentile	Mean	
	Hatchery	25	27	29	27	4,106
2010	Wild	25	28	31	28	977
	Hatchery	26	28	31	28	4,450
2011	Wild	27	29	31	29	1,433
	Hatchery	27	29	30	29	4,707
2012	Wild	26	28	30	28	1,482
	Hatchery	26	28	30	28	4,449
2013	Wild	24	26	29	27	1,106
	Hatchery	24	26	28	26	3,681
2014	Wild	25	27	29	27	1,329
	Hatchery	24	26	28	26	2,510
Average	Wild	25	27	29	27	930
	Hatchery	25	27	29	27	2,626



**Figure 5.9.** Proportion of wild and hatchery spring Chinook observed (using video) passing Tumwater Dam each week during their migration period May through September; data were pooled over survey years 1998-2014.

### Age at Maturity

Most of the wild and hatchery spring Chinook sampled during the period 1994-2013 in the Chiwawa River basin were age-4 fish (total age) (Table 5.29; Figure 5.10). On average, hatchery fish made up a higher percentage of age-3 Chinook than did wild fish. In contrast, a higher proportion of age-5 wild fish returned than did age-5 hatchery fish. Thus, wild fish tended to return at an older age than hatchery fish.

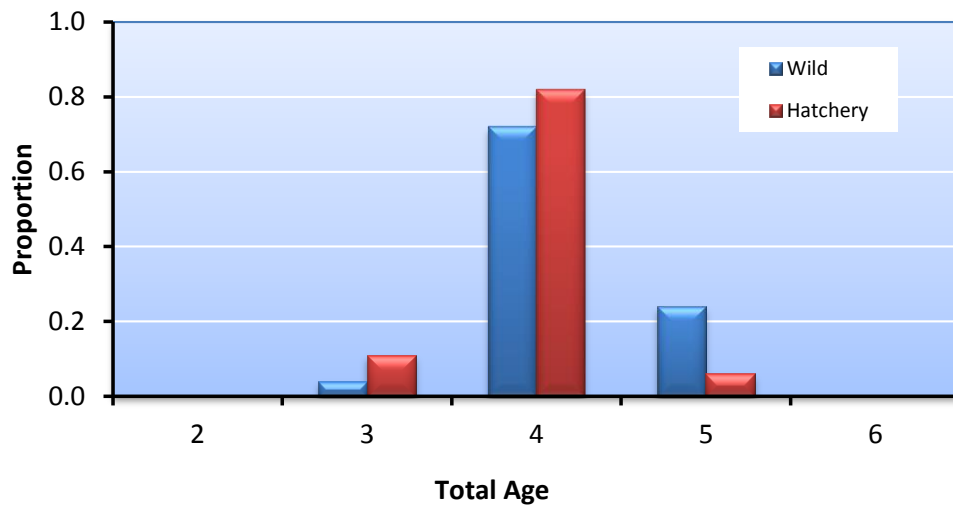
**Table 5.29.** Proportions of wild and hatchery spring Chinook of different ages (total age) sampled on spawning grounds in the Chiwawa River basin, 1994-2013.

Sample year	Origin	Total age					Sample size
		2	3	4	5	6	
1994	Wild	0.00	0.00	0.33	0.67	0.00	9
	Hatchery	0.00	0.20	0.00	0.80	0.00	5
1995	Wild	0.00	0.00	0.00	0.00	0.00	0
	Hatchery	0.00	0.00	1.00	0.00	0.00	5
1996	Wild	0.00	0.36	0.64	0.00	0.00	14
	Hatchery	0.00	0.83	0.17	0.00	0.00	6
1997	Wild	0.00	0.00	0.75	0.25	0.00	8
	Hatchery	0.00	0.00	1.00	0.00	0.00	9
1998	Wild	0.00	0.00	0.00	1.00	0.00	15
	Hatchery	0.00	0.00	0.13	0.88	0.00	8
1999	Wild	0.00	0.07	0.50	0.43	0.00	14
	Hatchery	0.00	0.00	0.00	1.00	0.00	1
2000	Wild	0.00	0.02	0.95	0.04	0.00	56
	Hatchery	0.00	0.50	0.50	0.00	0.00	52
2001	Wild	0.00	0.01	0.95	0.04	0.00	176
	Hatchery	0.00	0.02	0.98	0.00	0.00	571
2002	Wild	0.00	0.00	0.56	0.44	0.00	54
	Hatchery	0.00	0.00	0.91	0.09	0.00	129
2003	Wild	0.00	0.08	0.00	0.92	0.00	36
	Hatchery	0.00	0.19	0.03	0.78	0.00	32
2004	Wild	0.00	0.05	0.94	0.01	0.00	99
	Hatchery	0.00	0.42	0.58	0.00	0.00	78
2005	Wild	0.00	0.02	0.78	0.21	0.00	67
	Hatchery	0.00	0.04	0.96	0.00	0.00	324
2006	Wild	0.02	0.02	0.51	0.44	0.00	45
	Hatchery	0.01	0.04	0.78	0.18	0.00	196
2007	Wild	0.00	0.10	0.24	0.67	0.00	29
	Hatchery	0.00	0.35	0.59	0.06	0.00	221
2008	Wild	0.02	0.02	0.81	0.14	0.00	43
	Hatchery	0.00	0.07	0.89	0.05	0.00	340
2009	Wild	0.00	0.09	0.86	0.05	0.00	44
	Hatchery	0.00	0.24	0.75	0.02	0.00	196
2010	Wild	0.00	0.00	0.90	0.10	0.00	63
	Hatchery	0.00	0.07	0.91	0.02	0.00	127
2011	Wild	0.00	0.08	0.38	0.54	0.00	65
	Hatchery	0.00	0.26	0.45	0.30	0.00	112



Sample year	Origin	Total age					Sample size
		2	3	4	5	6	
2012	Wild	0.00	0.01	0.80	0.19	0.00	141
	Hatchery	0.00	0.03	0.96	0.02	0.00	243
2013	Wild	0.00	0.09	0.60	0.31	0.00	105
	Hatchery	0.00	0.13	0.78	0.10	0.00	290
Average	Wild	0.00	0.04	0.72	0.24	0.00	54
	Hatchery	0.00	0.11	0.83	0.06	0.00	147

### Spring Chinook Age Structure



**Figure 5.10.** Proportions of wild and hatchery spring Chinook of different total ages sampled at the Chiwawa Weir and on spawning grounds in the Chiwawa River basin for the combined years 1994-2014.

### Size at Maturity

On average, hatchery and wild spring Chinook of a given age differed slightly in length (Table 5.30). Differences were usually no more than 1-3 cm between hatchery and wild fish of the same age.

**Table 5.30.** Mean lengths (POH in cm;  $\pm 1SD$ ) and sample sizes (in parentheses) of different ages (total age) of male and female spring Chinook of wild and hatchery-origin sampled in the Chiwawa River basin, 1994-2013. Return years 2004-2014 include carcasses and live fish PIT-tag detections. In addition, 2005 and 2006 include fish released at the weir.

Return year	Total age	Mean length (cm)			
		Male		Female	
		Wild	Hatchery	Wild	Hatchery
1994	3				43 $\pm$ 0 (1)
	4			62 $\pm$ 3 (3)	
	5	76 $\pm$ 0 (1)		73 $\pm$ 2 (5)	
	6				

Return year	Total age	Mean length (cm)			
		Male		Female	
		Wild	Hatchery	Wild	Hatchery
1995	3				
	4		61 ±5 (5)		
	5				
	6				
1996	3	45 ±3 (5)	49 ±7 (10)		
	4	69 ±4 (6)	69 ±0 (1)	67 ±8 (2)	
	5				
	6				
1997	3				
	4	61 ±1 (2)	68 ±0 (1)	67 ±5 (3)	63 ±3 (8)
	5	67 ±5 (2)			
	6				
1998	3				
	4				54 ±0 (1)
	5	77 ±7 (8)	75 ±4 (4)	74 ±4 (7)	76 ±4 (3)
	6				
1999	3	44 ±0 (1)			
	4	61 ±0 (1)		64 ±3 (6)	
	5	76 ±5 (3)		72 ±5 (3)	66 ±0 (1)
	6				
2000	3		46 ±3 (17)		50 ±7 (3)
	4	60 ±8 (23)	62 ±5 (5)	61 ±5 (26)	62 ±3 (20)
	5	77 ±1 (2)			
	6				
2001	3	37 ±0 (1)	42 ±4 (11)	41 ±0 (1)	60 ±0 (1)
	4	63 ±5 (57)	65 ±5 (151)	62 ±4 (110)	63 ±4 (407)
	5	75 ±5 (2)	83 ±0 (1)	76 ±1 (5)	
	6				
2002	3				
	4	64 ±4 (14)	66 ±5 (46)	60 ±4 (15)	63 ±4 (71)
	5	80 ±6 (13)	75 ±5 (4)	72 ±3 (12)	73 ±6 (6)
	6				
2003	3	45 ±2 (3)	45 ±1 (6)		
	4		63 ±0 (1)		
	5	78 ±5 (12)	74 ±8 (11)	75 ±3 (19)	72 ±5 (14)
	6				
2004	3	42 ±3 (3)	44 ±5 (33)		
	4	63 ±7 (60)	66 ±5 (9)	63 ±4 (59)	63 ±6 (36)
	5			74 ±0 (1)	
	6				
2005	3		43 ±5 (48)		

Return year	Total age	Mean length (cm)			
		Male		Female	
		Wild	Hatchery	Wild	Hatchery
	4	61 ±5 (32)	65 ±5 (224)	62 ±4 (61)	62 ±4 (382)
	5	74 ±5 (6)	54±0 (1)	71 ±3 (11)	
	6				
2006	3	45 ±3 (3)	43 ±3 (73)		
	4	64 ±3 (7)	62 ±6 (91)	63 ±5 (41)	60 ±4 (227)
	5	74 ±6 (8)	75 ±6 (17)	71 ±4 (26)	71± 4 (37)
	6				
2007	3	39 ±3 (5)	45 ±6 (90)		50 ±3 (7)
	4	60 ±4 (4)	66 ±5 (45)	61 ±4 (10)	63 ±3 (142)
	5	78 ±6 (15)	76 ±5 (8)	74 ±3 (20)	73 ±5 (12)
	6				
2008	3	43 ±0 (1)	44 ±5 (22)		
	4	65 ±4 (9)	64 ±6 (73)	62 ±4 (26)	64 ±4 (229)
	5	65 ±5 (3)	79 ±5 (10)	73 ±3 (4)	72 ±3 (5)
	6				
2009	3	45 ±3 (8)	46 ±6 (68)		65 ±0 (1)
	4	64 ±4 (38)	65 ±5 (136)	63 ±3 (67)	64 ±4 (202)
	5	79 ±0 (1)		72 ±2 (4)	71 ±4 (10)
	6				
2010	3		46 ±4 (11)		65 ±3 (3)
	4	64 ±5 (31)	66 ±5 (74)	64 ±4 (82)	65 ±3 (196)
	5	77 ±4 (6)		73 ±5 (9)	73 ±6 (4)
	6				
2011	3	43 ±4 (133)	44 ±4 (1374)		53 ±4 (17)
	4	62 ±5 (137)	64 ±5 (169)	64 ±3 (94)	64 ±3 (258)
	5	80 ±5 (78)	79 ±4 (85)	75 ±3 (116)	75 ±3 (63)
	6				
2012	3	56 ±0 (1)	52 ±7 (7)		
	4	79 ± 6 (37)	80 ±6 (49)	79 ±3 (76)	78 ±4 (180)
	5	97 ±7 (11)	96 ±3 (4)	93 ±4 (16)	87 ±0 (1)
	6				
2013	3	45 ±4 (8)	43 ±4 (33)	35 ±0 (1)	49 ±12 (3)
	4	60 ±6 (29)	63 ±7 (41)	61 ±6 (34)	61 ±4 (181)
	5	74 ±5 (9)	71 ±2 (7)	71 ±3 (24)	69 ±5 (22)
	6				

### Contribution to Fisheries

Nearly all the harvest on hatchery-origin Chiwawa spring Chinook occurs within the Columbia River basin. Ocean catch records (Pacific Fishery Management Council) indicate that virtually no Upper Columbia spring Chinook are taken in ocean fisheries. Most of the harvest on

hatchery-origin Chiwawa spring Chinook occurs in the Lower Columbia River fisheries, which are managed by the states and tribes pursuant to management plans developed in *U.S. v Oregon*. The Lower Columbia River fisheries occur during what is referred to in *U.S. v Oregon* as the winter, spring, and summer seasons, which begin in February and ends 31 July of each year. The Tribal fishery occurs upstream from Bonneville Dam, but primarily in Zone 6, the area between Bonneville and McNary dams; the non-treaty commercial fisheries occur in Zones 1-5, which are downstream from Bonneville Dam. The non-treaty recreational (sport) fishery occurs in the lower mainstem. In 2014, a recreational fishery on spring Chinook occurred in Icicle Creek and on the lower Wenatchee River (up to 400 feet downstream from Dryden Dam).

The total number of hatchery-origin spring Chinook captured in different fisheries has been relatively low (Table 5.31). The largest harvests occurred on the 1997, 1998, and 2004-2008 brood years.

**Table 5.31.** Estimated number and percent (in parentheses) of hatchery-origin Chiwawa spring Chinook captured in different fisheries, brood years 1989-2008; NP = no hatchery program.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational <sup>a</sup> (sport)	
1989	3 (13)	5 (21)	0 (0)	16 (67)	24
1990	0 (0)	0 (0)	0 (0)	18 (100)	18
1991	0 (0)	3 (100)	0 (0)	0 (0)	3
1992	0 (0)	1 (100)	0 (0)	0 (0)	1
1993	3 (75)	1 (25)	0 (0)	0 (0)	4
1994	0 (0)	0 (0)	0 (0)	0 (0)	0
1995	NP	NP	NP	NP	NP
1996	0 (0)	2 (100)	0 (0)	0 (0)	2
1997	1 (0)	193 (51)	68 (18)	115 (31)	377
1998	10 (5)	47 (24)	12 (6)	126 (65)	195
1999	NP	NP	NP	NP	NP
2000	0 (0)	17 (74)	0 (0)	6 (26)	23
2001	36 (64)	8 (14)	1 (2)	11 (20)	56
2002	12 (17)	11 (15)	22 (31)	26 (37)	71
2003	18 (21)	29 (35)	11 (13)	26 (31)	84
2004	3 (1)	188 (40)	31 (7)	253 (53)	475
2005	18 (14)	31 (24)	6 (5)	74 (57)	129
2006	32 (4)	469 (60)	77 (10)	201 (26)	779
2007	14 (3)	180 (43)	74 (18)	151 (36)	419
2008	8 (1)	298 (21)	41 (3)	1,047 (75)	1,394
<b>Average</b>	<b>9 (12)</b>	<b>82 (42)</b>	<b>19 (6)</b>	<b>115 (35)</b>	<b>225</b>

<sup>a</sup> Includes the Wanapum fishery.

## Straying

Stray rates were determined by examining CWTs recovered on spawning grounds within and outside the Wenatchee River basin. Targets for strays based on return year (recovery year) within the Wenatchee River basin should be less than 10% and targets for strays outside the Wenatchee River basin should be less than 5%. The target for brood year stray rates should be less than 5%.

The percentage of the spawning escapement made up of hatchery-origin Chiwawa spring Chinook in non-target spawning areas within the Wenatchee River basin has been high in some years and exceeded the target of 10% (Table 5.32). Chiwawa spring Chinook have strayed into spawning areas on Nason Creek, the White River, the Little Wenatchee River, and the Upper Wenatchee River. On average, Chiwawa spring Chinook made up the highest percentage of the spawning escapement within Nason Creek and the Upper Wenatchee River. Stray rates of hatchery-origin Chiwawa spring Chinook do not appear to have declined with the change in source water that was implemented in 2006 for the Chiwawa rearing ponds.

**Table 5.32.** Number (No.) and percent (%) of the spawning escapement in other non-target spawning streams within the Wenatchee River basin that consisted of hatchery-origin Chiwawa spring Chinook, return years 1992-2013. For example, for return year 2001, 35.3% of the spring Chinook spawning escapement in Nason Creek consisted of hatchery-origin Chiwawa spring Chinook. Percent strays should be less than 10%.

Return year	Nason Creek		Icicle Creek		Peshastin Creek		Upper Wenatchee		White River		Little Wenatchee	
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%
1992	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1993	61	12.4	0	0.0	0	0.0	34	18.0	7	4.8	0	0.0
1994	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1995	0	0.0	0	0.0	0	0.0	2	66.7	0	0.0	0	0.0
1996	25	30.1	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1997	55	45.1	8	11.0	0	0.0	0	0.0	0	0.0	0	0.0
1998	3	4.7	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1999	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2000	45	16.7	0	0.0	0	0.0	31	31.0	0	0.0	6	27.3
2001	211	35.3	0	0.0	0	0.0	271	77.7	46	39.0	52	31.3
2002	188	31.2	10	2.0	0	0.0	60	45.8	14	16.3	21	24.4
2003	14	6.9	0	0.0	0	0.0	30	51.7	0	0.0	0	0.0
2004	139	27.4	0	0.0	0	0.0	54	39.1	6	9.1	0	0.0
2005	252	72.6	7	50.0	0	0.0	256	99.6	106	68.4	65	56.5
2006	131	48.3	13	14.4	0	0.0	28	58.3	9	16.4	12	32.4
2007	303	65.4	0	0.0	0	0.0	37	67.3	7	7.6	6	5.9
2008	381	67.4	48	23.4	29	78.4	258	85.4	30	57.7	52	81.3
2009	289	54.1	8	9.2	0	0.0	16	100.0	63	36.4	56	44.8
2010	272	66.3	58	13.7	11	78.6	86	84.3	23	31.9	59	71.1
2011	397	56.6	61	18.8	0	0.0	41	82.0	0	0.0	53	42.7
2012	398	59.1	49	13.0	7	36.8	98	82.4	45	32.1	15	21.4
2013	281	68.4	15	8.0	0	0.0	24	72.7	5	4.8	10	10.1
<b>Average</b>	<b>157</b>	<b>34.9</b>	<b>13</b>	<b>7.4</b>	<b>2</b>	<b>8.8</b>	<b>60</b>	<b>48.3</b>	<b>16</b>	<b>14.7</b>	<b>19</b>	<b>20.4</b>

Hatchery-origin Chiwawa spring Chinook have strayed into the Methow and Entiat basins (Table 5.33). Based on return year analyses, rates of hatchery-origin Chiwawa spring Chinook straying into these populations have been low in most years. However, during return years 2002, 2006, 2008-2009, and 2011-2013, Chiwawa spring Chinook made up more than 5% of the spawning escapement in the Entiat River basin. In some years, Chiwawa spring Chinook hatchery fish made up more than 20% of the spawning escapement in the Entiat River basin.

**Table 5.33.** Number and percent of spawning escapements within other non-target basins that consisted of hatchery-origin Chiwawa spring Chinook, return years 1992-2013. For example, for return year 2002, 9.2% of the spring Chinook spawning escapement in the Entiat River basin consisted of hatchery-origin Chiwawa spring Chinook. Percent strays should be less than 5%. NS = not sampled.

Return year	Methow River basin		Entiat River basin	
	Number	%	Number	%
1992	0	0.0	0	0.0
1993	0	0.0	0	0.0
1994	0	0.0	0	0.0
1995	0	0.0	0	0.0
1996	ns	ns	0	0.0
1997	0	0.0	0	0.0
1998	ns	ns	0	0.0
1999	0	0.0	0	0.0
2000	0	0.0	1	0.6
2001	0	0.0	1	0.2
2002	0	0.0	34	9.2
2003	0	0.0	6	2.3
2004	0	0.0	0	0.0
2005	10	0.7	15	4.2
2006	8	0.5	24	9.3
2007	9	0.8	4	1.6
2008	12	1.2	61	21.9
2009	9	0.3	15	5.4
2010	10	0.4	18	3.7
2011	51	1.7	190	31.9
2012	13	1.0	133	23.5
2013	9	0.8	24	10.1
<b>Average</b>	<b>7</b>	<b>0.4</b>	<b>24</b>	<b>5.6</b>

Based on brood year analyses, on average, about 31% of the hatchery returns have strayed into non-target spawning areas, exceeding the target of 5% (Table 5.34). Depending on brood year, percent strays into non-target spawning areas have ranged from 0-81%. In most years, few (<1%) have strayed into non-target hatchery programs. The change in source water that was implemented in 2006 for the Chiwawa rearing ponds does not appear to have decreased stray rates.

**Table 5.34.** Number and percent of hatchery-origin Chiwawa spring Chinook that homed to target spawning areas and the target hatchery program, and number and percent that strayed to non-target spawning areas and non-target hatchery programs, by brood years 1989-2008. Percent strays should be less than 5%.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1989	74	41.1	1	0.6	102	56.7	3	1.7
1990	0	0.0	1	100.0	0	0.0	0	0.0
1991	29	90.6	0	0.0	2	6.3	1	3.1
1992	2	6.5	4	12.9	25	80.6	0	0.0
1993	134	47.5	82	29.1	63	22.3	3	1.1
1994	4	19.0	14	66.7	3	14.3	0	0.0
1995	No program							
1996	58	75.3	7	9.1	12	15.6	0	0.0
1997	1,242	55.6	298	13.4	687	30.8	5	0.2
1998	553	55.8	109	11.0	329	33.2	0	0.0
1999	No program							
2000	149	42.1	115	32.5	90	25.4	0	0.0
2001	647	35.8	276	15.3	881	48.7	4	0.2
2002	314	44.3	238	33.6	156	22.0	1	0.1
2003	556	78.6	11	1.6	133	18.8	7	1.0
2004	1,198	47.4	203	8.0	1104	43.7	23	0.9
2005	822	59.3	139	10.0	415	29.9	10	0.7
2006	1,007	54.8	147	8.0	669	36.4	14	0.8
2007	510	57.8	60	6.8	294	33.3	19	2.2
2008	1,160	47.1	62	2.5	1,144	46.4	99	4.0
<b>Average</b>	<b>423</b>	<b>47.7</b>	<b>98</b>	<b>20.0</b>	<b>339</b>	<b>31.4</b>	<b>11</b>	<b>0.9</b>

\* Homing to the target hatchery includes Chiwawa hatchery spring Chinook that are captured and included as broodstock in the Chiwawa Hatchery program. These hatchery fish are typically collected at the Chiwawa weir and Tumwater Dam.

Recently, Ford et al. (2015) used parentage analysis to estimate rates of straying and homing of spring Chinook within the Wenatchee River basin. They found that stray rates of hatchery spring Chinook based on parentage analysis were consistent with rates estimated using physical tag recoveries (the latter estimates are shown in the tables above). They also found that stray rates among the major spawning tributaries were higher than stray rates of tagged fish to areas outside of the Wenatchee River basin (e.g., Entiat and Methow basins), which is consistent with the results shown in the tables above. Finally, the researchers noted that hatchery spring Chinook homed at a far lower rate than natural-origin fish. Rates of straying of natural-origin spring Chinook were affected by spawning tributary and by parental origin (i.e., progeny of naturally spawning hatchery-produced fish strayed at higher rates than progeny whose parents were of natural origin).

## Genetics

Genetic studies were conducted to determine the potential effects of the Chiwawa Supplementation Program on natural-origin spring Chinook in the upper Wenatchee River basin (Blankenship et al. 2007; the entire report is appended as Appendix J). Microsatellite DNA allele frequencies collected from temporally replicated natural and hatchery-origin spring Chinook were used to statistically assign individual fish to specific demes (locations) within the Wenatchee population. In addition, genetic effects of the hatchery program were assessed by examining relationships between census and effective population sizes ( $N_e$ ) from samples collected before and after supplementation.

Overall, this work showed that although allele frequencies within and between natural and hatchery-origin spring Chinook were significantly different, there was no evidence (i.e., robust signal) that the difference was the result of the hatchery program. Rather, the differences were more likely the result of life history characteristics. However, there was an increasing trend toward homogenization of the allele frequencies of the natural and hatchery-origin fish that comprised the broodstock, even though there was consistent year-to-year variation in allele frequencies among hatchery and natural-origin fish. In addition, there were no robust signals indicating that hatchery-origin hatchery broodstock, hatchery-origin natural spawners, natural-origin hatchery broodstock, and natural-origin natural spawners were substantially different from each other. Finally, the  $N_e$  estimate of 387 was only slightly larger than the pre-hatchery  $N_e$  (based on demographic data from 1989-1992), which means that the Chiwawa hatchery program has not reduced the  $N_e$  of the Wenatchee spring Chinook population.

Significant differences in allele frequencies were observed within and among major spawning areas in the Upper Wenatchee River basin. However, these differences made up only a very small portion of the overall variation, indicating genetic similarity among the major spawning areas. There was no evidence that the Chiwawa program has changed the genetic structure (allele frequency) of spring Chinook in Nason Creek and the White River, despite the presence of hatchery-origin spawners in both systems.

## Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For brood years 1989-1994, the approximate PNI Index was greater than or equal to 0.67 (Table 5.35). Since brood year 1994, the PNI has been less than 0.67.



**Table 5.35.** Approximate Proportionate Natural Influence (PNI) Index of the Chiwawa spring Chinook supplementation program for brood years 1989-2014. PNI was calculated as the proportion of naturally produced Chinook in the hatchery broodstock (pNOB) divided by the proportion of hatchery Chinook on the spawning grounds (pHOS) plus pNOB. NOS = number of natural-origin Chinook on the spawning grounds; HOS = number of hatchery-origin Chinook on the spawning grounds; NOB = number of natural-origin Chinook collected for broodstock; and HOB = number of hatchery-origin Chinook included in hatchery broodstock.

Brood year	Spawners			Broodstock			PNI	
	NOS	HOS	pHOS	NOB	HOB	pNOB		
1989	713	0	0.00	28	0	1.00	1.00	
1990	571	0	0.00	18	0	1.00	1.00	
1991	242	0	0.00	27	0	1.00	1.00	
1992	676	0	0.00	78	0	1.00	1.00	
1993	231	2	0.01	94	0	1.00	0.99	
1994	123	61	0.33	8	4	0.67	0.67	
1995	0	33	1.00	No Program				
1996	41	17	0.29	8	10	0.44	0.60	
1997	60	122	0.67	32	79	0.29	0.30	
1998	59	32	0.35	13	34	0.28	0.44	
1999	87	7	0.07	No Program				
2000	233	113	0.33	9	21	0.30	0.48	
2001	506	1,219	0.71	113	259	0.30	0.30	
2002	255	452	0.64	20	51	0.28	0.30	
2003	167	103	0.38	41	53	0.44	0.54	
2004	573	278	0.33	83	132	0.39	0.54	
2005	139	460	0.77	91	181	0.33	0.30	
2006	115	413	0.78	91	224	0.29	0.27	
2007	155	1,141	0.88	43	104	0.29	0.25	
2008	197	961	0.83	83	220	0.27	0.25	
2009	303	1,044	0.78	96	111	0.46	0.37	
2010	418	676	0.62	77	98	0.44	0.42	
2011	795	1,237	0.61	80	93	0.46	0.43	
2012	576	902	0.61	73	0	1.00	0.62	
2013	412	966	0.70	70	0	1.00	0.59	
2014	533	466	0.47	61	134	0.31	0.40	
<b>Average</b>	<b>315</b>	<b>412</b>	<b>0.47</b>	<b>51</b>	<b>70</b>	<b>0.51</b>	<b>0.50</b>	

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel time (arithmetic mean days) of hatchery spring Chinook from the Chiwawa River release site to McNary Dam, and smolt to

adult ratios (SARs) from release to detection at Bonneville Dam (Table 5.36).<sup>7</sup> Over the eight brood years for which PIT-tagged hatchery fish were released, survival rates from the Chiwawa River to McNary Dam ranged from 0.435 to 0.662; SARs from release to detection at Bonneville Dam ranged from 0.003 to 0.018. Average travel time from the Chiwawa River to McNary Dam ranged from 14 to 44 days. Although there is only one year in which a forced release was compared to a volitional release (brood year 2005), hatchery spring Chinook that were forced out of the Chiwawa Acclimation Facility had slightly higher survival rates and SARs, and a faster travel time to McNary Dam, than did the volitional release.

**Table 5.36.** Total number of Chiwawa hatchery spring Chinook released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2005-2012. Standard errors are shown in parentheses. NA = not available (i.e., not all the adults from the release groups have returned to the Columbia River).

Brood year	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2005	4,993 (forced)	0.662 (0.027)	22.9 (6.6)	0.008 (0.001)
2005	4,988 (volitional)	0.638 (0.027)	43.6 (6.9)	0.003 (0.001)
2006	9,894	0.619 (0.038)	30.6 (7.6)	0.011 (0.001)
2007	10,031	0.435 (0.019)	32.9 (7.7)	0.007 (0.001)
2008	10,006	0.631 (0.038)	39.9 (10.3)	0.018 (0.001)
2009	9,412	0.547 (0.044)	30.2 (6.7)	0.006 (0.001)
2010	5,020	0.548 (0.038)	18.9 (7.3)	NA
2011	9,987	0.458 (0.029)	14.2 (7.5)	NA
2012	5,061	0.478 (0.043)	30.9 (6.5)	NA

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). We calculated NORs with and without harvest. NORs without harvest include all returning fish that either returned to the basin or were collected as wild broodstock. NORs with harvest include all fish harvested and are based on a brood year harvest rates from the hatchery program. For brood years 1989-2008, NRR for spring Chinook in the Chiwawa averaged 1.11 (range, 0.01-4.40) if harvested fish were not include in the estimate and 1.22 (range, 0.01-4.81) if harvested fish were included in the estimate (Table 5.37). NRRs for more recent brood years will be calculated as soon as all tag recoveries and sampling rates have been loaded into the database.

<sup>7</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and were calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. These rates should be greater than the NRRs and greater than or equal to 5.30 (the calculated target value in Hillman et al. 2013). In nearly all years, HRRs were greater than NRRs, regardless if harvest was or was not included (Table 5.37). HRRs exceeded the estimated target value of 5.3 in 8 or 11 of the 18 years, depending on if harvested fish were or were not included in the estimates.

**Table 5.37.** Broodstock collected, spawning escapements, natural and hatchery-origin recruits (NOR and HOR), and natural and hatchery replacement rates (NRR and HRR; with and without harvest) for spring Chinook in the Chiwawa River basin, brood years 1989-2008; NP = no hatchery program.

Brood year	Broodstock Collected	Spawning Escapement	Harvest not included				Harvest included			
			HOR	NOR	HRR	NRR	HOR	NOR	HRR	NRR
1989	28	713	180	194	6.43	0.27	204	282	7.29	0.40
1990	19	571	1	34	0.05	0.06	19	40	1.00	0.07
1991	32	242	32	2	1.00	0.01	35	2	1.09	0.01
1992	113	676	31	46	0.27	0.07	32	48	0.28	0.07
1993	100	233	282	159	2.82	0.68	286	163	2.86	0.70
1994	13	184	21	37	1.62	0.20	21	38	1.62	0.21
1995	NP	33	--	66	--	2.00	--	69	--	2.09
1996	18	58	77	255	4.28	4.40	79	279	4.39	4.81
1997	120	182	2,232	714	18.60	3.92	2,609	792	21.74	4.35
1998	48	91	991	349	20.65	3.84	1,186	372	24.71	4.09
1999	NP	94	--	10	--	0.11	--	11	--	0.12
2000	48	346	354	699	7.38	2.02	377	733	7.85	2.12
2001	382	1,725	1,808	309	4.73	0.18	1,864	317	4.88	0.18
2002	84	707	709	244	8.44	0.35	780	254	9.29	0.36
2003	119	270	707	107	5.94	0.40	791	115	6.65	0.43
2004	296	858	2,528	276	8.54	0.32	3,003	298	10.15	0.35
2005	283	598	1,386	396	4.90	0.66	1,515	412	5.35	0.69
2006	398	529	1,837	967	4.62	1.83	2,616	1,219	6.57	2.30
2007	169	1,296	883	474	5.22	0.37	1,302	570	7.70	0.44
2008	329	1,158	2,465	726	7.49	0.63	3,859	816	11.73	0.70
<b>Average</b>	<b>144</b>	<b>528</b>	<b>918</b>	<b>303</b>	<b>6.28</b>	<b>1.11</b>	<b>1,143</b>	<b>342</b>	<b>7.51</b>	<b>1.22</b>

### Smolt-to-Adult Survivals

Smolt-to-adult survival ratios (SARs) were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery smolts released. Here, SARs were based on CWT returns. For the available brood years, SARs have ranged from 0.00036 to 0.01563 for hatchery spring Chinook (Table 5.38).

**Table 5.38.** Smolt-to-adult ratios (SARs) for Chiwawa hatchery spring Chinook, brood years 1989-2008.

Brood year	Number of tagged smolts released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1989	42,707	204	0.00478
1990	52,798	19	0.00036
1991	61,088	35	0.00057
1992	82,976	31	0.00037
1993	221,316	284	0.00128
1994	27,135	21	0.00077
1995	No hatchery program		
1996	12,767	67	0.00525
1997	259,585	2,549	0.00982
1998	71,571	1,119	0.01563
1999	No hatchery program		
2000	46,726	375	0.00803
2001	374,129	1,849	0.00494
2002	145,074	760	0.00524
2003	216,702	775	0.00358
2004	491,987	2,992	0.00608
2005	489,664	1,506	0.00308
2006	548,777	2,604	0.00475
2007	292,682	1,300	0.00444
2008	609,286	3,859	0.00633
<b>Average</b>	<b>224,832</b>	<b>1,131</b>	<b>0.00474</b>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

## 5.8 ESA/HCP Compliance

### Broodstock Collection

The collection of 2012 Brood Chiwawa River spring Chinook broodstock was consistent with the 2012 Upper Columbia River salmon and steelhead broodstock objectives and site-based broodstock collection protocols. Specifically, broodstock collection targeted hatchery and natural-origin fish at the Chiwawa Weir. In-season adjustments were made to the number of hatchery and natural-origin spring Chinook collected for broodstock as needed and were based on in-season escapement monitoring at Tumwater Dam and estimated Chiwawa run-escapement.

Trapping at the Chiwawa Weir began on 1 June 2012 and concluded on 6 August 2012. Broodstock collection targeted natural-origin spring Chinook and hatchery-origin spring

Chinook as needed to attain a minimum 33% natural-origin broodstock and a maximum 33% extraction of the estimated natural-origin return to the Chiwawa River.

The 2012 brood collection retained a total of 116 spring Chinook, including 75 natural-origin fish, representing a 65% natural-origin broodstock. The brood successfully met the minimum targeted 33% natural-origin composition.

At the Chiwawa Weir, the trap was operated passively, checked several times per day, and fish were processed once daily. Trapping at the Chiwawa Weir generally followed a four-up and three-down schedule, and operated only as needed to meet weekly collection objectives consistent with the 2012 collection protocol or as adjusted based on in-season run escapement monitoring and ESA Section 10 Permit 1196 requirements. All spring Chinook, steelhead, and bull trout that were captured were anesthetized with tricaine methanesulfonate (MS-222) and subject to water-to-water transfers during handling. All fish were allowed to fully recover before release.

The estimated broodstock extraction rate of natural-origin Chiwawa spring Chinook and overall extraction of spring Chinook upstream from Tumwater Dam comply with provisions of ESA Permit 1196 (expired).

No additional spring Chinook were handled and released as a function of maintaining, at minimum, 33% natural-origin spring Chinook in the broodstock. About 294 bull trout were captured and released. To minimize fallback or impingement on the weir, all spring Chinook and bull trout were released unharmed about 10 km upstream from the weir.

### **Hatchery Rearing and Release**

The rearing and release of 2012 brood Chiwawa spring Chinook was completed without incident. No mortality events occurred that exceeded 10% of the population. Fish were acclimated on Chiwawa River water with regulated amounts of Wenatchee River water to prevent frazzle ice formation during the winter months (see Section 5.2).

The release of 2012 brood Chiwawa spring Chinook smolts totaled 222,504 fish, representing 108.8% of program objectives and complied with the ESA Section 10 Permit 1196 program level of 204,452 smolts.

### **Hatchery Effluent Monitoring**

Per ESA Permits 1196 (expired), 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at the Chelan PUD Hatchery facilities during the period 1 January through 31 December 2014. NPDES monitoring and reporting for Chelan PUD Hatchery Programs during 2014 are provided in Appendix F.

### **Smolt and Emigrant Trapping**

Per ESA Section 10 Permit No. 1196 (expired) and 18121, the permit holders are authorized a direct take of up to 20% of the emigrating spring Chinook population during juvenile emigration monitoring and a lethal take not to exceed 2% of the fish captured (NMFS 2003). Based on the estimated wild spring Chinook population (smolt trap expansion) and hatchery juvenile spring Chinook population estimate (hatchery release data) for the Wenatchee River basin, the reported

spring Chinook encounters during 2014 emigration monitoring complied with take provisions in the Section 10 permit. Spring Chinook encounter and mortality rates for each trap site (including PIT tag mortalities) are detailed in Table 5.39. Additionally, juvenile fish captured at the trap locations were handled consistent with provisions in ESA Section 10 Permit 1196, Section B.

**Table 5.39.** Estimated take of Upper Columbia River spring Chinook resulting from juvenile emigration monitoring in the Wenatchee River basin, 2014.

Trap location	Population estimate			Number trapped			Total	Take allowed under Permit
	Wild <sup>a</sup>	Hatchery <sup>b</sup>	Sub-yearling <sup>c</sup>	Wild	Hatchery	Sub-yearling		
<b>Chiwawa Trap</b>								
Population	34,334	222,504	73,695	4,519	5,293	23,755	33,567	
Encounter rate	NA	NA	NA	0.1316	0.0237	0.3223	0.1016	0.20
Mortality <sup>e</sup>	NA	NA	NA	28	0	84	112	
Mortality rate	NA	NA	NA	0.0062	0.0000	0.0035	0.0033	0.02
<b>Lower Wenatchee Trap</b>								
Population	67,973	222,504	11,936,928	1,700	31,290	81,445	114,435	
Encounter rate	NA	NA	NA	0.025	0.1406	0.0068	0.0096	0.20
Mortality <sup>d</sup>	NA	NA	NA	17	12	250	279	
Mortality rate	NA	NA	NA	0.0100	0.0004	0.0031	0.0024	0.02
<b>Wenatchee River Basin Total</b>								
Population	67,393	222,504	11,936,928	6,219	36,583	105,200	148,002	
Encounter rate	NA	NA	NA	0.0922	0.1644	0.0088	0.0121	0.20
Mortality <sup>d</sup>	NA	NA	NA	45	12	84	141	
Mortality rate	NA	NA	NA	0.0072	0.0003	0.0008	0.0010	0.02

<sup>a</sup> Smolt population estimate derived from juvenile emigration trap data.

<sup>b</sup> 2012BY smolt release data for the Wenatchee River basin.

<sup>c</sup> Based on size, date of capture and location of capture, subyearling Chinook encountered at the Lower Wenatchee Trap are categorized as summer Chinook..

<sup>d</sup> Combined trapping and PIT tagging mortality.

## Spawning Surveys

Spring Chinook spawning ground surveys were conducted in the Wenatchee River basin during 2014, as authorized by ESA Section 10 Permits 18118, 18119, and 18121. Because of the difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical, and extreme caution was used to avoid established redds when wading was required.

## Spring Chinook Reproductive Success Study

ESA Section 10 Permit 1196 (expired) and new Section 10 Permits 18118, 18119, and 18121 specifically provide authorization to capture, anesthetize, biologically sample, PIT tag, and release adult spring Chinook at Tumwater Dam for reproductive success studies and general program monitoring. During 2010 through 2014, all spring Chinook passing Tumwater Dam were enumerated, anesthetize, biologically sampled, PIT tagged, and released (not including

hatchery-origin Chinook retained for broodstock) as a component of the reproductive success study (BPA Project No. 2003-039-00). Please refer to Ford et al. (2010, 2011, 2012, 2013 and 2014) for complete details on the methods and results of the spring Chinook reproductive success study for the period 2010-2014.





## SECTION 6: NASON CREEK SPRING CHINOOK

The goals of the Nason Creek spring Chinook salmon supplementation program are to conserve, aid in the recovery, and prevent the extinction of naturally spawning spring Chinook in Nason Creek, and to meet the mitigation responsibilities of Grant County PUD. In 1997, a spring Chinook captive-broodstock program was initiated for the Nason Creek population to reduce the risk of extinction. Improvements in adult escapement in Nason Creek have reduced the near-term risk of extinction and therefore the captive-broodstock program was discontinued. An adult-based supplementation program began with the collection of broodstock in 2013. The first releases of the program will take place from the Nason Creek Acclimation Facility in the spring of 2015.

In 2013, natural-origin adult spring Chinook were collected for broodstock at Tumwater Dam and from Nason Creek using tangle and dip nets. In 2014, all natural-origin broodstock were collected from Nason Creek using tangle and dip nets. While these brood collection methods were successful at collecting adults from the Nason Creek spawning aggregate, they were unable to collect the necessary number of adults to meet mitigation production goals in 2013 and 2014. The production goal for the Nason Creek program requires collection of 126 adult spring Chinook (64 natural-origin fish and 66 hatchery-origin fish). However, the Section 10 permit requirements restrict the number of natural-origin adults collected and cannot exceed 33% of the natural-origin spring Chinook estimates to Tumwater Dam.

Adult spring Chinook are spawned and reared at Eastbank Fish Hatchery. Juvenile spring Chinook are transferred from the hatchery to the Nason Creek Acclimation Facility in late September or early October. Fish are reared in 30-foot dual-drain circular tanks throughout winter at the Nason Creek Acclimation Facility. Yearling Chinook are released voluntarily during April and May the following year.

The current production goal is to release 223,670 smolts (125,000 for conservation and 98,670 for safety net). Juveniles released from the Nason facility will be 100% marked with CWTs and a minimum of 5,000 fish will be PIT tagged annually.

The following information focuses on results from monitoring the Nason Creek spring Chinook program. Information on spring Chinook collected throughout the Wenatchee River basin is presented in Section 5.

### 6.1 Broodstock Sampling

This section focuses on results from sampling 2013-2014 Nason Creek spring Chinook broodstock, which were collected in Nason Creek and at Tumwater Dam. Some information for the 2014 return is not available at this time (e.g., age structure and final origin determination). This information will be provided in the 2015 annual report.

#### Origin of Broodstock

Natural-origin adults made up most of the Nason Creek spring Chinook broodstock for brood year 2013 and only natural-origin adults made up the broodstock for brood year 2014 (Table 6.1). For brood year 2013, natural-origin adults were collected at Tumwater Dam (N = 3) and in Nason Creek (N = 19). For brood year 2014, natural-origin adults were targeted for collection at

Tumwater Dam during trapping operations and in Nason Creek using tangle nets. Natural-origin fish collected at Tumwater Dam were used for broodstock if they had been previously PIT tagged in Nason Creek. No fish were identified using this method at Tumwater Dam; however, 28 natural-origin fish were acquired using tangle nets.

**Table 6.1.** Numbers of wild and hatchery Nason Creek spring Chinook collected for broodstock, numbers that died before spawning, and numbers of Chinook spawned, 2013-2014. Unknown origin fish (i.e., undetermined by scale analysis, no CWT or fin clips, and no additional hatchery marks) were considered naturally produced. Mortality includes fish that died of natural causes typically near the end of spawning and were not needed for the program or were surplus fish killed at spawning.

Brood year	Wild spring Chinook					Hatchery spring Chinook					Total number spawned
	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	
2013	22	0	1	21	0	4	0	0	4	0	25
2014	28	2	5	21	0	0	0	0	0	0	21
<i>Average<sup>b</sup></i>	<i>25</i>	<i>1</i>	<i>3</i>	<i>21</i>	<i>0</i>	<i>2</i>	<i>0</i>	<i>0</i>	<i>2</i>	<i>0</i>	<i>23</i>

<sup>a</sup> Pre-spawn loss represents the number of fish that died during the holding period before spawning. Mortality is the number of fish that were surplus following spawning.

<sup>b</sup> Origin determinations should be considered preliminary pending scale analyses.

### Age/Length Data

Ages were determined from scales and/or coded wire tags (CWT) collected from broodstock. For the 2013 return, most adults were age-4 Chinook (Table 6.2).

**Table 6.2.** Percent of hatchery and wild spring Chinook of different ages (total age) collected from broodstock in 2013.

Return year	Origin	Total age			
		2	3	4	5
2013	Wild	0.0	14.3	85.7	0.0
	Hatchery	0.0	0.0	100.0	0.0
<i>Average</i>	<i>Wild</i>	0.0	14.3	85.7	0.0
	<i>Hatchery</i>	0.0	0.0	100.0	0.0

Length at age for Nason Creek wild spring Chinook are shown in Table 6.3.

**Table 6.3.** Mean fork length (cm) at age (total age) of hatchery and wild spring Chinook collected from broodstock in 2013; N = sample size and SD = 1 standard deviation.

Return year	Origin	Spring Chinook fork length (cm)											
		Age-2			Age-3			Age-4			Age-5		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
2013	Wild	-	0	-	56	3	2	75	18	6	-	0	-
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-
<i>Average</i>	<i>Wild</i>	-	0	-	56	3	2	75	18	6	-	0	-
	<i>Hatchery</i>	-	0	-	-	0	-	79	4	7	-	0	-

## Sex Ratios

Male spring Chinook in the 2013-2014 return years made up 50% and 60%, respectively, of the adults collected. This resulted in an overall male to female ratios of 1.00:1.00 and 1.50:1.00, respectively (Table 6.4).

**Table 6.4.** Numbers of male and female wild and hatchery spring Chinook collected for broodstock, 2013-2014. Ratios of males to females are also provided.

Return year	Number of wild spring Chinook			Number of hatchery spring Chinook			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
2013	12	10	1.20:1.00	1	3	0.33:1.00	1.00:1.00
2014	18	12	1.50:1.00	0	0	-	1.50:1.00
<b>Total</b>	<b>30</b>	<b>22</b>	<b>1.36:1.00</b>	<b>1</b>	<b>3</b>	<b>0.33:1.00</b>	<b>1.24:1.00</b>

## Fecundity

The mean fecundities for the 2013-2014 returns of Nason Creek spring Chinook ranged from 4,052-4,484 eggs per female (Table 6.5). These fecundities were less than the expected fecundity of 4,400 eggs per female assumed in the broodstock protocol.

**Table 6.5.** Mean fecundity of wild, hatchery, and all female spring Chinook collected for broodstock, 2013-2014.

Return year	Mean fecundity		
	Wild	Hatchery	Total
2013	4,047	4,069	4,052
2014	4,484	-	4,484
<b>Average</b>	<b>4,266</b>	<b>4,069</b>	<b>4,268</b>

## 6.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Based on the unfertilized egg-to-release survival standard of 81%, a total of 147,059 eggs are required to meet the program release goal of 125,000 smolts (Table 6.6). The green egg take for the 2013 and 2014 brood years was 34% and 30% of program goal, respectively. This was largely because of the low number of Nason Creek broodstock collected at Tumwater Dam and Nason Creek.

ESA Permit 18118 sets limits on the percentage of the total run and total number of natural-origin fish in the broodstock to meet the conservation program. Applying these criteria to the low total abundance of spring Chinook salmon to the Nason Creek basin, and the low abundance of natural-origin fish returning to the basin, has resulted in the program not meeting production goals.

**Table 6.6.** Numbers of eggs taken from spring Chinook broodstock, 2013-2014.

Return year	Number of eggs taken
2013	49,720
2014	44,844
<i>Average</i>	<i>47,282</i>

**Number of acclimation days**

There is currently no juvenile release information because the Nason Creek spring Chinook program started with return year 2013. Juveniles from the 2013 brood will be released in 2015 (Table 6.7).

**Table 6.7.** Number of days spring Chinook broods were acclimated and water source, brood year 2013; NA = not available.

Brood year	Release year	Transfer date	Release date	Number of days and water source	
				Total	Nason Creek
2013	2015	NA	NA	NA	NA

**Release Information**

**Numbers released**

There is currently no juvenile release information because the Nason Creek spring Chinook program started with return year 2013. Juveniles from the 2013 brood will be released in 2015 (Table 6.8).

**Table 6.8.** Numbers of spring Chinook smolts tagged and released from the hatchery, brood year 2013. The release target for Nason Creek spring Chinook is 125,000 smolts.

Brood year	Release year	Type of release	CWT mark rate	Number released that were PIT tagged	Number of smolts released	Total number of smolts released
2013	2015	NA	NA	NA	NA	NA

**Numbers tagged**

In 2014, a total of 20,234 Nason Creek spring Chinook from the 2013 brood were PIT tagged at Eastbank Hatchery on 18-22 August (Table 6.9). These fish were tagged in raceway #6. Fish were not fed during tagging or for two days before and after tagging. Fish averaged 95 mm in length and 11.0 g at time of tagging. These fish were transferred to the Nason Creek Acclimation Facility in October.

**Table 6.9.** Summary of PIT-tagging activities for Nason Creek hatchery spring Chinook, brood year 2013.

Brood year	Release year	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2013	2015	20,234	NA	NA	NA

**Fish size and condition at release**

There is currently no juvenile release information because the Nason Creek spring Chinook program started with return year 2013. Lengths and weights of juvenile spring Chinook from the 2013 brood will be measured in 2015 (Table 6.10).

**Table 6.10.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of spring Chinook smolts released from the hatchery, brood year 2013. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
2013	2015	NA	NA	NA	NA
<i>Average</i>		<i>NA</i>	<i>NA</i>	<i>NA</i>	<i>NA</i>
<i>Targets</i>		<i>155</i>	<i>9.0</i>	<i>37.8</i>	<i>24</i>

**Survival Estimates**

There is currently limited juvenile survival information because the Nason Creek spring Chinook program started with return year 2013. Survival of juveniles from the 2013 brood will be assessed in 2015 (Table 6.11).

**Table 6.11.** Hatchery life-stage survival rates (%) for spring Chinook, brood year 2013. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
2013	100.0	100.0	96.2	93.0	99.4	98.1	NA	NA	NA
<i>Average</i>	<i>100.0</i>	<i>100.0</i>	<i>96.2</i>	<i>93.0</i>	<i>99.4</i>	<i>98.1</i>	<i>NA</i>	<i>NA</i>	<i>NA</i>
<i>Standard</i>	<i>90.0</i>	<i>85.0</i>	<i>92.0</i>	<i>98.0</i>	<i>97.0</i>	<i>93.0</i>	<i>90.0</i>	<i>95.0</i>	<i>81.0</i>

**6.3 Disease Monitoring**

Results of 2014 adult broodstock bacterial kidney disease (BKD) monitoring indicated that most females (80%) had ELISA values less than 0.199. About 80% of the females had ELISA values less than 0.120, which would have required about 20% of the progeny to be reared at densities not to exceed 0.06 fish per pound (Table 6.12).

**Table 6.12.** Proportion of bacterial kidney disease (BKD) titer groups for the Nason Creek spring Chinook broodstock, brood years 2013-2014. Also included are the proportions to be reared at either 0.125 fish per pound or 0.060 fish per pound.

Brood year	Optical density values by titer group								Proportion at rearing densities (fish per pound, fpp)			
	Very Low (≤ 0.099)		Low (0.1-0.199)		Moderate (0.2-0.449)		High (≥ 0.450)		≤ 0.125 fpp (<0.119)		≤ 0.060 fpp (>0.120)	
	Wild	Hatch	Wild	Hatch	Wild	Hatch	Wild	Hatch	Wild	Hatch	Wild	Hatch
2013	0.7000	0.3333	0.3000	0.6666	--	--	--	--	0.9231	0.1000	0.0769	--
2014 <sup>a</sup>	0.5000	--	0.3000	--	--	--	0.2000	--	0.8000	--	0.2000	--
<i>Average</i>	0.6000	0.3333	0.3000	0.6666	--	--	0.2000	--	0.8616	0.1000	0.1385	--

<sup>a</sup>Origin determinations should be considered preliminary pending scale analyses.

## 6.4 Natural Juvenile Productivity

During 2014, juvenile spring Chinook were sampled at the Nason Creek trap.

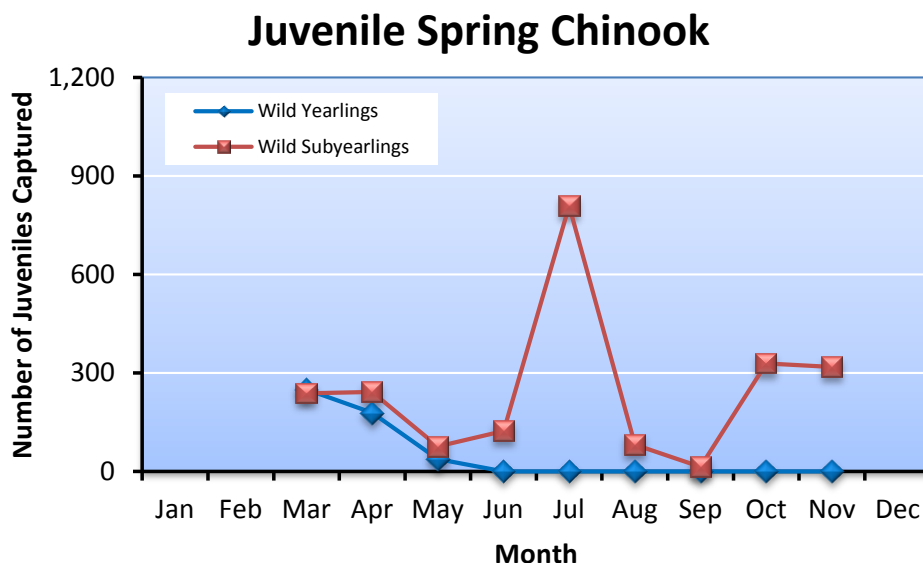
### Smolt and Emigrant Estimates

Numbers of spring Chinook smolts and emigrants were estimated at the Nason Creek trap in 2014. A complete description of trapping operations on Nason Creek can be found in Appendix K.

#### Nason Creek Trap

The Nason Creek Trap operated between 1 March and 30 November 2014. During that time period the trap was inoperable for 48 days because of low stream discharge or ice accumulation. Daily trap efficiencies were estimated from a flow-efficiency regression model. The daily number of fish captured was expanded by the estimated trap efficiency to estimate daily total emigration. In the event that a viable flow-efficiency regression could not be developed, a pooled efficiency was used to expand daily catch.

Wild yearling spring Chinook (2012 brood year) were primarily captured from March through May 2014 (Figure 6.1). Based on capture efficiencies estimated from the flow model, the total number of wild yearling Chinook emigrating from Nason Creek was 4,561 (±1,540). Combining the total number of subyearling spring Chinook (28,110) that emigrated during the fall of 2013 with the total number of yearling Chinook (4,561) that emigrated during 2014 resulted in a total emigrant estimate of 32,671 (±4,863) spring Chinook for the 2012 brood year (Table 6.13).



**Figure 6.1.** Monthly captures of wild subyearling and yearling spring Chinook at the Nason Creek Trap, 2014.

**Table 6.13.** Numbers of redds and juvenile spring Chinook at different life stages in the Nason Creek basin for brood years 2002-2013; ND = no data.

Brood year	Number of redds	Egg deposition <sup>a</sup>	Number of subyearling emigrants <sup>b</sup>	Number of smolts produced within Nason Creek basin	Number of emigrants
2002	294	1,368,276	ND	4,683	ND
2003	83	485,052	8,829	6,358	15,187
2004	169	811,031	11,822	2,597	14,419
2005	193	835,111	11,814	8,696	20,510
2006	152	657,248	4,144	7,798	11,942
2007	101	448,541	15,556	5,679	21,235
2008	336	1,542,912	23,182	3,611	26,793
2009	167	763,691	27,720	1,705	29,425
2010	188	811,032	8,491	3,535	12,026
2011	170	745,450	17,991	2,422	20,413
2012	413	1,744,099	28,110	4,561	32,671
2013	212	999,792	30,078	--	--
<b>Average</b>	<b>207</b>	<b>934,353</b>	<b>17,067</b>	<b>4,695</b>	<b>20,462</b>

<sup>a</sup> Egg deposition is calculated as the number of redds times the fecundity of both wild and hatchery spring Chinook salmon (from Table 5.5).

<sup>b</sup> Subyearling emigrants does not include fry that left the watershed before 1 July.

Wild subyearling spring Chinook (2013 brood year) were captured between 3 March and 30 November 2014. Because a viable flow-efficiency regression model could not be established at the new downstream trap location (July-November), a pooled estimate was employed as a

temporary method of expansion. Based on this pooled efficiency, the total number of wild subyearling Chinook from the Nason Creek basin was 30,078 ( $\pm 32,238$ ).

Yearling spring Chinook sampled in 2014 averaged 90 mm in length, 7.5 g in weight, and had a mean condition of 1.03 (Table 6.14). These size estimates were less than the overall mean of yearling spring Chinook sampled in previous years (overall means, 92 mm, 8.5 g, and condition of 1.06). Subyearling spring Chinook sampled in 2014 at the Nason Creek Trap averaged 69 mm in length, averaged 3.8 g, and had a mean condition of 1.05 (Table 6.14). These sizes were less than the overall mean of subyearling spring Chinook sampled in previous years (overall means, 75 mm, 4.9 g, and condition of 1.06).

**Table 6.14.** Mean fork length (mm), weight (g), and condition factor of subyearling and yearling spring Chinook collected in the Nason Creek Trap, 2004-2014. Numbers in parentheses indicate 1 standard deviation.

Sample year	Life stage	Sample size <sup>a</sup>	Mean size		
			Length (mm)	Weight (g)	Condition (K)
2004	Subyearling	656	82 (7)	5.9 (1.7)	1.04 (0.11)
	Yearling	323	92 (8)	8.2 (2.3)	1.04 (0.08)
2005	Subyearling	872	76 (9)	4.8 (1.7)	1.02 (0.13)
	Yearling	276	94 (7)	8.7 (2.0)	1.04 (0.12)
2006	Subyearling	1422	73 (9)	3.9 (1.9)	0.92 (0.16)
	Yearling	362	91 (7)	7.5 (1.8)	0.98 (0.11)
2007	Subyearling	609	78 (14)	5.9 (2.6)	1.15 (0.16)
	Yearling	678	88 (9)	7.4 (2.4)	1.05 (0.13)
2008	Subyearling	1,001	75 (14)	5.0 (2.5)	1.10 (0.11)
	Yearling	881	96 (6)	9.5 (2.0)	1.06 (0.09)
2009	Subyearling	2,147	72 (11)	4.4 (2.1)	1.08 (0.08)
	Yearling	162	96 (8)	9.6 (2.4)	1.08 (0.09)
2010	Subyearling	3,032	81 (11)	6.2 (2.3)	1.13 (0.10)
	Yearling	366	97 (7)	10.2 (2.3)	1.10 (0.09)
2011	Subyearling	1,064	72 (13)	4.7 (2.5)	1.13 (0.12)
	Yearling	150	89 (10)	7.7 (1.8)	1.09 (0.12)
2012	Subyearling	2,141	78 (11)	5.3 (2.0)	1.05 (0.09)
	Yearling	363	93 (6)	9.3 (2.2)	1.11 (0.08)
2013	Subyearling	4,408	70 (11)	3.8 (1.7)	1.03 (0.10)
	Yearling	239	91 (7)	7.9 (2.1)	1.03 (0.07)
2014	Subyearling	1,543	69 (12)	3.8 (2.3)	1.05 (0.06)
	Yearling	464	90 (7)	7.5 (1.8)	1.03 (0.06)
<i>Average</i>	<i>Subyearling</i>	<i>1,718</i>	<i>75 (4)</i>	<i>4.9 (0.9)</i>	<i>1.06 (0.07)</i>
	<i>Yearling</i>	<i>388</i>	<i>92 (3)</i>	<i>8.5 (1.0)</i>	<i>1.06 (0.04)</i>

<sup>a</sup> Sample size represents the number of fish that were measured for both length and weight.



## Freshwater Productivity

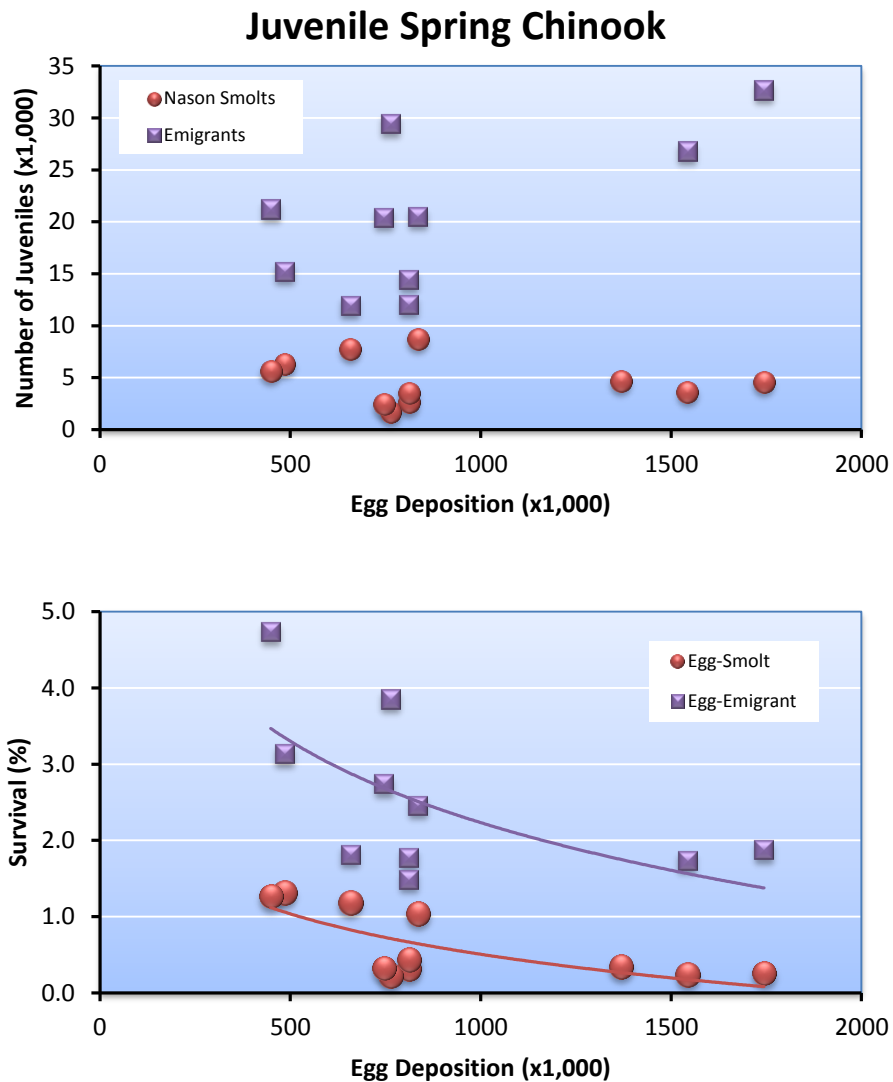
Both productivity and survival estimates for different life stages of spring Chinook in the Nason Creek watershed are provided in Table 6.15. Estimates for brood year 2012 fall within the ranges estimated over the period of brood years 2002-2012. During that period, freshwater productivities ranged from 10-77 smolts/redd and 64-210 emigrants/redd. Survivals during the same period ranged from 0.2-1.3% for egg-smolt and 1.5-4.7% for egg-emigrants.

**Table 6.15.** Productivity (fish/redd) and survival (%) estimates for different juvenile life stages of spring Chinook in the Nason Creek watershed for brood years 2002-2013; ND = no data. These estimates were derived from data in Table 6.13.

Brood year	Smolts/Redd <sup>a</sup>	Emigrants/ Redd	Egg-Smolt <sup>a</sup> (%)	Egg-Emigrant (%)
2002	16	ND	0.3	ND
2003	77	183	1.3	3.1
2004	15	85	0.3	1.8
2005	45	106	1.0	2.5
2006	51	79	1.2	1.8
2007	56	210	1.3	4.7
2008	11	80	0.2	1.7
2009	10	176	0.2	3.9
2010	19	64	0.4	1.5
2011	14	120	0.3	2.7
2012	11	79	0.3	1.9
<i>Average</i>	<i>30</i>	<i>107</i>	<i>0.6</i>	<i>2.3</i>

<sup>a</sup>These estimates include Nason Creek smolts produced only within the Nason Creek basin.

Seeding level (egg deposition) explained most of the variability in productivity and survival of juvenile spring Chinook in the Nason Creek watershed. That is, for estimates based on smolts produced within the Nason Creek watershed, survival and productivity decreased as seeding levels increased (Figure 6.2). This suggests that density dependence regulates juvenile productivity and survival within the Nason Creek watershed.



**Figure 6.2.** Relationships between seeding levels (egg deposition) and juvenile life-stage survivals and productivities for Nason Creek spring Chinook, brood years 2002-2012. Nason Creek smolts are smolts produced only in the Nason Creek watershed.

### 6.5 Spawning Surveys

Surveys for spring Chinook redds were conducted during August through September, 2014, in the Chiwawa River (including Rock, Phelps, Big Meadow, and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek (including Ingalls Creek), Upper Wenatchee River (including Chiwaukum Creek), Little Wenatchee River, and White River (including the Napeequa River and Panther Creek). See Section 5.5 for a complete coverage of spring Chinook redd surveys in the Wenatchee River basin. In the following section we describe the number and distribution of redds within the Nason Creek basin.

#### Redd Counts and Distribution

A total of 115 spring Chinook redds were counted in Nason Creek in 2014 (Table 6.16; see Table 5.19 for the complete time series of redd counts). This is lower than the average of 146 redds

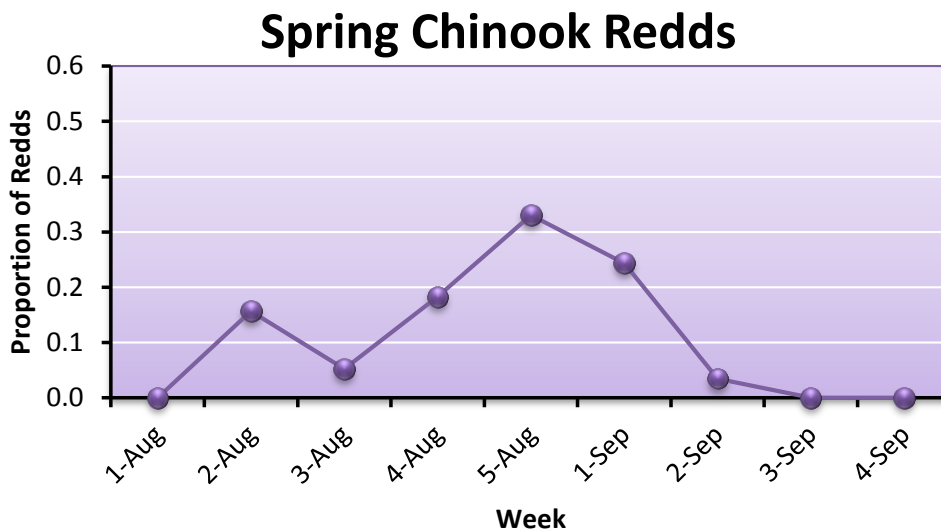
counted during the period 1989-2014 in Nason Creek. Redds were not distributed evenly among the four reaches in Nason Creek. Most were located in Reaches 1, 3, and 4 (Table 6.16).

**Table 6.16.** Numbers and proportions of spring Chinook redds counted within different reaches within Nason Creek during August through September, 2014. See Table 2.8 for description of survey reaches.

Stream/watershed	Reach	Number of redds	Proportion of redds within stream/watershed
Nason	Nason 1 (N1)	42	0.37
	Nason 2 (N2)	11	0.10
	Nason 3 (N3)	36	0.31
	Nason 4 (N4)	26	0.23
<i>Total</i>		<i>115</i>	<i>1.00</i>

### Spawn Timing

Spring Chinook began spawning during the second week of August in Nason Creek and peaked the fifth week of August (Figure 6.3). Spawning in Nason Creek ended the third week of September.



**Figure 6.3.** Proportion of spring Chinook redds counted during different weeks within Nason Creek, August through September 2014.

### Spawning Escapement

Spawning escapement for spring Chinook was calculated as the number of redds times the male-to-female ratio (i.e., fish per redd expansion factor) estimated from broodstock and fish sampled at adult trapping sites. The estimated fish per redd ratio for spring Chinook upstream from Tumwater in 2014 was 2.06 (based on sex ratios estimated at Tumwater Dam). Multiplying this ratio by the number of redds counted in Nason Creek resulted in a total spawning escapement of

237 spring Chinook. The estimated total spawning escapement of spring Chinook in 2014 was less than the overall average of 325 spring Chinook in Nason Creek (see Table 5.22).

## 6.6 Carcass Surveys

Surveys for spring Chinook carcasses were conducted during August through September, 2014, in the Chiwawa River (including Rock, Phelps, and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek, Upper Wenatchee River, Little Wenatchee River, and White River (including the Napeequa River and Panther Creek). In 2014, 68 spring Chinook carcasses were sampled in Nason Creek. Most of these were sampled in Reaches 1 and 3. The number of carcasses sampled in 2014 was less than the overall average of 153 carcasses sampled during the period 1996-2014. See Section 5.6 for a complete coverage of spring Chinook carcass surveys in the Wenatchee River basin.

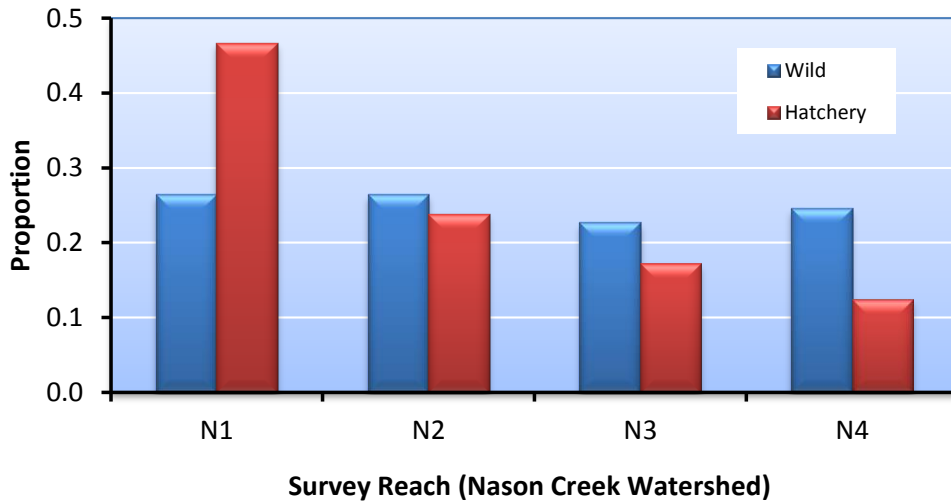
In the Nason Creek watershed, the spatial distribution of hatchery and wild fish was not equal among survey reaches (Table 6.17). In 2014, more wild fish were collected during surveys than hatchery fish (these numbers may change after analysis of CWTs). A similar percentage of hatchery and wild fish were found in the lower reaches (N1 and N2; Mouth to Highway 2). In upstream reaches, more wild fish were observed than hatchery fish. This general trend was also apparent in the pooled data (Figure 6.4). It should be noted that the hatchery fish spawning in Nason Creek are strays from the Chiwawa spring Chinook Program. Nason Creek hatchery fish will return to Nason Creek beginning in 2016 as age-3 fish.

**Table 6.17.** Numbers of wild and hatchery spring Chinook carcasses sampled within different reaches in the Nason Creek watershed, 1999-2014. See Table 2.8 for description of survey reaches.

Survey year	Origin	Survey Reach				Total
		N-1	N-2	N-3	N-4	
1999	Wild	2	3	0	0	5
	Hatchery	0	0	0	0	0
2000	Wild	19	21	0	9	49
	Hatchery	11	9	0	1	21
2001	Wild	25	22	0	41	88
	Hatchery	91	54	0	22	167
2002	Wild	16	34	0	37	87
	Hatchery	33	29	0	35	97
2003	Wild	6	19	0	22	47
	Hatchery	3	9	0	3	15
2004	Wild	29	33	18	24	104
	Hatchery	42	26	11	3	82
2005	Wild	19	6	11	7	43
	Hatchery	130	17	22	4	173
2006	Wild	24	17	28	9	78
	Hatchery	50	31	17	14	112
2007	Wild	2	13	8	6	29
	Hatchery	54	77	26	15	172
2008	Wild	14	13	16	10	53

Survey year	Origin	Survey Reach				Total
		N-1	N-2	N-3	N-4	
	Hatchery	102	39	36	13	190
2009	Wild	1	12	10	16	39
	Hatchery	25	21	20	23	89
2010	Wild	3	6	6	4	19
	Hatchery	47	29	30	16	122
2011	Wild	8	11	11	5	35
	Hatchery	22	12	21	8	63
2012	Wild	24	11	65	7	107
	Hatchery	95	37	70	23	225
2013	Wild	4	2	9	8	23
	Hatchery	51	12	28	27	118
2014	Wild	20	4	12	2	38
	Hatchery	24	2	3	0	29
Average	Wild	14	14	12	13	53
	Hatchery	49	25	18	13	105

### Spring Chinook Carcass Distribution



**Figure 6.4.** Distribution of wild and hatchery produced carcasses in different reaches in the Nason Creek watershed, 1999-2014. Reach codes are described in Table 2.8.

## 6.7 Life History Monitoring

Life history characteristics of spring Chinook were assessed by examining carcasses on spawning grounds and fish collected at broodstock collection sites, and by reviewing tagging data and fisheries statistics.

### Migration Timing

See Section 5.7 for a description of migration timing of spring Chinook at Tumwater Dam.

### Age at Maturity

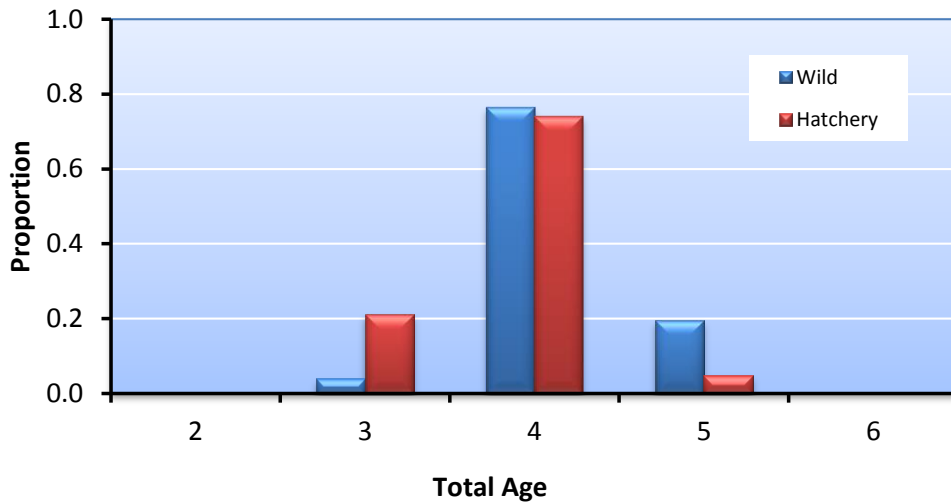
Most of the wild and hatchery spring Chinook sampled during the period 1999-2014 in the Nason Creek watershed were age-4 fish (total age) (Table 6.18; Figure 6.5). Until 2014, hatchery fish made up a higher percentage of age-3 Chinook than did wild fish. As in other years, a higher proportion of age-5 wild fish returned than did age-5 hatchery fish. Thus, wild fish tended to return at an older age than hatchery fish.

**Table 6.18.** Numbers of wild and hatchery spring Chinook of different ages (total age) sampled on spawning grounds in the Nason Creek watershed, 1999-2014.

Sample year	Origin	Total age					Sample size
		2	3	4	5	6	
1999	Wild	0	0	5	0	0	5
	Hatchery	0	0	0	0	0	0
2000	Wild	0	1	45	0	0	46
	Hatchery	0	18	3	0	0	21
2001	Wild	0	0	63	13	0	76
	Hatchery	0	5	159	3	0	167
2002	Wild	0	0	58	23	0	81
	Hatchery	0	0	85	11	0	96
2003	Wild	0	4	3	36	0	43
	Hatchery	0	3	1	5	0	9
2004	Wild	0	1	101	1	0	103
	Hatchery	0	57	23	2	0	82
2005	Wild	0	1	25	17	0	43
	Hatchery	0	3	170	0	0	173
2006	Wild	0	0	60	18	0	78
	Hatchery	0	12	78	22	0	112
2007	Wild	0	0	18	11	0	29
	Hatchery	0	123	40	9	0	172
2008	Wild	0	2	46	4	0	52
	Hatchery	0	21	163	6	0	190
2009	Wild	0	1	36	2	0	39
	Hatchery	0	19	65	4	0	88
2010	Wild	0	1	18	0	0	19
	Hatchery	0	5	116	1	0	122
2011	Wild	0	3	24	8	0	35
	Hatchery	0	33	17	13	0	63
2012	Wild	0	1	89	17	0	107
	Hatchery	0	25	198	2	0	225
2013	Wild	0	0	16	7	0	23
	Hatchery	0	22	92	5	0	119

Sample year	Origin	Total age					Sample size
		2	3	4	5	6	
2014	Wild	0	12	17	2	0	31
	Hatchery	0	8	19	0	0	27
Average	Wild	0	2	39	10	0	51
	Hatchery	0	22	77	5	0	104

### Spring Chinook Age Structure



**Figure 6.5.** Proportions of wild and hatchery spring Chinook of different total ages sampled on spawning grounds in the Nason Creek watershed for the combined years 1999-2014.

### Size at Maturity

On average, hatchery and wild spring Chinook of a given age differed little in length (Table 6.19). Differences were usually no more than 1-3 cm between hatchery and wild fish of the same age.

**Table 6.19.** Mean lengths (POH in cm;  $\pm 1SD$ ) and sample sizes (in parentheses) of different ages (total age) of male and female spring Chinook of wild and hatchery-origin sampled in the Nason Creek watershed, 1999-2014.

Return year	Total age	Mean length (cm)			
		Male		Female	
		Wild	Hatchery	Wild	Hatchery
1999	3	0	0	0	0
	4	71 $\pm$ 2 (2)	0	64 $\pm$ 2 (3)	0
	5	0	0	0	0
	6	0	0	0	0
2000	3	46 $\pm$ 0 (1)	44 $\pm$ 4 (14)	0	52 $\pm$ 10 (4)
	4	62 $\pm$ 4 (19)	0	63 $\pm$ 3 (25)	60 $\pm$ 1 (3)
	5	0	0	0	0

Return year	Total age	Mean length (cm)			
		Male		Female	
		Wild	Hatchery	Wild	Hatchery
	6	0	0	0	0
2001	3	0	47 ±12 (5)	0	0
	4	65 ±4 (21)	66 ±5 (36)	63 ±4 (42)	63 ±4 (123)
	5	81 ±5 (3)	0	72 ±3 (10)	71 ±7 (3)
	6	0	0	0	0
2002	3	0	0	0	0
	4	62 ±6 (24)	66 ±5 (35)	63 ±4 (34)	62 ±5 (50)
	5	77 ±4 (12)	81 ±7 (8)	75 ±3 (11)	71 ±5 (3)
	6	0	0	0	0
2003	3	44 ±7 (3)	43 ±5 (3)	0	0
	4	58 ±7 (2)	79 ±0 (1)	67 ±0 (1)	0
	5	75 ±9 (11)	81 ±6 (2)	72 ±6 (25)	71 ±2 (3)
	6	0	0	0	0
2004	3	46 ±0 (1)	43 ±4 (56)	0	0
	4	61 ±4 (35)	60 ±3 (6)	61 ±3 (66)	62 ±4 (17)
	5	0	0	81 ±0 (1)	73 ±4 (2)
	6	0	0	0	0
2005	3	37 ±0 (1)	41 ±7 (3)	0	0
	4	59 ±6 (8)	63 ±4 (54)	61 ±3 (17)	61 ±3 (116)
	5	73 ±5 (4)	0	71 ±1 (13)	0
	6	0	0	0	0
2006	3	0	41 ±3 (12)	0	0
	4	60 ±5 (26)	62 ±3 (29)	61 ±3 (34)	59 ±4 (49)
	5	72 ±5 (10)	73 ±5 (6)	69 ±4 (8)	70 ±4 (16)
	6	0	0	0	0
2007	3	0	44 ±4 (122)	0	51 ±0 (1)
	4	62 ±4 (6)	60 ±7 (13)	63 ±4 (12)	61 ±4 (27)
	5	77 ±5 (7)	67 ±5 (3)	68 ±2 (4)	70 ±2 (6)
	6	0	0	0	0
2008	3	51 ±21 (2)	45 ±5 (20)	0	45 ±0 (1)
	4	60 ±5 (15)	63 ±4 (42)	61 ±3 (31)	63 ±3 (121)
	5	0	77 ±2 (3)	71 ±3 (4)	64 ±7 (3)
	6	0	0	0	0
2009	3	41 ±0 (1)	46 ±5 (18)	0	65 ±0 (1)
	4	60 ±5 (12)	63 ±4 (19)	60 ±3 (24)	61 ±4 (46)
	5	0	71 ±1 (2)	72 ±4 (2)	73 ±3 (2)
	6	0	0	0	0
2010	3	44 ±0 (1)	45 ±5 (5)	0	0
	4	62 ±5 (7)	63 ±4 (42)	61 ±3 (10)	62 ±4 (74)
	5	0	75 ±0 (1)	0	0
	6	0	0	0	0



Return year	Total age	Mean length (cm)			
		Male		Female	
		Wild	Hatchery	Wild	Hatchery
2011	3	48 ±11 (3)	43 ±4 (31)	0	48 ±2 (2)
	4	61 ±5 (11)	59 ±11 (6)	60 ±5 (12)	63 ±5 (11)
	5	79 ±2 (3)	73 ±3 (6)	75 ±4 (5)	70 ±3 (7)
	6	0	0	0	0
2012	3	41 ±0 (1)	42 ±3 (24)	0	0
	4	61 ±7 (35)	60 ±5 (45)	61 ±4 (54)	60 ±4 (151)
	5	77 ±4 (6)	0	66 ±5 (11)	70 ±3 (2)
	6	0	0	0	0
2013	3	0	42 ±4 (21)	0	0
	4	60 ±6 (5)	62 ±4 (23)	60 ±4 (10)	60 ±4 (69)
	5	71 ±0 (1)	75 ±0 (1)	68 ±3 (6)	70 ±4 (4)
	6	0	0	0	0
2014	3	44 ±5 (12)	47 ±5 (8)	0	0
	4	67 ±4 (6)	58 ±3 (8)	63 ±3 (11)	60 ±3 (11)
	5	0	0	65 ±1 (2)	0
	6	0	61 ±8 (2)	69 ±13 (2)	0

### Contribution to Fisheries

Because the Nason Creek program began in 2013, there will be no harvest information on Nason Creek hatchery spring Chinook until about 2017.

### Straying

Stray rates will be determined by examining CWTs and PIT tags recovered on spawning grounds within and outside the Wenatchee River basin. Targets for strays based on return year (recovery year) within the Wenatchee River basin should be less than 10% and targets for strays outside the Wenatchee River basin should be less than 5%. The target for brood year stray rates should be less than 5%. Straying of Nason Creek spring Chinook will be estimated beginning in 2016 or 2017 when the 2013 brood fish return.

### Genetics

Because the Nason Creek spring Chinook program began in 2013 with the collection of broodstock, there are no studies that examine the effects of the program on the genetics of natural-origin spring Chinook in the Wenatchee River basin. However, genetic studies were conducted to determine the potential effects of the Chiwawa Supplementation Program on natural-origin spring Chinook in the upper Wenatchee River basin (Blankenship et al. 2007; the entire report is appended as Appendix J). This work included the analysis of Nason Creek spring Chinook. Researchers collected microsatellite DNA allele frequencies from temporally replicated natural and hatchery-origin spring Chinook to statistically assign individual fish to specific demes (locations) within the Wenatchee population.

Significant differences in allele frequencies were observed within and among major spawning areas in the Upper Wenatchee River basin. However, these differences made up only a very

small portion of the overall variation, indicating genetic similarity among the major spawning areas. There was no evidence that the Chiwawa program has changed the genetic structure (allele frequency) of spring Chinook in Nason Creek and the White River, despite the presence of hatchery-origin spawners in both systems.

### Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For brood years 1989-2012, when no brood stock were collected for the Nason Creek Program, the approximate PNI Index values ranged from 0.26 to 1.00 (Table 6.20). During this period, PNI Index values varied over time because of Chiwawa spring Chinook straying into Nason Creek. For brood years 2013-2014, a period when brood stock was collected for the Nason Creek Program, approximate PNI Index for the Nason Creek Program ( $PNI_N$ ) was 1.00. If hatchery strays are included in the PNI Index calculation ( $PNI_{N+S}$ ), values were less than 0.67 (Table 6.20).

**Table 6.20.** Approximate Proportionate Natural Influence (PNI) Index of hatchery spring Chinook spawning in Nason Creek. See notes below the table for description of each metric.

Brood year	Spawners					Broodstock			$PNI_N$	$PNI_{N+S}$
	NOS	$HOS_N$	$HOS_S$	$pHOS_N$	$pHOS_{N+S}$	$NOB_N$	$HOB_N$	pNOB		
1989	222	0	0	0.00	0.00	0	0	1.00	--	1.00
1990	231	0	0	0.00	0.00	0	0	1.00	--	1.00
1991	156	0	0	0.00	0.00	0	0	1.00	--	1.00
1992	181	0	0	0.00	0.00	0	0	1.00	--	1.00
1993	430	0	61	0.00	0.12	0	0	1.00	--	0.89
1994	60	0	0	0.00	0.00	0	0	0.67	--	1.00
1995	18	0	0	0.00	0.00	0	0	0.00	--	1.00
1996	58	0	25	0.00	0.30	0	0	0.44	--	0.59
1997	67	0	55	0.00	0.45	0	0	0.29	--	0.39
1998	61	0	3	0.00	0.05	0	0	0.28	--	0.85
1999	22	0	0	0.00	0.00	0	0	0.00	--	1.00
2000	189	0	81	0.00	0.30	0	0	0.30	--	0.50
2001	257	0	341	0.00	0.57	0	0	0.30	--	0.34
2002	313	0	290	0.00	0.48	0	0	0.28	--	0.37
2003	152	0	50	0.00	0.25	0	0	0.44	--	0.64
2004	297	0	210	0.00	0.41	0	0	0.39	--	0.49
2005	81	0	266	0.00	0.77	0	0	0.33	--	0.30

Brood year	Spawners					Broodstock			PNI <sub>N</sub>	PNI <sub>N+S</sub>
	NOS	HOS <sub>N</sub>	HOS <sub>S</sub>	pHOS <sub>N</sub>	pHOS <sub>N+S</sub>	NOB <sub>N</sub>	HOB <sub>N</sub>	pNOB		
2006	117	0	154	0.00	0.57	0	0	0.29	--	0.34
2007	83	0	380	0.00	0.82	0	0	0.29	--	0.26
2008	139	0	426	0.00	0.75	0	0	0.27	--	0.26
2009	163	0	371	0.00	0.69	0	0	0.46	--	0.40
2010	59	0	351	0.00	0.86	0	0	0.44	--	0.34
2011	250	0	452	0.00	0.64	0	0	0.46	--	0.42
2012	220	0	474	0.00	0.68	0	0	1.00	--	0.60
<b>Average*</b>	<b>159</b>	<b>0</b>	<b>166</b>	<b>0.00</b>	<b>0.36</b>	<b>0</b>	<b>0</b>	<b>0.50</b>	<b>--</b>	<b>0.62</b>
2013	70	0	339	0.00	0.83	21	4	0.84	1.00	0.55
2014	169	0	68	0.00	0.29	21	0	1.00	1.00	0.52
<b>Average**</b>	<b>120</b>	<b>0</b>	<b>204</b>	<b>0.00</b>	<b>0.56</b>	<b>21</b>	<b>2</b>	<b>0.92</b>	<b>1.00</b>	<b>0.54</b>

HOS<sub>N</sub> = hatchery-origin spawners in Nason Creek from the Nason Creek spring Chinook Supplementation Program.

pHOS<sub>N</sub> = proportion of hatchery-origin spawners from Nason Creek spring Chinook Supplementation Program.

HOS<sub>S</sub> = stray hatchery-origin spawners in Nason Creek.

pHOS<sub>S</sub> = proportion of stray hatchery-origin spawners.

NOB<sub>N</sub> = natural-origin broodstock spawned in the Nason Creek spring Chinook Supplementation Program.

HOB<sub>N</sub> = hatchery-origin broodstock spawned in the Nason Creek spring Chinook Supplementation Program.

pNOB = proportion of hatchery-origin broodstock. Because of the high incidence of strays to Nason Creek from the Chiwawa River spring Chinook program, pNOB values from the Chiwawa program were used to estimate PNI values during the period from 1989 to 2012 (*italicized*). The weighting for those years was 100% based on the Chiwawa program broodstock selection, because there have been no hatchery returns from the Nason Creek spring Chinook program (see Table 5.1 for Chiwawa broodstock selection).

PNI<sub>N</sub> = pNOB/(pNOB + pNOS<sub>N</sub>); where pNOB is weighted 100% toward broodstock collection from the Nason Creek spring Chinook Supplementation Program. The purpose for this PNI value is to track the performance of the Nason Creek Program apart from the influence of strays and broodstock collection outside the Nason Creek watershed.

PNI<sub>N+S</sub> = pNOB/(pNOB + pHOS<sub>N+S</sub>); where pNOB is weighted by the proportion of HOS<sub>N</sub> and HOS<sub>S</sub> observed in Nason Creek. This PNI value tracks the combined influence of broodstock selection from the Nason Program and/or Chiwawa Program according to the proportion of HOS that return to Nason Creek from those programs.

\* Average for the period 1989-2012, a period when no brood stock were collected for the Nason Creek Program.

\*\* Average for the period 2013-present, a period when brood stock was collected for the Nason Creek Program.

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). We calculated NORs with and without harvest. NORs without harvest include all returning fish that either returned to the basin or were collected as wild broodstock. NORs with harvest include all fish harvested and are based on brood-year harvest rates from the Chiwawa Hatchery program. For brood years 1989-2008, NRR for spring Chinook in Nason Creek averaged 0.90 (range, 0.05-5.48) if harvested fish were not include in the estimate and 0.99 (range, 0.05-5.84) if harvested fish were included in the estimate (Table 6.21). NRRs for more recent brood years will be calculated as soon as all tag recoveries and sampling rates have been loaded into the database.

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and will be calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. These rates should be greater than the NRRs. HRRs will be calculated beginning with the return of 2013 brood fish.

**Table 6.21.** Spawning escapements, natural-origin recruits (NOR), and natural replacement rates (NRR; with and without harvest) for spring Chinook in the Nason Creek watershed, brood years 1989-2008.

Brood year	Spawning Escapement	Harvest not included		Harvest included	
		NOR	NRR	NOR	NRR
1989	222	171	0.77	249	1.12
1990	231	15	0.06	18	0.08
1991	156	21	0.13	23	0.15
1992	181	47	0.26	49	0.27
1993	491	133	0.27	137	0.28
1994	60	3	0.05	3	0.05
1995	18	22	1.22	23	1.28
1996	83	229	2.76	250	3.01
1997	122	306	2.51	339	2.78
1998	64	351	5.48	374	5.84
1999	22	14	0.64	15	0.68
2000	270	337	1.25	354	1.31
2001	598	77	0.13	79	0.13
2002	603	123	0.20	128	0.21
2003	202	63	0.31	67	0.33
2004	507	131	0.26	141	0.28
2005	347	155	0.45	161	0.46
2006	271	118	0.44	149	0.55
2007	463	209	0.45	254	0.55
2008	565	239	0.42	270	0.48
<i>Average</i>	<i>274</i>	<i>138</i>	<i>0.90</i>	<i>154</i>	<i>0.99</i>

### Smolt-to-Adult Survivals

Smolt-to-adult survival ratios (SARs) will be calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery smolts released. SARs will be calculated with the return of the 2013 brood fish.

## 6.8 ESA/HCP Compliance

### Broodstock Collection

The first broodstock were collected in 2013.

### Hatchery Rearing and Release

The first broodstock were collected in 2013.

### Hatchery Effluent Monitoring

Per ESA Permits 1196, 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination

Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January through 31 December 2014. NPDES monitoring and reporting for PUD Hatchery Programs during 2014 are provided in Appendix F.

### Smolt and Emigrant Trapping

Per ESA Section 10 Permit No. 1196, 18118, 18120, and 18121 the permit holders are authorized a direct take of 20% of the emigrating spring Chinook population during juvenile emigration monitoring and a lethal take not to exceed 2% of the fish captured (NMFS 2003). Based on the estimated wild spring Chinook population (smolt trap expansion) and hatchery juvenile spring Chinook population estimate (hatchery release data) for the Wenatchee River basin, the reported spring Chinook encounters during 2013 emigration monitoring complied with take provisions in the Section 10 permit. Spring Chinook encounter and mortality rates for each trap site (including PIT tag mortalities) are detailed in Table 6.22. Additionally, juvenile fish captured at the trap locations were handled consistent with provisions in ESA Section 10 Permit 1196, 18118, 18120, and 18121, Section B. Table 6.22 does not include incidental or direct take associated with the Nason Creek smolt trap operated by the Yakama Nation.

**Table 6.22.** Estimated take of Upper Columbia River spring Chinook resulting from juvenile emigration monitoring in the Wenatchee River basin, 2014.

Trap location	Population estimate			Number trapped			Total	Take allowed under Permit
	Wild <sup>a</sup>	Hatchery <sup>b</sup>	Sub-yearling <sup>c</sup>	Wild	Hatchery	Sub-yearling		
<b>Chiwawa Trap</b>								
Population	34,334	222,504	73,695	4,519	5,293	23,755	33,567	
Encounter rate	NA	NA	NA	0.1316	0.0237	0.3223	0.1016	0.20
Mortality <sup>c</sup>	NA	NA	NA	28	0	84	112	
Mortality rate	NA	NA	NA	0.0062	0.0000	0.0035	0.0033	0.02
<b>Lower Wenatchee Trap</b>								
Population	67,973	222,504	11,936,928	1,700	31,290	81,445	114,435	
Encounter rate	NA	NA	NA	0.025	0.1406	0.0068	0.0096	0.20
Mortality <sup>d</sup>	NA	NA	NA	17	12	250	279	
Mortality rate	NA	NA	NA	0.0100	0.0004	0.0031	0.0024	0.02
<b>Wenatchee River Basin Total</b>								
Population	67,393	222,504	11,936,928	6,219	36,583	105,200	148,002	
Encounter rate	NA	NA	NA	0.0922	0.1644	0.0088	0.0121	0.20
Mortality <sup>d</sup>	NA	NA	NA	45	12	84	141	
Mortality rate	NA	NA	NA	0.0072	0.0003	0.0008	0.0010	0.02

<sup>a</sup> Smolt population estimate derived from juvenile emigration trap data.

<sup>b</sup> 2012 BYsmolt release data for the Wenatchee River basin.

<sup>c</sup> Based on size, date of capture and location of capture, subyearling Chinook encountered at the Lower Wenatchee Trap are categorized as summer Chinook..

<sup>d</sup> Combined trapping and PIT tagging mortality.

### Spawning Surveys

Spring Chinook spawning ground surveys were conducted in the Wenatchee River basin during 2014, as authorized by ESA Section 10 Permits 18118, 18119, and 18121. Because of the

difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical, and extreme caution was used to avoid established redds when wading was required.

### **Spring Chinook Reproductive Success Study**

ESA Section 10 Permit 1196 (expired) and new Section 10 Permits 18118, 18119, and 18121 specifically provide authorization to capture, anesthetize, biologically sample, PIT tag, and release adult spring Chinook at Tumwater Dam for reproductive success studies and general program monitoring. During 2010 through 2014, all spring Chinook passing Tumwater Dam were enumerated, anesthetized, biologically sampled, PIT tagged, and released (not including hatchery-origin and natural-origin Chinook retained for broodstock) as a component of the reproductive success study (BPA Project No. 2003-039-00). Please refer to Ford et al. (2010, 2011, 2012, 2013, and 2014) for complete details on the methods and results of the spring Chinook reproductive success study for the period 2010-2014.

## SECTION 7: WHITE RIVER SPRING CHINOOK

The White River spring Chinook salmon captive brood program began in 1997 with goals to conserve, aid in the recovery, and prevent the extinction of naturally spawning spring Chinook in the White River, and to meet the mitigation responsibilities of Grant County PUD. Collection of eggs or juveniles from the White River (brood years 1997-2009) made up the first-generation ( $F_1$ ) component of the White River captive brood program. Initially, rearing occurred at AquaSeed in Rochester, Washington, but transitioned to the Little White Salmon National Fish Hatchery near Cook, Washington, in 2006. The  $F_1$  component was reared to maturation and spawned within the hatchery. The resulting progeny ( $F_2$ ) were then reared in the hatchery until final acclimation and release in the upper Wenatchee Basin. The first large release of  $F_2$  juveniles was in 2008. The last release of juveniles from the captive brood program will occur in 2015.

The production goal for the White River captive brood program following the 2013 hatchery recalculation is to release 74,556 yearling smolts into the upper Wenatchee River basin at 18-24 fish per pound. Fish lengths and weights for the recent broods have been manipulated to evaluate different approaches to reduce precocious maturation. All of the fish are marked with CWTs. In addition, since 2008, juvenile spring Chinook have been PIT tagged annually.

Since its inception, the captive brood program has undergone several adaptive changes designed to improve program success. These changes included: (1) use of a pedigree approach to reduce the use of stray fish in the broodstock, (2) transfer of fish from AquaSeed to the Little White Salmon National Fish Hatchery to improve fish quality, (3) injection of hormones into  $F_1$  females to improve maturation of eggs, (4) manipulation of diet and ration for the  $F_2$  fish to reduce precocious maturation of males, (5) use of temporary tanks and natural enclosures during acclimation to improve homing, and (6) trucking fish around Lake Wenatchee to improve survival.

The following information focuses on results from monitoring the White River spring Chinook program. More detailed information on the White River program can be found in Lauer et al. (2012). Information on spring Chinook collected throughout the Wenatchee River basin is presented in Section 5.

### 7.1 Captive Brood Collection

The captive brood program was designed to provide a rapid, short-term demographic boost to the White River spring Chinook spawning aggregate, which was at a high risk of local extinction (Lauer et al. 2012). This section describes the collection of broodstock for the White River program.

#### Brood Collection and Rearing

A primary objective of the White River program was to collect progeny of naturally spawning spring Chinook in the White River. The progeny (eggs or juveniles) make up the first-generation ( $F_1$ ) of the captive brood program. However, strays from the Chiwawa supplementation program made this a challenge. As a result, researchers attempted to identify the origin of spawners on redds in the White River and then focused egg and juvenile collection efforts on those redds that had the highest likelihood of being produced from White River parents. During most years, this

limited the number of redds from which eggs or juveniles could be collected. Starting with brood year 2006, a pedigree approach was adopted to improve the likelihood that eggs or juveniles used in the captive brood program were of White River origin.

During 1997 to 2009, first-generation broodstock for the captive brood program originated from about 10,353 natural-origin eggs and juveniles collected from 122 redds in the White River. Broodstock from brood year 1997 were trapped as parr with nets in the fall of 1998. Broodstock from brood year 2006 were trapped as fry with nets in the spring of 2007. It was assumed that the parr and fry in close proximity of known redds were produced from those redds, and origin was confirmed with pedigree analyses. All other brood years were collected as eggs in the fall using redd pumping techniques. Broodstock collection levels were calculated based on the following assumptions and the known number of suitable redds each year (Tonseth and Maitland 2011):

1. 150,000 smolt target/0.70 (green egg to release survival) = 214,000 green eggs
2. 214,000 green eggs/1,500 eggs per female = 143 females/0.50 (sex ratio) = 286 fish
3. 286 fish/0.30 (eyed egg to maturity survival) = 953 eyed eggs
4. 953 eyed eggs/**X** redds = **Y** eyed-eggs per redd

Eyed eggs or juveniles collected in the White River were transported to Aquaseed (brood years 1997 to 2007) or to the Little White Salmon Hatchery (brood years 2008 to 2009) and reared to adults. Table 7.1 summarizes the collection of eyed eggs or juveniles for the captive brood program.

**Table 7.1.** Numbers of eyed eggs or juvenile brood stock collected for the White River captive brood program, brood years 1997-2009 (2009 was the last year for broodstock collection). Also shown are the number of redds that were sampled for eggs or juveniles and the hatchery in which the fish were reared (LWSFH = Little White Salmon Fish Hatchery); NS = no sample.

Brood year	Number of eyed eggs collected	Number of juvenile Chinook collected	Number of redds sampled	Rearing facility
1997	0	527 (parr)	8	Aquaseed
1998	182	0	4	Aquaseed
1999	NS	NS	NS	--
2000	272	0	NS	Aquaseed
2001	NS	NS	NS	--
2002	167	0	3	Aquaseed
2003	250	0	8	Aquaseed
2004	1,216	0	10	Aquaseed
2005	2,733	0	21	Aquaseed/LWSFH <sup>1</sup>
2006	0	1,487 (fry)	29	Aquaseed/ LWSFH <sup>2</sup>
2007	1,153	0	13	Aquaseed/ LWSFH <sup>3</sup>
2008	933	0	11	LWSFH
2009	1,433	0	15	LWSFH
<b>Average</b>	<b>927</b>	<b>1,007</b>	<b>12</b>	

<sup>1</sup> Fish were transferred on 30 June and 2 July 2008 and 20 January 2009.



<sup>2</sup> Fish were transferred on 21 October and 13 November 2008.

<sup>3</sup> Fish were transferred on 26 September and 21 October 2008.

## 7.2 Hatchery Spawning and Release

### Captive Brood Spawning

As noted above, eyed eggs or juveniles collected in the White River were transported to Aquaseed (for brood years 1997-2007) or to the Little White Salmon Hatchery (for brood years 2008-2009) and reared to adults (Lauver et al. 2012). After rearing broodstock to maturity in captivity, adult spring Chinook are spawned and their progeny are grown to smolt size for release into the White River.

During spawning, eggs and sperm are collected and those gametes are crossed based on a 2x2 factorial spawning matrix. That is, each female is spawned with two males and each male is spawned with two females. Using pedigree analysis, spawning crosses are arranged to maximize genetic diversity. Because incomplete ripening of ova has been an issue in the program, implementation of hormone treatments began in 2011 to facilitate ripening. In addition, following spawning, milt from excess males was collected for cryopreservation. Based on a pilot study, the cryopreserved milt was relatively ineffective at fertilizing eggs, so it was not used widely in the program. There are no plans to use the cryopreserved milt in the future. Table 7.2 shows the ages of first-generation males and females spawned for the captive brood program.

**Table 7.2.** Total ages of first-generation (F<sub>1</sub>) male and female spring Chinook spawned for the White River captive brood program, spawning years 2001-2011; NA = not available.

Spawning year	Sex	Total age				Total
		2	3	4	5	
2001	Female	0	0	3	0	3
	Male	0	2	0	0	2
2002	Female	0	0	4	4	8
	Male	10	0	0	0	10
2003	Female	0	5	0	0	5
	Male	0	2	0	0	2
2004	Female	0	0	2	0	2
	Male	4	0	0	0	4
2005	Female	0	85*	0	0	85
	Male	90	1	0	0	91
2006	Female	2	104	110	0	216
	Male	104	6	0	0	110
2007	Female	0	21	118	1	140
	Male	113	7	0	0	120
2008	Female	0	58	0	0	58
	Male	NA	NA	NA	NA	NA
2009	Female	0	0	119	0	119

Spawning year	Sex	Total age				Total
		2	3	4	5	
	Male	65	54	0	0	119
2010	Female	0	0	42	0	42
	Male	22	23	0	0	45
2011	Female	0	0	0	150	150
	Male	0	148	2	0	150
Average	Female	0	19	36	14	75
	Male	41	24	0	0	65

\* Included some unknown number of second-generation females.

## Release Information

### Numbers released

Several different acclimation and release scenarios have been conducted since 1997. Acclimation scenarios have involved naturalized features such as in-channel enclosures, stream-side tanks supplied with pass-through surface water, and net pens in Lake Wenatchee near the mouth of the White River. Release scenarios have included on-site releases from tanks, in-channel enclosures, and net pens in Lake Wenatchee. In 2010, acclimated fish were towed in net pens to the mouth of the lake and released there. In 2011, tank and net-pen acclimated fish were loaded into transport trucks and released into the Wenatchee River. In addition, subyearling and yearling Chinook with no acclimation have been released from transport trucks directly into Lake Wenatchee and the White River. A total of 944,591 second-generation (F<sub>2</sub>) juvenile spring Chinook have been released from the captive brood program. Table 7.3 summarizes the acclimation and release history of F<sub>2</sub> spring Chinook released into the upper Wenatchee River basin.

**Table 7.3.** Numbers of White River juvenile spring Chinook released and their acclimation histories for brood years 2002-2013.

Brood year	Acclimation site	Acclimation vessel	Number of smolts released	Release scenario	Release date	Number of acclimation days
2002	WR RM 11.5	Tanks	2,589	White River	4/22/2004	17
2003	WR RM 11.5	Tanks	2,096	White River	5/2/2005	47
2004	WR RM 11.5	Tanks	1,639	White River	4/4/2006	0
2005	Lake Wen	Net Pens	69,032	Lake Wen	5/2/2007	34
2006	NA	NA	139,644*	White River	4/17, 4/25/2007	0
	NA	NA	142,033	White River	3/18, 3/20/2008	0
2007	Lake Wen	Net Pens	87,671	Lake Wen	5/5/2009	35-40
	None	None	44,172	Lake Wen	4/1/2009	0
2008	WR Bridge	Eddy Pen	10,156	Escape	~4/12/2010	~10
	Lake Wen	Net Pens	38,400	Mouth of lake	5/5, 5/6/2010	38-41
2009	WR RM 11.5	Side Channel	12,000	Escape	~3/31/2011	~7

Brood year	Acclimation site	Acclimation vessel	Number of smolts released	Release scenario	Release date	Number of acclimation days
	WR RM 11.5	Tanks	10,000	White River	5/12/2011	49
	WR Bridge	Tanks	28,000	White River	5/14/2011	51
	WR Bridge	Tanks		Wen River	5/13/2011	50
	WR Bridge	Eddy Pen	14,596	Escape	~3/27/2011	~3
	Lake Wen	Net Pens	48,000	Wen River	5/14/2011	46
	Lake Wen	Net Pens		Wen River	5/14/2011	44
2010	WR Bridge	Tanks	18,850	Wen River	5/9/2012	44
2011	WR Bridge	Tanks	42,000	Wen & White R	5/6, 5/7, 5/8/13	49, 50, 51
	Lake Wen	Net Pens	105,000	Wen River	5/8, 5/13, 5/14/13	51, 56, 57
2012	WR Bridge	Tanks	42,000	Wen River	5/6/14	50
	Lake Wen	Net Pens	55,713	Wen River	5/8/14	49
2013	WR Bridge	Tanks	31,000	Wen River	5/4/15	56

\* Subyearling release.

**Numbers tagged**

Brood years 2005 and 2007-2013 spring Chinook were tagged with a CWT in their peduncle. None of these fish were adipose fin clipped.<sup>8</sup> Subyearling fish from the 2006 brood year were tagged with half of a CWT in their snouts. Yearling fish from the 2006 brood year were tagged with CWTs in the peduncle. None of these fish were adipose fin clipped. In addition, beginning in 2008 (brood year 2006), 303,207 juvenile spring Chinook have been PIT tagged before release. Table 7.4 identifies the number of second-generation (F<sub>2</sub>) juvenile spring Chinook tagged with PIT tags.

**Table 7.4.** Numbers of second-generation (F<sub>2</sub>) White River spring Chinook smolts tagged and released in the upper Wenatchee River basin, brood years 2002-2013.

Brood year	Acclimation site	Acclimation vessel	Release scenario	CWT mark rate	Number released that were PIT tagged	Number of smolts released
2002	WR RM 11.5	Tanks	White River	0.00	0	2,589
2003	WR RM 11.5	Tanks	White River	0.00	0	2,096
2004	WR RM 11.5	Tanks	White River	0.00	0	1,639
2005	Lake Wen	Net Pens	Lake Wen	1.00	0	69,032
2006	NA	NA	White River	0.00	29,881	139,644*
	NA	NA	White River	0.00		142,033
2007	Lake Wen	Net Pens	Lake Wen	1.00	29,863	87,671

<sup>8</sup> Given that juvenile spring Chinook were tagged with CWTs in the peduncle and were not ad-clipped, it is possible that field crews missed hatchery-origin adults on the spawning grounds because they did not know they were supposed to sample fish with adipose fins. Thus, this bias in carcass sampling may bias derived metrics such as spawning distribution of hatchery and natural-origin fish, spawn timing of hatchery and natural-origin fish, age at maturity, size at maturity, contributions to fisheries, HOR, NOR, HRR, NRR, PNI, straying, and SARs.

Brood year	Acclimation site	Acclimation vessel	Release scenario	CWT mark rate	Number released that were PIT tagged	Number of smolts released
	None	None	Lake Wen	1.00	9,957	44,172
2008	WR Bridge	Eddy Pen	Escape	1.00	38,148	10,156
	Lake Wen	Net Pens	Lake Mouth	1.00		38,400
2009	WR RM 11.5	Side Channel	Escape	1.00	41,886	12,000
	WR RM 11.5	Tanks	White River	1.00		10,000
	WR Bridge	Tanks	White River	1.00		28,000
	WR Bridge	Tanks	Wen River	1.00		14,596
	WR Bridge	Eddy Pen	Escape	1.00		48,000
	Lake Wen	Net Pens	Wen River	1.00		
	Lake Wen	Net Pens	Wen River	1.00		
2010	WR Bridge	Tanks	Wen River	1.00	12,283	18,850
2011	WR Bridge	Tanks	Wen & White	1.00	2,490	42,000
	Lake Wen	Net Pens	Wen River	1.00	51,697	105,000
2012	WR Bridge	Tanks	Wen River	1.00	52,097	42,000
	Lake Wen	Net Pens	Wen River	1.00		55,713
2013	WR Bridge	Tanks	Wen River	1.00	34,905	31,000

### *Fish size and condition at release*

Table 7.5 summarizes the size and condition of second-generation White River juvenile spring Chinook released in the upper Wenatchee River basin.

**Table 7.5.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of second-generation White River (WR) juvenile spring Chinook released in the upper Wenatchee River basin, brood years 2002-2013. Size targets are provided in the last row of the table. NA = not available.

Brood year	Acclimation site	Release scenario	Fork length (mm)		Mean weight	
			Mean	CV	Grams (g)	Fish/pound
2002	WR RM 11.5	White River	NA	NA	NA	NA
2003	WR RM 11.5	White River	166	12.4	53.7	8
2004	WR RM 11.5	White River	207	11.6	117.7	4
2005	Lake Wen	Lake Wen	145	9.7	36.9	31
2006	NA	White River	NA	NA	NA	NA
	NA	White River	NA	NA	NA	NA
2007	Lake Wen	Lake Wen	135	7.8	29.2	29
	None	Lake Wen	NA	NA	NA	NA
2008	WR Bridge	Escape	--	--	--	--
	Lake Wen	Mouth of lake	138	10.0	32.5	14

Brood year	Acclimation site	Release scenario	Fork length (mm)		Mean weight	
			Mean	CV	Grams (g)	Fish/pound
2009	WR RM 11.5	Escape	--	--	--	--
	WR RM 11.5	White River	134	8.7	29.3	16
	WR Bridge	White River	138	9.3	28.6	16
	WR Bridge	Wen River	NA	NA	NA	NA
	WR Bridge	Escape	--	--	--	--
	Lake Wen	Wen River	140	8.9	31.6	14
	Lake Wen	Wen River	142	9.8	39.3	12
2010	WR Bridge	Wen River	125	8.0	22.8	20
2011	WR Bridge	Wen & White	130	8.4	24.1	19
	Lake Wen	Wen River	128	8.2	24.0	19
2012	WR Bridge	Wen River	131	8.1	24.2	18.8
	Lake Wen	Wen River	NA	NA	NA	NA
2013	WR Bridge	Wen River	132	8.7	24.5	19
<i>Average</i>			142	9.3	37.0	17

### Post-Release Survival

We used PIT-tagged fish to estimate survival rates of released second-generation (F<sub>2</sub>) White River spring Chinook smolts to McNary Dam, and smolt to adult ratios (SARs) from release to detection at Bonneville Dam.<sup>9</sup> Based on the available data, post-release survival has been low for fish released into the White River and Lake Wenatchee (Table 7.6). In contrast, survival of fish released in the Wenatchee River tends to be higher than those released in the White River or in Lake Wenatchee. These results suggest that high mortality in Lake Wenatchee may explain why adult returns of program fish have been consistently poor; however, other factors such as high precocious maturation may also contribute to the estimated low survival (e.g., see Ford et al. 2015). Because of uncertain release times for the different release scenarios, travel times could not be calculated for the different groups.

**Table 7.6.** Survival of second-generation (F<sub>2</sub>) White River spring Chinook smolts to McNary Dam and SARs to Bonneville Dam for different release scenarios, brood years 2006-2013. Values in parentheses represent the standard error of the estimate. NA = not available (i.e., not all the fish from the release groups have returned to the Columbia River).

Brood year	Release scenario	Number of Chinook released with PIT tags	Survival to McNary Dam (d)	SAR to Bonneville Dam (%)
2006	White River	29,881	0.037 (0.008)	0.000 (0.000)
2007	Lake Wen Pens	29,863	0.096 (0.010)	0.000 (--)
	Lake Wenatchee	9,957	0.080 (0.015)	0.000 (--)

<sup>9</sup> It is important to point out that because of fish size differences among rearing net pens, tanks, or raceways, fish PIT tagged in one pen, tank, or raceway may not represent untagged fish rearing in other pens, tanks, or raceways.

Brood year	Release scenario	Number of Chinook released with PIT tags	Survival to McNary Dam (d)	SAR to Bonneville Dam (%)
2008	Lake Wenatchee	38,146	0.065 (0.010)	0.001 (0.000)
2009	White and Wenatchee rivers	19,913	0.269 (0.027)	0.002 (0.000)
	White River	21,829	0.055 (0.013)	0.000 (0.000)
2010	Wenatchee River	12,283	0.267 (0.017)	NA
2011	Wenatchee River	2,490	0.385 (0.042)	NA
	White and Wenatchee rivers	51,697	0.434 (0.010)	NA
2012	Wenatchee River	52,440	0.351 (0.013)	NA

## 7.3 Disease Monitoring

### First-Generation Health Maintenance

First-generation ( $F_1$ ) adults are fed an azithromycin-medicated feed in the spring to prevent bacterial kidney disease (BKD), which is a common affliction of spring Chinook salmon. As needed, fish receive a dose of 20 mg/kg of body weight. The fish also receive formalin treatments as needed throughout the year to prevent and treat fungus infections. This is especially important during the pre-spawning period when individual fish are maturing in preparation for spawning. Formalin treatments are conducted three times per week and consist of one hour of flow-through at a concentration of 167 parts per million (ppm).

### Second-Generation Health Maintenance

Following fertilization and initial incubation in September, second-generation ( $F_2$ ) eggs are shocked in October. Eggs are treated with a 1,667 ppm formalin solution in a 15-minute flow-through treatment three times a week to prevent fungus growth. Formalin treatments end after hatching, and water flow is increased from three to five gallons per minute. Dead and deformed fry are removed before relocating the fry to nursery tanks in late January or early February. Fry are then relocated to raceways in July, where they remain until transfer to the White River for acclimation the following March. Coded-wire tagging is typically conducted in July, and PIT tagging occurs the following January or February, just before the fish are transferred to acclimation facilities on the White River in March.

## 7.4 Natural Juvenile Productivity

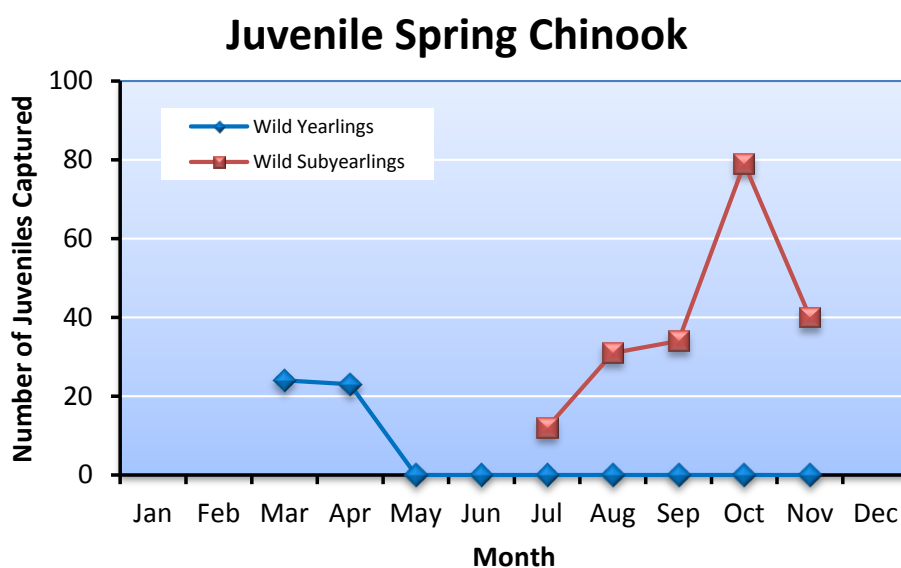
Juvenile productivity estimation began with the monitoring of emigration of spring Chinook in the White River in 2007 (Lauver et al. 2012). A five-foot diameter rotary screw trap is operated annually from about 1 March through November. The purpose of the program is to estimate the number and timing of subyearlings and yearling spring Chinook emigrating from the White River basin.

### Smolt and Emigrant Estimates

In 2014, the White River Trap operated between 1 March and 30 November 2014. During that time period the trap was inoperable for 20 days because of ice or debris accumulation, unsafe working conditions, or for maintenance and repairs. Daily trap efficiencies were estimated by

conducting mark-recapture trials. The daily number of fish captured was expanded by the estimated trap efficiency to estimate daily total emigration. In the event that trap efficiencies could not be assessed because of low numbers of juvenile Chinook trapped, a composite model based on efficiency trials from previous years was used to calculate abundance. Daily captures of fish and results of mark-recapture efficiency tests at the White River trap are reported in Appendix L.

Wild yearling spring Chinook (2012 brood year) were primarily captured from March through April 2014 (Figure 7.1). Based on a composite regression model, the total number of wild yearling Chinook emigrating from the White River was 3,995 ( $\pm 3,616$ ). Combining the total number of subyearling spring Chinook ( $3,905 \pm 1,456$ ) that emigrated during the fall of 2013 with the total number of yearling Chinook (3,995) that emigrated during 2014 resulted in a total emigrant estimate of 7,900 ( $\pm 3,898$ ) spring Chinook for the 2012 brood year (Table 7.7).



**Figure 7.1.** Monthly captures of wild subyearling (parr) and yearling spring Chinook at the White River Trap, 2014.

**Table 7.7.** Numbers of redds and juvenile spring Chinook at different life stages in the White River basin for brood years 2005-2013; ND = no data.

Brood year	Number of redds	Egg deposition <sup>a</sup>	Number of subyearling emigrants <sup>b</sup>	Number of smolts produced within White River basin	Number of emigrants
2005	86	372,122	ND	4,856	ND
2006	31	134,044	642	2,004	2,646
2007	20	88,820	2,293	3,399	5,692
2008	31	142,352	5,552	5,193	10,745
2009	54	246,942	2,485	2,939	5,424
2010	33	142,362	1,859	4,121	5,980
2011	20	87,700	3,128	1,659	4,787

Brood year	Number of redds	Egg deposition <sup>a</sup>	Number of subyearling emigrants <sup>b</sup>	Number of smolts produced within White River basin	Number of emigrants
2012	86	363,178	3,905	3,995	7,900
2013	54	254,664	2,482	--	--
<b>Average<sup>c</sup></b>	<b>46</b>	<b>203,576</b>	<b>2,793</b>	<b>3,521</b>	<b>6,168</b>

<sup>a</sup> Egg deposition is calculated as the number of redds times the fecundity of both wild and hatchery spring Chinook salmon (from Table 5.5).

<sup>b</sup> Subyearling emigrants do not include fry that left the watershed before 1 July.

<sup>c</sup> Average is based on the entire time series of data, not just the period 2006 through 2012.

Wild subyearling spring Chinook (2013 brood year) were captured between 7 July and 25 November 2014, with peak catch during October (Figure 7.1). Based on a composite regression model, the total number of wild subyearling Chinook emigrating from the White River was 2,482 ( $\pm 851$ ).

Yearling spring Chinook sampled in 2014 averaged 94 mm in length, 9.4 g in weight, and had a mean condition of 1.11 (Table 7.8). Length and weight estimates for these fish were less than the overall mean of yearling spring Chinook sampled in previous years (overall means, 99 mm and 10.9 g), while the estimated condition equaled the overall mean (overall mean, 1.11). Subyearling spring Chinook parr sampled in 2014 at the White River Trap averaged 86 mm in length, averaged 7.5 g, and had a mean condition of 1.10 (Table 7.8). Length and weight estimates for these fish were less than the overall mean of subyearling spring Chinook sampled in previous years (overall means, 90 mm and 8.3 g), while the estimated condition was similar to the overall mean (overall mean, 1.09).

**Table 7.8.** Mean fork length (mm), weight (g), and condition factor of subyearling (parr) and yearling spring Chinook collected in the White River Trap, 2007-2014. Numbers in parentheses indicate 1 standard deviation.

Sample year	Life stage	Sample size <sup>a</sup>	Mean size		
			Length (mm)	Weight (g)	Condition (K)
2007	Subyearling	33	95 (12)	9.8 (4.1)	1.07 (0.11)
	Yearling	173	93 (9)	8.6 (2.2)	1.03 (0.09)
2008	Subyearling	202	95 (9)	9.4 (2.5)	1.08 (0.13)
	Yearling	105	100 (12)	11.3 (3.3)	1.07 (0.13)
2009	Subyearling	499	85 (11)	7.1 (2.6)	1.09 (0.11)
	Yearling	274	104 (6)	12.5 (2.6)	1.11 (0.10)
2010	Subyearling	168	87 (13)	7.8 (3.1)	1.12 (0.11)
	Yearling	346	100 (7)	11.2 (2.4)	1.12 (0.09)
2011	Subyearling	145	94 (9)	9.3 (2.5)	1.10 (0.10)
	Yearling	64	99 (8)	11.3 (2.8)	1.14 (0.09)
2012	Subyearling	285	91 (10)	8.9 (2.7)	1.13 (0.09)
	Yearling	179	98 (8)	10.9 (2.8)	1.14 (0.08)
2013	Subyearling	444	84 (12)	6.6 (2.5)	1.05 (0.09)



Sample year	Life stage	Sample size <sup>a</sup>	Mean size		
			Length (mm)	Weight (g)	Condition (K)
2014	Yearling	20	102 (7)	12.3 (3.0)	1.12 (0.14)
	Subyearling	185	86 (14)	7.5 (3.3)	1.10 (0.11)
	Yearling	43	94 (7)	9.4 (2.2)	1.11 (0.13)
Average	Subyearling	245	90 (5)	8.3 (1.2)	1.09 (0.03)
	Yearling	151	99 (4)	10.9 (1.3)	1.11 (0.04)

<sup>a</sup> Sample size represents the number of fish that were measured for both length and weight.

### Freshwater Productivity

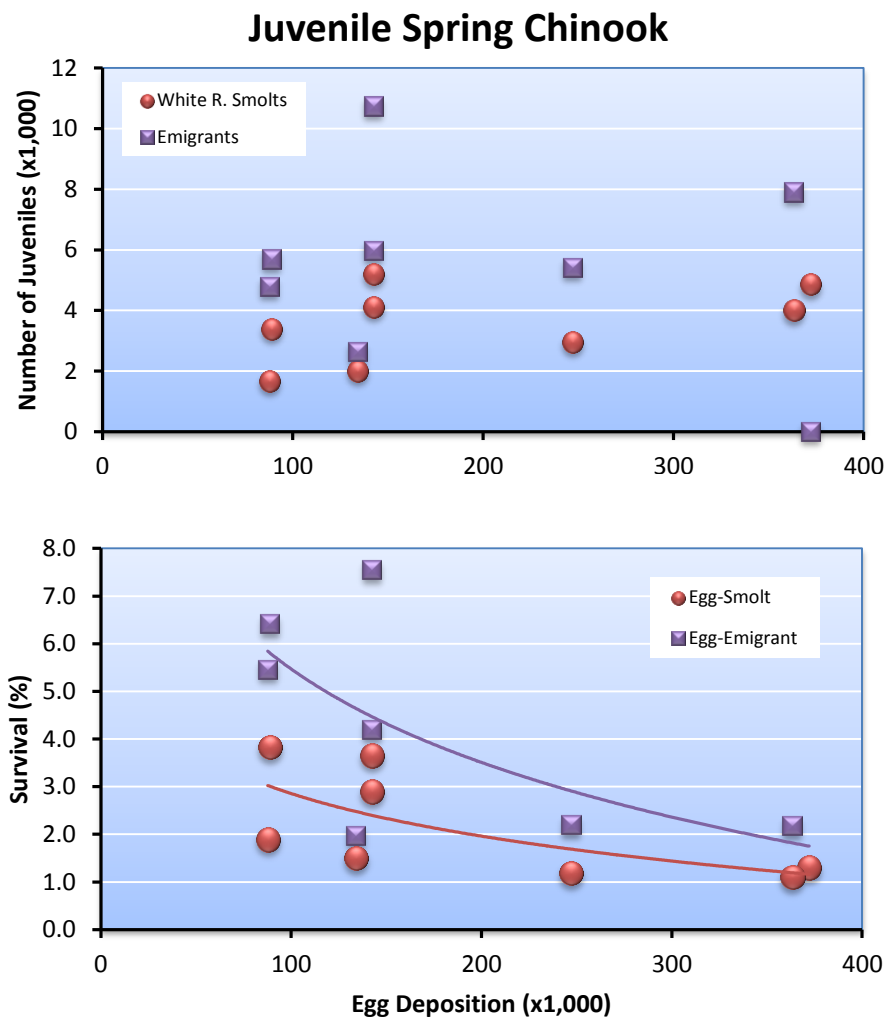
Both productivity and survival estimates for different life stages of spring Chinook in the White River basin are provided in Table 7.9. Estimates for brood year 2012 fall near the lower end of the range of productivity and survival estimates for brood years 2005-2012. During that period, freshwater productivities ranged from 46-170 smolts/redd and 85-347 emigrants/redd. Survivals during the same period ranged from 1.1-3.8% for egg-smolt and 2.0-7.5% for egg-emigrants.

**Table 7.9.** Productivity (fish/redd) and survival (%) estimates for different juvenile life stages of spring Chinook in the White River basin for brood years 2005-2013. These estimates were derived from data in Table 7.7.

Brood year	Smolts/Redd <sup>a</sup>	Emigrants/ Redd	Egg-Smolt <sup>a</sup> (%)	Egg-Emigrant (%)
2005	56	ND	1.3	ND
2006	65	85	1.5	2.0
2007	170	285	3.8	6.4
2008	168	347	3.6	7.5
2009	54	100	1.2	2.2
2010	125	181	2.9	4.2
2011	83	239	1.9	5.5
2012	46	92	1.1	2.2
<b>Average</b>	<b>96</b>	<b>190</b>	<b>2.2</b>	<b>4.3</b>

<sup>a</sup> These estimates include White River smolts produced only within the White River basin.

Seeding level (egg deposition) explained part of the variability in productivity and survival of juvenile spring Chinook in the White River basin. That is, for estimates based on smolts produced within the White River basin, survival and productivity decreased as seeding levels increased (Figure 7.2). This suggests that density dependence in part regulates juvenile productivity and survival within the White River basin.



**Figure 7.2.** Relationships between seeding levels (egg deposition) and juvenile life-stage survivals and productivities for White River spring Chinook, brood years 2005-2012. White River smolts are smolts produced only within the White River basin.

### 7.5 Spawning Surveys

Surveys for spring Chinook redds were conducted during August through September, 2014, in the Chiwawa River (including Rock, Phelps, Big Meadow, and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek (including Ingalls Creek), Upper Wenatchee River (including Chiwaukum Creek), Little Wenatchee River, and White River (including the Napeequa River and Panther Creek). See Section 5.5 for a complete coverage of spring Chinook redd surveys in the Wenatchee River basin. In the following section we describe the number and distribution of redds within the White River basin.

#### Redd Counts and Distribution

A total of 26 spring Chinook redds were counted in the White River basin in 2014 (Table 7.10; see Table 5.19 for the complete time series of redd counts). This is lower than the average of 34 redds counted during the period 1989-2014 in the White River. Redds were not distributed

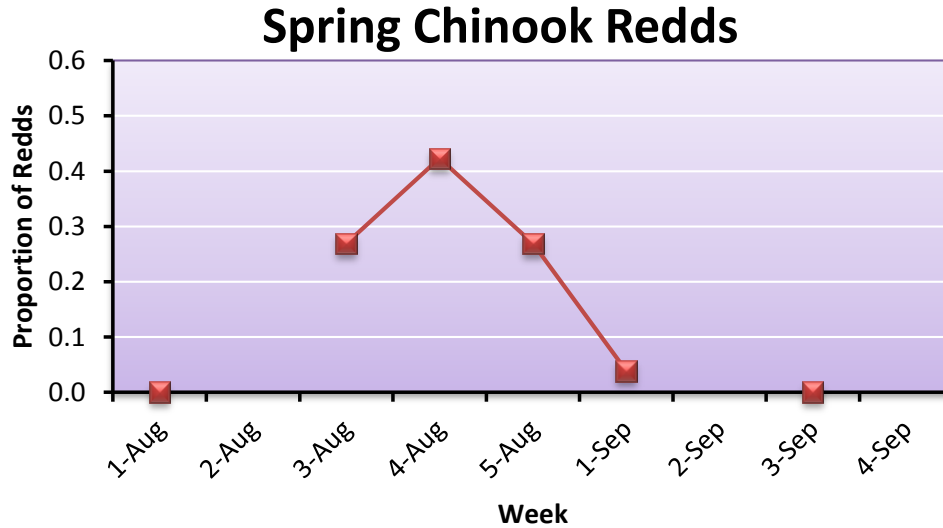
evenly among the six survey areas in the White River basin. Most were located in Reach 3 (Napeequa River to Grasshopper Meadows) in the White River (Table 7.10).

**Table 7.10.** Numbers and proportions of spring Chinook redds counted within different survey areas within the White River basin during August through September, 2014. See Table 2.8 for description of survey reaches.

Stream/watershed	Reach	Number of redds	Proportion of redds within stream/watershed
White River	White 2 (H2)	1	0.04
	White 3 (H3)	22	0.85
	White 4 (H4)	0	0.00
	Napeequa 1 (Q1)	2	0.08
	Panther 1 (T1)	1	0.04
<i>Total</i>		<i>26</i>	<i>1.00</i>

### Spawn Timing

Spring Chinook began spawning during the second week of August in the White River and peaked the four week of August (Figure 7.3). Spawning in the White River ended the second or third week of September.



**Figure 7.3.** Proportion of spring Chinook redds counted during different weeks within the White River basin, August through September 2014.

### Spawning Escapement

Spawning escapement for spring Chinook was calculated as the number of redds times the male-to-female ratio (i.e., fish per redd expansion factor) estimated from broodstock and fish sampled at adult trapping sites. The estimated fish per redd ratio for spring Chinook upstream from

Tumwater in 2014 was 2.06 (based on sex ratios estimated at Tumwater Dam). Multiplying this ratio by the number of redds counted in the White River basin resulted in a total spawning escapement of 54 spring Chinook. The estimated total spawning escapement of spring Chinook in 2014 was less than the overall average of 74 spring Chinook in the White River basin (see Table 5.22).

## 7.6 Carcass Surveys

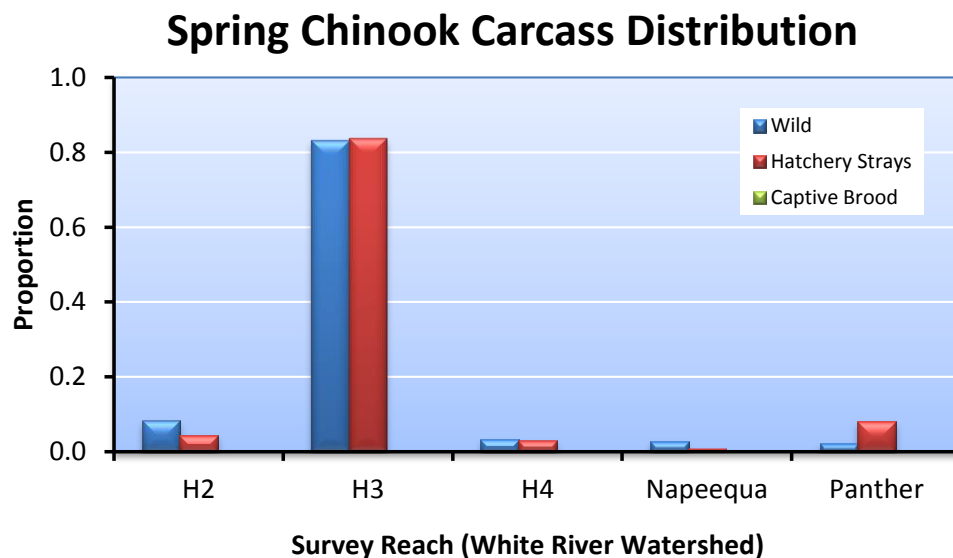
Surveys for spring Chinook carcasses were conducted during August through September, 2014, in the Chiwawa River (including Rock, Phelps, and Chikamin creeks), Nason Creek, Icicle Creek, Peshastin Creek, Upper Wenatchee River, Little Wenatchee River, and White River (including the Napeequa River and Panther Creek). In 2014, eight spring Chinook carcasses were sampled in the White River basin. All of these were sampled in Reach 3. The total number of carcasses sampled in 2014 was less than the overall average of 17 carcasses sampled during the period 1996-2014. See Section 5.6 for a complete coverage of spring Chinook carcass surveys in the Wenatchee River basin.

In the White River basin, the spatial distribution of hatchery strays (primarily from the Chiwawa Spring Chinook program) and wild spring Chinook was similar among survey reaches (Table 7.11). In 2014, all carcasses were observed in the reach between the Napeequa River and Grasshopper Meadows (Reach 3) (Table 7.11). Over the years, spring Chinook have spawned more often in this reach than in other reaches (Figure 7.4). At this time, only two captive brood carcasses have been identified on the spawning grounds, both in Reach 3 in 2013. This may be because captive brood returns were not adipose-fin clipped and therefore any returns from the captive brood program may have been included inadvertently with wild fish.

**Table 7.11.** Numbers of wild, hatchery strays, and captive brood spring Chinook carcasses sampled within different reaches in the White River basin, 2000-2014. See Table 2.8 for description of survey reaches.

Survey year	Origin	Survey Reach					Total
		H-2	H-3	H-4	Napeequa	Panther	
2000	Wild	1	0	0	0	0	1
	Hatchery Strays	0	0	0	0	0	0
2001	Wild	5	40	5	3	1	54
	Hatchery Strays	1	19	3	1	2	26
2002	Wild	3	15	0	0	0	18
	Hatchery Strays	0	6	0	0	1	7
2003	Wild	0	6	0	0	0	6
	Hatchery Strays	0	1	1	0	0	2
2004	Wild	1	9	1	0	0	11
	Hatchery Strays	0	1	0	0	1	2
2005	Wild	1	10	0	1	0	12
	Hatchery Strays	3	37	0	0	0	40
	Captive Brood	0	0	0	0	0	0
2006	Wild	2	16	0	1	0	19
	Hatchery Strays	0	6	0	0	0	6
	Captive Brood	0	0	0	0	0	0

Survey year	Origin	Survey Reach					Total
		H-2	H-3	H-4	Napeequa	Panther	
2007	Wild	1	6	0	0	2	9
	Hatchery Strays	0	4	0	0	0	4
	Captive Brood	0	0	0	0	0	0
2008	Wild	1	3	0	0	1	5
	Hatchery Strays	2	5	0	0	1	8
	Captive Brood	0	0	0	0	0	0
2009	Wild	0	9	0	0	0	9
	Hatchery Strays	0	8	0	0	3	11
	Captive Brood	0	0	0	0	0	0
2010	Wild	0	4	0	0	0	4
	Hatchery Strays	0	7	0	0	0	7
	Captive Brood	0	0	0	0	0	0
2011	Wild	0	4	0	0	0	4
	Hatchery Strays	0	0	0	0	0	0
	Captive Brood	0	0	0	0	0	0
2012	Wild	0	13	0	0	0	13
	Hatchery Strays	0	8	0	0	0	8
	Captive Brood	0	0	0	0	0	0
2013	Wild	0	8	0	0	0	8
	Hatchery Strays	0	10	0	0	3	13
	Captive Brood	0	2	0	0	0	2
2014	Wild	0	6	0	0	0	6
	Hatchery Strays	0	2	0	0	0	2
	Captive Brood	0	0	0	0	0	0
<i>Average</i>	<i>Wild</i>	<i>1</i>	<i>10</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>12</i>
	<i>Hatchery Strays</i>	<i>0</i>	<i>8</i>	<i>0</i>	<i>0</i>	<i>1</i>	<i>9</i>
	<i>Captive Brood</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>0</i>



**Figure 7.4.** Distribution of wild, hatchery strays, and captive brood produced carcasses in different reaches in the White River basin, 2000-2014. Reach codes are described in Table 2.8.

## 7.7 Life History Monitoring

Life history characteristics of spring Chinook were assessed by examining carcasses on spawning grounds and fish collected at broodstock collection sites, and by reviewing tagging data and fisheries statistics.

### Migration Timing

See Section 5.7 for a description of migration timing of spring Chinook at Tumwater Dam.

### Age at Maturity

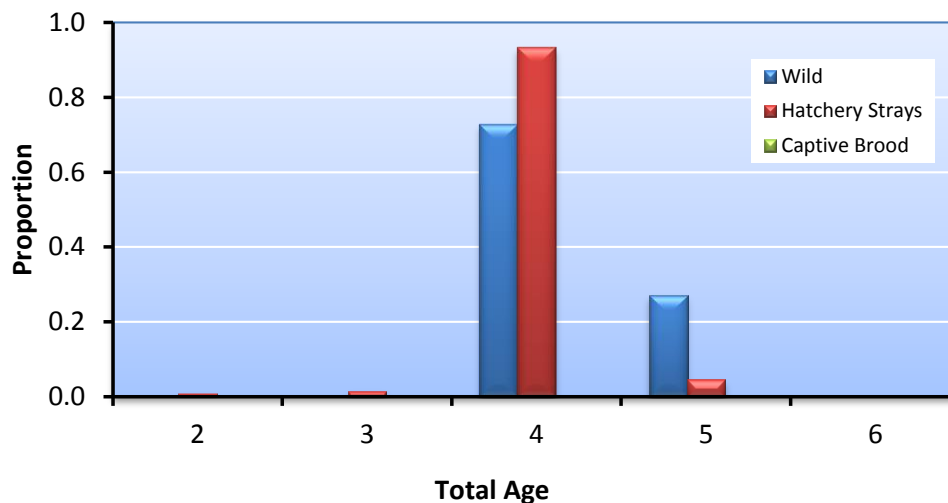
Most of the wild and hatchery stray spring Chinook sampled during the period 2000-2014 in the White River basin were age-4 fish (total age) (Table 7.12; Figure 7.5). Hatchery strays made up a higher percentage of age-3 Chinook than did wild fish. In contrast, a higher proportion of age-5 wild fish returned than did age-5 hatchery strays. Thus, wild fish tended to return at an older age than hatchery strays. At this time, only two captive brood carcasses have been identified on the spawning grounds; one was age-4 the other was age-5.

**Table 7.12.** Numbers of wild, hatchery strays, and captive brood spring Chinook of different ages (total age) sampled on spawning grounds in the White River basin, 2001-2014.

Sample year	Origin	Total age					Sample size
		2	3	4	5	6	
2001	Wild	0	0	47	0	0	47
	Hatchery Strays	0	0	27	0	0	27
2002	Wild	0	0	7	11	0	18
	Hatchery Strays	0	0	6	1	0	7
2003	Wild	0	0	0	6	0	6

Sample year	Origin	Total age					Sample size
		2	3	4	5	6	
	Hatchery Strays	0	0	0	1	0	1
2004	Wild	0	0	9	0	0	9
	Hatchery Stray	0	0	2	0	0	2
2005	Wild	0	0	12	0	0	12
	Hatchery Strays	0	0	40	0	0	40
	Captive Brood	0	0	0	0	0	0
2006	Wild	0	0	7	12	0	19
	Hatchery Strays	0	0	3	3	0	6
	Captive Brood	0	0	0	0	0	0
2007	Wild	0	0	1	8	0	9
	Hatchery Strays	0	2	2	0	0	4
	Captive Brood	0	0	0	0	0	0
2008	Wild	0	0	4	1	0	5
	Hatchery Strays	0	0	8	0	0	8
	Captive Brood	0	0	0	0	0	0
2009	Wild	0	0	8	1	0	9
	Hatchery Strays	1	0	10	0	0	11
	Captive Brood	0	0	0	0	0	0
2010	Wild	0	0	4	0	0	4
	Hatchery Strays	0	0	6	0	0	6
	Captive Brood	0	0	0	0	0	0
2011	Wild	0	0	0	4	0	4
	Hatchery Strays	0	0	0	0	0	0
	Captive Brood	0	0	0	0	0	0
2012	Wild	0	0	13	0	0	13
	Hatchery Strays	0	0	8	0	0	8
	Captive Brood	0	0	0	0	0	0
2013	Wild	0	0	6	2	0	8
	Hatchery Strays	0	0	11	1	0	12
	Captive Brood	0	0	1	1	0	2
2014	Wild	0	0	5	1	0	6
	Hatchery Strays	0	0	2	0	0	2
	Captive Brood	0	0	0	0	0	0
<i>Average</i>	<i>Wild</i>	<i>0</i>	<i>0</i>	<i>9</i>	<i>3</i>	<i>0</i>	<i>12</i>
	<i>Hatchery Strays</i>	<i>0</i>	<i>0</i>	<i>9</i>	<i>0</i>	<i>0</i>	<i>10</i>
	<i>Captive Brood</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>0</i>	<i>0</i>

### Spring Chinook Age Structure



**Figure 7.5.** Proportions of wild, hatchery strays, and captive brood spring Chinook of different total ages sampled on spawning grounds in the White River basin for the combined years 2000-2014.

### Size at Maturity

On average, hatchery strays and wild spring Chinook of a given age differed little in length (Table 7.13). Differences were usually no more than 1-4 cm between hatchery strays and wild fish of the same age. At this time, only two captive brood carcasses have been identified on the spawning grounds; both were females.

**Table 7.13.** Mean lengths (POH in cm;  $\pm 1SD$ ) and sample sizes (in parentheses) of different ages (total age) of male and female spring Chinook of wild, hatchery strays, and captive brood origin sampled in the White River basin, 2001-2014.

Return year	Total age	Mean length (cm)					
		Male			Female		
		Wild	Hatchery stray	Captive brood	Wild	Hatchery stray	Captive brood
2001	3	0	0	0	0	0	0
	4	65 $\pm$ 3 (17)	66 $\pm$ 4 (5)	0	63 $\pm$ 3 (30)	63 $\pm$ 4 (21)	0
	5	0	0	0	0	0	0
	6	0	0	0	0	0	0
2002	3	0	0	0	0	0	0
	4	66 $\pm$ 0 (1)	69 $\pm$ 0 (1)	0	63 $\pm$ 4 (6)	59 $\pm$ 6 (5)	0
	5	75 $\pm$ 11 (2)	0	0	72 $\pm$ 3 (9)	72 $\pm$ 0 (1)	0
	6	0	0	0	0	0	0
2003	3	0	0	0	0	0	0
	4	0	0	0	0	0	0
	5	0	0	0	75 $\pm$ 5 (6)	73 $\pm$ 0 (1)	0
	6	0	0	0	0	0	0
2004	3	0	0	0	0	0	0
	4	68 $\pm$ 3 (3)	0	0	63 $\pm$ 3 (6)	59 $\pm$ 2 (2)	0



Return year	Total age	Mean length (cm)					
		Male			Female		
		Wild	Hatchery stray	Captive brood	Wild	Hatchery stray	Captive brood
	5	0	0	0	0	0	0
	6	0	0	0	0	0	0
2005	3	0	0	0	0	0	0
	4	64 ±5 (3)	62 ±7 (5)	0	63 ±5 (8)	62 ±4 (33)	0
	5	0	0	0	0	0	0
	6	0	0	0	0	0	0
2006	3	0	0	0	0	0	0
	4	65 ±2 (3)	0	0	61 ±4 (4)	60 ±2 (3)	0
	5	69 ±4 (4)	0	0	67 ±5 (8)	70 ±5 (3)	0
	6	0	0	0	0	0	0
2007	3	0	49 ±5 (2)	0	0	0	0
	4	0	0	0	58 ±0 (1)	66 ±2 (2)	0
	5	75 ±5 (3)	0	0	75 ±1 (5)	0	0
	6	0	0	0	0	0	0
2008	3	0	0	0	0	0	0
	4	56 ±0 (1)	61 ±0 (1)	0	63 ±8 (2)	61 ±2 (7)	0
	5	0	0	0	75 ±0 (1)	0	0
	6	0	0	0	0	0	0
2009	3	0	0	0	0	0	0
	4	61 ±5 (3)	68 ±4 (2)	0	63 ±2 (5)	62 ±2 (8)	0
	5	0	0	0	78 ±0 (1)	0	0
	6	0	0	0	0	0	0
2010	3	0	0	0	0	0	0
	4	0	67 ±0 (1)	0	60 ±3 (3)	61 ±6 (5)	0
	5	0	0	0	0	0	0
	6	0	0	0	0	0	0
2011	3	0	0	0	0	0	0
	4	0	0	0	0	0	0
	5	0	0	0	73 ±5 (4)	0	0
	6	0	0	0	0	0	0
2012	3	0	0	0	0	0	0
	4	47 ±0 (1)	0	0	62 ±4 (12)	60 ±4 (8)	0
	5	0	0	0	0	0	0
	6	0	0	0	0	0	0
2013	3	0	0	0	0	0	0
	4	64 ±4 (3)	60 ±4 (2)	0	61 ±2 (3)	61 ±4 (7)	63 ±0 (1)
	5	0	0	0	67 ±1 (2)	71 ±0 (1)	71 ±0 (1)
	6	0	0	0	0	0	0
2014	3	0	0	0	0	0	0
	4	0	54 ±0 (1)	0	60 ±2 (5)	58 ±0 (1)	0
	5	0	0	0	74 ±0 (1)	0	0

Return year	Total age	Mean length (cm)					
		Male			Female		
		Wild	Hatchery stray	Captive brood	Wild	Hatchery stray	Captive brood
	6	0	0	0	0	0	0

### Contribution to Fisheries

No White River spring Chinook from the captive brood program tagged with CWTs or PIT tags have been recaptured (or reported) in ocean or Columbia River (tribal, commercial, or recreational) fisheries.

### Straying

Stray rates of White River spring Chinook from the captive brood program were determined by examining the locations where PIT-tagged Chinook demonstrating anadromy (based on detections at Bonneville Dam) were last detected. PIT tagging of White River spring Chinook began with release year 2008, which allows estimation of stray rates by brood return. Targets for strays based on return year (recovery year) within the Wenatchee River basin should be less than 10% and targets for strays outside the Wenatchee River basin should be less than 5%. The target for brood year stray rates should be less than 5%.

Based on PIT-tag analyses, on average, about 46% of the White River spring Chinook returns were last detected in streams outside the White River (Table 7.14). The numbers in Table 7.14 should be considered rough estimates because they are not based on confirmed spawning (only last detections) and the numbers have not been adjusted for detection efficiencies, which currently do not exist for most PIT-tag detection arrays in tributaries. In addition, last detections in adult fishways (i.e., Bonneville, Rock Island, and Tumwater dams) were not included, nor were detections in areas outside the distribution of known spring Chinook spawning (i.e., Lower and Middle Wenatchee River). All fish reported in Table 7.14 are at least age-3 fish (total age) and some of them may not have migrated to the ocean, but rather resided completely in freshwater.

**Table 7.14.** Number and percent of White River spring Chinook from the captive brood program that homed to target spawning areas on the White River and the target hatchery program (Little White Salmon Fish Hatchery), and number and percent that strayed to non-target spawning areas and hatchery programs for brood years 2006 to 2009. Only PIT-tagged fish demonstrating anadromy were included in the analysis. Estimates were based on last detections of PIT-tagged spring Chinook. Percent strays should be less than 5%.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
2006	1	100.0	0	0.0	0	0.0	0	0
2007	0	0.0	0	0.0	0	0.0	0	0
2008	0	0.0	0	0.0	15	100.0	0	0
2009	4	14.3	0	0.0	25	85.7	0	0
<b>Average</b>	<b>1</b>	<b>28.6</b>	<b>0</b>	<b>0.0</b>	<b>10</b>	<b>46.4</b>	<b>0</b>	<b>0</b>

\* Homing to the target hatchery includes White River hatchery spring Chinook that are captured and included as broodstock in the White River Hatchery program.

The percentage of the PIT-tagged White River spring Chinook from the captive brood program that were last detected in different watersheds within and outside the Wenatchee River basin are shown in Table 7.15. On average, a small percentage of the PIT-tagged White River spring Chinook homed to the White River. Relatively high percentages of them were last detected in the Little Wenatchee River, Nason Creek, and the Chiwawa River. Few have strayed into spawning areas outside the Wenatchee River basin.

**Table 7.15.** Number and percent (in parentheses) of PIT-tagged White River spring Chinook from the captive brood program that were last detected in different tributaries within the Wenatchee River basin, return years 2010-2014. Only PIT-tagged fish demonstrating anadromy were included in the analysis.

Return year	Homing	Straying							
	White River	Chiwawa River	Chiwaukum Creek	Icicle Creek	Little Wenatchee	Nason Creek	Peshastin Creek	Upper Wenatchee	Entiat River
2010	1 (100.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
2011	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (50.0)	1 (50.0)	0 (0.0)	0 (0.0)	0 (0.0)
2012	2 (16.7)	1 (8.3)	0 (0.0)	0 (0.0)	8 (66.7)	1 (8.3)	0 (0.0)	0 (0.0)	0 (0.0)
2013	2 (6.7)	8 (26.7)	1 (3.3)	2 (6.7)	7 (23.3)	8 (26.7)	0 (0.0)	2 (6.7)	0 (0.0)
2014	4 (8.3)	17 (35.4)	0 (0.0)	1 (2.1)	3 (6.3)	17 (35.4)	0 (0.0)	5 (10.4)	1 (2.1)
<i>Average</i>	<i>2 (26.3)</i>	<i>5 (14.1)</i>	<i>0 (0.7)</i>	<i>1 (1.8)</i>	<i>4 (29.3)</i>	<i>5 (24.1)</i>	<i>0 (0.0)</i>	<i>1 (3.4)</i>	<i>0 (0.4)</i>

## Genetics

At this time, there are no studies that examine the effects of the White River captive brood program on the genetics of natural-origin spring Chinook in the Wenatchee River basin. However, genetic studies were conducted to determine the potential effects of the Chiwawa Supplementation Program on natural-origin spring Chinook in the upper Wenatchee River basin (Blankenship et al. 2007; the entire report is appended as Appendix J). This work included the analysis of White River spring Chinook. Researchers collected microsatellite DNA allele frequencies from temporally replicated natural and hatchery-origin spring Chinook to statistically assign individual fish to specific demes (locations) within the Wenatchee population.

Significant differences in allele frequencies were observed within and among major spawning areas in the Upper Wenatchee River basin. However, these differences made up only a very small portion of the overall variation, indicating genetic similarity among the major spawning areas. There was no evidence that the Chiwawa program has changed the genetic structure (allele frequency) of spring Chinook in the White River, despite the presence of hatchery-origin spawners in both systems.

## Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment

relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For brood years 1989-2000, approximate PNI Index values ranged from 0.95 to 1.00 (Table 7.16). For brood years 2001-2013, the approximate PNI Index for the White River Program (PNI<sub>W</sub>) averaged 0.90 (range, 0.65-1.00). If hatchery strays are included in the PNI Index calculation (PNI<sub>W+S</sub>), values averaged 0.70 (range, 0.44-1.00) (Table 7.16).

**Table 7.16.** Approximate Proportionate Natural Influence (PNI) Index of hatchery spring Chinook spawning in the White River. See notes below the table for description of each metric.

Brood year	Spawners					Broodstock			PNI <sub>W</sub>	PNI <sub>W+S</sub>
	NOS	HOS <sub>W</sub>	HOS <sub>S</sub>	pHOS <sub>W</sub>	pHOS <sub>W+S</sub>	NOB <sub>N</sub>	HOB <sub>N</sub>	pNOB		
1989	145	0	0	0.00	0.00	0	0	1.00	--	1.00
1990	49	0	0	0.00	0.00	0	0	1.00	--	1.00
1991	49	0	0	0.00	0.00	0	0	1.00	--	1.00
1992	78	0	0	0.00	0.00	0	0	1.00	--	1.00
1993	138	0	7	0.00	0.05	0	0	0.99	--	0.95
1994	7	0	0	0.00	0.00	0	0	0.67	--	1.00
1995	5	0	0	0.00	0.00	0	0	1.00	--	1.00
1996	30	0	0	0.00	0.00	0	0	0.60	--	1.00
1997	33	0	0	0.00	0.00	0	0	0.30	--	1.00
1998	11	0	0	0.00	0.00	0	0	0.44	--	1.00
1999	3	0	0	0.00	0.00	0	0	1.00	--	1.00
2000	22	0	0	0.00	0.00	0	0	0.48	--	1.00
<b>Average*</b>	<b>48</b>	<b>0</b>	<b>1</b>	<b>0.00</b>	<b>0.00</b>	<b>0</b>	<b>0</b>	<b>0.79</b>	<b>--</b>	<b>1.00</b>
2001	111	0	55	0.00	0.33	5	0	1.00	1.00	0.48
2002	60	0	26	0.00	0.30	18	0	1.00	1.00	0.77
2003	31	0	5	0.00	0.14	7	0	1.00	1.00	0.88
2004	54	0	12	0.00	0.18	6	0	1.00	1.00	0.85
2005	38	11	106	0.07	0.75	103	73	0.59	0.89	0.44
2006	41	5	9	0.09	0.25	191	135	0.59	0.87	0.70
2007	62	23	7	0.25	0.33	254	6	0.98	0.80	0.75
2008	20	2	30	0.04	0.62	116	0	1.00	0.96	0.62
2009	81	29	63	0.17	0.53	238	0	1.00	0.85	0.65
2010	27	22	23	0.31	0.63	90	0	1.00	0.76	0.61
2011	83	0	0	0.00	0.00	306	0	1.00	1.00	1.00
2012	89	10	45	0.07	0.38	390	0	1.00	0.93	0.72
2013	44	55	5	0.53	0.58	383	0	1.00	0.65	0.63
<b>Average**</b>	<b>57</b>	<b>12</b>	<b>30</b>	<b>0.12</b>	<b>0.39</b>	<b>162</b>	<b>16</b>	<b>0.94</b>	<b>0.90</b>	<b>0.70</b>

HOS<sub>W</sub> = hatchery-origin spawners in White River from the White River spring Chinook Supplementation Program.

pHOS<sub>W</sub> = proportion of hatchery-origin spawners from White River spring Chinook Supplementation Program.

HOS<sub>S</sub> = stray hatchery-origin spawners in the White River.

pHOS<sub>S</sub> = proportion of stray hatchery-origin spawners.

NOB<sub>W</sub> = natural origin broodstock spawned for the White River spring Chinook Supplementation Program.

**HOB<sub>w</sub>** = hatchery-origin broodstock spawned in the White River spring Chinook Supplementation Program.

**pNOB** = proportion of hatchery-origin broodstock. Because of the high incidence of strays to the White River from the Chiwawa River spring Chinook program, pNOB values from the Chiwawa program were used to estimate PNI values during the period from 1989 to 2000 (*italicized*). The weighting for those years was 100% based on the Chiwawa program broodstock selection, because there have been no hatchery returns from the White River spring Chinook program during this period (see Table 5.1 for Chiwawa broodstock selection).

**PNI<sub>w</sub>** = pNOB/(pNOB + pNOS<sub>N</sub>); where pNOB is weighted 100% toward broodstock collection from the White River spring Chinook Supplementation Program. The intent of this PNI value is to track the performance of the White River Program apart from the influence of strays and broodstock collection outside the White River watershed.

**PNI<sub>w+s</sub>** = pNOB/(pNOB + pHOS<sub>N+S</sub>); where pNOB is weighted by the proportion of HOS<sub>w</sub> and HOS<sub>s</sub> observed in the White River. This PNI value tracks the combined influence of broodstock selection from the White River Program and/or Chiwawa Program according to the proportion of HOS that return to the White River from those programs.

\* Average for the period 1989-2000.

\*\* Average for the period 2001-present.

## Natural and Hatchery Replacement Rates

In general, natural replacement rates (NRR) are calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). We calculated NORs with and without harvest. NORs include all returning fish that either returned to the basin or were collected as wild broodstock. For brood years 1989-2008, NRR for spring Chinook in the White River basin averaged 1.09 (range, 0.00-4.91) if harvested fish were not include in the estimate and 1.21 (range, 0.00-5.27) if harvested fish were included in the estimate (Table 7.17). NRRs for more recent brood years will be calculated as soon as all tag recoveries and sampling rates have been loaded into the database.

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and are calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. For brood years 2006-2008, hatchery replacement rates averaged 0.08 (range, 0.02-0.20) (Table 7.17). Only for brood year 2007 was HRR greater than the NRR.

**Table 7.17.** Numbers of brood stock spawned, spawning escapements, hatchery origin recruits (HOR), natural-origin recruits (NOR), hatchery replacement rates (HRR), and natural replacement rates (NRR) with and without harvest for spring Chinook in the White River basin, brood years 1989-2008.

Brood year	Brood stock spawned	Spawning Escapement	Harvest not included				Harvest included			
			HOR <sup>1</sup>	NOR <sup>2</sup>	HRR <sup>1</sup>	NRR <sup>2</sup>	NOR <sup>3</sup>	NOR <sup>4</sup>	HRR <sup>3</sup>	NRR <sup>4</sup>
1989	--	145	--	81	--	0.56	--	118	--	0.81
1990	--	49	--	2	--	0.04	--	2	--	0.04
1991	--	49	--	3	--	0.06	--	3	--	0.06
1992	--	78	--	30	--	0.38	--	32	--	0.41
1993	--	145	--	44	--	0.30	--	45	--	0.31
1994	--	7	--	1	--	0.14	--	1	--	0.14
1995	--	5	--	9	--	1.80	--	9	--	1.80
1996	--	30	--	15	--	0.50	--	16	--	0.53
1997	--	33	--	148	--	4.48	--	164	--	4.97
1998	--	11	--	54	--	4.91	--	58	--	5.27
1999	--	3	--	0	--	0.00	--	0	--	0.00
2000	--	22	--	54	--	2.45	--	57	--	2.59

Brood year	Brood stock spawned	Spawning Escapement	Harvest not included				Harvest included			
			HOR <sup>1</sup>	NOR <sup>2</sup>	HRR <sup>1</sup>	NRR <sup>2</sup>	NOR <sup>3</sup>	NOR <sup>4</sup>	HRR <sup>3</sup>	NRR <sup>4</sup>
2001	5	166	--	64	--	0.39	--	66	--	0.40
2002	18	86	--	70	--	0.81	--	73	--	0.85
2003	7	36	--	11	--	0.31	--	12	--	0.33
2004	6	66	--	25	--	0.38	--	27	--	0.41
2005	176	155	--	72	--	0.46	--	75	--	0.48
2006	326	55	5	110	0.02	2.00	5	139	0.02	2.53
2007	260	92	0	0	0.00	0.00	0	0	0.00	0.00
2008	116	52	30	100	0.26	1.92	30	113	0.26	2.17
<b>Average</b>	<b>114</b>	<b>64</b>	<b>13</b>	<b>45</b>	<b>0.09</b>	<b>1.09</b>	<b>13</b>	<b>51</b>	<b>0.09</b>	<b>1.21</b>

<sup>1</sup>HOR and HRR values represented here are detections of PIT-tag hatchery fish detected at Tumwater Dam. These values have not been expanded based on the untagged proportion of fish released from the White River spring Chinook Program or the sampling rate at Tumwater Dam.

<sup>2</sup>NOR and NRR values represented here are based on carcasses recovery in the White River adjusted by H:W ratios and age composition and expanded to the escapement in the White River.

<sup>3</sup>Harvest rates on hatchery-origin White River spring Chinook have not yet been estimated but will be expanded based on harvest rates observed for Chiwawa spring Chinook.

<sup>4</sup>Expanded NORs for harvest were based on harvest rates from Chiwawa River spring Chinook.

### Smolt-to-Adult Survivals

Smolt-to-adult survival ratios (SARs) were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery smolts released. SARs were based on both CWT returns and PIT-tag detections. For the available brood years, SARs have ranged from 0.00001 to 0.00025 based on CWTs and from 0.00008 to 0.00022 based on PIT-tag detections (Table 7.18).

**Table 7.18.** Smolt-to-adult ratios (SARs) for White River spring Chinook from the captive brood program, brood years 2005-2008.

Brood year	CWTs		PIT-tags	
	Recoveries	SAR	Detections	SAR
2005	16	0.00025	NA	NA
2006	9	0.00003	6	0.00020
2007	1	0.00001	3	0.00008
2008	NA	NA	9	0.00022
<b>Average</b>	<b>8.7</b>	<b>0.00010</b>	<b>6</b>	<b>0.00017</b>

## 7.8 ESA/HCP Compliance

### Brood Collection

The last collection of eggs or fry for this program occurred in 2010 (brood year 2009). From 2011 to 2013, the White River Captive Brood Program has operated without ESA permit coverage. The hatchery program will sunset with the last release of juveniles in 2015 (brood year 2013).

## Hatchery Rearing, Spawning, and Release

From 2011 to 2013, the White River Captive Brood Program has operated without ESA permit coverage. The hatchery program will sunset with the last release of juveniles in 2015 (brood year 2013). Release of juveniles in 2013 and 2014 were consistent with the terms and conditions of Section 10(a)(1)(A) Permit 18120.

## Hatchery Effluent Monitoring

Per ESA Permits 1196 (expired), 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January through 31 December 2014. NPDES monitoring and reporting for Grant PUD Hatchery Programs during 2014 are provided in Appendix F.

This report does not cover hatchery rearing of the White River Captive Brood Program (adults and juveniles) at the Little White Salmon National Fish Hatchery, operated by the U.S. Fish and Wildlife Service.

## Smolt and Emigrant Trapping

Per ESA Section 10 Permit No. 1196 (expired), 18118, 18120, and 18121, the permit holders are authorized a direct take of 20% of the emigrating spring Chinook population during juvenile emigration monitoring and a lethal take not to exceed 2% of the fish captured (NMFS 2003). Based on the estimated wild spring Chinook population (smolt trap expansion) and hatchery juvenile spring Chinook population estimate (hatchery release data) for the Wenatchee River basin, the reported spring Chinook encounters during 2014 emigration monitoring complied with take provisions in the Section 10 permit. Spring Chinook encounter and mortality rates for each trap site (including PIT tag mortalities) are detailed in Table 7.19. Additionally, juvenile fish captured at the trap locations were handled consistent with provisions in ESA Section 10 Permit 1196 (expired), 18118, 18120, and 18121, Section B. Table 7.19 does not include incidental or direct take associated with the White River smolt trap operated by the Yakama Nation.

**Table 7.19.** Estimated take of Upper Columbia River spring Chinook resulting from juvenile emigration monitoring in the Wenatchee River basin, 2014.

Trap location	Population estimate			Number trapped			Total	Take allowed under Permit
	Wild <sup>a</sup>	Hatchery <sup>b</sup>	Sub-yearling <sup>c</sup>	Wild	Hatchery	Sub-yearling		
<b>Chiwawa Trap</b>								
Population	34,334	222,504	73,695	4,519	5,293	23,755	33,567	
Encounter rate	NA	NA	NA	0.1316	0.0237	0.3223	0.1016	0.20
Mortality <sup>c</sup>	NA	NA	NA	28	0	84	112	
Mortality rate	NA	NA	NA	0.0062	0.0000	0.0035	0.0033	0.02
<b>Lower Wenatchee Trap</b>								
Population	67,973	222,504	11,936,928	1,700	31,290	81,445	114,435	
Encounter rate	NA	NA	NA	0.025	0.1406	0.0068	0.0096	0.20
Mortality <sup>d</sup>	NA	NA	NA	17	12	250	279	
Mortality rate	NA	NA	NA	0.0100	0.0004	0.0031	0.0024	0.02

Trap location	Population estimate			Number trapped			Total	Take allowed under Permit
	Wild <sup>a</sup>	Hatchery <sup>b</sup>	Sub-yearling <sup>c</sup>	Wild	Hatchery	Sub-yearling		
<b>Wenatchee River Basin Total</b>								
Population	67,393	222,504	11,936,928	6,219	36,583	105,200	148,002	
Encounter rate	NA	NA	NA	0.0922	0.1644	0.0088	0.0121	0.20
Mortality <sup>d</sup>	NA	NA	NA	45	12	84	141	
Mortality rate	NA	NA	NA	0.0072	0.0003	0.0008	0.0010	0.02

<sup>a</sup> Smolt population estimate derived from juvenile emigration trap data.

<sup>b</sup> 2012 BY smolt release data for the Wenatchee River basin.

<sup>c</sup> Based on size, date of capture and location of capture, subyearling Chinook encountered at the Lower Wenatchee Trap are categorized as summer Chinook..

<sup>d</sup> Combined trapping and PIT tagging mortality.

### Spawning Surveys

Spring Chinook spawning ground surveys were conducted in the Wenatchee River basin during 2014, as authorized by ESA Section 10 Permits 18118, 18119, and 18121. Because of the difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical, and extreme caution was used to avoid established redds when wading was required.

### Spring Chinook Reproductive Success Study

ESA Section 10 Permit 1196 (expired) and new Section 10 Permits 18118, 18119, and 18121 specifically provide authorization to capture, anesthetize, biologically sample, PIT tag, and release adult spring Chinook at Tumwater Dam for reproductive success studies and general program monitoring. During 2010 through 2014, all spring Chinook passing Tumwater Dam were enumerated, anesthetized, biologically sampled, PIT tagged, and released (not including hatchery-origin and natural-origin Chinook retained for broodstock) as a component of the reproductive success study (BPA Project No. 2003-039-00). Please refer to Ford et al. (2010, 2011, 2012, 2013, and 2014) for complete details on the methods and results of the spring Chinook reproductive success study for the period 2010-2014.



## SECTION 8: WENATCHEE SUMMER CHINOOK

The goal of summer Chinook salmon supplementation in the Wenatchee Basin is to use artificial production to replace adult production lost because of mortality at Rock Island, Wanapum, and Priest Rapids dams, while not reducing the natural production or long-term fitness of summer Chinook in the basin. The Rock Island Fish Hatchery Complex began operation in 1989 under funding from Chelan PUD and subsequently Grant PUD began cost-sharing the program in 2012. The Complex operated originally through the Rock Island Settlement Agreement, but since 2004 has operated under the Anadromous Fish Agreement and Habitat Conservation Plans as well as the Priest Rapids Project Salmon and Steelhead Settlement Agreement.

Adult summer Chinook are collected for broodstock from the run-at-large at the right and left-bank traps at Dryden Dam, and at Tumwater Dam if the weekly quotas cannot be achieved at Dryden Dam. Prior to 2012, the goal was to collect up to 492 natural-origin adult summer Chinook for the Wenatchee program for an annual release of 864,000 smolts. In 2011, the Hatchery Committees reevaluated the amount of hatchery compensation needed to achieve NNI. Based on that evaluation, the goal of the program was revised. The current goal (beginning in 2012) is to collect up to 256 adult natural-origin summer Chinook for an annual release of 500,001 smolts. Broodstock collection occurs from about 1 July through 15 September with trapping occurring up to 24 hours per day, seven days a week. If natural-origin broodstock collection falls short of expectation, hatchery-origin adults can be collected to make up the difference.

Adult summer Chinook are spawned and reared at Eastbank Fish Hatchery. Juvenile summer Chinook are transferred from the hatchery to Dryden Acclimation Pond in March. They are released from the pond in late April to early May.

Before 2012, the production goal for the Wenatchee summer Chinook supplementation program was to release 864,000 yearling smolts into the Wenatchee River at ten fish per pound. Beginning with the 2012 brood, the revised production goal is to release 500,001 yearling smolts into the Wenatchee River at 10 and 15 fish per pound. Targets for fork length and weight are 163 mm (CV = 9.0) and 45.4 g, respectively. Over 90% of these fish are marked with CWTs. In addition, since 2009, about 10,000 juvenile summer Chinook have been PIT tagged annually.

### 8.1 Broodstock Sampling

This section focuses on results from sampling 2012-2013 Wenatchee summer Chinook broodstock, which were collected at Dryden and Tumwater dams. Complete information is not currently available for the 2014 brood (this information will be provided in the 2015 annual report).

#### Origin of Broodstock

Consistent with the broodstock collection protocol, both the 2012 and 2013 broodstock consisted primarily of natural-origin (adipose fin present and no CWT) summer Chinook (Table 8.1). Less than 1% of the 2013 broodstock was comprised of hatchery-origin fish (hatchery-origin was determined by examination of scales and/or CWTs).

**Table 8.1.** Numbers of wild and hatchery summer Chinook collected for broodstock, numbers that died before spawning, and numbers of Chinook spawned in the Wenatchee River basin, 1989-2013. Unknown origin fish (i.e., undetermined by scale analysis, no CWT or fin clips, and no additional hatchery marks) were considered naturally produced. Mortality includes fish that died of natural causes typically near the end of spawning and were not needed for the program and surplus fish killed at spawning.

Brood year	Wild summer Chinook					Hatchery summer Chinook					Total number spawned
	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	
1989	346	29	27	290	0	0	0	0	0	0	290
1990	87	6	24	57	0	0	0	0	0	0	57
1991	128	9	14	105	0	0	0	0	0	0	105
1992	341	48	19	274	0	0	0	0	0	0	274
1993	480	28	46	406	0	44	0	0	44	0	450
1994	363	29	1	333	0	55	1	0	54	0	387
1995	382	15	4	363	0	16	0	0	16	0	378
1996	331	34	34	263	0	3	0	0	3	0	266
1997	225	14	6	205	0	15	1	1	13	0	218
1998	378	40	39	299	0	94	4	12	78	0	377
1999	250	7	1	242	0	238	1	1	236	0	478
2000	298	18	5	275	0	194	7	7	180	0	455
2001	311	41	60	210	0	182	8	38	136	0	346
2002	469	28	32	409	0	13	1	2	10	0	419
2003	488	90	61	337	0	8	1	0	7	0	344
2004	494	24	46	424	0	2	0	0	2	0	426
2005	491	29	19	397	46	3	0	0	3	0	400
2006	483	29	21	433	0	5	1	0	4	0	437
2007	415	53	99	263	0	4	0	1	3	0	266
2008	400	11	11	378	0	72	2	1	69	0	447
2009	482	22	8	452	0	9	1	0	8	0	460
2010	427	14	25	388	0	7	2	0	5	0	393
2011	398	11	11	376	0	7	0	0	7	0	405
<i>Average<sup>b</sup></i>	<i>368</i>	<i>27</i>	<i>27</i>	<i>312</i>	<i>2</i>	<i>42</i>	<i>1</i>	<i>3</i>	<i>38</i>	<i>0</i>	<i>351</i>
2012	273	5	1	267	0	1	0	0	1	0	268
2013	256	12	10	234	0	2	0	0	2	0	236
<i>Average<sup>c</sup></i>	<i>265</i>	<i>9</i>	<i>6</i>	<i>251</i>	<i>0</i>	<i>2</i>	<i>0</i>	<i>0</i>	<i>2</i>	<i>0</i>	<i>252</i>

<sup>a</sup> Pre-spawn loss represents the number of fish that died during the holding period before spawning. Mortality is the number of fish that were surplus following spawning.

<sup>a</sup> This average represents the program before recalculation in 2011.

<sup>b</sup> This average represents the current program, which began in 2012.

### Age/Length Data

Ages of summer Chinook broodstock were determined from analysis of scales and/or CWTs. Broodstock collected from the 2012 return consisted primarily of age-4 and age-5 natural-origin Chinook (96%). Age-3 natural-origin fish made up 3% of the broodstock (Table 8.2). The one hatchery Chinook included in the broodstock was an age-5 fish.

Broodstock collected from the 2013 return consisted primarily of age-4 and age-5 natural-origin Chinook (86%). Age-3 and age-6 natural-origin fish made up 12% and 2% of the broodstock,

respectively (Table 8.2). The two hatchery Chinook included in the broodstock were age-4 and age-5 fish.

**Table 8.2.** Percent of hatchery and wild Wenatchee summer Chinook of different ages (total age) collected from broodstock in the Wenatchee River basin, 1991-2013.

Return Year	Origin	Total age				
		2	3	4	5	6
1991	Wild	0.0	4.6	36.8	57.5	1.1
	Hatchery	0.0	0.0	0.0	0.0	0.0
1992	Wild	0.0	2.6	40.4	50.9	6.1
	Hatchery	0.0	0.0	0.0	0.0	0.0
1993	Wild	0.0	1.5	35.7	60.4	2.3
	Hatchery	0.0	0.0	93.2	6.8	0.0
1994	Wild	0.0	1.0	33.7	64.3	1.0
	Hatchery	0.0	0.0	1.9	98.1	0.0
1995	Wild	0.0	3.3	19.2	76.3	1.2
	Hatchery	0.0	0.0	0.0	0.0	100.0
1996	Wild	0.0	4.6	40.1	53.3	2.0
	Hatchery	0.0	0.0	33.3	66.7	0.0
1997	Wild	0.0	2.3	42.6	53.2	1.9
	Hatchery	0.0	26.7	66.7	6.7	0.0
1998	Wild	0.0	5.5	34.7	58.6	1.2
	Hatchery	0.0	5.3	68.1	20.2	6.4
1999	Wild	0.5	1.9	39.0	56.3	2.3
	Hatchery	0.0	1.3	23.2	72.2	3.4
2000	Wild	2.6	6.3	24.6	66.5	0.0
	Hatchery	0.0	24.2	14.9	42.8	18.0
2001	Wild	0.3	16.6	53.6	27.7	1.7
	Hatchery	0.0	6.1	80.5	10.4	3.0
2002	Wild	0.7	8.4	61.6	28.5	0.7
	Hatchery	0.0	0.0	41.7	58.3	0.0
2003	Wild	0.9	2.8	31.4	64.8	0.0
	Hatchery	0.0	12.5	25.0	62.5	0.0
2004	Wild	0.2	3.6	10.1	83.9	2.1
	Hatchery	0.0	0.0	50.0	50.0	0.0
2005	Wild	0.0	4.3	53.5	35.1	7.1
	Hatchery	0.0	0.0	0.0	100.0	0.0
2006	Wild	0.9	0.9	14.9	82.1	1.1
	Hatchery	0.0	0.0	0.0	80.0	20.0
2007	Wild	3.1	15.0	18.7	46.6	16.6
	Hatchery	0.0	0.0	0.0	100.0	0.0

Return Year	Origin	Total age				
		2	3	4	5	6
2008	Wild	0.5	6.4	65.5	26.0	1.6
	Hatchery	0.0	2.9	13.0	69.6	14.5
2009	Wild	1.1	6.9	45.8	46.8	0.0
	Hatchery	0.0	0.0	11.1	88.9	0.0
2010	Wild	1.0	6.3	66.1	26.6	0.0
	Hatchery	0.0	0.0	62.5	37.5	0.0
2011	Wild	0.8	8.2	50.3	40.4	0.3
	Hatchery	0.0	42.9	14.3	42.9	0.0
2012	Wild	0.0	3.5	47.2	49.2	0.0
	Hatchery	0.0	0.0	0.0	100.0	0.0
2013	Wild	0.0	12.1	57.1	29.1	1.6
	Hatchery	0.0	0.0	50.0	50.0	0.0
<i>Average</i>	<i>Wild</i>	<i>0.5</i>	<i>5.6</i>	<i>40.1</i>	<i>51.5</i>	<i>2.3</i>
	<i>Hatchery</i>	<i>0.0</i>	<i>5.3</i>	<i>36.8</i>	<i>50.6</i>	<i>7.2</i>

Mean lengths of natural-origin summer Chinook of a given age differed little between return years 2012 and 2013 (Table 8.3). The two hatchery fish that were included in broodstock were 2 and 9 cm smaller than their natural counterparts in the 2013 brood (Table 8.3).

**Table 8.3.** Mean fork length (cm) at age (total age) of hatchery and wild Wenatchee summer Chinook collected from broodstock in the Wenatchee River basin, 1991-2013; N = sample size and SD = 1 standard deviation.

Return year	Origin	Summer Chinook fork length (cm)														
		Age-2			Age-3			Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
1991	Wild	-	0	-	-	4	-	-	32	-	-	50	-	-	1	-
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-	-	0	-
1992	Wild	-	0	-	66	3	10	69	46	5	81	58	3	87	7	1
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-	-	0	-
1993	Wild	-	0	-	68	6	10	84	138	9	98	235	6	100	9	6
	Hatchery	-	0	-	-	0	-	79	41	8	101	3	8	-	0	-
1994	Wild	-	0	-	74	3	5	86	101	8	96	193	7	106	3	7
	Hatchery	-	0	-	-	0	-	75	1	-	90	53	8	-	0	-
1995	Wild	-	0	-	66	11	8	85	64	7	97	255	6	106	4	7
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-	91	16	8
1996	Wild	-	0	-	69	14	5	86	121	6	97	161	6	104	6	5
	Hatchery	-	0	-	-	0	-	63	1	-	96	2	4	-	0	-
1997	Wild	-	0	-	54	5	10	85	92	7	98	115	6	97	4	9
	Hatchery	-	0	-	46	4	2	74	10	4	98	1	-	-	0	-
1998	Wild	-	0	-	66	19	9	85	119	7	99	201	7	106	4	7

Return year	Origin	Summer Chinook fork length (cm)														
		Age-2			Age-3			Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
	Hatchery	-	0	-	53	5	2	77	64	8	95	19	8	98	6	8
1999	Wild	42	1	-	65	4	6	86	83	6	97	120	7	103	5	8
	Hatchery	-	0	-	52	3	6	79	55	7	90	171	6	100	8	6
2000	Wild	43	7	3	60	17	7	84	67	5	98	181	6	-	0	-
	Hatchery	-	0	-	53	47	7	76	29	8	93	83	7	102	35	9
2001	Wild	48	1	-	66	48	7	88	155	7	97	80	6	102	5	3
	Hatchery	-	0	-	51	10	3	75	132	8	91	17	8	100	5	8
2002	Wild	51	3	3	64	37	8	89	270	7	100	125	7	99	7	5
	Hatchery	-	0	-	-	0	-	78	5	8	95	7	5	-	0	-
2003	Wild	41	4	2	58	13	4	87	144	8	100	297	7	-	0	-
	Hatchery	-	0	-	40	1	-	78	2	4	101	5	8	-	0	-
2004	Wild	51	1	-	69	17	5	84	47	8	99	392	6	109	10	7
	Hatchery	-	0	-	-	0	-	84	1	-	108	1	-	-	0	-
2005	Wild	-	0	-	68	20	7	86	247	8	95	162	6	101	33	6
	Hatchery	-	0	-	-	0	-	-	0	-	90	3	9	-	0	-
2006	Wild	44	4	7	63	4	11	88	66	7	99	363	6	96	5	7
	Hatchery	-	0	-	-	0	-	-	0	-	99	4	7	100	1	-
2007	Wild	44	12	5	65	58	7	89	72	8	99	180	7	102	64	6
	Hatchery	-	0	-	-	0	-	-	0	-	90	4	5	-	0	-
2008	Wild	46	2	3	69	24	7	90	247	6	98	98	7	105	6	9
	Hatchery	-	0	-	63	2	14	81	9	7	93	48	6	99	10	5
2009	Wild	46	5	5	68	31	8	89	207	8	101	209	6	-	0	-
	Hatchery	-	0	-	61	4	7	81	1	-	98	8	14	-	0	-
2010	Wild	45	4	4	70	26	9	89	273	7	99	110	6	-	0	-
	Hatchery	-	0	-	-	0	-	72	5	8	88	3	7	-	0	-
2011	Wild	49	3	3	66	30	7	88	183	7	98	147	7	114	1	-
	Hatchery	-	0	-	55	3	2	90	1	-	81	3	5	-	0	-
2012	Wild	-	0	-	71	9	4	87	120	7	96	125	7	-	0	-
	Hatchery	-	0	-	-	0	-	-	0	-	83	1	-	-	0	-
2013	Wild	-	0	-	72	30	3	87	141	7	98	72	7	97	4	6
	Hatchery	-	0	-	-	0	-	79	1	-	96	1	-	-	0	-
Average	Wild	<b>46</b>	<b>3</b>	<b>4</b>	<b>66</b>	<b>19</b>	<b>7</b>	<b>86</b>	<b>132</b>	<b>7</b>	<b>97</b>	<b>171</b>	<b>6</b>	<b>102</b>	<b>8</b>	<b>6</b>
	Hatchery	-	<b>0</b>	-	<b>53</b>	<b>5</b>	<b>5</b>	<b>78</b>	<b>17</b>	<b>7</b>	<b>94</b>	<b>20</b>	<b>7</b>	<b>99</b>	<b>5</b>	<b>7</b>

### Sex Ratios

Male summer Chinook in the 2012 broodstock made up about 50% of the adults collected, resulting in an overall male to female ratio of 1.02:1.00 (Table 8.4.). In 2013, males made just under 50% of the adults collected, resulting in an overall male to female ratio of 0.98:1.00 (Table 8.4). The ratios in 2012 and 2013 were nearly equal to the 1:1 ratio goal in the broodstock protocol.

**Table 8.4.** Numbers of male and female wild and hatchery summer Chinook collected for broodstock in the Wenatchee River basin, 1989-2013. Ratios of males to females are also provided.

Return year	Number of wild summer Chinook			Number of hatchery summer Chinook			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
1989	166	180	0.92:1.00	0	0	-	0.92:1.00
1990	45	39	1.15:1.00	0	0	-	1.15:1.00
1991	60	68	0.88:1.00	0	0	-	0.88:1.00
1992	154	187	0.82:1.00	0	0	-	0.82:1.00
1993	208	228	0.91:1.00	35	9	3.89:1.00	1.03:1.00
1994	158	179	0.88:1.00	24	31	0.77:1.00	0.87:1.00
1995	169	213	0.79:1.00	1	15	0.07:1.00	0.75:1.00
1996	150	181	0.83:1.00	2	1	2.00:1.00	0.84:1.00
1997	104	121	0.86:1.00	15	0	-	0.98:1.00
1998	211	167	1.26:1.00	64	30	2.13:1.00	1.40:1.00
1999	130	120	1.08:1.00	108	130	0.83:1.00	0.95:1.00
2000	153	145	1.06:1.00	112	82	1.37:1.00	1.17:1.00
2001	187	124	1.51:1.00	132	50	2.64:1.00	1.83:1.00
2002	266	203	1.31:1.00	5	8	0.63:1.00	1.28:1.00
2003	270	218	1.24:1.00	5	3	1.67:1.00	1.24:1.00
2004	230	264	0.87:1.00	1	1	1.00:1.00	0.87:1.00
2005	291	200	1.46:1.00	2	1	2.00:1.00	1.46:1.00
2006	237	246	0.96:1.00	1	4	0.25:1.00	0.95:1.00
2007	239	176	1.36:1.00	2	2	1.00:1.00	1.35:1.00
2008	208	192	1.08:1.00	29	43	0.67:1.00	1.01:1.00
2009	223	236	0.94:1.00	25	7	3.57:1.00	1.02:1.00
2010	217	198	1.10:1.00	5	2	2.50:1.00	1.12:1.00
2011	198	200	0.99:1.00	4	3	1.33:1.00	0.99:1.00
2012	138	135	1.02:1.00	1	0	-	1.03:1.00
2013	127	130	0.98:1.00	1	1	1.00:1.00	0.98:1.00
<b>Total</b>	<b>4539</b>	<b>4350</b>	<b>1.04:1.00</b>	<b>574</b>	<b>423</b>	<b>1.36:1.00</b>	<b>1.07:1.00</b>

### Fecundity

Fecundities for the 2012 and 2013 returns of summer Chinook averaged 4,801 and 4,990 eggs per female, respectively (Table 8.5). These values are close to the overall average of 5,143 eggs per female. Mean observed fecundities for the 2012 and 2013 returns were near the expected fecundity of 5,099 eggs per female assumed in the broodstock protocol.

**Table 8.5.** Mean fecundity of wild, hatchery, and all female summer Chinook collected for broodstock in the Wenatchee River basin, 1989-2013; NA = not available.

Return year	Mean fecundity		
	Wild	Hatchery	Total
1989*	NA	NA	5,280
1990*	NA	NA	5,436
1991*	NA	NA	4,333
1992*	NA	NA	5,307
1993*	NA	NA	5,177
1994*	NA	NA	5,899
1995*	NA	NA	4,402
1996*	NA	NA	4,941
1997	5,385	5,272	5,390
1998	5,393	4,825	5,297
1999	5,036	4,942	4,987
2000	5,464	5,403	5,441
2001	5,280	4,647	5,097
2002	5,502	5,027	5,484
2003	5,357	5,696	5,361
2004	5,372	6,681	5,377
2005	5,045	6,391	5,053
2006	5,126	5,633	5,133
2007	5,124	4,510	5,115
2008	5,147	4,919	5,108
2009	5,308	4,765	5,291
2010	4,971	3,323	4,963
2011	4,943	2,983	4,913
2012	4,801	NA	4,801
2013	4,987	5,272	4,990
<b>Average</b>	<b>5,191</b>	<b>5,018</b>	<b>5,143</b>

\* Individual fecundities were not tracked with females until 1997.

## 8.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Based on the unfertilized egg-to-release survival standard of 81%, a total of 1,066,667 eggs were required to meet the program release goal of 864,000 smolts for brood years 1989-2011. An evaluation of the program in 2011 determined that 617,285 eggs are needed to meet the revised release goal of 500,001 smolts. This revised goal will begin with brood year 2012. From 1989 to 2011, the egg take goal was reached in seven of those years (Table 8.6). The egg take in 2012

exceeded the revised goal of 617,285 eggs, while the egg take in 2013 was lower than the revised goal.

**Table 8.6.** Numbers of eggs taken from Wenatchee summer Chinook broodstock, 1989-2013.

Return year	Number of eggs taken
1989	829,012
1990	163,109
1991	247,000
1992	827,911
1993	1,133,852
1994	999,364
1995	949,531
1996	756,000
1997	554,617
1998	854,997
1999	1,182,130
2000	1,113,159
2001	733,882
2002	1,049,255
2003	901,095
2004	1,311,051
2005	883,669
2006	1,190,757
2007	655,201
2008	1,145,330
2009	1,217,028
2010	947,875
2011	959,202
<i>Average (1989-2011)</i>	<i>895,871</i>
2012	633,677
2013	578,513
<i>Average (2012-present)</i>	<i>606,095</i>

### *Number of acclimation days*

The 2012 brood Wenatchee summer Chinook were transferred to Dryden Acclimation Pond between 17 and 27 March 2014. These fish received 34-44 days of acclimation on Wenatchee River water before being released on 30 April 2014 (Table 8.7).



**Table 8.7.** Number of days Wenatchee summer Chinook were acclimated at Dryden Acclimation Pond, brood years 1989-2012. Numbers in parenthesis represents the number of days fish reared at Chiwawa Acclimation Facility.

Brood year	Release year	Transfer date	Release date	Number of days
1989	1991	2-Mar	7-May	66
1990	1992	19-Feb	2-May	73
1991	1993	10-Mar	8-May	59
1992	1994	1-Mar	6-May	66
1993	1995	3-Mar	1-May	59
1994	1996	2-Oct	6-May	217 (154)
		5-Mar	6-May	62
1995	1997	16-Oct	8-May	205 (139)
		27-Feb	8-May	70
1996	1998	6-Oct	28-Apr	204 (142)
		25-Feb	28-Apr	62
1997	1999	23-Feb	27-Apr	63
1998	2000	5-Mar	1-May	57
1999	2001	8-Mar	23-Apr	46
2000	2002	1-Mar	6-May	66
2001	2003	19-Feb	23-Apr	63
2002	2004	5-Mar	23-Apr	49
2003	2005	15-Mar	25-Apr	41
2004	2006	25-Mar	27-Apr	33
2005	2007	15-Mar	30-Apr	46
2006	2008	11-14-Mar	28-Apr	45-48
2007	2009	30-31-Mar	29-Apr	29-30
2008	2010	9-12, 15, 22-Mar	28-Apr	38-51
2009	2011	15-18, 21-Mar, 22-Apr	26-Apr	5-43
2010	2012	26-30-Mar	25-Apr	26-30
2011	2013	25-29-Mar	24-Apr	26-30
2012	2014	17-27-Mar	30-Apr	34-44

## Release Information

### Numbers released

The 2012 Wenatchee summer Chinook program achieved 110% of the 500,001 target goal with about 550,877 fish being released in 2014 (Table 8.8).

**Table 8.8.** Numbers of Wenatchee summer Chinook smolts released from the hatchery, 1989-2012. Up to 2012, the release target for Wenatchee summer Chinook was 864,000 smolts. Beginning in 2012, the release target is 500,001 smolts.

Brood year	Release year	CWT mark rate	Number released with PIT tags	Number of smolts released
1989	1991	0.2013	0	720,000
1990	1992	0.9597	0	124,440
1991	1993	0.9957	0	191,179
1992	1994	0.9645	0	627,331
1993	1995	0.9881	0	900,429
1994	1996	0.9697	0	797,350
1995	1997	0.9725	0	687,439
1996	1998	0.9758	0	600,127
1997	1999	0.9913	0	438,223
1998	2000	0.9869	0	649,612
1999	2001	0.9728	0	1,005,554
2000	2002	0.9723	0	929,496
2001	2003	0.9868	0	604,668
2002	2004	0.9644	0	835,645
2003	2005	0.9778	0	653,764
2004	2006	0.9698	0	892,926
2005	2007	0.9596	0	644,182
2006	2008	0.9676	0	51,550 <sup>a</sup>
		0.9676	0	899,107
2007	2009	0.9768	0	456,805
2008	2010	0.9664	10,035	888,811
2009	2011	0.9767	29,930	843,866
2010	2012	0.9964	0	792,746
2011	2013	0.9904	5,020	827,709
<b>Average (1989-2011)</b>		<b>0.9761</b>	<b>1,956</b>	<b>667,085</b>
2012	2014	0.9700	19,911	550,877
<b>Average (2012-present)</b>		<b>0.9700</b>	<b>19,911</b>	<b>550,877</b>

<sup>a</sup> Represents high ELISA group planted directly in the Wenatchee River at Leavenworth Boat Launch.

### Numbers tagged

The 2012 brood Wenatchee summer Chinook were 99.0% CWT and adipose fin-clipped (Table 8.8).

In 2014, a total of 10,320 Wenatchee summer Chinook (brood year 2013) were tagged at Eastbank Hatchery in November and December. These fish were tagged in raceways #11 and #13. Those tagged in raceway #13 were designated as the “small-size fish” group (n = 5,160), while those tagged in raceway #11 were designated as the “big-size fish” group (n = 5,160). The two size groups were developed to identify techniques that maximize performance of hatchery-origin summer yearling Chinook salmon. This is part of the NOAA Fisheries, Grant PUD, and Chelan PUD size-target study. Fish were not fed during tagging or for two days before and after tagging. Fish in the small-size fish group averaged 102 mm in length and 13.8 g at time of tagging, while those in the big-size fish group averaged 95 mm in length and 10.8 g.

An additional 10,321 Wenatchee summer Chinook were tagged at Eastbank Hatchery in September 2014. These fish were tagged in water reuse circular ponds #1 and #2. Those tagged in circular pond #2 were designated as the “small-size fish” group (n = 5,161), while those tagged in circular pond #1 were designated as the “big-size fish” group (n = 5,160). This is also part of the size-target study. Fish were not fed during tagging or for two days before and after tagging. Fish in the small-size fish group averaged 75 mm in length and 6.0 g at time of tagging, while those in the big-size fish group averaged 75 mm in length and 6.0 g.

Table 8.9 summarizes the number of hatchery summer Chinook that have been PIT-tagged and released into the Wenatchee River.

**Table 8.9.** Summary of PIT-tagging activities for Wenatchee hatchery summer Chinook, brood years 2008-2013.

Brood year	Release year	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2008	2010	10,100	64	1	10,035
2009	2011	10,108 (Control)	140	3	9,965
		10,100 (R1)	129	0	9,971
		10,099 (R2)	105	0	9,994
2010	2012	0	0	0	0
2011	2013	5,100	80	0	5,020
2012	2014 (Raceway)	5,150 (small-size)	90	12	5,048
		5,153 (big-size)	379	34	4,740
2012	2014 (Reuse Circular)	5,150 (small-size)	109	0	5,041
		5,151 (big-size)	69	0	5,082

### *Fish size and condition at release*

About 550,877 summer Chinook from the 2012 brood were force-released from Dryden Acclimation Pond on 30 April 2014. Size at release was 96.9% and 89.6% of the target fork length and weight goals, respectively. This brood year was under the target CV for length (Table 8.10). One possible reason the fish did not meet the size targets is because of the size-target study. However, since the program began, Wenatchee summer Chinook have not met the target

length and CV values. The target weight (fish/pound or FPP) of juvenile fish has been met occasionally.

**Table 8.10.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of Wenatchee summer Chinook smolts released from the hatchery, brood years 1989-2012; NA = not available. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (cm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1989	1991	158	13.7	45.4	10
1990	1992	155	14.2	45.4	10
1991	1993	156	15.5	42.3	11
1992	1994	152	13.1	40.1	10
1993	1995	149	NA	34.9	13
1994	1996	138	NA	21.7	21
1995	1997	149	12.2	42.5	11
1996	1998	151	16.6	43.2	10
1997	1999	154	10.1	42.8	11
1998	2000	166	9.7	53.1	9
1999	2001	137	16.1	29.0	16
2000	2002	148	14.6	37.1	12
2001	2003	148	NA	38.9	12
2002	2004	146	15.1	37.3	14
2003	2005	147	13.2	36.5	12
2004	2006	147	10.7	35.4	13
2005	2007	153	16.3	40.6	11
2006	2008	136	21.5	29.2	16
2007	2009	163	21.6	49.7	9
2008	2010	166	15.0	52.0	9
2009	2011	152	15.9	39.0	12
2010	2012	154	17.2	43.1	11
2011	2013	149	13.8	41.4	11
<i>Average (1989-2011)</i>		<i>151</i>	<i>14.8</i>	<i>40.0</i>	<i>12</i>
<i>Targets (1989-2011)</i>		<i>176</i>	<i>9.0</i>	<i>45.4</i>	<i>10</i>
2012	2014	158	12.6	40.7	11
<i>Average (2012-present)</i>		<i>158</i>	<i>12.6</i>	<i>40.7</i>	<i>11</i>
<i>Targets (2012-present)<sup>a</sup></i>		<i>163</i>	<i>9.0</i>	<i>45.4</i>	<i>10</i>

<sup>a</sup> For brood year 2012, the fish per pound (fpp) targets were 10 fpp and 15 fpp.

### Survival Estimates

Overall survival of the 2012 brood Wenatchee summer Chinook from green (unfertilized) egg to release was higher than the standard set for the program. This was in part because of a high ponding to release survival (Table 8.11).

**Table 8.11.** Hatchery life-stage survival rates (%) for Wenatchee summer Chinook, brood years 1989-2012. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
1989	90.0	93.4	90.9	97.0	99.7	99.3	98.5	99.4	86.9
1990	89.7	95.6	80.9	96.6	99.6	99.2	97.7	98.8	76.3
1991	88.2	98.3	86.9	96.1	99.3	98.5	94.9	98.1	77.4
1992	84.3	92.2	79.8	97.8	99.9	99.9	97.1	98.1	75.8
1993	92.4	95.9	84.2	97.5	99.6	99.3	96.7	98.8	79.4
1994	90.7	95.3	83.7	100	99.2	97.0	95.3	98.4	79.8
1995	94.7	98.2	86.0	100	96.7	96.4	74.9	90.8	72.4
1996	84.6	96.1	84.1	100	97.9	97.7	94.4	97.7	79.4
1997	89.3	98.3	82.6	97.3	97.1	96.9	98.3	98.2	79.0
1998	85.3	94.6	80.9	98.3	99.4	98.6	95.6	99.8	76.0
1999	98.4	98.3	90.4	97.9	98.1	97.9	96.2	99.4	85.1
2000	93.0	96.6	88.3	98.0	99.6	99.3	96.5	98.9	83.5
2001	87.4	91.5	90.6	97.7	99.8	99.6	93.1	93.3	82.4
2002	93.8	94.1	85.1	99.8	98.1	97.6	93.7	96.5	79.6
2003	77.4	85.1	80.5	98.1	99.6	99.1	91.9	93.5	72.6
2004	92.8	97.8	85.7	87.8	99.9	99.6	86.6	92.1	65.1
2005	97.3	89.6	83.5	98.0	99.7	99.4	89.1	99.5	72.9
2006	92.4	95.2	85.6	98.4	99.3	98.4	94.8	97.2	79.8
2007	73.6	97.5	73.7	97.9	99.5	98.7	96.6	99.1	69.7
2008	96.6	97.9	90.4	97.3	99.4	98.7	88.2	89.6	77.6
2009	95.1	95.6	92.0	99.6	97.3	97.3	84.8	98.2	78.1
2010	94.7	97.8	96.1	99.3	97.6	97.1	87.2	90.3	83.2
2011	98.0	96.4	92.3	97.9	99.5	98.9	95.9	97.3	86.7
2012	97.8	97.2	92.3	98.1	99.7	99.1	96.1	97.3	86.9
<i>Average</i>	<i>97.0</i>	<i>95.4</i>	<i>86.1</i>	<i>97.8</i>	<i>99.0</i>	<i>98.5</i>	<i>93.1</i>	<i>96.7</i>	<i>78.6</i>
<i>Standard</i>	<i>90.0</i>	<i>85.0</i>	<i>92.0</i>	<i>98.0</i>	<i>97.0</i>	<i>93.0</i>	<i>90.0</i>	<i>95.0</i>	<i>81.0</i>

### 8.3 Disease Monitoring

Rearing of the 2012 brood Wenatchee summer Chinook was similar to previous years with fish being held on well water before being transferred to Dryden Acclimation Pond for final acclimation in March 2014. Fish were transferred to Dryden Acclimation Pond from 17-27 March. Increased mortality caused by external fungus and bacterial cold water disease began to occur during the acclimation period at Dryden Acclimation Pond at which time a formalin treatment was initiated to prevent the fungus from proliferating in combination with initiation of an early release.

Results of the 2014 adult broodstock bacterial kidney disease (BKD) monitoring indicated that most females (94.6%) had ELISA values less than 0.199. The seven females that had an ELISA value greater than 0.120 were not included in the program and the eggs were culled. All remaining females had ELISA values less than 0.120, which means that none of the progeny needed to be reared at densities less than 0.06 fish per pound (Table 8.12).

**Table 8.12.** Proportion of bacterial kidney disease (BKD) titer groups for the Wenatchee summer Chinook broodstock, brood years 1997-2014. Also included are the proportions to be reared at either 0.125 fish per pound or 0.060 fish per pound.

Brood year <sup>a</sup>	Optical density values by titer group				Proportion at rearing densities (fish per pound, fpp)	
	Very Low (≤ 0.099)	Low (0.1-0.199)	Moderate (0.2-0.449)	High (≥ 0.450)	≤ 0.125 fpp (<0.119)	≤ 0.060 fpp (>0.120)
1997	0.7714	0.0857	0.0381	0.1048	0.8095	0.1905
1998	0.3067	0.2393	0.1656	0.2883	0.4479	0.5521
1999	0.9590	0.0123	0.0123	0.0164	0.9713	0.0287
2000	0.6268	0.1053	0.1627	0.1053	0.7321	0.2679
2001	0.6513	0.0263	0.0987	0.2237	0.6776	0.3224
2002	0.7868	0.0457	0.0711	0.0964	0.8325	0.1675
2003	0.9825	0.0000	0.0058	0.0117	0.9825	0.0175
2004	0.9593	0.0081	0.0163	0.0163	0.9675	0.0325
2005	0.9833	0.0056	0.0000	0.0111	0.9833	0.0167
2006	0.9134	0.0563	0.0000	0.0303	0.9351	0.0649
2007	0.9535	0.0078	0.0078	0.0310	0.9535	0.0465
2008	0.9868	0.0088	0.0044	0.0000	0.9868	0.0132
2009	0.9957	0.0000	0.0000	0.0043	0.9957	0.0043
2010	0.9897	0.0025	0.0000	0.0025	0.9949	0.0051
2011	0.9585	0.0363	0.0000	0.0052	0.9896	0.0104
2012	0.9697	0.0303	0.0000	0.0000	1.0000	0.0000
2013	0.8120	0.1790	0.0000	0.0090	0.8890	0.1110
2014	0.9462	0.0154	0.0000	0.0385	0.9462	0.0538
<b>Average</b>	<b>0.8640</b>	<b>0.0480</b>	<b>0.0324</b>	<b>0.0553</b>	<b>0.8942</b>	<b>0.1058</b>

<sup>a</sup> Individual ELISA samples were not collected before the 1997 brood.

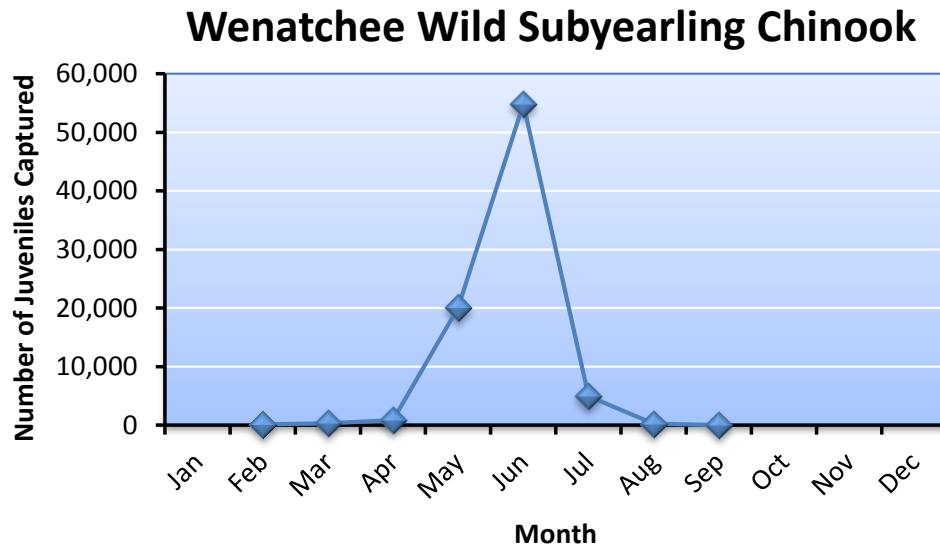
## 8.4 Natural Juvenile Productivity

During 2014, juvenile summer Chinook were sampled at the Lower Wenatchee Trap located near the Town of Cashmere. Because the Lower Wenatchee Trap began operation in a new location in 2013, the historic flow-discharge relationships are invalid and new models to estimate trap efficiency must be developed for all species. Until the new models are developed (2-3 years) all estimates of juvenile abundance should be considered preliminary.

## Emigrant Estimates

### Lower Wenatchee Trap

The Lower Wenatchee Trap operated between 12 February and 7 September 2014. During that time period the trap was inoperable for 12 days because of high river flows, debris, snow/ice, mechanical failure, or major hatchery releases. During the seven-month sampling period, a total of 81,445 wild subyearling Chinook were captured at the Lower Wenatchee Trap. Based on nineteen capture efficiencies estimated from the flow model, the total number of wild subyearling Chinook that emigrated past the Lower Wenatchee Trap was 11,936,928 ( $\pm 2,448,536$ ). Because 172 summer Chinook redds were observed downstream from the trap in 2013, the total number of summer Chinook emigrating from the Wenatchee River in 2014 was expanded using the ratio of the number of redds downstream from the trap to the number upstream from the trap. This resulted in a total summer Chinook emigrant estimate of 12,605,925 fish. Most of these fish emigrated during early June (Figure 8.1). Monthly captures and mortalities of all fish collected at the Lower Wenatchee Trap are reported in Appendix B.



**Figure 8.1.** Numbers of wild subyearling Chinook captured at the Lower Wenatchee Trap during February through October, 2014.

## 8.5 Spawning Surveys

Surveys for Wenatchee summer Chinook redds were conducted from late 15 September to 17 November 2014 in the Wenatchee River and Icicle Creek.

### Redd Counts

A total count of summer Chinook redds was estimated in 2014 based on weekly census surveys conducted in the Wenatchee River. Redds were counted in Icicle Creek when feasible (Table 8.13).

**Table 8.13.** Numbers of redds counted in the Wenatchee River basin, 1989-2014; ND = no data. From 1989-2013, numbers of redds were based on “peak counts.” Total counts in those years were based on expanded peak counts. Since 2014, numbers of redds were based on census surveys.

Survey year	Redd counts		Total count
	Wenatchee River	Icicle Creek	
1989	3,331	ND	4,215
1990	2,479	ND	3,103
1991	2,180	ND	2,748
1992	2,328	ND	2,913
1993	2,334	ND	2,953
1994	2,426	ND	3,077
1995	1,872	ND	2,350
1996	1,435	ND	1,814
1997	1,388	ND	1,739
1998	1,660	ND	2,230
1999	2,188	ND	2,738
2000	2,022	ND	2,540
2001	2,857	ND	3,550
2002	5,419	ND	6,836
2003	4,281	ND	5,268
2004	4,003	ND	4,874
2005	2,895	ND	3,538
2006	7,165	68	8,896
2007	1,857	13	1,970
2008	2,338	23	2,800
2009	2,667	21	3,441
2010	2,553	11	3,261
2011	2,583	9	3,078
2012	2,301	2	2,504
2013	2,875	42	3,241
2014	3,383	75	3,458
<i>Average</i>			<b>3,428</b>

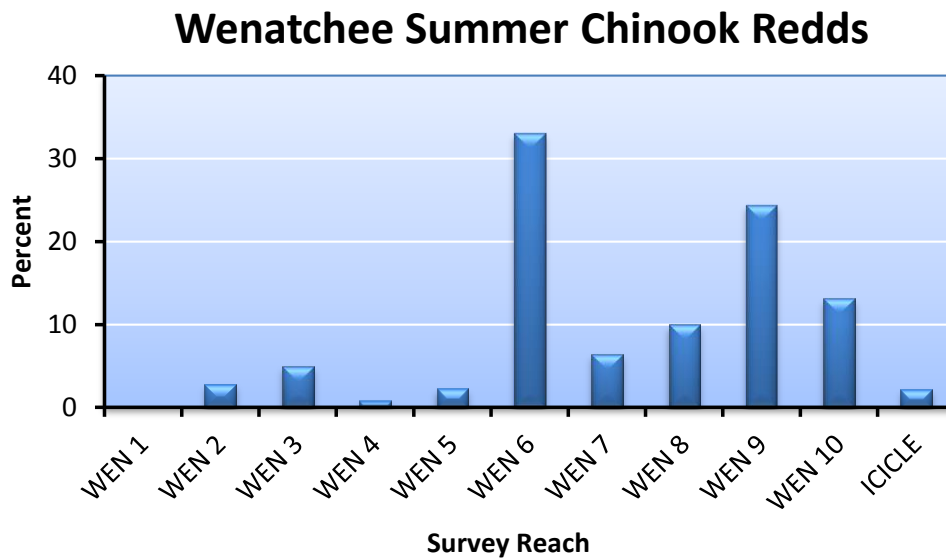
### Redd Distribution

Summer Chinook redds were not evenly distributed among reaches within the Wenatchee River basin in 2014 (Table 8.14; Figure 8.2). Most of the spawning occurred upstream from the Leavenworth Bridge in Reaches 6, 9, and 10. The highest density of redds occurred in Reach 6 near the confluence of the Icicle River. In September, the counting of redds downstream from the confluence of Chiwaukum Creek (Reaches 1-8) was hampered because of recruitment of suspended sediments from the Chiwaukum Complex fire. High flows and turbidity also hampered redd surveys in November.



**Table 8.14.** Total numbers of summer Chinook redds counted in different reaches in the Wenatchee River basin during September through mid-November, 2014. Reach codes are described in Table 2.10.

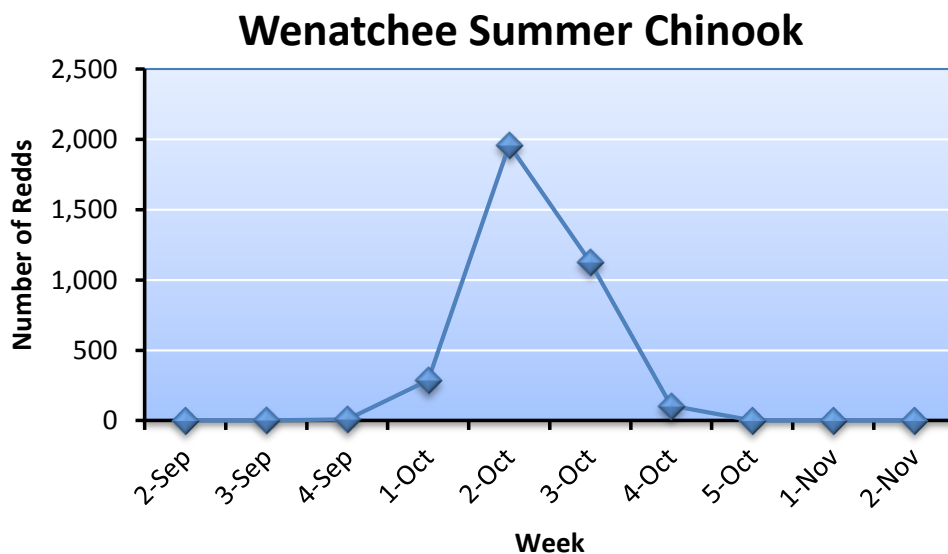
Survey reach	Total redd count
Wenatchee 1 (W1)	3
Wenatchee 2 (W2)	98
Wenatchee 3 (W3)	172
Wenatchee 4 (W4)	27
Wenatchee 5 (W5)	78
Wenatchee 6 (W6)	1,144
Wenatchee 7 (W7)	222
Wenatchee 8 (W8)	344
Wenatchee 9 (W9)	842
Wenatchee 10 (W10)	453
Icicle Creek (I1)	75
<b>Totals</b>	<b>3,458</b>



**Figure 8.2.** Percent of the total number of summer Chinook redds counted in different reaches in the Wenatchee River basin during September through early-November, 2014. Reach codes are described in Table 2.10.

### Spawn Timing

In 2014, spawning in the Wenatchee River began during the third week in September, peaked in the second week of October, and ended in the fourth week of October (Figure 8.3).



**Figure 8.3.** Number of new summer Chinook redds counted during different weeks in the Wenatchee River, September through mid-November 2014.

### Spawning Escapement

Spawning escapement for Wenatchee summer Chinook was calculated as the total number of redds (expanded peak counts for return years 1989-2013) times the fish per redd ratio estimated from broodstock and fish sampled at adult trapping sites. The estimated fish per redd ratio for summer Chinook in 2014 was 3.02. Multiplying this ratio by the number of redds counted in the Wenatchee River basin resulted in a total spawning escapement of 10,443 summer Chinook (Table 8.15).

**Table 8.15.** Spawning escapements for summer Chinook in the Wenatchee River basin, return years 1989-2014. Number of redds is based on expanded peak redd counts.

Return year	Fish/Redd	Redds	Total spawning escapement
1989	3.40	4,215	14,331
1990	3.50	3,103	10,861
1991	3.70	2,748	10,168
1992	4.00	2,913	11,652
1993	3.20	2,953	9,450
1994	3.30	3,077	10,154
1995	3.30	2,350	7,755
1996	3.40	1,814	6,168
1997	3.40	1,739	5,913
1998	2.40	2,230	5,352
1999	2.00	2,738	5,476
2000	2.17	2,540	5,512
2001	3.20	3,550	11,360
2002	2.30	6,836	15,723

Return year	Fish/Redd	Redds	Total spawning escapement
2003	2.24	5,268	11,800
2004	2.15	4,874	10,479
2005	2.46	3,538	8,703
2006	2.00	8,896	17,792
2007	2.33	1,970	4,590
2008	2.32	2,800	6,496
2009	2.42	3,441	8,327
2010	2.29	3,261	7,468
2011	3.20	3,078	9,850
2012	3.41	2,504	8,539
2013	3.15	3,241	10,209
2014	3.02	3,458	10,443
<b>Average</b>	<b>2.86</b>	<b>3,428</b>	<b>10,443</b>

## 8.6 Carcass Surveys

Surveys for Wenatchee summer Chinook carcasses were conducted during late September to early November 2014 in the Wenatchee River and Icicle Creek.

### Number sampled

A total of 1,723 summer Chinook carcasses were sampled during October through early November in the Wenatchee River basin in 2014 (Table 8.16).

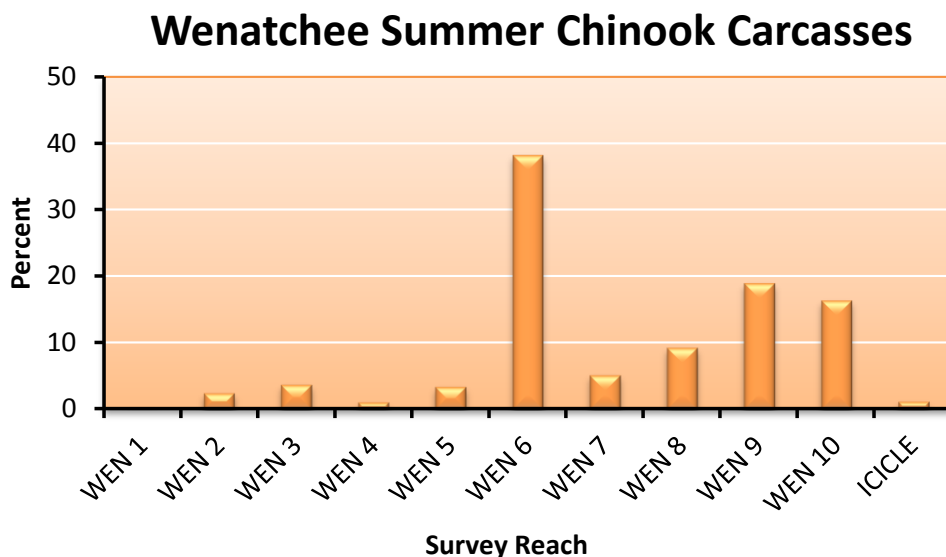
**Table 8.16.** Numbers of summer Chinook carcasses sampled within each survey reach in the Wenatchee River basin, 1993-2014. Reach codes are described in Table 2.10.

Survey year	Number of summer Chinook carcasses											Total
	W-1	W-2	W-3	W-4	W-5	W-6	W-7	W-8	W-9	W-10	Icicle	
1993	68	151	696	13	82	150	215	41	0	0	0	<b>1,416</b>
1994	0	6	25	1	21	50	20	49	131	1	0	<b>304</b>
1995	0	10	14	0	0	117	50	37	20	0	0	<b>248</b>
1996	0	5	84	42	10	206	27	37	43	0	0	<b>454</b>
1997	1	47	127	5	29	312	8	80	70	13	0	<b>692</b>
1998	6	81	159	4	1	270	32	395	354	65	0	<b>1,367</b>
1999	0	169	112	16	35	932	68	146	185	79	0	<b>1,742</b>
2000	8	118	178	9	85	693	82	121	172	208	0	<b>1,674</b>
2001	0	49	138	31	0	338	36	124	101	94	0	<b>911</b>
2002	0	249	189	0	205	848	0	341	564	166	6	<b>2,568</b>
2003	6	369	195	72	149	768	66	266	537	58	40	<b>2,526</b>
2004	8	157	193	177	173	1,086	103	346	493	409	16	<b>3,161</b>
2005	8	85	106	39	46	709	70	140	353	258	7	<b>1,821</b>
2006	22	140	160	64	112	953	435	343	703	658	18	<b>3,608</b>

Survey year	Number of summer Chinook carcasses											
	W-1	W-2	W-3	W-4	W-5	W-6	W-7	W-8	W-9	W-10	Icicle	Total
2007	3	15	49	10	26	475	38	38	96	91	8	<b>849</b>
2008	10	34	63	38	36	676	47	42	106	144	8	<b>1,204</b>
2009	11	29	43	32	27	389	16	58	240	175	6	<b>1,026</b>
2010	3	31	98	57	122	681	135	49	124	194	15	<b>1,509</b>
2011	5	88	126	19	38	1,332	77	45	211	289	9	<b>2,239</b>
2012	8	82	95	22	40	600	53	62	173	183	0	<b>1,318</b>
2013	3	100	149	22	109	767	5	60	353	265	14	<b>1,847</b>
2014	3	42	64	18	59	659	89	160	327	282	20	<b>1,723</b>
<i>Average</i>	<i>8</i>	<i>94</i>	<i>139</i>	<i>31</i>	<i>64</i>	<i>591</i>	<i>76</i>	<i>135</i>	<i>243</i>	<i>165</i>	<i>8</i>	<i>1,555</i>

### Carcass Distribution and Origin

Summer Chinook carcasses were not evenly distributed among reaches within the Wenatchee River basin in 2014 (Table 8.16; Figure 8.4). Most of the carcasses in the Wenatchee River basin were found upstream from the Leavenworth Bridge. The highest percentage of carcasses (38%) was sampled in Reach 6 near the confluence of the Icicle River. As with redd surveys, in September, carcass surveys downstream from the confluence of Chiwaukum Creek (Reaches 1-8) were hampered because of recruitment of suspended sediments from the Chiwaukum Complex fire. High flows and turbidity also limited carcass surveys in November.



**Figure 8.4.** Percent of summer Chinook carcasses sampled within different reaches in the Wenatchee River basin during September through mid-November, 2014. Reach codes are described in Table 2.10.

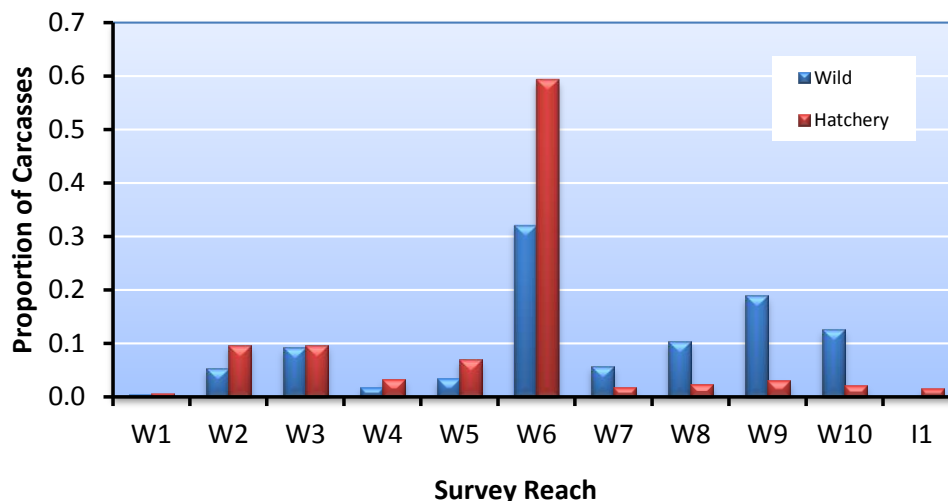
Numbers of wild and hatchery-origin summer Chinook carcasses sampled in 2014 will be available after analysis of CWTs and scales. Based on the available data (1993-2013), most fish, regardless of origin, were found in Reach 6 (Leavenworth Bridge to Icicle Road Bridge) (Table 8.17). In general, a larger percentage of wild fish were found in the upper reaches than were hatchery fish (Figure 8.5). In contrast, a larger percentage of hatchery fish were found in reaches downstream from the Icicle Road Bridge.

**Table 8.17.** Numbers of wild and hatchery summer Chinook carcasses sampled within different reaches in the Wenatchee River basin, 1993-2013.

Survey year	Origin	Survey reach											Total
		W-1	W-2	W-3	W-4	W-5	W-6	W-7	W-8	W-9	W-10	Icicle	
1993	Wild	59	146	660	12	82	133	213	40	0	0	0	<b>1,345</b>
	Hatchery	9	5	36	1	0	17	2	1	0	0	0	<b>71</b>
1994	Wild	0	2	18	1	19	36	20	49	130	1	0	<b>276</b>
	Hatchery	0	4	7	0	2	14	0	0	1	0	0	<b>28</b>
1995	Wild	0	4	11	0	0	105	50	35	20	0	0	<b>225</b>
	Hatchery	0	6	3	0	0	12	0	2	0	0	0	<b>23</b>
1996	Wild	0	5	82	40	9	196	27	37	43	0	0	<b>439</b>
	Hatchery	0	0	2	2	1	10	0	0	0	0	0	<b>15</b>
1997	Wild	1	38	112	5	22	266	8	80	69	13	0	<b>614</b>
	Hatchery	0	9	15	0	7	46	0	0	1	0	0	<b>78</b>
1998	Wild	6	62	124	3	1	191	29	374	327	62	0	<b>1,179</b>
	Hatchery	0	19	35	1	0	79	3	21	27	3	0	<b>188</b>
1999	Wild	0	88	70	8	18	600	58	137	169	75	0	<b>1,223</b>
	Hatchery	0	81	42	8	17	332	10	9	16	4	0	<b>519</b>
2000	Wild	5	78	115	8	57	485	75	110	167	200	0	<b>1,300</b>
	Hatchery	3	40	63	1	28	208	7	11	5	8	0	<b>374</b>
2001	Wild	0	37	100	9	0	245	32	122	97	91	0	<b>733</b>
	Hatchery	0	12	38	22	0	93	4	2	4	3	0	<b>178</b>
2002	Wild	0	151	127	0	103	479	0	330	558	161	3	<b>1,912</b>
	Hatchery	0	98	62	0	102	369	0	11	6	5	3	<b>656</b>
2003	Wild	5	261	147	32	111	519	62	252	498	57	15	<b>1,959</b>
	Hatchery	1	108	48	40	38	249	4	14	39	1	25	<b>567</b>
2004	Wild	7	124	163	120	112	749	90	316	481	399	11	<b>2,572</b>
	Hatchery	1	33	30	56	61	337	13	30	12	10	5	<b>588</b>
2005	Wild	4	49	78	24	26	399	66	125	336	244	0	<b>1,351</b>
	Hatchery	4	36	28	15	20	310	4	15	17	14	7	<b>470</b>
2006	Wild	15	91	122	44	75	688	388	309	646	593	5	<b>2,976</b>
	Hatchery	7	49	38	20	37	265	47	34	57	65	13	<b>632</b>
2007	Wild	1	7	24	1	10	197	34	30	95	81	3	<b>483</b>
	Hatchery	2	8	25	9	16	278	4	8	1	10	5	<b>366</b>
2008	Wild	7	15	38	24	21	361	41	31	98	133	2	<b>771</b>
	Hatchery	3	19	25	14	15	315	6	11	8	11	6	<b>433</b>
2009	Wild	6	22	32	23	19	288	13	55	236	173	4	<b>871</b>

Survey year	Origin	Survey reach											Total	
		W-1	W-2	W-3	W-4	W-5	W-6	W-7	W-8	W-9	W-10	Icicle		
2010	Hatchery	5	7	11	9	8	101	3	3	4	2	2	155	
	Wild	2	22	62	44	64	477	125	47	121	192	0	1,156	
2011	Hatchery	1	9	36	13	58	204	10	2	3	2	15	353	
	Wild	4	46	75	11	25	914	74	45	211	287	3	1,695	
2012	Hatchery	1	42	51	7	13	418	3	0	0	2	6	543	
	Wild	4	49	72	13	24	490	47	62	173	182	0	1,116	
2013	Hatchery	4	33	23	9	16	110	6	0	0	1	0	202	
	Wild	1	63	89	16	69	374	5	59	340	261	2	1,279	
Average	Hatchery	2	52	60	6	40	395	0	1	12	4	12	585	
	Wild	6	65	111	21	41	390	69	126	229	153	2	1,213	
		Hatchery	2	32	32	11	23	198	6	8	10	7	5	334

### Wenatchee Summer Chinook



**Figure 8.5.** Distribution of wild and hatchery produced carcasses in different reaches in the Wenatchee River basin, 1993-2013. Reach codes are described in Table 2.10.

### Sampling Rate

If escapement is based on total numbers of redds, then about 16% of the total spawning escapement of summer Chinook in the Wenatchee River basin was sampled in 2014 (Table 8.18). Sampling rates among survey reaches varied from 9 to 25%. Carcass surveys in 2014 were significantly influenced by poor weather conditions and unseasonable high flows.

**Table 8.18.** Number of redds and carcasses, total spawning escapement, and sampling rates for summer Chinook in the Wenatchee River basin, 2014.

Sampling reach	Total number of redds	Total number of carcasses	Total spawning escapement	Sampling rate
Wenatchee 1 (W1)	3	3	9	0.33
Wenatchee 2 (W2)	98	42	296	0.14
Wenatchee 3 (W3)	172	64	519	0.12
Wenatchee 4 (W4)	27	18	82	0.22
Wenatchee 5 (W5)	78	59	236	0.25
Wenatchee 6 (W6)	1,144	659	3,455	0.19
Wenatchee 7 (W7)	222	89	670	0.13
Wenatchee 8 (W8)	344	160	1,039	0.15
Wenatchee 9 (W9)	842	327	2,543	0.13
Wenatchee 10 (W10)	453	282	1,368	0.21
Icicle Creek (I1)	75	20	227	0.09
<b>Total</b>	<b>3,458</b>	<b>1,723</b>	<b>10,443</b>	<b>0.16</b>

### Length Data

Mean lengths (POH, cm) of male and female summer Chinook carcasses sampled during surveys in the Wenatchee River basin in 2014 are provided in Table 8.19. The average size of males and females sampled in the Wenatchee River basin were 66 cm and 69 cm, respectively.

**Table 8.19.** Mean lengths (postorbital-to-hypural length; cm) and standard deviations (in parentheses) of male and female summer Chinook carcasses sampled in different streams/watersheds in the Wenatchee River basin, 2014.

Stream/watershed	Mean length (cm)	
	Male	Female
Wenatchee 1 (W1)	66.3 (2.3)	--
Wenatchee 2 (W2)	64.5 (12.3)	68.6 (3.3)
Wenatchee 3 (W3)	60.7 (10.3)	66.5 (6.7)
Wenatchee 4 (W4)	63.7 (9.1)	71.8 (4.8)
Wenatchee 5 (W5)	66.0 (10.2)	69.7 (3.6)
Wenatchee 6 (W6)	64.6 (8.5)	69.2 (4.8)
Wenatchee 7 (W7)	63.1 (10.2)	70.5 (6.2)
Wenatchee 8 (W8)	66.0 (10.4)	67.2 (5.2)
Wenatchee 9 (W9)	68.7 (7.6)	71.0 (5.0)
Wenatchee 10 (W10)	66.3 (8.1)	67.9 (5.1)
Icicle Creek (I1)	--	--
<b>Total</b>	<b>65.5 (9.0)</b>	<b>69.2 (5.2)</b>

## 8.7 Life History Monitoring

Life history characteristics of Wenatchee summer Chinook were assessed by examining carcasses on spawning grounds and fish collected or examined at broodstock collection sites, and by reviewing tagging data and fisheries statistics.

### Migration Timing

Migration timing of hatchery and wild Wenatchee summer Chinook was determined from broodstock data and stock assessment data collected at Dryden Dam. Sampling at Dryden Dam occurs from early July through mid-October. On average, during the early part of the migration, hatchery summer Chinook arrived about one week later than wild Chinook (Table 8.20). This pattern carried through the migration distribution of summer Chinook at Dryden Dam. By the end of the migration, hatchery fish pass Dryden Dam about five weeks after 90% of the wild fish passed the dam.

**Table 8.20.** The week that 10%, 50% (median), and 90% of the wild and hatchery summer Chinook salmon passed Dryden Dam, 2007-2014. The average week is also provided. Migration timing is based on collection of summer Chinook broodstock at Dryden Dam.

Survey year	Origin	Wenatchee Summer Chinook Migration Time (week)				Sample size
		10 Percentile	50 Percentile	90 Percentile	Mean	
2007	Wild	28	31	37	31	274
	Hatchery	30	33	41	35	305
2008	Wild	29	31	40	32	219
	Hatchery	32	37	41	37	576
2009	Wild	27	29	41	31	469
	Hatchery	28	34	42	35	382
2010	Wild	30	33	35	32	403
	Hatchery	29	30	33	30	268
2011	Wild	30	31	34	32	293
	Hatchery	32	34	39	35	304
2012	Wild	30	32	39	33	247
	Hatchery	31	37	41	36	366
2013	Wild	28	30	34	31	494
	Hatchery	29	33	39	33	570
2014	Wild	29	31	37	32	512
	Hatchery	29	32	40	33	338
<i>Average</i>	<i>Wild</i>	<i>28</i>	<i>31</i>	<i>36</i>	<i>32</i>	<i>2,911</i>
	<i>Hatchery</i>	<i>29</i>	<i>33</i>	<i>41</i>	<i>34</i>	<i>3,109</i>

### Age at Maturity

Because hatchery summer Chinook are released after one year of rearing and natural-origin summer Chinook migrate primarily as age-0 fish, total ages will differ between hatchery and



natural-origin Chinook (see Hillman et al. 2011). Therefore, in this section, we evaluated age at maturity by comparing differences in salt (ocean) ages between the two groups.

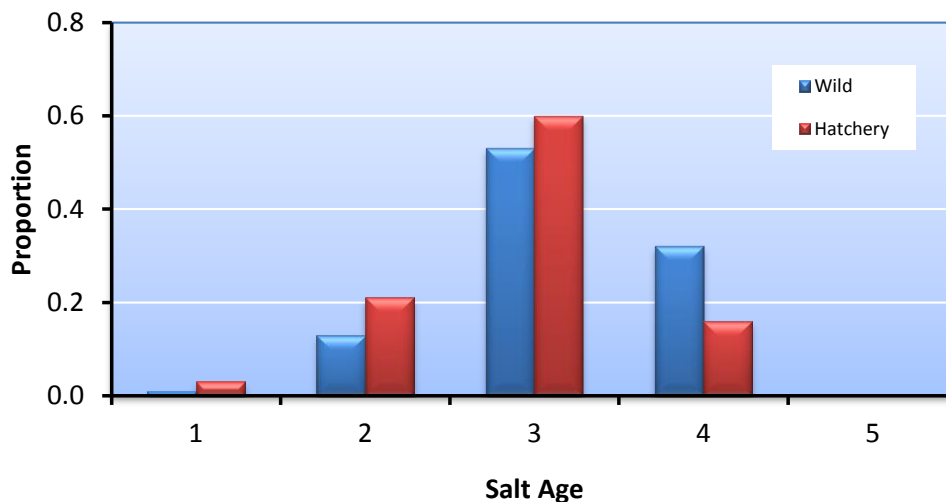
Most of the wild and hatchery summer Chinook sampled during the period 1993-2013 in the Wenatchee River basin were salt age-3 fish (Table 8.21; Figure 8.6). Over the survey years, a higher percentage of salt age-4 wild Chinook returned to the basin than did salt age-4 hatchery Chinook. In contrast, a higher proportion of salt age-1 and 2 hatchery fish returned than did salt age-1 and 2 wild fish. Thus, a higher percentage of wild fish returned at an older age than did hatchery fish.

**Table 8.21.** Proportions of wild and hatchery summer Chinook of different salt (ocean) ages sampled on spawning grounds in the Wenatchee River basin, 1993-2013.

Sample year	Origin	Salt age					Sample size
		1	2	3	4	5	
1993	Wild	0.02	0.24	0.62	0.12	0.00	1,224
	Hatchery	0.03	0.91	0.03	0.03	0.00	64
1994	Wild	0.02	0.21	0.45	0.32	0.00	257
	Hatchery	0.00	0.14	0.86	0.00	0.00	21
1995	Wild	0.02	0.15	0.65	0.18	0.00	216
	Hatchery	0.00	0.00	0.05	0.95	0.00	21
1996	Wild	0.01	0.25	0.66	0.08	0.00	512
	Hatchery	0.00	0.33	0.33	0.29	0.05	21
1997	Wild	0.01	0.24	0.57	0.18	0.00	561
	Hatchery	0.05	0.20	0.67	0.08	0.00	75
1998	Wild	0.02	0.23	0.66	0.09	0.00	1,041
	Hatchery	0.03	0.49	0.38	0.10	0.00	187
1999	Wild	0.01	0.34	0.55	0.10	0.00	1,087
	Hatchery	0.01	0.15	0.79	0.05	0.00	510
2000	Wild	0.02	0.20	0.64	0.15	0.00	1,181
	Hatchery	0.07	0.11	0.66	0.15	0.00	342
2001	Wild	0.01	0.16	0.74	0.08	0.00	653
	Hatchery	0.05	0.76	0.14	0.04	0.00	181
2002	Wild	0.00	0.14	0.62	0.24	0.00	1,744
	Hatchery	0.01	0.16	0.80	0.02	0.00	646
2003	Wild	0.01	0.07	0.51	0.41	0.00	1,653
	Hatchery	0.05	0.07	0.75	0.12	0.00	530
2004	Wild	0.00	0.12	0.32	0.54	0.01	2,233
	Hatchery	0.08	0.57	0.25	0.10	0.00	566
2005	Wild	0.00	0.12	0.75	0.13	0.00	1,190
	Hatchery	0.02	0.09	0.86	0.03	0.00	450
2006	Wild	0.00	0.02	0.27	0.71	0.00	2,972
	Hatchery	0.02	0.16	0.24	0.57	0.00	299

Sample year	Origin	Salt age					Sample size
		1	2	3	4	5	
2007	Wild	0.01	0.09	0.31	0.53	0.07	480
	Hatchery	0.00	0.15	0.75	0.07	0.03	275
2008	Wild	0.01	0.06	0.76	0.17	0.00	767
	Hatchery	0.02	0.12	0.76	0.11	0.00	329
2009	Wild	0.01	0.07	0.51	0.41	0.00	797
	Hatchery	0.10	0.36	0.49	0.05	0.00	132
2010	Wild	0.01	0.18	0.65	0.16	0.00	1,068
	Hatchery	0.00	0.49	0.47	0.03	0.00	294
2011	Wild	0.01	0.11	0.60	0.29	0.00	1,533
	Hatchery	0.06	0.04	0.90	0.01	0.00	472
2012	Wild	0.00	0.04	0.48	0.48	0.00	1,017
	Hatchery	0.00	0.03	0.88	0.08	0.03	200
2013	Wild	0.01	0.10	0.61	0.28	0.00	1,148
	Hatchery	0.00	0.01	0.15	0.83	0.00	487
Average	Wild	0.01	0.13	0.53	0.32	0.00	1,111
	Hatchery	0.03	0.21	0.60	0.16	0.00	291

### Wenatchee Summer Chinook



**Figure 8.6.** Proportions of wild and hatchery summer Chinook of different salt (ocean) ages sampled at broodstock collection sites and on spawning grounds in the Wenatchee River basin for the combined years 1993-2013.

#### Size at Maturity

On average, hatchery summer Chinook were about 4 cm smaller than wild summer Chinook sampled in the Wenatchee River basin (Table 8.22). This is likely because a higher percentage of

hatchery fish returned as salt age-2 and 3 fish than did wild fish. In contrast, a higher percentage of wild fish returned as salt age-4 fish than did hatchery fish. Analyses for the five-year reports will compare sizes of hatchery and wild fish of the same age groups and sex.

**Table 8.22.** Mean lengths (POH; cm) and variability statistics for wild and hatchery summer Chinook sampled in the Wenatchee River basin, 1993-2013; SD = 1 standard deviation.

Sample year	Origin	Sample size	Summer Chinook length (POH; cm)			
			Mean	SD	Minimum	Maximum
1993 <sup>a</sup>	Wild	1,344	73	8	33	94
	Hatchery	68	61	9	37	83
1994 <sup>a</sup>	Wild	276	73	8	31	89
	Hatchery	25	70	8	54	85
1995 <sup>a</sup>	Wild	225	75	7	48	87
	Hatchery	23	74	7	57	85
1996 <sup>a</sup>	Wild	210	74	7	43	92
	Hatchery	9	66	12	52	84
1997	Wild	614	74	8	29	99
	Hatchery	79	69	10	29	83
1998	Wild	1,179	73	8	28	97
	Hatchery	188	67	10	37	87
1999	Wild	1,217	72	8	29	95
	Hatchery	518	71	8	26	94
2000	Wild	1,301	71	10	24	94
	Hatchery	369	69	11	33	91
2001	Wild	728	70	9	30	93
	Hatchery	178	63	10	28	86
2002	Wild	1,911	72	8	39	94
	Hatchery	656	71	8	34	95
2003	Wild	1,943	74	9	24	105
	Hatchery	554	69	10	26	97
2004	Wild	2,570	72	9	32	98
	Hatchery	584	59	11	25	91
2005	Wild	1,352	69	7	41	92
	Hatchery	469	69	8	39	91
2006	Wild	3,249	74	6	29	99
	Hatchery	350	71	9	35	90
2007	Wild	566	73	9	29	92
	Hatchery	269	70	7	45	87
2008	Wild	836	69	8	29	89
	Hatchery	363	70	9	24	94
2009	Wild	872	71	8	30	94

Sample year	Origin	Sample size	Summer Chinook length (POH; cm)			
			Mean	SD	Minimum	Maximum
	Hatchery	153	64	11	32	84
2010	Wild	1,147	68	8	32	92
	Hatchery	351	65	10	25	87
2011	Wild	1,698	68	8	33	101
	Hatchery	541	66	9	34	85
2012	Wild	1,116	70	7	29	91
	Hatchery	202	60	7	40	79
2013	Wild	1,279	66	9	24	95
	Hatchery	584	67	7	5	85
<i>Pooled</i>	<i>Wild</i>	<i>1,221</i>	<i>71</i>	<i>8</i>	<i>32</i>	<i>94</i>
	<i>Hatchery</i>	<i>311</i>	<i>67</i>	<i>9</i>	<i>34</i>	<i>88</i>

<sup>a</sup> These years include sizes reported in annual reports. The data contained in the WDFW database do not include all these data.

### Contribution to Fisheries

Most of the harvest on hatchery-origin Wenatchee summer Chinook occurred in the ocean (Table 8.23). Ocean harvest has made up 47% to 100% of all hatchery Wenatchee summer Chinook harvested. Total harvest on early brood years (1990-1996 and 2007) was lower than for brood years 1997-2008.

**Table 8.23.** Estimated number and percent (in parentheses) of hatchery-origin Wenatchee summer Chinook captured in different fisheries, brood years 1989-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
1989	1,510 (51)	1,432 (48)	0 (0)	20 (1)	2,962
1990	30 (100)	0 (0)	0 (0)	0 (0)	30
1991	30 (63)	0 (0)	0 (0)	18 (38)	48
1992	147 (79)	39 (21)	0 (0)	0 (0)	186
1993	35 (58)	25 (42)	0 (0)	0 (0)	60
1994	641 (91)	62 (9)	2 (0)	0 (0)	706
1995	561 (98)	9 (2)	5 (1)	0 (0)	575
1996	195 (96)	3 (1)	0 (0)	6 (3)	205
1997	2,991 (95)	49 (2)	12 (0)	106 (3)	3,158
1998	4,985 (92)	128 (2)	15 (0)	287 (5)	5,415
1999	1,550 (84)	168 (9)	21 (1)	104 (6)	1,843
2000	7,959 (73)	1,248 (11)	447 (4)	1,224 (11)	10,878
2001	1,062 (60)	238 (13)	106 (6)	364 (21)	1,770
2002	1,489 (56)	557 (21)	189 (7)	430 (16)	2,665
2003	816 (50)	484 (30)	89 (5)	257 (16)	1,646
2004	409 (47)	218 (25)	70 (8)	167 (19)	864

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
2005	1,334 (58)	481 (21)	186 (8)	287 (13)	2,288
2006	3,812 (52)	1,969 (27)	406 (6)	1,142 (16)	7,329
2007	212 (60)	81 (23)	8 (2)	53 (15)	354
2008	3,874 (60)	1,010 (16)	225 (3)	1,364 (21)	6,473
<i>Average</i>	<i>1,682 (71)</i>	<i>410 (16)</i>	<i>89 (3)</i>	<i>291 (10)</i>	<i>2,473</i>

### Straying

Stray rates were determined by examining CWTs recovered on spawning grounds within and outside the Wenatchee River basin. Targets for strays based on return year (recovery year) and brood year should be less than 5%.

Hatchery-origin Wenatchee summer Chinook have strayed into the Entiat, Chelan, Methow, and Okanogan River basins and into the Hanford Reach (Table 8.24). In five different years, Wenatchee summer Chinook strays have made up more than 5% of the spawning escapement in the Chelan Tailrace. They have made up more than 5% of the spawning escapement in the Entiat River basin in seven different years and in the Methow River basin in eight different years. Few have strayed into the Okanogan River basin or into the Hanford Reach.

**Table 8.24.** Number and percent of spawning escapements within other non-target basins that consisted of hatchery-origin Wenatchee summer Chinook, return years 1994-2011. For example, for return year 2000, 3% of the summer Chinook escapement in the Methow River basin consisted of hatchery-origin Wenatchee summer Chinook. Percent strays should be less than 5%.

Return year	Methow		Okanogan		Chelan		Entiat		Hanford Reach	
	Number	%	Number	%	Number	%	Number	%	Number	%
1994	0	0.0	75	1.9	--	--	--	--	--	--
1995	0	0.0	0	0.0	--	--	--	--	--	--
1996	0	0.0	0	0.0	--	--	--	--	--	--
1997	0	0.0	0	0.0	--	--	--	--	--	--
1998	25	3.7	0	0.0	0	0.0	0	0.0	0	0.0
1999	20	2.0	3	0.1	0	0.0	0	0.0	13	0.0
2000	36	3.0	13	0.4	0	0.0	0	0.0	0	0.0
2001	163	5.9	57	0.5	30	3.0	0	0.0	0	0.0
2002	153	3.3	53	0.4	40	6.9	74	14.8	0	0.0
2003	80	2.0	24	0.7	44	10.5	132	19.1	26	0.0
2004	113	5.2	42	0.6	30	7.1	0	0.0	0	0.0
2005	245	9.6	67	0.8	51	9.7	49	13.4	0	0.0
2006	170	6.2	12	0.1	12	2.9	61	10.6	0	0.0
2007	127	9.3	5	0.1	9	4.8	18	7.3	20	0.1
2008	87	4.5	24	0.3	10	2.0	31	9.7	0	0.0
2009	101	5.7	13	0.2	2	0.3	12	4.8	0	0.0

Return year	Methow		Okanogan		Chelan		Entiat		Hanford Reach	
	Number	%	Number	%	Number	%	Number	%	Number	%
2010	208	8.3	35	0.6	55	4.9	34	7.8	0	0.0
2011	258	8.8	5	0.1	78	6.1	4	0.9	0	0.0
<b>Average</b>	<b>99</b>	<b>4.3</b>	<b>24</b>	<b>0.4</b>	<b>26</b>	<b>4.2</b>	<b>30</b>	<b>6.3</b>	<b>4</b>	<b>0.0</b>

Based on brood year analyses, on average, about 10% of the hatchery-origin Wenatchee summer Chinook returns have strayed into non-target spawning areas, exceeding the target of 5% (Table 8.25). Depending on brood year, percent strays into non-target spawning areas have ranged from 0-19%. In addition, on average, about 5% have strayed into non-target hatchery programs, but straying into non-target programs has declined over time.

**Table 8.25.** Number and percent of hatchery-origin Wenatchee summer Chinook that homed to target spawning areas and the target hatchery program, and number and percent that strayed to non-target spawning areas and non-target hatchery programs, by brood years 1989-2008. Percent stays should be less than 5%.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1989	1,352	62.9	60	2.8	75	3.5	662	30.8
1990	74	84.1	1	1.1	0	0.0	13	14.8
1991	15	65.2	0	0.0	0	0.0	8	34.8
1992	375	84.8	7	1.6	0	0.0	60	13.6
1993	67	72.8	9	9.8	4	4.3	12	13.0
1994	890	71.8	207	16.7	61	4.9	81	6.5
1995	748	74.8	139	13.9	48	4.8	65	6.5
1996	261	70.4	42	11.3	53	14.3	15	4.0
1997	3,609	83.0	171	3.9	397	9.1	170	3.9
1998	1,790	78.2	11	0.5	416	18.2	72	3.1
1999	507	79.7	0	0.0	121	19.0	8	1.3
2000	2,745	82.3	0	0.0	545	16.3	44	1.3
2001	521	80.9	0	0.0	114	17.7	9	1.4
2002	1,521	84.6	10	0.6	259	14.4	8	0.4
2003	1,268	88.6	42	2.9	112	7.8	9	0.6
2004	497	84.2	3	0.5	72	12.2	18	3.1
2005	1,126	83.7	1	0.1	193	14.3	25	1.9
2006	2,693	79.6	0	0.0	612	18.1	78	2.3
2007	99	79.8	0	0.0	22	17.7	3	2.4
2008	3,224	85.3	0	0.0	419	11.1	135	3.6
<b>Average</b>	<b>1,169</b>	<b>78.8</b>	<b>35</b>	<b>3.3</b>	<b>176</b>	<b>10.4</b>	<b>75</b>	<b>7.5</b>

\* Homing to the target hatchery includes Wenatchee hatchery summer Chinook that are captured and included as broodstock in the Wenatchee Hatchery program. These hatchery fish are typically collected at Dryden and Tumwater dams.

## Genetics

Genetic studies were conducted to investigate relationships among temporally replicated collections of summer Chinook from the Wenatchee River, Methow River, and Okanogan River in the upper Columbia River basin (Kassler et al. 2011; the entire report is appended as Appendix M). Samples from the Eastbank Hatchery – Wenatchee stock, Eastbank Hatchery – Methow/Okanogan (MEOK) stock, and Wells Hatchery were also included in the analysis. Samples of natural and hatchery-origin summer Chinook were analyzed and compared to determine if the supplementation program has affected the genetic structure of these populations. The study also calculated the effective number of breeders for collection locations of natural and hatchery-origin summer Chinook from 1993 and 2008.

In general, population differentiation was not observed among the temporally replicated collection locations. A single collection from the Okanogan River (1993) was the only collection showing statistically significant differences. The effective number of breeders was not statistically different from the early collection in 1993 in comparison to the late collection in 2008. Overall, these analyses revealed a lack of differentiation among the temporal replicates from the same locations and among the collection from different locations, suggesting the populations have been homogenized or that there has been substantial gene flow among populations. Additional comparisons among summer-run and fall-run Chinook populations in the upper Columbia River were conducted to determine if there was any differentiation between Chinook with different run timing. These analyses revealed pairwise  $F_{ST}$  values that were less than 0.01 for the collections of summer Chinook to collections of fall Chinook from Hanford Reach, lower Yakima River, Priest Rapids, and Umatilla. Collections of fall Chinook from Crab Creek, Lyons Ferry Hatchery, Marion Drain, and Snake River had pairwise  $F_{ST}$  values that were higher in comparison to the collections of summer Chinook. The consensus clustering analysis did not provide good statistical support to the groupings, but did show relationships among collections based on geographic proximity. Overall the summer and fall run Chinook that have historically been spawned together were not differentiated while fall Chinook from greater geographic distances were differentiated.

## Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For all brood years the approximate PNI Index has been greater than or equal to 0.67 (Table 8.26). This suggests that the natural environment has a greater influence on adaptation of Wenatchee summer Chinook than does the hatchery environment.

**Table 8.26.** Approximate Proportionate Natural Influence (PNI) Index of the Wenatchee summer Chinook supplementation program for brood years 1989-2013. PNI was calculated as the proportion of naturally produced Chinook in the hatchery broodstock (pNOB) divided by the proportion of hatchery Chinook on the spawning grounds (pHOS) plus pNOB. NOS = number of natural-origin Chinook on the spawning grounds; HOS = number of hatchery-origin Chinook on the spawning grounds; NOB = number of natural-origin Chinook collected for broodstock; and HOB = number of hatchery-origin Chinook included in hatchery broodstock.

Brood year	Spawners			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
1989	14,331	0	0	290	0	1.00	1.00
1990	10,861	0	0	57	0	1.00	1.00
1991	10,168	0	0	105	0	1.00	1.00
1992	11,652	0	0	274	0	1.00	1.00
1993	8,849	600	0.06	406	44	0.9	0.94
1994	8,476	1,678	0.17	333	54	0.86	0.83
1995	6,862	894	0.12	363	16	0.96	0.89
1996	6,004	165	0.03	263	3	0.99	0.97
1997	5,408	505	0.09	205	13	0.94	0.91
1998	4,611	741	0.14	299	78	0.79	0.85
1999	4,101	1,375	0.25	242	236	0.51	0.67
2000	4,462	1,051	0.19	275	180	0.6	0.76
2001	9,414	1,946	0.17	210	136	0.61	0.78
2002	11,892	3,831	0.24	409	10	0.98	0.80
2003	10,025	1,775	0.15	337	7	0.98	0.87
2004	9,220	1,259	0.12	424	2	1.00	0.89
2005	6,862	1,841	0.21	397	3	0.99	0.83
2006	16,060	1,732	0.1	433	4	0.99	0.91
2007	3,173	1,417	0.31	263	3	0.99	0.76
2008	4,794	1,702	0.26	378	69	0.85	0.77
2009	7,113	1,214	0.15	452	8	0.98	0.87
2010	5,879	1,589	0.21	388	5	0.99	0.83
2011	8,155	1,695	0.17	376	7	0.98	0.85
2012	7,327	1,212	0.14	267	1	1.00	0.88
2013	7,449	2,760	0.27	234	2	0.99	0.79
<b>Average</b>	<b>8,126</b>	<b>1,239</b>	<b>0.14</b>	<b>307</b>	<b>35</b>	<b>0.92</b>	<b>0.87</b>

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel time (arithmetic mean days) of hatchery summer Chinook from the Wenatchee River release site to McNary Dam, and smolt to



adult ratios (SARs) from release to detection at Bonneville Dam (Table 8.27).<sup>10</sup> Over the four brood years for which PIT-tagged hatchery fish were released, survival rates from the Wenatchee River to McNary Dam ranged from 0.619 to 0.910; SARs from release to detection at Bonneville Dam ranged from 0.004 to 0.017. Average travel time from the Wenatchee River to McNary Dam ranged from 11 to 29 days.

Much of the variation in survival rates and travel time resulted from releases of different experimental groups (Table 8.27). For example, brood year 2009 was split into three groups (control group, long-term raceway group (R1), and short-term raceway group (R2)). In this case, the control group appeared to have a higher survival rate but a longer travel time from release to McNary Dam than did the two treatment groups. SARs varied little among the three groups.

Another experiment was conducted with brood year 2012. That brood year was split into four different treatment groups (small-size fish in raceway, large-size fish in raceway, small-size fish in circular, and large-size fish in circular). In general, releases from the circulars appeared to have higher survival rates to McNary Dam and faster travel times. Large-size fish from the circulars appeared to have the highest survival rates and fastest travel time.

**Table 8.27.** Total number of Wenatchee hatchery summer Chinook released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2008-2012. Standard errors are shown in parentheses. NA = not available (i.e., not all the fish from the release groups have returned to the Columbia River).

Brood year	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2008	10,035	0.847 (0.054)	28.9 (9.6)	0.017 (0.001)
2009	9,965 (Control)	0.702 (0.039)	19.3 (10.3)	0.006 (0.001)
	9,971 (R1)	0.646 (0.030)	16.4 (8.8)	0.005 (0.001)
	9,994 (R2)	0.648 (0.031)	16.0 (8.4)	0.004 (0.001)
2010	0	--	--	--
2011	5,018	0.753 (0.070)	20.9 (8.9)	NA
2012 (Raceway)	5,047 (small size)	0.724 (0.066)	18.9 (9.2)	NA
	4,740 (large size)	0.619 (0.061)	16.9 (8.6)	NA
2012 (Circular)	5,041 (small size)	0.784 (0.060)	11.8 (5.0)	NA
	5,082 (large size)	0.910 (0.077)	11.1 (4.6)	NA

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in

<sup>10</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

Hillman et al. 2012). We calculated NORs with and without harvest. NORs without harvest include all returning fish that either returned to the basin or were collected as wild broodstock. NORs with harvest include all fish harvested and are based on a brood year harvest rates from the hatchery program. For brood years 1989-2007, NRR for summer Chinook in the Wenatchee averaged 0.98 (range, 0.16-2.95) if harvested fish were not include in the estimate and 2.87 (range, 0.34-9.97) if harvested fish were included in the estimate (Table 8.28). NRRs for more recent brood years will be calculated as soon as all tag recoveries and sampling rates have been loaded into the database.

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and were calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. These rates should be greater than the NRRs and greater than or equal to 5.30 (the calculated target value in Hillman et al. 2013). HRRs exceeded NRRs in 14 of the 19 years of data, regardless if harvest was or was not included in the estimate (Table 8.28). Hatchery replacement rates for Wenatchee summer Chinook have exceeded the estimated target value of 5.30 in four or eight of the 19 years of data depending on if harvest was or was not included in the estimate.

**Table 8.28.** Broodstock collected, spawning escapements, natural and hatchery-origin recruits (NOR and HOR), and natural and hatchery replacement rates (NRR and HRR; with and without harvest) for summer Chinook in the Wenatchee River basin, brood years 1989-2007.

Brood year	Broodstock Collected	Spawning Escapement	Harvest not included				Harvest included			
			HOR	NOR	HRR	NRR	HOR	NOR	HRR	NRR
1989	346	14,331	2,149	9,182	6.21	0.64	5,111	21,810	14.77	1.52
1990	87	10,861	88	9,597	1.01	0.88	118	12,986	1.36	1.20
1991	128	10,168	23	5,562	0.18	0.55	71	17,167	0.55	1.69
1992	341	11,652	442	5,865	1.30	0.50	628	8,403	1.84	0.72
1993	524	9,450	92	5,388	0.18	0.57	152	8,906	0.29	0.94
1994	418	10,154	1,239	4,219	2.96	0.42	1,945	6,644	4.65	0.65
1995	398	7,755	1,000	5,329	2.51	0.69	1,575	8,459	3.96	1.09
1996	334	6,168	371	4,440	1.11	0.72	576	6,948	1.72	1.13
1997	240	5,913	4,347	9,770	18.11	1.65	7,505	16,903	31.27	2.86
1998	472	5,352	2,289	15,795	4.85	2.95	7,704	53,361	16.32	9.97
1999	488	5,476	636	12,062	1.30	2.20	2,479	47,302	5.08	8.64
2000	492	5,512	3,334	3,885	6.78	0.70	14,212	16,603	28.89	3.01
2001	493	11,360	644	19,209	1.31	1.69	2,414	72,487	4.90	6.38
2002	482	15,723	1,798	4,955	3.73	0.32	4,463	12,388	9.26	0.79
2003	496	11,800	1,431	1,847	2.89	0.16	3,077	3,989	6.20	0.34
2004	496	10,479	590	7,429	1.19	0.71	1,454	18,434	2.93	1.76
2005	494	8,703	1,345	5,177	2.72	0.59	3,633	14,068	7.35	1.62
2006	488	17,792	3,383	6,802	6.93	0.38	10,712	21,594	21.95	1.21
2007	419	4,590	124	10,761	0.30	2.34	478	41,548	1.14	9.05
<b>Average</b>	<b>402</b>	<b>9,644</b>	<b>1,333</b>	<b>7,751</b>	<b>3.45</b>	<b>0.98</b>	<b>3,595</b>	<b>21,579</b>	<b>8.66</b>	<b>2.87</b>

## Smolt-to-Adult Survivals

Smolt-to-adult survival ratios (SARs) were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery smolts released. Here, SARs were based on CWT returns. For the available brood years, SARs have ranged from 0.00037 to 0.01554 for hatchery summer Chinook in the Wenatchee River basin (Table 8.29).

**Table 8.29.** Smolt-to-adult ratios (SARs) for Wenatchee hatchery summer Chinook, brood years 1989-2008.

Brood year	Number of tagged smolts released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1989	144,905	1,027	0.00709
1990	119,214	115	0.00096
1991	190,371	71	0.00037
1992	605,055	613	0.00101
1993	210,626	152	0.00072
1994	452,340	1,920	0.00424
1995	668,409	1,541	0.00231
1996	585,590	568	0.00097
1997	480,418	7,465	0.01554
1998	641,109	7,631	0.01190
1999	988,328	2,457	0.00249
2000	903,368	13,858	0.01534
2001	596,618	2,400	0.00402
2002	805,919	4,333	0.00538
2003	639,381	3,029	0.00474
2004	603,942	1,439	0.00238
2005	631,492	3,586	0.00568
2006	931,880	10,532	0.01130
2007	453,719	478	0.00105
2008	859,401	9,945	0.01157
<b>Average</b>	<b>575,604</b>	<b>3,658</b>	<b>0.00545</b>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

## 8.8 ESA/HCP Compliance

### Broodstock Collection

Per the 2012 broodstock collection protocol, 274 natural-origin (adipose fin present) summer Chinook adults were targeted for collection at Dryden and Tumwater dams. The actual 2012 collection totaled 274 summer Chinook (273 natural-origin and one hatchery-origin; the

hatchery-origin fish were not direct collections, but rather adipose present non-wired fish with a hatchery scale pattern) in combination from Dryden and Tumwater dams. Trapping began 1 July and ended 4 September 2012.

Summer Chinook and steelhead broodstock collections occurred concurrently at Dryden Dam; therefore, steelhead and spring Chinook encounters at Dryden Dam during Wenatchee summer Chinook broodstock collection were attributable to steelhead broodstock collections authorized under ESA Permit 1395 take authorizations. No steelhead or spring Chinook takes were associated with the Wenatchee summer Chinook collection.

Consistent with impact minimization measures in ESA Permit 1347, all ESA-listed species handled during summer Chinook broodstock collection were subject to water-to-water transfers or anesthetized if removed from the water during handling.

### **Hatchery Rearing and Release**

The 2012 Wenatchee summer Chinook program released an estimated 550,877 smolts, representing 110% of the 500,001 programmed production, and was right at the 10% overage allowance identified in ESA permit 1347.

### **Hatchery Effluent Monitoring**

Per ESA Permits 1196, 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January through 31 December 2014. NPDES monitoring and reporting for Chelan PUD Hatchery Programs during 2014 are provided in Appendix F.

### **Smolt and Emigrant Trapping**

ESA-listed spring Chinook and steelhead were encountered during operation of the Lower Wenatchee Trap. ESA takes are reported in the steelhead (Section 3.8) and spring Chinook (Section 5.8) sections and are not repeated here.

### **Spawning Surveys**

Summer Chinook spawning ground surveys conducted in the Wenatchee River basin during 2014 were consistent with ESA Section 10 Permit No. 1347. Because of the difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical, and extreme caution was used to avoid established redds when wading was required.

## SECTION 9: METHOW SUMMER CHINOOK

The original goal of summer Chinook salmon supplementation in the Methow Basin was in part to use artificial production to replace adult production lost because of mortality at Wells, Rocky Reach, and Rock Island dams<sup>11</sup>, while not reducing the natural production or long-term fitness of summer Chinook in the basin. The Rock Island Fish Hatchery Complex began operation in 1989 under funding from Chelan PUD. The Complex operated originally through the Rock Island Settlement Agreement, but since 2004 has operated under the Anadromous Fish Agreement and Habitat Conservation Plans. Beginning with broodstock collection in 2012, Grant PUD took over the summer Chinook salmon supplementation program in the Methow Basin. Grant PUD constructed a new overwinter acclimation facility adjacent to the Carlton Acclimation Pond and the first fish released from this facility was 2014. The new facility includes eight, 30-foot diameter dual-drain circular tanks.

Presently, adult summer Chinook are collected for broodstock from the run-at-large at the west-ladder trapping facility at Wells Dam. Prior to 2012, the goal was to collect up to 222 natural-origin adult summer Chinook for the Methow program. In 2011, the Hatchery Committees reevaluated that amount of hatchery compensation needed to achieve NNI. Based on that evaluation, the goal of the program was revised. The current goal (beginning in 2012) is to collect up to 102 natural-origin summer Chinook for the Methow program. Broodstock collection occurs from about 1 July through 15 September with trapping occurring no more than 16 hours per day, three days a week. If natural-origin broodstock collection falls short of expectation, hatchery-origin adults can be collected to make up the difference.

Adult summer Chinook are spawned and reared at Eastbank Fish Hatchery. Juvenile summer Chinook were transferred from the hatchery to Carlton Acclimation Pond in March until overwinter acclimation was initiated with the 2013 brood year. They are now released from the new facility in late April to early May.

Before 2012, the production goal for the Methow summer Chinook supplementation program was to release 400,000 yearling smolts into the Methow River at ten fish per pound. Beginning with the 2012 brood, the revised goal is to release 200,000 yearling smolts at 15 fish per pound. Targets for fork length and weight are 163 mm (CV = 9.0) and 45.4 g, respectively. Over 90% of these fish are marked with CWTs. In addition, since 2009, juvenile summer Chinook have been PIT tagged annually.

### 9.1 Broodstock Sampling

This section focuses on results from sampling 2012-2013 Methow summer Chinook broodstock that were collected in the West Ladder of Wells Dam in 2012 and 2013. Summer Chinook adults collected at Wells Dam are also used in the Okanogan/Similkameen supplementation program.

---

<sup>11</sup> The majority of the production at Carlton Acclimation Pond is initial production, which terminated in 2013, and is not necessarily tied to hydro facility mortality. The balance of the production is the result of a swap between spring and summer Chinook. That is, Chelan PUD is currently producing summer Chinook at Carlton for Douglas PUD in exchange for Douglas PUD producing spring Chinook at the Methow Fish Hatchery for Chelan PUD.

Complete information is not currently available for the 2014 return (this information will be provided in the 2015 annual report).

### Origin of Broodstock

Both 2012 and 2013 broodstock consisted almost entirely of natural-origin (adipose fin present) summer Chinook (Table 9.1). These fish were used for both the Methow and Okanogan supplementation programs. In 2012, to meet production goals, hatchery-origin adults were collected in concert with natural-origin fish. About 4% of the 2013 broodstock were comprised of hatchery-origin fish (hatchery-origin was determined by examination of scales and CWTs).

**Table 9.1.** Numbers of wild and hatchery summer Chinook collected for broodstock, numbers that died before spawning, and numbers of Chinook spawned for the Methow/Okanogan programs during 1989-2012. Numbers of broodstock collected from 2013 to present are only for the Methow summer Chinook Program. Unknown origin fish (i.e., undetermined by scale analysis, no CWT or fin clips, and no additional hatchery marks) were considered naturally produced. Mortality includes fish that died of natural causes typically near the end of spawning and were not needed for the program and surplus fish killed at spawning.

Brood year	Wild summer Chinook					Hatchery summer Chinook					Total number spawned
	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	Number collected	Prespawn loss <sup>a</sup>	Mortality	Number spawned	Number released	
1989 <sup>b</sup>	1,419	72	-	1,297	-	341	17	-	312	-	1,609
1990 <sup>b</sup>	864	34	-	828	-	214	8	-	206	-	1,034
1991 <sup>b</sup>	1,003	59	-	924	-	341	20	-	314	-	1,238
1992 <sup>b</sup>	312	6	-	297	-	428	9	-	406	-	703
1993 <sup>b</sup>	813	48	-	681	-	464	28	-	388	-	1,069
1994	385	33	11	341	12	266	15	7	244	1	585
1995	254	13	10	173	58	351	28	9	240	74	413
1996	316	15	11	290	0	234	2	9	223	0	513
1997	214	11	5	198	0	308	24	20	264	0	462
1998	239	28	58	153	0	348	18	119	211	0	364
1999	248	5	19	224	0	307	2	16	289	0	513
2000	184	15	5	164	0	373	17	17	339	0	503
2001	135	8	36	91	0	423	29	128	266	0	357
2002	270	2	21	247	0	285	11	33	241	0	488
2003	449	14	53	381	0	112	2	9	101	0	482
2004	541	23	12	506	0	17	0	1	16	0	522
2005	551	29	76	391	55	12	2	0	9	1	400
2006	579	50	10	500	19	12	2	0	10	0	510
2007	504	22	26	456	0	19	0	2	17	0	473
2008	418	5	9	404	0	41	0	0	41	0	445
2009	553	31	15	507	0	5	5	0	0	0	507
2010	503	13	6	484	0	8	0	0	8	0	492
2011	498	18	13	467	0	30	4	0	26	0	493
<b>Average<sup>c</sup></b>	<b>380</b>	<b>19</b>	<b>22</b>	<b>332</b>	<b>8</b>	<b>175</b>	<b>9</b>	<b>21</b>	<b>141</b>	<b>4</b>	<b>473</b>
2012	125	5	0	98	22	3	0	0	1	2	99
2013	98	1	0	97	0	4	0	0	4	0	101
<b>Average<sup>d</sup></b>	<b>112</b>	<b>3</b>	<b>0</b>	<b>98</b>	<b>11</b>	<b>4</b>	<b>0</b>	<b>0</b>	<b>3</b>	<b>1</b>	<b>100</b>

<sup>a</sup> Pre-spawn loss represents the number of fish that died during the holding period before spawning. Mortality is the number of fish that were surplus following spawning.

<sup>b</sup> Number of fish spawned and collected during these years included fish retained from the right- and left-bank ladder traps at Wells Dam and fish collected from the volunteer channel. There was no distinction made between fish collected at trap locations and program (i.e., aggregated population used for Wells, Methow, and Okanogan summer Chinook programs).

<sup>c</sup> This average represents broodstock collected for the combined Methow and Okanogan programs. Because of bias from aggregating the spawning population from 1989-1993, averages are based on adult numbers collected from 1994-2011.

<sup>d</sup> This average represents broodstock collected only for the Methow program.

### Age/Length Data

Ages of summer Chinook broodstock were determined from analysis of scales and/or CWTs. Broodstock collected from the 2012 return consisted primarily of age-4 and 5 natural-origin Chinook (95.1%) and age-5 hatchery-origin Chinook. Age-3 natural-origin fish made up 3.9% of the broodstock (Table 9.2).

Broodstock collected from the 2013 return consisted primarily of age-4 and 5 natural-origin Chinook (84.8%) and age-5 hatchery-origin Chinook (100%). Age-3 natural-origin Chinook made up 15.2% of the broodstock (Table 9.2).

**Table 9.2.** Percent of hatchery and wild summer Chinook of different ages (total age) collected from broodstock for the Methow/Okanogan programs, 1991-2013.

Return Year	Origin	Total age				
		2	3	4	5	6
1991	Wild	0.5	6.8	35.1	55.4	2.2
	Hatchery	0.5	5.1	36.2	49.0	9.2
1992	Wild	0.0	13.0	36.2	50.7	0.0
	Hatchery	0.0	0.0	0.0	0.0	0.0
1993	Wild	0.0	3.9	75.3	20.8	0.0
	Hatchery	0.0	1.0	85.7	13.3	0.0
1994	Wild	3.1	9.7	26.3	60.3	0.6
	Hatchery	0.0	14.7	11.2	74.0	0.0
1995	Wild	0.0	4.6	15.3	75.6	4.6
	Hatchery	0.0	0.4	13.0	25.6	61.0
1996	Wild	0.0	8.4	56.7	30.4	4.6
	Hatchery	0.0	3.0	31.0	47.0	19.0
1997	Wild	0.5	9.4	53.0	35.1	2.0
	Hatchery	0.0	20.6	11.1	61.8	6.5
1998	Wild	1.1	12.1	56.3	30.5	0.0
	Hatchery	2.1	18.9	56.2	16.0	6.8
1999	Wild	4.7	5.1	53.7	36.0	0.5
	Hatchery	0.3	3.5	29.3	65.0	1.9
2000	Wild	0.6	14.0	28.7	56.1	0.6
	Hatchery	0.0	27.0	14.3	54.3	4.3
2001	Wild	0.0	23.5	58.8	11.8	5.9
	Hatchery	1.8	21.1	64.6	10.1	2.4
2002	Wild	0.4	17.4	65.6	16.6	0.0
	Hatchery	0.0	2.4	39.4	58.3	0.0

Return Year	Origin	Total age				
		2	3	4	5	6
2003	Wild	0.7	3.9	65.8	29.5	0.0
	Hatchery	0.0	5.6	18.7	70.1	5.6
2004	Wild	0.6	15.4	11.6	72.2	0.2
	Hatchery	0.0	6.7	53.3	33.3	6.7
2005	Wild	0.0	17.1	69.9	11.0	1.9
	Hatchery	0.0	10.0	40.0	50.0	0.0
2006	Wild	1.7	3.0	41.0	52.9	1.5
	Hatchery	0.0	16.7	25.0	50.0	8.3
2007	Wild	1.8	15.3	8.2	70.3	4.4
	Hatchery	0.0	0.0	21.1	57.9	21.1
2008	Wild	0.3	17.9	67.1	13.3	1.4
	Hatchery	0.0	7.2	62.7	47.7	2.4
2009	Wild	1.3	10.1	68.7	19.9	0.0
	Hatchery	0.0	0.0	16.7	83.3	0.0
2010	Wild	0.2	16.2	51.0	32.6	0.0
	Hatchery	0.0	12.5	50.0	25.0	12.5
2011	Wild	0.1	7.1	75.5	17.0	0.0
	Hatchery	0.0	30.0	20.0	40.0	0.0
2012	Wild	0.0	3.9	49.0	46.1	1.0
	Hatchery	0.0	0.0	0.0	100.0	0.0
2013	Wild	0.0	15.2	70.7	14.1	0.0
	Hatchery	0.0	0.0	50.0	50.0	0.0
<i>Average</i>	<i>Wild</i>	<i>0.8</i>	<i>11.0</i>	<i>49.5</i>	<i>37.3</i>	<i>1.4</i>
	<i>Hatchery</i>	<i>0.2</i>	<i>9.0</i>	<i>32.6</i>	<i>47.0</i>	<i>7.3</i>

Mean lengths of natural-origin summer Chinook of a given age differed little between 2012 and 2013 (Table 9.3). Average fork lengths for age-5 natural-origin adults were 5 cm longer than that of age-5 hatchery fish (Table 9.3). Differences in hatchery-origin and natural-origin fish were hard to discern given the small sample size of hatchery-origin fish (i.e., few hatchery fish were included in the broodstock).

**Table 9.3.** Mean fork length (cm) at age (total age) of hatchery and wild Methow/Okanogan summer Chinook collected from broodstock for the Methow/Okanogan programs, 1991-2013; N = sample size and SD = 1 standard deviation.

Return year	Origin	Summer Chinook fork length (cm)														
		Age-2			Age-3			Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
1991	Wild	47	1	-	68	15	6	82	78	10	94	123	8	97	5	5
	Hatchery	47	1	-	49	10	6	78	71	5	91	96	8	96	18	6
1992	Wild	-	0	-	55	9	5	69	25	6	78	35	6	-	0	-



Return year	Origin	Summer Chinook fork length (cm)														
		Age-2			Age-3			Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
	Hatchery	-	0	-	-	0	-	-	0	-	-	0	-	-	0	-
1993	Wild	-	0	-	72	3	4	86	58	7	98	16	5	-	0	-
	Hatchery	-	0	-	42	1	-	75	84	8	88	13	6	-	0	-
1994	Wild	42	10	6	50	31	7	80	84	9	93	193	8	104	2	13
	Hatchery	-	0	-	49	38	5	76	29	7	88	191	7	-	0	-
1995	Wild	-	0	-	67	6	8	79	20	9	96	99	5	94	6	5
	Hatchery	-	0	-	52	1	-	73	32	9	89	63	9	95	150	7
1996	Wild	-	0	-	68	22	9	83	149	8	95	79	7	101	12	5
	Hatchery	-	0	-	52	7	10	77	72	7	90	109	8	100	44	6
1997	Wild	31	1	-	60	19	7	85	107	8	96	71	7	98	4	11
	Hatchery	-	0	-	45	63	5	72	34	9	92	189	7	97	20	7
1998	Wild	39	2	1	59	23	6	83	107	7	96	58	7	-	0	-
	Hatchery	43	7	6	50	64	6	74	190	7	92	54	8	98	23	5
1999	Wild	38	10	3	64	11	8	82	115	7	96	76	6	104	1	-
	Hatchery	37	1	-	53	11	9	75	92	6	91	204	6	98	6	5
2000	Wild	39	1	-	66	23	7	83	47	6	96	92	5	95	1	-
	Hatchery	-	0	-	54	100	7	78	53	8	92	201	6	99	16	6
2001	Wild	-	0	-	63	4	12	88	10	9	90	2	4	94	1	-
	Hatchery	41	9	3	55	107	9	79	327	8	93	51	7	101	12	9
2002	Wild	56	1	-	65	44	7	88	166	6	100	42	7	-	0	-
	Hatchery	-	0	-	45	6	5	76	100	7	95	148	5	-	0	-
2003	Wild	43	3	6	61	16	6	87	268	7	99	120	6	-	0	-
	Hatchery	-	0	-	55	6	9	73	20	8	91	75	7	102	6	9
2004	Wild	51	3	5	67	78	6	81	59	6	97	367	7	99	1	-
	Hatchery	-	0	-	52	1	-	70	8	5	97	5	8	109	1	-
2005	Wild	-	0	-	68	89	6	83	363	7	94	57	6	101	10	7
	Hatchery	-	0	-	55	1	-	70	4	4	89	5	4	-	0	-
2006	Wild	38	9	3	54	16	4	69	221	6	77	286	5	78	8	4
	Hatchery	-	0	-	42	2	1	62	3	2	69	6	6	76	1	-
2007	Wild	39	8	5	53	69	5	67	37	6	78	317	5	77	20	7
	Hatchery	-	0	-	-	0	-	54	4	2	75	11	5	78	4	3
2008	Wild	41	1	-	55	62	4	69	233	6	76	46	4	82	5	3
	Hatchery	-	0	-	59	6	9	67	52	5	73	23	6	79	2	8
2009	Wild	38	7	5	54	54	5	72	367	5	79	106	5	-	0	-
	Hatchery	-	0	-	-	0	-	59	1	-	71	5	7	-	0	-
2010	Wild	43	1	-	54	78	5	71	246	5	78	157	5	-	0	-
	Hatchery	-	0	-	57	1	-	67	4	5	79	2	1	89	1	-
2011	Wild	43	2	3	66	32	8	87	338	7	97	76	5	-	0	-
	Hatchery	-	0	-	63	9	11	78	9	6	92	12	9	-	0	-
2012	Wild	-	0	-	70	10	3	84	62	5	96	54	6	-	0	-

Return year	Origin	Summer Chinook fork length (cm)														
		Age-2			Age-3			Age-4			Age-5			Age-6		
		Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD	Mean	N	SD
	Hatchery	-	0	-	-	0	-	-	0	-	90	1	-	-	0	-
2013	Wild	-	0	-	72	14	5	86	65	7	97	13	5	-	0	-
	Hatchery	-	0	-	-	0	-	76	2	6	92	2	0	-	0	-
Average	Wild	42	3	4	62	32	6	80	140	7	91	108	6	94	3	7
	Hatchery	42	1	5	52	19	7	72	52	6	87	64	6	94	13	6

## Sex Ratios

Male summer Chinook in the 2012 broodstock made up about 49.0% of the adults collected, resulting in an overall male to female ratio of 0.96:1.00 (Table 9.4.). In 2013, males made up about 51.0% of the adults collected, resulting in an overall male to female ratio of 1.04:1.00 (Table 9.4). The ratios for 2012 and 2013 broodstock were above and below the assumed 1:1 ratio goal in the broodstock protocol, respectively.

**Table 9.4.** Numbers of male and female wild and hatchery summer Chinook collected for broodstock at Wells Dam for the Methow/Okanogan programs, 1991-2013. Ratios of males to females are also provided.

Return year	Number of wild summer Chinook			Number of hatchery summer Chinook			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
1989 <sup>a</sup>	752	667	1.13:1.00	181	160	1.13:1.00	1.13:1.00
1990 <sup>a</sup>	381	482	0.79:1.00	95	120	0.79:1.00	0.79:1.00
1991 <sup>a</sup>	443	559	0.79:1.00	151	191	0.79:1.00	0.79:1.00
1992 <sup>a</sup>	349	318	1.10:1.00	38	35	1.09:1.00	1.10:1.00
1993 <sup>a</sup>	513	300	1.71:1.00	293	171	1.71:1.00	1.71:1.00
1994	205	180	1.14:1.00	165	101	1.63:1.00	1.32:1.00
1995	103	149	0.69:1.00	158	197	0.80:1.00	0.75:1.00
1996	178	138	1.29:1.00	132	102	1.29:1.00	1.29:1.00
1997	102	112	0.91:1.00	174	134	1.30:1.00	1.12:1.00
1998	130	109	1.19:1.00	263	85	3.09:1.00	2.03:1.00
1999	138	110	1.25:1.00	161	146	1.10:1.00	1.17:1.00
2000	82	102	0.80:1.00	243	130	1.87:1.00	1.40:1.00
2001	89	46	1.93:1.00	311	112	2.78:1.00	2.53:1.00
2002	166	104	1.60:1.00	149	136	1.10:1.00	1.31:1.00
2003	255	194	1.31:1.00	61	51	1.20:1.00	1.29:1.00
2004	263	278	0.95:1.00	12	5	2.40:1.00	0.97:1.00
2005	365	186	1.96:1.00	6	6	1.00:1.00	1.93:1.00
2006	287	292	0.98:1.00	9	3	3.00:1.00	1.00:1.00
2007	228	276	0.83:1.00	11	8	1.38:1.00	0.84:1.00
2008	210	208	1.01:1.00	13	28	0.46:1.00	0.94:1.00
2009	261	292	0.89:1.00	2	3	0.67:1.00	0.89:1.00

Return year	Number of wild summer Chinook			Number of hatchery summer Chinook			Total M/F ratio
	Males (M)	Females (F)	M/F	Males (M)	Females (F)	M/F	
2010	248	255	0.97:1.00	5	3	1.67:1.00	0.98:1.00
2011	236	262	0.90:1.00	23	7	3.29:1.00	0.96:1.00
2012	50	53	0.94:1.00	1	0	-	0.96:1.00
2013	49	49	1.00:1.00	3	1	3.00:1.00	1.04:1.00
<b>Total<sup>b</sup></b>	<b>3645</b>	<b>3395</b>	<b>1.07:1.00</b>	<b>1902</b>	<b>1258</b>	<b>1.51:1.00</b>	<b>1.19:1.00</b>

<sup>a</sup> Numbers and male to female ratios were derived from the aggregate population collected at Wells Fish Hatchery volunteer channel and left- and right-ladder traps at Wells Dam.

<sup>b</sup> Total values were derived from 1994-present data to exclude aggregate population bias from 1989-1993 returns.

### Fecundity

Fecundities for the 2012 and 2013 summer Chinook broodstock averaged 4,470 and 4,717 eggs per female, respectively (Table 9.5). These values are close to the overall average of 4,943 eggs per female. Mean observed fecundities for the 2012 and 2013 returns were slightly below the expected fecundity of 5,000 eggs per female assumed in the broodstock protocol.

**Table 9.5.** Mean fecundity of wild, hatchery, and all female summer Chinook collected for broodstock at Wells Dam for the Methow/Okanogan programs, 1989-2013; NA = not available.

Return year	Mean fecundity		
	Wild	Hatchery	Total
1989*	NA	NA	4,750
1990*	NA	NA	4,838
1991*	NA	NA	4,819
1992*	NA	NA	4,804
1993*	NA	NA	4,849
1994*	NA	NA	5,907
1995*	NA	NA	4,930
1996*	NA	NA	4,870
1997	5,166	5,296	5,237
1998	5,043	4,595	4,833
1999	4,897	4,923	4,912
2000	5,122	5,206	5,170
2001	5,040	4,608	4,735
2002	5,306	5,258	5,279
2003	5,090	4,941	5,059
2004	5,130	5,118	5,130
2005	4,545	4,889	4,553
2006	4,854	4,824	4,854
2007	5,265	5,093	5,260
2008	4,814	4,588	4,787
2009	5,115	-	5,115
2010	5,124	4,717	5,116

Return year	Mean fecundity		
	Wild	Hatchery	Total
2011	4,594	3,915	4,578
2012	4,470	--	4,470
2013	4,700	5,490	4,717
<i>Average</i>	<i>4,957</i>	<i>4,897</i>	<i>4,943</i>

\* Individual fecundities were not assigned to females until 1997 brood.

## 9.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Based on the unfertilized egg-to-release survival standard of 81%, a total of 493,827 eggs were needed to meet the program release goal of 400,000 smolts for brood years 1989-2011. An evaluation of the program in 2011 determined that 246,913 eggs are needed to meet the revised release goal of 200,000 smolts. This revised goal began with brood year 2012. From 1989 through 2011, the egg take goal was reached in eight of those years (Table 9.6). From 2012 to present, the egg take goal was not reached in any year, but the numbers were close to the goal (Table 9.6).

**Table 9.6.** Numbers of eggs taken from summer Chinook broodstock collected at Wells Dam for the Methow/Okanogan programs, 1989-2014.

Return year	Number of eggs taken
1989	482,800
1990	464,097
1991	586,594
1992	486,260
1993	531,490
1994	595,390
1995	491,000
1996	448,000
1997	401,162
1998	389,346
1999	483,726
2000	403,268
2001	279,272
2002	466,530
2003	473,681
2004	537,210
2005	305,826
2006	509,334
2007	549,802

Return year	Number of eggs taken
2008	441,778
2009	560,602
2010	505,188
2011	488,747
<i>Average (1989-2011)</i>	<i>473,091</i>
2012	245,245
2013	231,136
2014	223,839
<i>Average (2012-present)</i>	<i>233,407</i>

### *Number of acclimation days*

Rearing of the 2012 brood Methow summer Chinook was similar to previous years with fish being held on well water before being transferred to Carlton Acclimation Pond for final acclimation on Methow River water in March 2014 (Table 9.7). Groups of the 1994 and 1995 broods were reared for longer durations at the Methow Fish Hatchery on Methow River water.

**Table 9.7.** Number of days Methow summer Chinook were acclimated at Carlton Acclimation Pond, brood years 1989-2012.

Brood year	Release year	Transfer date	Release date	Number of days
1989	1991	15-Mar	6-May	52
1990	1992	26-Feb	28-Apr	61
1991	1993	10-Mar	23-Apr	44
1992	1994	4-Mar	21-Apr	48
1993	1995	18-Mar	2-May	45
1994	1996	25-Sep	28-Apr	215
		19-Mar	28-Apr	40
1995	1997	22-Oct	8-Apr	168
		19-Mar	22-Apr	34
1996	1998	9-Mar	14-Apr	36
1997	1999	10-Mar	20-Apr	41
1998	2000	19-Mar	2-May	44
1999	2001	18-Mar	18-Apr	31
2000	2002	28-Mar	1-May	34
2001	2003	27-Mar	24-Apr	28
2002	2004	16-Mar	24-Apr	39
2003	2005	18-Mar	21-Apr	34

Brood year	Release year	Transfer date	Release date	Number of days
2004	2006	12-Mar	22-Apr	41
2005	2007	12-Mar	15-Apr – 8-May	34-57
2006	2008	4-7-Mar	16-Apr – 2 May	40-59
2007	2009	18-24-Mar	21-Apr	28-34
2008	2010	4-5, 8-9-Mar	4-21-Apr	33-50
2009	2011	25, 29, 31-Mar & 4-Apr	11-25-Apr	8-31
2010	2012	19-21, 24-Mar	23-24-Apr	31-37
2011	2013	13-21-Mar	15-23-Apr	25-41
2012	2014	19-21-Mar	7-Apr – 14 May	18-57

## Release Information

### *Numbers released*

The 2012 brood Methow summer Chinook program achieved 99% of the 200,000 target goal with about 197,391 fish being released volitionally from the circular ponds on 7-14 May 2014 (Table 9.8).

**Table 9.8.** Numbers of Methow summer Chinook smolts released from the hatchery, brood years 1989-2012. Beginning with the 2014 release, the release target for Methow summer Chinook is 200,000 smolts.

Brood year	Release year	CWT mark rate	Number of smolts released
1989	1991	0.8529	420,000
1990	1992	0.9485	391,650
1991	1993	0.6972	540,900
1992	1994	0.9752	402,641
1993	1995	0.4623	433,375
1994	1996	0.9851	406,560
1995	1997	0.9768	353,182
1996	1998	0.9221	298,844
1997	1999	0.9884	384,909
1998	2000	0.9429	205,269
1999	2001	0.9955	424,363
2000	2002	0.9928	336,762
2001	2003	0.9902	248,595
2002	2004	0.9913	399,975
2003	2005	0.9872	354,699
2004	2006	0.9848	400,579
2005	2007	0.9897	263,723
2006	2008	0.9783	419,734

Brood year	Release year	CWT mark rate	Number of smolts released
2007	2009	0.9837	433,256
2008	2010	0.9394	397,554
2009	2011	0.9862	404,956
2010	2012	0.9962	439,000
2011	2013	0.9734	436,092
<i>Average (1989-2011)</i>		<i>0.9365</i>	<i>382,462</i>
2012	2014	0.9987	197,391
<i>Average (2012-present)</i>		<i>0.9987</i>	<i>197,391</i>

### Numbers tagged

The 2012 brood Methow summer Chinook were 99.9% CWT and adipose fin-clipped (Table 9.8).

A total of 10,159 Methow summer Chinook (brood 2013) were PIT tagged at Eastbank Hatchery on 25-28 August 2014. These fish were tagged in raceways #1, #2, #7, and #8. Fish were not fed during tagging or for two days before and after tagging. Fish averaged 72 mm in length and 5.3 g at time of tagging. These fish will be released in spring 2015.

Table 9.9 summarizes the number of hatchery summer Chinook that have been PIT-tagged and released into the Methow River.

**Table 9.9.** Summary of PIT-tagging activities for Methow hatchery summer Chinook, brood years 2008-2012.

Brood year	Release year	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2008	2010	10,100	4	0	10,096
2009	2011	5,050	17	9	5,024
2010	2012	0	0	0	0
2011	2013	0	0	0	0
2012	2014	10,099	41	7	10,051

### Fish size and condition at release

A volitional release of yearling smolts took place between 7 April and 14 May 2014. Size at release from the acclimated population was 96.9% and 91.6% of the respective target fork length and weight goals (Table 9.10). This brood year exceeded the target CV for length by 34.4%.

**Table 9.10.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of Methow summer Chinook smolts released from the hatchery, brood years 1991-2012. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1991	1993	152	13.6	40.3	11
1992	1994	145	16.0	37.2	12
1993	1995	154	8.6	37.1	12
1994	1996	163	8.2	48.2	9
1995	1997	141	9.6	37.0	12
1996	1998	199	13.1	105.1	4
1997	1999	153	7.6	39.5	12
1998	2000	164	8.7	51.7	9
1999	2001	153	9.3	41.5	11
2000	2002	170	10.2	54.2	8
2001	2003	167	7.4	52.7	9
2002	2004	148	13.1	35.7	13
2003	2005	148	10.1	35.5	13
2004	2006	142	9.8	31.1	15
2005	2007	158	15.0	42.2	11
2006	2008	156	18.0	42.8	11
2007	2009	138	21.0	32.1	14
2008	2010	155	14.2	42.0	11
2009	2011	170	15.8	56.9	8
2010	2012	145	16.7	34.5	13
2011	2013	160	13.0	43.6	6
<i>Average</i>		<i>156</i>	<i>12.3</i>	<i>44.8</i>	<i>11</i>
<i>Targets</i>		<i>163</i>	<i>9.0</i>	<i>45.4</i>	<i>10</i>
2012	2014	158	12.1	41.6	11
<i>Average</i>		<i>158</i>	<i>12.1</i>	<i>41.6</i>	<i>11</i>
<i>Targets</i>		<i>163</i>	<i>9.0</i>	<i>45.4</i>	<i>15</i>

### Survival Estimates

Overall survival of the Methow summer Chinook from green (unfertilized) egg-to-release was above the standard set for the program (Table 9.11). High hatchery survival can be attributed to exceeding the survival standards set for the program at almost every life stage.



**Table 9.11.** Hatchery life-stage survival rates (%) for Methow summer Chinook, brood years 1989-2012. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
1989 <sup>a</sup>	89.8	99.5	89.9	96.7	99.7	99.4	73.3	98.5	87.0
1990 <sup>a</sup>	93.9	99.0	84.9	97.1	81.2	80.6	97.7	99.5	84.4
1991 <sup>a</sup>	93.1	95.5	88.2	98.0	99.4	99.1	97.5	99.6	92.2
1992 <sup>a</sup>	96.9	99.0	87.8	98.0	99.9	99.9	90.9	98.3	82.8
1993 <sup>a</sup>	82.2	99.4	85.4	97.6	99.8	99.5	92.0	99.4	81.5
1994	96.1	90.0	86.6	100.0	98.1	97.4	73.1	99.1	68.3
1995	91.9	96.2	98.2	84.1	96.5	96.2	92.7	89.6	71.9
1996	95.4	98.1	83.2	100.0	97.7	96.9	86.5	89.0	66.7
1997	91.9	94.6	86.1	98.4	98.7	98.3	98.8	99.7	95.9
1998	84.0	96.2	54.1	98.0	99.4	98.9	96.6	99.9	52.7
1999	98.8	98.7	92.9	96.9	98.0	97.6	96.9	99.9	87.7
2000	90.5	96.9	89.2	98.1	98.5	98.3	94.6	94.4	83.5
2001	96.2	92.3	89.1	97.6	97.2	97.1	97.5	99.8	89.0
2002	97.1	98.1	88.3	99.9	97.7	97.5	96.7	99.9	85.7
2003	96.7	97.5	82.8	98.2	99.7	99.2	93.7	99.9	74.9
2004	93.6	98.2	84.0	97.8	99.6	99.2	98.3	98.5	74.6
2005	97.0	89.6	88.0	95.5	99.6	98.9	96.6	99.9	86.2
2006	92.9	89.5	86.3	98.3	99.6	98.7	97.2	99.5	82.4
2007	92.6	99.6	84.1	98.5	99.7	99.5	98.9	99.8	81.9
2008	99.6	97.9	91.9	99.5	99.3	98.9	98.5	99.9	90.0
2009 <sup>b</sup>	93.6	93.5	91.0	97.7	99.7	99.2	98.8	100.0	87.9
2010 <sup>c</sup>	96.5	100.0	91.1	100.0	96.4	96.1	95.4	99.5	86.9
2011	94.9	96.4	93.8	97.8	99.7	99.1	98.6	99.9	90.4
2012	94.3	94.2	93.1	97.8	99.4	99.0	97.0	98.3	88.3
<b>Average</b>	<b>93.7</b>	<b>96.2</b>	<b>87.1</b>	<b>97.6</b>	<b>98.1</b>	<b>97.7</b>	<b>94.1</b>	<b>98.4</b>	<b>82.2</b>
<b>Standard</b>	<b>90.0</b>	<b>85.0</b>	<b>92.0</b>	<b>98.0</b>	<b>97.0</b>	<b>93.0</b>	<b>90.0</b>	<b>95.0</b>	<b>81.0</b>

<sup>a</sup> Survival rates were calculated from aggregate population collected at Wells Fish Hatchery volunteer channel and left- and right-ladder traps at Wells Dam.

<sup>b</sup> Survival rates were calculated from aggregate collections at Wells east fish ladder for the Methow and Okanogan/Similkameen programs. About 41% of the total fish collected were used to estimate survival rates.

<sup>c</sup> Survival rates were calculated from aggregate collections at Wells West Ladder for the Methow and Similkameen programs. About 71% of the total fish collected were used to estimate survival rates.

### 9.3 Disease Monitoring

Results of adult broodstock bacterial kidney disease (BKD) monitoring indicated that most females had ELISA values less than 0.199. Just less than 94% of females had ELISA values less than 0.120, which means about 6% of the progeny needed to be reared at densities not to exceed 0.06 fish per pound (Table 9.12).

**Table 9.12.** Proportion of bacterial kidney disease (BKD) titer groups for the Methow/Okanogan summer Chinook broodstock, brood years 1997-2014. Also included are the proportions to be reared at either 0.125 fish per pound or 0.060 fish per pound.

Brood year <sup>a</sup>	Optical density values by titer group				Proportion at rearing densities (fish per pound, fpp)	
	Very Low (≤ 0.099)	Low (0.1-0.199)	Moderate (0.2-0.449)	High (≥ 0.450)	≤ 0.125 fpp (<0.119)	≤ 0.060 fpp (>0.120)
1997	0.6267	0.1333	0.0622	0.1778	0.6844	0.3156
1998	0.9632	0.0184	0.0123	0.0061	0.9816	0.0184
1999	0.9444	0.0198	0.0238	0.0119	0.9643	0.0357
2000	0.7476	0.0952	0.0238	0.1333	0.8000	0.2000
2001	0.9801	0.0199	0.0000	0.0000	1.0000	0.0000
2002	0.9567	0.0130	0.0130	0.0173	0.9740	0.0260
2003	0.9620	0.0127	0.0169	0.0084	0.9747	0.0253
2004	0.9585	0.0151	0.0075	0.0189	0.9736	0.0264
2005	0.9884	0.0000	0.0000	0.0116	0.9884	0.0116
2006	0.9962	0.0038	0.0000	0.0000	0.9962	0.0038
2007	0.9202	0.0266	0.0152	0.0380	0.9354	0.0646
2008	1.0000	0.0000	0.0000	0.0000	1.0000	0.0000
2009	0.9891	0.0073	0.0037	0.0000	0.9927	0.0073
2010	0.9960	0.0040	0.0000	0.0000	1.0000	0.0000
2011	0.9766	0.0140	0.0000	0.0093	0.9860	0.0140
2012	0.9341	0.0440	0.0110	0.0110	0.9780	0.0220
2013	0.8776	0.1224	0.0000	0.0000	0.9388	0.0612
2014	0.9170	0.0210	0.0210	0.0420	0.9381	0.0630
<b>Average</b>	<b>0.9297</b>	<b>0.0317</b>	<b>0.0117</b>	<b>0.0270</b>	<b>0.9503</b>	<b>0.0497</b>

<sup>a</sup> Individual ELISA samples were not collected before the 1997 brood.

### 9.4 Natural Juvenile Productivity

During 2014, juvenile summer Chinook were sampled at the Methow Trap located near RM 18.6. Trapping has occurred in this location since 2004.

#### Emigrant Estimates

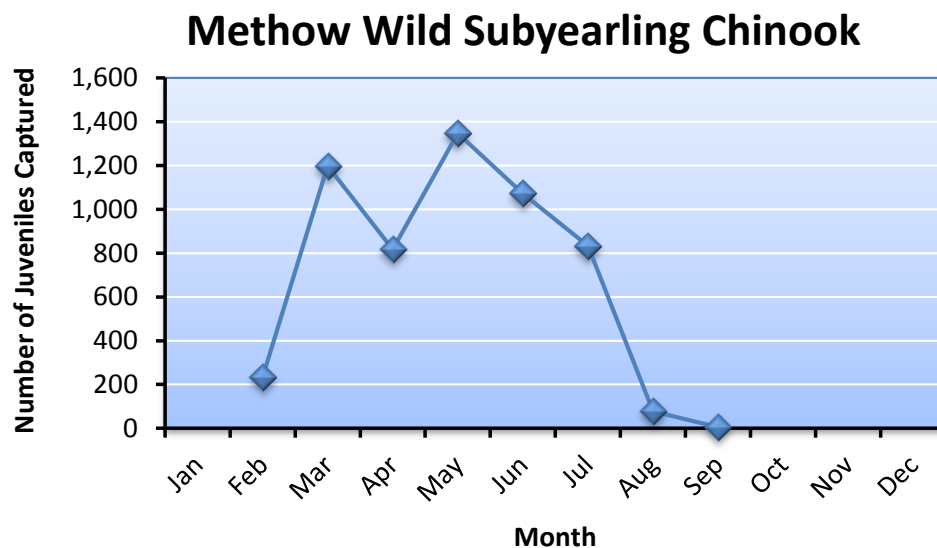
##### *Methow Trap*

On the Methow River, WDFW used traps with cone diameters of 2.4 m and 1.5 m to increase trap efficiency over a greater range of river discharge. Large variation in discharge and channel

configuration required the use of two trapping positions. The 1.5-m trap was deployed in the lower position at discharges less than 45.3 m<sup>3</sup>/s. At discharges greater than 45.3 m<sup>3</sup>/s, the 2.4-m trap was installed and operated in tandem with the 1.5 m trap.

A pooled-efficiency model estimated the total number of emigrants when the trap was operated in the low trapping position. A flow-efficiency model estimated the total number of emigrants when the trap was operated in the upper trapping position. The pooled-efficiency estimate was based on six mark-recapture release groups in 2014. The flow-efficiency estimate was based on 12 mark-recapture release groups that were conducted over the period 2008-2011.

The Methow Trap operated at night between 21 February and 29 November 2014. During that time period the trap was inoperable for 24 days because of high river flows, fires, landslides, or snow/ice. During the ten-month sampling period, a total of 5,586 wild subyearling Chinook were captured at the Methow Trap. Based on the pooled-efficiency model and the flow efficiency model, the total number of wild subyearling Chinook that emigrated past the Methow Trap in 2014 was 473,625 ( $\pm 923,267$ ). Because 516 summer Chinook redds were observed downstream from the trap in 2013, the total number of summer Chinook emigrating from the Methow River in 2014 was expanded using the ratio of the number of redds downstream from the trap to the number upstream from the trap. This resulted in a total summer Chinook emigrant estimate of 709,066 fish. Most of these fish emigrated during late May (Figure 9.1).



**Figure 9.1.** Numbers of wild subyearling Chinook captured at the Methow Trap during February through September, 2014.

## 9.5 Spawning Surveys

Surveys for Methow summer Chinook redds were conducted from late September to mid-November 2014 in the Methow River. Total redd counts (not peak counts) were conducted in the river (see Appendix N for more details).

## Redd Counts

A total of 591 summer Chinook redds were counted in the Methow River in 2014 (Table 9.13). This was lower than the overall average of 675 redds.

**Table 9.13.** Total number of redds counted in the Methow River, 1989-2014.

Survey year	Total redd count
1989	149*
1990	418*
1991	153
1992	107
1993	154
1994	310
1995	357
1996	181
1997	205
1998	225
1999	448
2000	500
2001	675
2002	2,013
2003	1,624
2004	973
2005	874
2006	1,353
2007	620
2008	599
2009	692
2010	887
2011	941
2012	960
2013	1,551
2014	591
<i>Average</i>	<i>675</i>

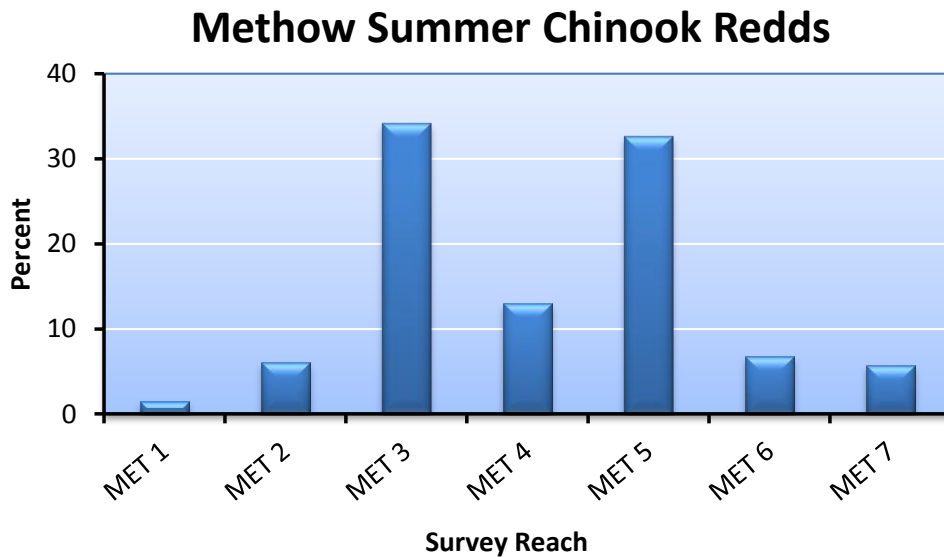
\* Total counts based on expanded aerial counts.

## Redd Distribution

Summer Chinook redds were not evenly distributed among the seven reaches in the Methow River. Most redds (67%) were located between Carlton and Twisp and between MVID and Winthrop (Reaches 3 and 5) (Table 9.14; Figure 9.2). Unlike in past years, few summer Chinook spawned downstream from Carlton (Reaches 1 and 2) because of recruitment of suspended sediments from the Carlton Complex fire. High flows also hampered redd surveys in November.

**Table 9.14.** Total number of summer Chinook redds counted in different reaches on the Methow River during September through early November, 2014. Reach codes are described in Table 2.11.

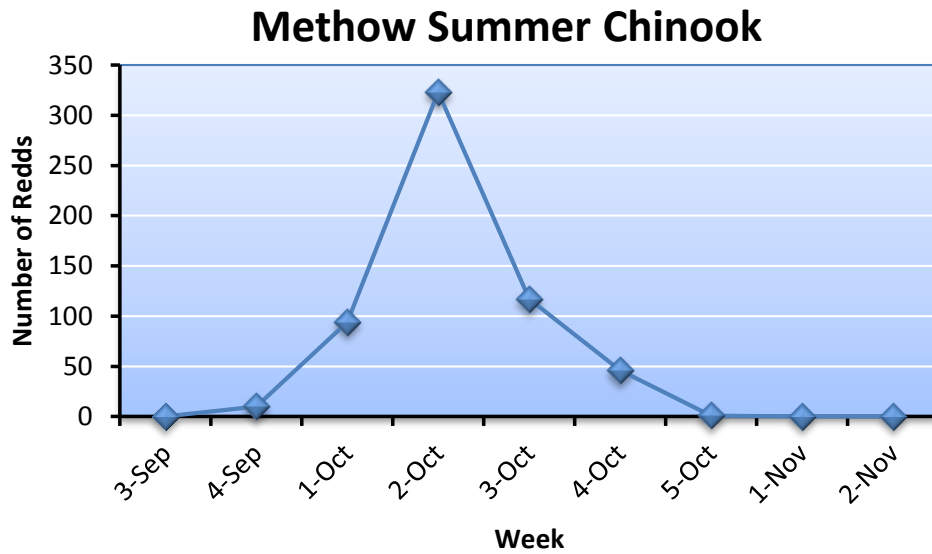
Survey reach	Total redd count	Percent
Methow 1 (M1)	9	1.5
Methow 2 (M2)	36	6.1
Methow 3 (M3)	202	34.2
Methow 4 (M4)	77	13.0
Methow 5 (M5)	193	32.7
Methow 6 (M6)	40	6.8
Methow 7 (M7)	34	5.8
<i>Totals</i>	<i>591</i>	<i>100.0</i>



**Figure 9.2.** Percent of the total number of summer Chinook redds counted in different reaches on the Methow River during September through mid-November, 2014. Reach codes are described in Table 2.11.

### Spawn Timing

Spawning in 2014 began the last week of September, peaked the second week of October, and ended after the last week of October (Figure 9.3). Stream temperatures in the Methow River, when spawning began, varied from 9.5-12.0°C. Peak spawning occurred during the first week of October in the upper reaches of the Methow River and one or two weeks later in the lower reaches.



**Figure 9.3.** Number of new summer Chinook redds counted during different weeks in the Methow River, September through mid-November 2014.

### Spawning Escapement

Spawning escapement for Methow summer Chinook was calculated as the total number of redds times the fish per redd ratio estimated from fish sampled at Wells Dam. The estimated fish per redd ratio for Methow summer Chinook in 2014 was 2.75. Multiplying this ratio by the number of redds counted in the Methow River resulted in a total spawning escapement of 1,625 summer Chinook (Table 9.15).

**Table 9.15.** Spawning escapements for summer Chinook in the Methow River for return years 1989-2014.

Return year	Fish/Redd	Redds	Total spawning escapement
1989*	3.30	149	492
1990*	3.40	418	1,421
1991*	3.70	153	566
1992*	4.30	107	460
1993*	3.30	154	508
1994*	3.50	310	1,085
1995*	3.40	357	1,214
1996*	3.40	181	615
1997*	3.40	205	697
1998	3.00	225	675
1999	2.20	448	986
2000	2.40	500	1,200
2001	4.10	675	2,768
2002	2.30	2,013	4,630
2003	2.42	1,624	3,930

Return year	Fish/Redd	Redds	Total spawning escapement
2004	2.25	973	2,189
2005	2.93	874	2,561
2006	2.02	1,353	2,733
2007	2.20	620	1,364
2008	3.25	599	1,947
2009	2.54	692	1,758
2010	2.81	887	2,492
2011	3.10	941	2,917
2012	3.07	960	2,947
2013	2.31	1,551	3,583
2014	2.75	591	1,625
<i>Average</i>	<i>2.98</i>	<i>675</i>	<i>1,822</i>

\* Spawning escapement was calculated using the "Modified Meekin Method" (i.e., 3.1 x jack multiplier).

## 9.6 Carcass Surveys

Surveys for Methow summer Chinook carcasses were conducted during late September to mid-November 2014 in the Methow River (see Appendix N for more details).

### Number sampled

A total of 487 summer Chinook carcasses were sampled during September through mid-November in the Methow River (Table 9.16). This was lower than the overall average of 507 carcasses sampled since 1991.

**Table 9.16.** Numbers of summer Chinook carcasses sampled within each survey reach on the Methow River, 1991-2014. Reach codes are described in Table 2.11.

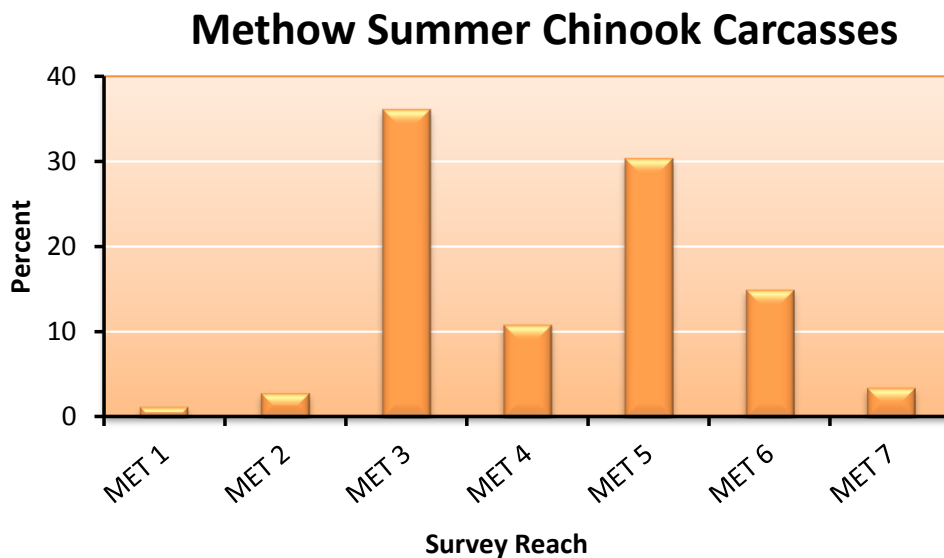
Survey year	Number of summer Chinook carcasses							Total
	M-1	M-2	M-3	M-4	M-5	M-6	M-7	
1991	0	12	8	4	2	0	0	26
1992	8	8	19	0	17	1	0	53
1993	19	25	14	2	5	0	0	65
1994 <sup>a</sup>	43	33	20	5	13	0	0	114
1995	14	33	58	7	7	0	0	119
1996	6	30	46	5	2	0	0	89
1997	6	12	38	2	19	1	0	78
1998	90	84	99	17	30	0	0	320
1999	47	144	232	32	37	12	2	506
2000	62	118	105	9	99	5	0	398
2001	392	275	88	14	76	11	1	857
2002	551	318	518	164	219	34	10	1,814
2003	115	268	317	115	128	5	0	948
2004	40	173	187	82	92	2	1	577

Survey year	Number of summer Chinook carcasses							
	M-1	M-2	M-3	M-4	M-5	M-6	M-7	Total
2005	154	173	182	42	112	3	0	<b>666</b>
2006	121	148	110	56	144	3	1	<b>583</b>
2007	142	132	108	27	53	0	0	<b>462</b>
2008	64	128	197	33	57	3	0	<b>482</b>
2009	144	158	159	36	94	0	0	<b>591</b>
2010	105	180	184	38	63	5	1	<b>576</b>
2011	56	134	201	78	83	5	1	<b>558</b>
2012	127	154	169	75	82	14	7	<b>628</b>
2013	296	287	385	90	100	7	5	<b>1,170</b>
2014	6	14	176	53	148	73	17	<b>487</b>
<i>Average</i>	<i>109</i>	<i>127</i>	<i>151</i>	<i>41</i>	<i>70</i>	<i>8</i>	<i>2</i>	<i>507</i>

<sup>a</sup> An additional 113 carcasses were sampled, but reach was not identified.

### Carcass Distribution and Origin

Summer Chinook carcasses were not evenly distributed among reaches within the Methow River in 2014 (Table 9.15; Figure 9.4). Most of the carcasses were found between Carlton and Twisp and between MVID and Winthrop (Reaches 3 and 5). Unlike in past years, few summer Chinook carcasses were found downstream from Carlton (Reaches 1 and 2) because of high turbidity levels associated with the Carlton Complex fire. High flows also hampered carcass surveys in November.



**Figure 9.4.** Percent of summer Chinook carcasses sampled within different reaches on the Methow River during September through mid-November, 2014. Reach codes are described in Table 2.11.



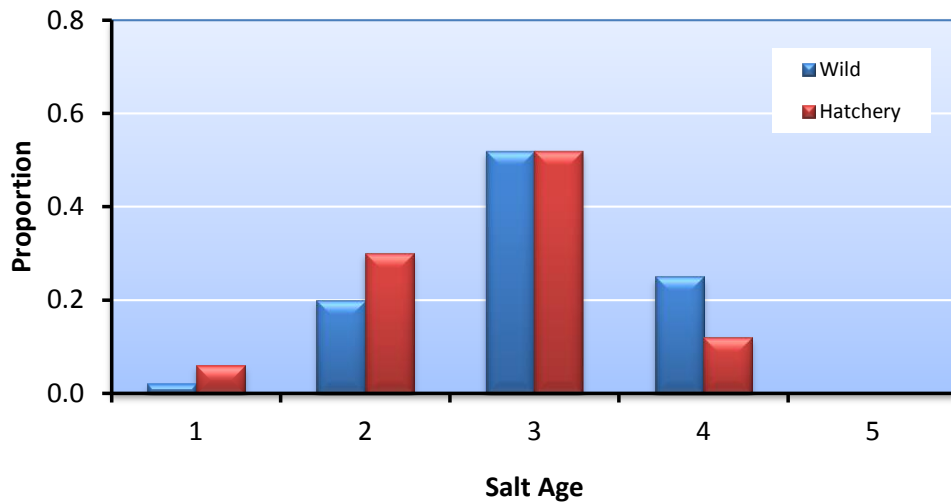
Numbers of wild and hatchery-origin summer Chinook carcasses sampled in 2014 will be available after analysis of CWTs and scales. Based on the available data (1991-2013), hatchery and wild summer Chinook carcasses were not distributed equally among the reaches in the Methow River (Table 9.17). A larger percentage of hatchery carcasses occurred in the lower reaches, while a larger percentage of wild summer Chinook carcasses occurred in upstream reaches (Figure 9.5).

**Table 9.17.** Numbers of wild and hatchery summer Chinook carcasses sampled within different reaches on the Methow River, 1991-2013.

Survey year	Origin	Survey reach							Total
		M-1	M-2	M-3	M-4	M-5	M-6	M-7	
1991	Wild	0	12	8	4	2	0	0	26
	Hatchery	0	0	0	0	0	0	0	0
1992	Wild	8	8	19	0	17	1	0	53
	Hatchery	0	0	0	0	0	0	0	0
1993	Wild	11	18	9	0	3	0	0	41
	Hatchery	8	7	5	2	2	0	0	24
1994	Wild	23	18	9	5	10	0	0	65
	Hatchery	20	15	11	0	3	0	0	49
1995	Wild	7	9	33	7	6	0	0	62
	Hatchery	7	24	25	0	1	0	0	57
1996	Wild	1	23	35	4	2	0	0	65
	Hatchery	5	7	11	1	0	0	0	24
1997	Wild	5	8	31	1	17	0	0	62
	Hatchery	1	4	7	1	2	1	0	16
1998	Wild	42	48	71	11	25	0	0	197
	Hatchery	48	36	28	6	5	0	0	123
1999	Wild	32	87	130	15	24	4	2	294
	Hatchery	15	57	102	17	13	8	0	212
2000	Wild	25	85	85	8	83	3	0	289
	Hatchery	37	33	20	1	16	2	0	109
2001	Wild	62	118	56	10	70	11	1	328
	Hatchery	330	157	32	4	6	0	0	529
2002	Wild	138	177	380	140	197	34	9	1,075
	Hatchery	413	141	138	24	22	0	1	739
2003	Wild	33	146	188	76	92	3	0	538
	Hatchery	82	122	129	39	36	2	0	410
2004	Wild	16	120	155	65	78	1	0	435
	Hatchery	24	53	32	17	14	1	1	142
2005	Wild	62	99	133	33	107	3	0	437
	Hatchery	92	74	49	9	5	0	0	229
2006	Wild	52	82	67	44	109	2	1	357
	Hatchery	69	66	43	12	35	1	0	226

Survey year	Origin	Survey reach							Total
		M-1	M-2	M-3	M-4	M-5	M-6	M-7	
2007	Wild	35	58	59	16	40	0	0	208
	Hatchery	107	74	49	11	13	0	0	254
2008	Wild	13	62	146	27	52	2	0	302
	Hatchery	51	66	51	6	5	1	0	180
2009	Wild	45	87	103	27	84	0	0	346
	Hatchery	99	71	56	9	10	0	0	245
2010	Wild	33	79	101	24	53	5	1	296
	Hatchery	72	101	83	14	10	0	0	280
2011	Wild	21	56	87	54	56	5	1	280
	Hatchery	35	78	114	24	27	0	0	278
2012	Wild	59	53	96	58	74	13	7	355
	Hatchery	73	101	73	17	8	1	0	273
2013	Wild	110	128	178	67	64	7	5	559
	Hatchery	186	160	208	23	36	0	0	613
Average	Wild	36	69	95	30	55	4	1	290
	Hatchery	77	63	55	10	12	1	0	218

### Methow Summer Chinook



**Figure 9.5.** Distribution of wild and hatchery produced carcasses in different reaches on the Methow River, 1993-2013. Reach codes are described in Table 2.11.

#### Sampling Rate

Overall, 30% of the total spawning escapement of summer Chinook in the Methow River basin was sampled in 2014 (Table 9.18). Sampling rates among survey reaches varied from 14 to 66%.

**Table 9.18.** Number of redds and carcasses, total spawning escapement, and sampling rates for summer Chinook in the Methow River basin, 2014. Reach codes are described in Table 2.11.

Survey reach	Total number of redds	Total number of carcasses	Total spawning escapement	Sampling rate
Methow 1 (M1)	9	6	25	0.24
Methow 2 (M2)	36	14	99	0.14
Methow 3 (M3)	202	176	556	0.32
Methow 4 (M4)	77	53	212	0.25
Methow 5 (M5)	193	148	531	0.28
Methow 6 (M6)	40	73	110	0.66
Methow 7 (M7)	34	17	94	0.18
<b>Total</b>	<b>591</b>	<b>487</b>	<b>1,625</b>	<b>0.30</b>

### Length Data

Mean lengths (POH, cm) of male and female summer Chinook carcasses sampled during surveys on the Methow River in 2014 are provided in Table 9.19. The average size of males and females sampled in the Methow River were 65 cm and 68 cm, respectively.

**Table 9.19.** Mean lengths (postorbital-to-hypural length; cm) and standard deviations (in parentheses) of male and female summer Chinook carcasses sampled in different reaches on the Methow River, 2014. Reach codes are described in Table 2.11.

Stream/watershed	Mean length (cm)	
	Male	Female
Methow 1 (M1)	52.0 (12.0)	71.0 (0.0)
Methow 2 (M2)	65.3 (10.4)	66.4 (7.5)
Methow 3 (M3)	63.7 (9.5)	68.0 (5.1)
Methow 4 (M4)	65.1 (7.6)	69.5 (5.4)
Methow 5 (M5)	64.8 (7.9)	68.7 (5.6)
Methow 6 (M6)	67.2 (8.4)	69.0 (4.7)
Methow 7 (M7)	66.8 (5.7)	65.6 (4.4)
<b>Total</b>	<b>64.7 (8.8)</b>	<b>68.4 (5.3)</b>

## 9.7 Life History Monitoring

Life history characteristics of Methow summer Chinook were assessed by examining carcasses on spawning grounds and fish collected or examined at broodstock collection sites, and by reviewing tagging data and fisheries statistics.

### Migration Timing

Migration timing of hatchery and wild Methow/Okanogan summer Chinook was determined from broodstock data collected at Wells Dam. Counting of summer/fall Chinook at Wells Dam occurs from 29 June to 15 November. Broodstock collection at the Dam occurs from early July

(week 27) to mid-September (week 37) (Table 2.1). Based on broodstock sampling in 2014, hatchery summer Chinook generally arrived at Wells Dam earlier than did wild summer Chinook (Table 9.20). This was true throughout most of the migration period. In contrast, there was little difference in migration timing between wild and hatchery summer Chinook when data were pooled for the 2007-2014 survey period.

**Table 9.20.** The week that 10%, 50% (median), and 90% of the wild and hatchery summer Chinook salmon passed Wells Dam, 2007-2014. The average week is also provided. Migration timing is based on collection of summer Chinook broodstock at Wells Dam.

Survey year	Origin	Methow/Okanogan Summer Chinook Migration Time (week)				Sample size
		10 Percentile	50 Percentile	90 Percentile	Mean	
2007	Wild	27	30	34	30	485
	Hatchery	27	30	33	30	433
2008	Wild	28	30	34	30	542
	Hatchery	28	30	36	31	884
2009	Wild	27	29	34	30	585
	Hatchery	27	29	33	29	708
2010	Wild	27	29	33	29	377
	Hatchery	27	29	32	29	801
2011	Wild	30	32	36	32	516
	Hatchery	30	32	35	33	1223
2012	Wild	28	30	34	31	192
	Hatchery	28	31	34	31	591
2013	Wild	27	30	33	30	229
	Hatchery	27	30	33	30	282
2014	Wild	27	31	40	32	316
	Hatchery	27	30	35	30	208
<i>Average</i>	<i>Wild</i>	<i>27</i>	<i>30</i>	<i>35</i>	<i>31</i>	<i>3,242</i>
	<i>Hatchery</i>	<i>28</i>	<i>30</i>	<i>35</i>	<i>31</i>	<i>5,130</i>

### Age at Maturity

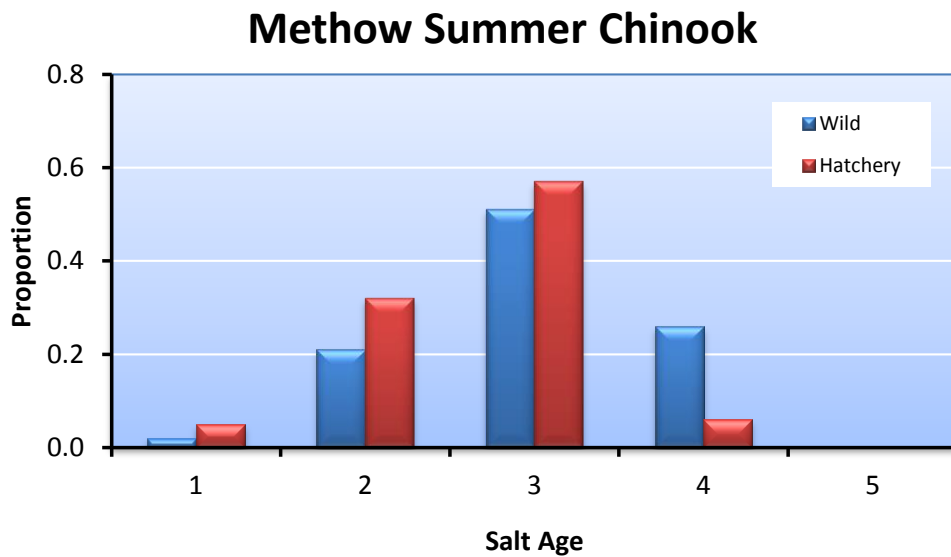
Because hatchery summer Chinook are released after one year of rearing and natural-origin summer Chinook migrate primarily as age-0 fish, total ages will differ between hatchery and natural-origin Chinook (see Hillman et al. 2011). Therefore, in this section, we evaluated age at maturity by comparing differences in salt (ocean) ages between the two groups.

Most of the wild and hatchery summer Chinook sampled during the period 1993-2013 in the Methow River were salt age-3 fish (Table 9.21; Figure 9.6). A higher percentage of salt age-4 wild Chinook returned to the basin than did salt age-4 hatchery Chinook. In contrast, a higher proportion of salt age-1 and 2 hatchery fish returned than did salt age-1 and 2 wild fish. Thus, a higher percentage of wild fish returned at an older age than did hatchery fish.

**Table 9.21.** Proportions of wild and hatchery summer Chinook of different salt (ocean) ages sampled on spawning grounds in the Methow River, 1993-2013.

Sample year	Origin	Salt age						Sample size
		1	2	3	4	5	6	
1993	Wild	0.05	0.08	0.76	0.11	0.00	0.00	38
	Hatchery	0.00	1.00	0.00	0.00	0.00	0.00	20
1994	Wild	0.03	0.26	0.51	0.20	0.00	0.00	101
	Hatchery	0.00	0.07	0.93	0.00	0.00	0.00	111
1995	Wild	0.00	0.09	0.70	0.20	0.00	0.00	54
	Hatchery	0.02	0.04	0.44	0.51	0.00	0.00	55
1996	Wild	0.04	0.30	0.54	0.13	0.00	0.00	56
	Hatchery	0.00	0.05	0.50	0.41	0.05	0.00	22
1997	Wild	0.00	0.22	0.51	0.27	0.00	0.00	55
	Hatchery	0.13	0.06	0.56	0.25	0.00	0.00	16
1998	Wild	0.09	0.38	0.45	0.09	0.00	0.00	188
	Hatchery	0.02	0.52	0.41	0.04	0.00	0.00	123
1999	Wild	0.01	0.51	0.43	0.05	0.00	0.00	252
	Hatchery	0.00	0.07	0.90	0.03	0.00	0.00	210
2000	Wild	0.01	0.09	0.75	0.16	0.00	0.00	257
	Hatchery	0.10	0.16	0.62	0.11	0.00	0.00	97
2001	Wild	0.02	0.20	0.72	0.07	0.00	0.00	292
	Hatchery	0.10	0.60	0.26	0.04	0.00	0.00	526
2002	Wild	0.01	0.17	0.61	0.21	0.00	0.00	1,003
	Hatchery	0.01	0.41	0.57	0.01	0.00	0.00	734
2003	Wild	0.01	0.11	0.50	0.37	0.00	0.00	478
	Hatchery	0.02	0.03	0.90	0.04	0.00	0.00	399
2004	Wild	0.00	0.09	0.35	0.56	0.00	0.00	394
	Hatchery	0.07	0.28	0.30	0.35	0.00	0.00	141
2005	Wild	0.11	0.74	0.14	0.01	0.00	0.00	410
	Hatchery	0.06	0.26	0.65	0.02	0.00	0.00	220
2006	Wild	0.00	0.02	0.33	0.64	0.00	0.00	356
	Hatchery	0.01	0.19	0.50	0.30	0.00	0.00	164
2007	Wild	0.03	0.09	0.24	0.59	0.05	0.00	208
	Hatchery	0.07	0.09	0.75	0.09	0.01	0.00	213
2008	Wild	0.01	0.14	0.71	0.13	0.01	0.00	298
	Hatchery	0.10	0.45	0.30	0.15	0.00	0.00	138
2009	Wild	0.00	0.11	0.41	0.48	0.00	0.00	317
	Hatchery	0.17	0.26	0.53	0.04	0.00	0.00	242
2010	Wild	0.01	0.16	0.59	0.24	0.00	0.00	269
	Hatchery	0.01	0.69	0.29	0.02	0.00	0.00	247

Sample year	Origin	Salt age						Sample size
		1	2	3	4	5	6	
2011	Wild	0.02	0.09	0.60	0.30	0.00	0.00	255
	Hatchery	0.16	0.10	0.74	0.01	0.00	0.00	261
2012	Wild	0.03	0.24	0.53	0.21	0.00	0.00	315
	Hatchery	0.09	0.71	0.16	0.04	0.00	0.00	243
2013	Wild	0.02	0.25	0.62	0.11	0.00	0.00	533
	Hatchery	0.02	0.18	0.79	0.01	0.00	0.00	570
Average	Wild	0.02	0.21	0.51	0.26	0.00	0.00	292
	Hatchery	0.05	0.32	0.57	0.06	0.00	0.00	226



**Figure 9.6.** Proportions of wild and hatchery summer Chinook of different salt (ocean) ages sampled at broodstock collection sites and on spawning grounds in the Methow River for the combined years 1993-2013.

### Size at Maturity

On average, hatchery summer Chinook were about 4 cm smaller than wild summer Chinook sampled in the Methow River basin (Table 9.22). This is likely because a higher percentage of wild fish returned as salt age-4 fish than did hatchery fish. Future analyses will compare sizes of hatchery and wild fish of the same age groups and sex.

**Table 9.22.** Mean lengths (POH; cm) and variability statistics for wild and hatchery summer Chinook sampled in the Methow River basin, 1993-2013; SD = 1 standard deviation.

Survey year	Origin	Sample size	Summer Chinook length (POH; cm)			
			Mean	SD	Minimum	Maximum
1993 <sup>a</sup>	Wild	41	74	9	51	89
	Hatchery	24	62	8	36	80
1994 <sup>a</sup>	Wild	112	69	8	35	87
	Hatchery	114	67	5	43	77
1995	Wild	62	74	6	52	88
	Hatchery	56	73	7	46	85
1996	Wild	64	70	11	34	91
	Hatchery	23	72	7	58	85
1997	Wild	62	76	9	35	90
	Hatchery	16	68	15	33	87
1998	Wild	196	67	10	38	97
	Hatchery	123	63	10	37	87
1999	Wild	292	66	8	43	99
	Hatchery	212	66	7	26	89
2000	Wild	288	74	8	37	89
	Hatchery	109	68	12	24	87
2001	Wild	328	67	10	29	86
	Hatchery	529	63	10	31	87
2002	Wild	1,075	70	8	37	94
	Hatchery	739	67	9	33	87
2003	Wild	538	71	8	35	88
	Hatchery	410	69	8	35	89
2004	Wild	435	73	7	38	89
	Hatchery	142	65	12	34	85
2005	Wild	437	69	8	45	86
	Hatchery	229	64	9	36	79
2006	Wild	438	73	7	35	92
	Hatchery	149	69	8	38	91
2007	Wild	249	72	11	33	89
	Hatchery	219	69	9	22	84
2008	Wild	384	69	8	30	90
	Hatchery	210	63	15	23	86
2009	Wild	363	71	9	32	88
	Hatchery	228	63	12	30	83
2010	Wild	296	69	8	33	90
	Hatchery	280	62	9	39	81

Survey year	Origin	Sample size	Summer Chinook length (POH; cm)			
			Mean	SD	Minimum	Maximum
2011	Wild	280	70	9	31	89
	Hatchery	278	64	11	26	82
2012	Wild	355	68	8	36	85
	Hatchery	273	59	9	21	81
2013	Wild	559	65	9	31	89
	Hatchery	613	66	8	27	83
<i>Pooled</i>	<i>Wild</i>	<i>6,854</i>	<i>70</i>	<i>9</i>	<i>29</i>	<i>99</i>
	<i>Hatchery</i>	<i>4,976</i>	<i>66</i>	<i>10</i>	<i>21</i>	<i>91</i>

<sup>a</sup> These years include sizes reported in annual reports. The data contained in the WDFW database do not include all these data.

### Contribution to Fisheries

Most of the harvest on hatchery-origin Methow summer Chinook occurred in the Ocean (Table 9.23). Ocean harvest has made up 13% to 99% of all hatchery-origin Methow summer Chinook harvested. Brood years 1989, 1998, 26, and 2008 provided the largest harvests, while brood years 1996 and 1999 provided the lowest.

**Table 9.23.** Estimated number and percent (in parentheses) of hatchery-origin Methow summer Chinook captured in different fisheries, brood years 1989-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
1989	1,041 (52)	884 (44)	0 (0)	66 (3)	1,991
1990	53 (56)	41 (44)	0 (0)	0 (0)	94
1991	10 (17)	49 (83)	0 (0)	0 (0)	59
1992	17 (55)	14 (45)	0 (0)	0 (0)	31
1993	14 (58)	8 (33)	2 (8)	0 (0)	24
1994	153 (81)	34 (18)	1 (1)	1 (1)	189
1995	77 (99)	0 (0)	1 (1)	0 (0)	78
1996	12 (92)	1 (8)	0 (0)	0 (0)	13
1997	214 (88)	7 (3)	0 (0)	21 (9)	242
1998	1,755 (83)	101 (5)	14 (1)	234 (11)	2,104
1999	2 (13)	13 (87)	0 (0)	0 (0)	15
2000	357 (71)	88 (17)	27 (5)	33 (7)	505
2001	319 (52)	97 (16)	43 (7)	160 (26)	619
2002	271 (48)	96 (17)	61 (11)	137 (24)	565
2003	58 (58)	17 (17)	7 (7)	18 (18)	100
2004	132 (49)	55 (20)	16 (6)	68 (25)	271
2005	295 (54)	137 (25)	50 (9)	66 (12)	548
2006	1,127 (48)	811 (35)	100 (4)	314 (13)	2,352
2007	205 (60)	69 (20)	16 (5)	54 (16)	344



Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
2008	1,280 (57)	349 (15)	43 (2)	592 (26)	2,264
<i>Average</i>	<i>370 (60)</i>	<i>144 (28)</i>	<i>19 (3)</i>	<i>88 (10)</i>	<i>620</i>

## Straying

Stray rates were determined by examining CWTs recovered on spawning grounds within and outside the Methow River basin. Targets for strays based on return year (recovery year) and brood year should be less than 5%.

Few hatchery-origin Methow summer Chinook have strayed into basins outside the Methow (Table 9.24). Although hatchery-origin Methow summer Chinook have strayed into the Wenatchee River basin, Okanogan River basin, Entiat River basin, Chelan tailrace, and Hanford Reach, they have made up less than 1% of the spawning escapement within those areas.

**Table 9.24.** Number and percent of spawning escapements within other non-target basins that consisted of hatchery-origin Methow summer Chinook, return years 1994-2011. For example, for return year 2002, 0.4% of the summer Chinook escapement in the Okanogan River basin consisted of hatchery-origin Methow summer Chinook. Percent strays should be less than 5%.

Return year	Wenatchee		Okanogan		Chelan		Entiat		Hanford Reach	
	Number	%	Number	%	Number	%	Number	%	Number	%
1994	0	0.0	72	1.8	-	-	-	-	-	-
1995	0	0.0	9	0.3	-	-	-	-	-	-
1996	0	0.0	0	0.0	-	-	-	-	-	-
1997	0	0.0	0	0.0	-	-	-	-	-	-
1998	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1999	0	0.0	6	0.2	0	0.0	0	0.0	7	0.0
2000	0	0.0	3	0.1	0	0.0	0	0.0	0	0.0
2001	0	0.0	0	0.0	0	0.0	0	0.0	7	0.0
2002	0	0.0	54	0.4	0	0.0	0	0.0	0	0.0
2003	0	0.0	1	0.0	6	1.4	0	0.0	0	0.0
2004	0	0.0	7	0.1	3	0.7	0	0.0	0	0.0
2005	0	0.0	24	0.3	0	0.0	0	0.0	0	0.0
2006	0	0.0	12	0.1	0	0.0	0	0.0	0	0.0
2007	0	0.0	17	0.4	2	1.1	1	0.4	0	0.0
2008	0	0.0	12	0.2	0	0.0	0	0.0	0	0.0
2009	0	0.0	14	0.2	0	0.0	0	0.0	0	0.0
2010	6	0.1	44	0.7	22	2.0	0	0.0	0	0.0
2011	0	0.0	45	0.5	8	0.6	0	0.0	0	0.0
<i>Average</i>	<i>0</i>	<i>0.0</i>	<i>18</i>	<i>0.3</i>	<i>3</i>	<i>0.4</i>	<i>0</i>	<i>0.0</i>	<i>0</i>	<i>0.0</i>

Based on brood year analyses, on average, about 3% of the returns have strayed into non-target spawning areas, falling within the acceptable level of less than 5% (Table 9.25). Depending on brood year, percent strays into non-target spawning areas have ranged from 0-11.9%. Few (<1% on average) have strayed into non-target hatchery programs.

**Table 9.25.** Number and percent of hatchery-origin Methow summer Chinook that homed to target spawning areas and the target hatchery program, and number and percent that strayed to non-target spawning areas and non-target hatchery programs, by brood years 1989-2008. Percent stays should be less than 5%.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1989	773	55.7	459	33.0	81	5.8	76	5.5
1990	199	70.6	81	28.7	0	0.0	2	0.7
1991	82	65.6	43	34.4	0	0.0	0	0.0
1992	68	63.0	40	37.0	0	0.0	0	0.0
1993	25	65.8	10	26.3	3	7.9	0	0.0
1994	419	79.7	94	17.9	13	2.5	0	0.0
1995	126	81.8	28	18.2	0	0.0	0	0.0
1996	57	93.4	4	6.6	0	0.0	0	0.0
1997	379	93.8	7	1.7	18	4.5	0	0.0
1998	1,653	94.7	32	1.8	60	3.4	0	0.0
1999	18	100.0	0	0.0	0	0.0	0	0.0
2000	239	93.0	4	1.6	14	5.4	0	0.0
2001	272	88.3	6	1.9	29	9.4	1	0.3
2002	315	95.2	4	1.2	12	3.6	0	0.0
2003	131	99.2	1	0.8	0	0.0	0	0.0
2004	194	85.5	6	2.6	27	11.9	0	0.0
2005	373	90.5	13	3.2	23	5.6	3	0.7
2006	1,317	91.4	15	1.0	109	7.6	0	0.0
2007	134	98.5	2	1.5	0	0.0	0	0.0
2008	1,871	97.9	13	0.7	25	1.3	3	0.2
<b>Average</b>	<b>434</b>	<b>85.2</b>	<b>44</b>	<b>11.0</b>	<b>21</b>	<b>3.4</b>	<b>4</b>	<b>0.4</b>

\* Homing to the target hatchery includes Methow hatchery summer Chinook that are captured and included as broodstock in the Methow Hatchery program. These hatchery fish are typically collected at Wells Dam.

## Genetics

Genetic studies were conducted to investigate relationships among temporally replicated collections of summer Chinook from the Wenatchee River, Methow River, and Okanogan River in the upper Columbia River basin (Kassler et al. 2011; the entire report is appended as Appendix M). Samples from the Eastbank Hatchery – Wenatchee stock, Eastbank Hatchery – Methow/Okanogan (MEOK) stock, and Wells Hatchery were also included in the analysis.

Samples of natural and hatchery-origin summer Chinook were analyzed and compared to determine if the supplementation program has affected the genetic structure of these populations. The study also calculated the effective number of breeders for collection locations of natural and hatchery-origin summer Chinook from 1993 and 2008.

In general, population differentiation was not observed among the temporally replicated collection locations. A single collection from the Okanogan River (1993) was the only collection showing statistically significant differences. The effective number of breeders was not statistically different from the early collection in 1993 in comparison to the late collection in 2008. Overall, these analyses revealed a lack of differentiation among the temporal replicates from the same locations and among the collection from different locations, suggesting the populations have been homogenized or that there has been substantial gene flow among populations. Additional comparisons among summer-run and fall-run Chinook populations in the upper Columbia River were conducted to determine if there was any differentiation between Chinook with different run timing. These analyses revealed pairwise  $F_{ST}$  values that were less than 0.01 for the collections of summer Chinook to collections of fall Chinook from Hanford Reach, lower Yakima River, Priest Rapids, and Umatilla. Collections of fall Chinook from Crab Creek, Lyons Ferry Hatchery, Marion Drain, and Snake River had pairwise  $F_{ST}$  values that were higher in comparison to the collections of summer Chinook. The consensus clustering analysis did not provide good statistical support to the groupings, but did show relationships among collections based on geographic proximity. Overall the summer and fall run Chinook that have historically been spawned together were not differentiated while fall Chinook from greater geographic distances were differentiated.

### Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For brood years 1993-2003, the approximate PNI Index was generally less than 0.67 (Table 9.26). However, since brood year 2003, the approximate PNI Index has generally been greater than 0.67; brood years 2011 and 2013 had PNI Index values of 0.66.

**Table 9.26.** Approximate Proportionate Natural Influence (PNI) Index of the Methow summer Chinook supplementation program for brood years 1989-2013. PNI was calculated as the proportion of naturally produced Chinook in the hatchery broodstock (pNOB) divided by the proportion of hatchery Chinook on the spawning grounds (pHOS) plus pNOB. NOS = number of natural-origin Chinook on the spawning grounds; HOS = number of hatchery-origin Chinook on the spawning grounds; NOB = number of natural-origin Chinook collected for broodstock; and HOB = number of hatchery-origin Chinook included in hatchery broodstock.

Brood year	Spawners			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
1989	492	0	0.00	1,297	312	0.81	1.00
1990	1,421	0	0.00	828	206	0.80	1.00
1991	566	0	0.00	924	314	0.75	1.00
1992	460	0	0.00	297	406	0.42	1.00
1993	314	194	0.38	681	388	0.64	0.63
1994	596	489	0.45	341	244	0.58	0.56
1995	596	618	0.51	173	240	0.42	0.45
1996	435	180	0.29	287	155	0.65	0.69
1997	529	168	0.24	197	265	0.43	0.64
1998	437	238	0.35	153	211	0.42	0.55
1999	573	413	0.42	224	289	0.44	0.51
2000	861	339	0.28	164	337	0.33	0.54
2001	1,122	1,646	0.59	12	345	0.03	0.05
2002	2,572	2,058	0.44	247	241	0.51	0.54
2003	2,307	1,623	0.41	381	101	0.79	0.66
2004	1,622	567	0.26	506	16	0.97	0.79
2005	1,672	889	0.35	391	9	0.98	0.74
2006	2,039	694	0.25	500	10	0.98	0.80
2007	764	600	0.44	456	17	0.96	0.69
2008	1,293	654	0.34	359	86	0.81	0.70
2009	1,093	665	0.38	503	4	0.99	0.72
2010	1,326	1,166	0.47	484	8	0.98	0.68
2011	1,503	1,414	0.48	467	26	0.95	0.66
2012	1,593	1,354	0.46	98	1	0.99	0.68
2013	1,807	1,776	0.50	97	4	0.96	0.66
<b>Average</b>	<b>1,120</b>	<b>710</b>	<b>0.33</b>	<b>403</b>	<b>169</b>	<b>0.70</b>	<b>0.68</b>

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel time (arithmetic mean days) of hatchery summer Chinook from the Methow River release site to McNary Dam, and smolt to

adult ratios (SARs) from release to detection at Bonneville Dam (Table 9.27).<sup>12</sup> Over the three brood years for which PIT-tagged hatchery fish were released, survival rates from the Methow River to McNary Dam ranged from 0.485 to 0.747; SARs from release to detection at Bonneville Dam ranged from 0.002 to 0.016. Average travel time from the Methow River to McNary Dam ranged from 17 to 39 days.

**Table 9.27.** Total number of Methow hatchery summer Chinook released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2008-2012. Standard errors are shown in parentheses. NA = not available (i.e., not all the fish from the release groups have returned to the Columbia River).

Brood year	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2008	10,094	0.747 (0.055)	39.1 (13.0)	0.016 (0.001)
2009	5,020	0.485 (0.037)	30.2 (11.1)	0.002 (0.001)
2010	0	--	--	--
2011	0	--	--	--
2012	9,801	0.545 (0.046)	17.0 (8.1)	NA

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). We calculated NORs with and without harvest. NORs without harvest include all returning fish that either returned to the basin or were collected as wild broodstock. NORs with harvest include all fish harvested and are based on a brood year harvest rates from the hatchery program. For brood years 1989-2007, NRR for summer Chinook in the Methow averaged 1.15 (range, 0.10-4.90) if harvested fish were not include in the estimate and 2.36 (range, 0.18-10.84) if harvested fish were included in the estimate (Table 9.28). NRRs for more recent brood years will be calculated as soon as all tag recoveries and sampling rates have been loaded into the database.

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and were calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. These rates should be greater than the NRRs and greater than or equal to 5.30 (the calculated target value in Hillman et al. 2013). HRRs exceeded NRRs in 11 out of the 19 years of data, regardless if harvest was or was not included in the estimate (Table 9.28). Hatchery replacement rates for Methow summer Chinook have exceeded the estimated target value of 5.30 in three of the 19 years of data, regardless if harvest was or was not included in the estimate.

<sup>12</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

**Table 9.28.** Broodstock collected, spawning escapements, natural and hatchery-origin recruits (NOR and HOR), and natural and hatchery replacement rates (NRR and HRR; with and without harvest) for wild summer Chinook in the Methow River basin, brood years 1989-2007.

Brood year	Broodstock Collected	Spawning Escapement	Harvest not included				Harvest included			
			HOR	NOR	HRR	NRR	HOR	NOR	HRR	NRR
1989	202	492	1,389	631	6.88	1.28	3,380	1,528	16.73	3.11
1990	202	1,421	282	979	1.40	0.69	376	1,311	1.86	0.92
1991	266	566	125	288	0.47	0.51	184	427	0.69	0.75
1992	214	460	108	614	0.50	1.33	139	792	0.65	1.72
1993	234	508	82	431	0.35	0.85	132	703	0.56	1.38
1994	260	1,085	526	545	2.02	0.50	715	743	2.75	0.68
1995	242	1,214	154	1,201	0.64	0.99	232	1,809	0.96	1.49
1996	220	615	61	445	0.28	0.72	74	541	0.34	0.88
1997	209	697	404	1,494	1.93	2.14	646	2,398	3.09	3.44
1998	235	675	1,745	3,308	7.43	4.90	3,849	7,319	16.38	10.84
1999	222	986	18	2,863	0.08	2.90	33	5,253	0.15	5.33
2000	222	1,200	257	808	1.16	0.67	762	2,405	3.43	2.00
2001	223	2,768	308	2,877	1.38	1.04	927	8,718	4.16	3.15
2002	222	4,630	331	1,072	1.49	0.23	896	2,921	4.04	0.63
2003	224	3,930	132	397	0.59	0.10	232	698	1.04	0.18
2004	223	2,189	227	1,646	1.02	0.75	498	3,618	2.23	1.65
2005	225	2,561	412	1,159	1.83	0.45	960	2,708	4.27	1.06
2006	236	2,733	1,441	1,714	6.11	0.63	3,793	4,522	16.07	1.65
2007	209	1,364	136	1,503	0.65	1.10	480	5,330	2.30	3.91
<b>Average</b>	<b>226</b>	<b>1,584</b>	<b>428</b>	<b>1,262</b>	<b>1.91</b>	<b>1.15</b>	<b>964</b>	<b>2,829</b>	<b>4.30</b>	<b>2.36</b>

### Smolt-to-Adult Survivals

Smolt-to-adult survival ratios (SARs) were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery smolts released. Here, SARs were based on CWT returns. For the available brood years, SARs have ranged from 0.00008 to 0.01883 for hatchery summer Chinook in the Methow River basin (Table 9.29).

**Table 9.29.** Smolt-to-adult ratios (SARs) for Methow summer Chinook, brood years 1989-2008.

Brood year	Number of tagged smolts released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1989	358,237	2,869	0.00801
1990	371,483	359	0.00097
1991	377,097	129	0.00034
1992	392,636	138	0.00035
1993	200,345	62	0.00031
1994	400,488	710	0.00177

Brood year	Number of tagged smolts released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1995	344,974	229	0.00066
1996	289,880	73	0.00025
1997	380,430	642	0.00169
1998	202,559	3,815	0.01883
1999	422,473	33	0.00008
2000	334,337	761	0.00228
2001	246,159	923	0.00375
2002	310,846	893	0.00287
2003	353,495	232	0.00066
2004	394,490	495	0.00125
2005	262,496	958	0.00365
2006	417,795	3,785	0.00906
2007	426,188	479	0.00112
2008	373,234	3,962	0.01062
<b>Average</b>	<b>342,982</b>	<b>1,077</b>	<b>0.00343</b>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

## 9.8 ESA/HCP Compliance

### Broodstock Collection

Summer Chinook adults collected at Wells Dam are used primarily for the Methow supplementation programs. On an as needed basis, adults collected at Wells Dam may be used to augment adult collections for the Okanogan summer Chinook supplementation program. Per the 2012 broodstock collection protocol, 108 natural-origin (adipose fin present) adults were targeted for collection between 1 July and 15 September at the West Ladder of Wells Dam. Actual collections occurred between 2 July and 10 September and totaled 128 summer Chinook (including 20 for the Okanogan program that were subsequently released when they were identified as being in excess of program needs). ESA Permit 1347 provides authorization to collect Methow and Okanogan summer Chinook at Wells Dam three days per week and up to 16 hours per day from July through November. During 2012, broodstock collection activities were accomplished within the allowable trapping days authorized under ESA Permit 1347.

Collection of Methow and Okanogan summer Chinook broodstock at Wells Dam occurred concurrently with collection of summer steelhead for the Wells steelhead program authorized under ESA Section 10 Permit 1395. Encounters with steelhead and spring Chinook during Methow and Okanogan summer Chinook broodstock collections did not result in takes that were outside those authorized in Permit 1347 and in Permit 1395 for the Wells Steelhead program. Steelhead encountered during summer Chinook collections that were not required for steelhead

broodstock were passed at the trap site and were not physically handled. Any spring Chinook encountered during summer Chinook broodstock activities were also passed without handling.

### **Hatchery Rearing and Release**

The 2012 brood Methow/Okanogan summer Chinook reared throughout their juvenile life-stages at Eastbank Fish Hatchery and the Carlton Acclimation Pond without incident (see Section 9.2). The 2012 brood smolt release totaled 197,391 summer Chinook, representing 98.7% of the 200,000 production objective and was compliant with the 10% overage allowable in ESA Section 10 Permit 1347.

### **Hatchery Effluent Monitoring**

Per ESA Permits 1196, 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January 2014 through 31 December 2014. NPDES monitoring and reporting for PUD Hatchery Programs during 2014 are provided in Appendix F.

### **Spawning Surveys**

Summer Chinook spawning ground surveys conducted in the Methow River basin during 2014 were consistent with ESA Section 10 Permit No. 1347. Because of the difficulty of quantifying the level of take associated with spawning ground surveys, the Permit does not specify a take level associated with these activities, even though it does authorize implementation of spawning ground surveys. Therefore, no take levels are reported. However, to minimize potential effects to established redds, wading was restricted to the extent practical, and extreme caution was used to avoid established redds when wading was required.



## SECTION 10: OKANOGAN/SIMILKAMEEN SUMMER CHINOOK

The goal of summer Chinook salmon supplementation in the Okanogan Basin is to use artificial production to replace adult production lost because of mortality at Wells, Rocky Reach, and Rock Island dams, while not reducing the natural production or long-term fitness of summer Chinook in the basin. The Rock Island Fish Hatchery Complex began operation in 1989 under funding from Chelan PUD. The Complex operated originally through the Rock Island Settlement Agreement, but since 2004 has operated under the Anadromous Fish Agreement and Habitat Conservation Plans.

Before 2012, adult summer Chinook were collected for broodstock from the run-at-large at the east ladder trapping facility at Wells Dam. Since then, the Colville Tribes collect broodstock using purse seines in the Okanogan and Columbia rivers. The goal was to collect up to 334 adult summer Chinook for the Okanogan program. Broodstock collection occurred from about 7 July through 15 September with trapping occurring no more than 16 hours per day, three days a week. If natural-origin broodstock collection fell short of expectation, hatchery-origin adults could be collected to make up the difference.

Before 2012, adult summer Chinook were spawned and reared at Eastbank Fish Hatchery. Juvenile summer Chinook were transferred from the hatchery to Similkameen Acclimation Pond in October. In addition, since 2005, about 20% (100,000) of the juveniles were transferred to Bonaparte Pond. Chinook were released from the ponds in April to early May.

Prior to 2012, the production goal for the Okanogan summer Chinook supplementation program was to release 576,000 yearling smolts into the Similkameen and Okanogan rivers at ten fish per pound. Beginning with the 2012 brood, the revised production goal is to release 166,569 yearling smolts into the rivers. Targets for fork length and weight are 176 mm (CV = 9.0) and 45.4 g, respectively. Over 90% of these fish are marked with CWTs. In addition, since 2009, juvenile summer Chinook have been PIT tagged annually.

The Colville Tribes began monitoring the Okanogan/Similkameen summer Chinook program in 2013. Their monitoring results will be published in annual reports to Bonneville Power Administration (BPA). The purpose of retaining this section is to provide readers with monitoring data collected with Chelan PUD funding through brood year 2012. Thus, this section tracks the status and life histories of summer Chinook up to and including brood year 2012. Results from monitoring brood year 2013 and beyond will be included in annual reports to BPA.

### 10.1 Broodstock Sampling

Summer Chinook broodstock for the Okanogan/Similkameen and Methow programs was typically collected at the East and West Ladders of Wells Dam. In 2012, broodstock was also collected at the mouth of the Okanogan River via purse seine. In 2012, a total of 81 summer Chinook (79 wild Chinook and two hatchery Chinook)<sup>13</sup> were spawned for the Okanogan

---

<sup>13</sup> It is important to point out that some summer Chinook were used for both the Methow and Okanogan programs in 2012 because of the availability of ripe adults at the time of spawning. In addition, some eyed-eggs were split between the two programs

program. Refer to Section 9.1 for information on the origin, age and length, sex ratios, and fecundity of summer Chinook broodstock collected at Wells Dam prior to 2013.

## 10.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Based on the unfertilized egg-to-release survival standard of 81%, a total of 711,111 eggs were required to meet the program release goal of 576,000 smolts through the 2011 brood year. An evaluation of the program in 2012 determined that 205,134 eggs were needed to meet the revised release goal of 166,569 smolts. This revised goal began with brood year 2012. From 1989 through 2012, the egg take goal was reached in 13 of those years (Table 10.1).

**Table 10.1.** Numbers of eggs taken from summer Chinook broodstock for the Okanogan program during 1989-2012. From 1989-2011, broodstock were collected at Wells Dam. In 2012, broodstock were collected in purse seines in the Okanogan River.

Return year	Number of eggs taken
1989	724,200
1990	696,144
1991	879,892
1992	729,389
1993	797,234
1994	893,086
1995	736,500
1996	672,000
1997	601,744
1998	584,018
1999	725,589
2000	645,403
2001	418,907
2002	718,599
2003	710,521
2004	805,814
2005	452,928
2006	757,350
2007	824,703
2008	662,668
2009	840,902
2010	726,979
2011	683,419
<i>Average (1989-2011)</i>	<i>708,173</i>
2012	201,295

Return year	Number of eggs taken
Average (2012-present)	201,295

### Number of acclimation days

Summer Chinook were released volitionally from Similkameen Pond as yearling smolts. Transfer dates, release dates, and the number of acclimation days for Okanogan summer Chinook are shown in Table 10.2.

**Table 10.2.** Number of days Okanogan summer Chinook broods were acclimated at Similkameen and Bonaparte ponds, brood years 1989-2012.

Brood year	Release year	Rearing facility	Transfer date	Release date	Number of days
1989	1991	Similkameen	29-Oct	7-May	190
1990	1992	Similkameen	5-Nov	25-Apr	171
1991	1993	Similkameen	1-Nov	9-Apr	159
1992	1994	Similkameen	2-Nov	1-Apr	150
			26-Feb	1-Apr	34
1993	1995	Similkameen	24-Oct	1-Apr	159
			24-Feb	1-Apr	36
1994	1996	Similkameen	30-Oct	6-Apr	158
			14-Mar	6-Apr	23
1995	1997	Similkameen	1-Oct	1-Apr	182
1996	1998	Similkameen	10-Oct	15-Mar	156
1997	1999	Similkameen	7-Oct	19-Apr	194
1998	2000	Similkameen	5-Oct	19-Apr	196
1999	2001	Similkameen	5-Oct	18-Apr	195
2000	2002	Similkameen	10-Oct	8-Apr	180
2001	2003	Similkameen	1-Oct	29-Apr	210
2002	2004	Similkameen	9-Nov	23-Apr	165
2003	2005	Similkameen	19-Oct	28-Apr	191
2004	2006	Similkameen	26-Oct	23-Apr	179
2005	2007	Bonaparte	6-Nov	11-Apr	156
		Similkameen	25-Oct	18-Apr – 9-May	179-200
2006	2008	Similkameen	15-17-Oct	16-Apr – 7-May	182-205
2007	2009	Bonaparte	3-4-Nov	10-22-Apr	157-170
		Similkameen	20-24-Oct	14-Apr – 9-May	172-201
2008	2010	Bonaparte	2-4-Nov	19-Apr – 5-May	167-185

Brood year	Release year	Rearing facility	Transfer date	Release date	Number of days
		Similkameen	26-28-Oct	19-Apr – 14-May	176-201
2009	2011	Bonaparte	8-9-Nov	12-Apr	155-156
		Similkameen	25-27-Oct	13-Apr – 5-May	169-193
2010	2012	Bonaparte	No program	No program	No program
		Similkameen	25-27 Oct	16-Apr – 7-May	173-196
2011	2013	Bonaparte	No program	No program	No program
		Similkameen	23-26 Oct	16-Apr – 8-May	175-197
2012	2014	Bonaparte	No program	No program	No program
		Similkameen	28-30 Oct	15 Apr – 5 May	167-189

## Release Information

### Numbers released

The 2012 Okanogan summer Chinook program achieved 68.4% of the 166,569 target goal with about 114,000 fish being released volitionally into the Similkameen River (Table 10.3).

**Table 10.3.** Numbers of Okanogan summer Chinook smolts released from the Similkameen and Bonaparte ponds, brood years 1989-2012; NA = not available. For brood years 1998-2012, the release target was 576,000 smolts. Since brood year 2013, the release target for Okanogan summer Chinook is 114,000 smolts.

Brood year	Release year	Rearing facility	CWT mark rate	Number of smolts released
1989	1991	Similkameen	0.5732	352,600
1990	1992	Similkameen	0.6800	540,000
1991	1993	Similkameen	0.5335	675,500
1992	1994	Similkameen	0.9819	548,182
1993	1995	Similkameen	0.6470	586,000
1994	1996	Similkameen	0.4176	536,299
1995	1997	Similkameen	0.9785	587,000
1996	1998	Similkameen	0.9769	507,913
1997	1999	Similkameen	0.9711	589,591
1998	2000	Similkameen	0.9825	293,191
1999	2001	Similkameen	0.9689	630,463
2000	2002	Similkameen	0.9928	532,453
2001	2003	Similkameen	0.9877	26,642
2002	2004	Similkameen	0.9204	388,589
2003	2005	Similkameen	0.9929	579,019
2004	2006	Similkameen	0.9425	703,359
2005	2007	Bonaparte	0	0 (assumed)

Brood year	Release year	Rearing facility	CWT mark rate	Number of smolts released
		Similkameen	0.9862	275,919
2006	2008	Similkameen	0.9878	604,035
2007	2009	Bonaparte	0.9920	102,099
		Similkameen	0.9914	513,039
2008	2010	Bonaparte	0.9947	175,729
		Similkameen	0.9947	343,628
2009	2011	Bonaparte	0.9981	151,382
		Similkameen	0.9953	524,521
2010	2012	Similkameen	0.9886	617,950
2011	2013	Similkameen	0.9956	627,978
<i>Average (1989-2011)</i>		<i>Bonaparte</i>	<i>0.7462</i>	<i>143,070</i>
		<i>Similkameen</i>	<i>0.8907</i>	<i>503,647</i>
2012	2014	Bonaparte	No program	No program
		Similkameen	0.9939	114,000
<i>Average (2012-present)</i>		<i>Bonaparte</i>	<i>No program</i>	<i>No program</i>
		<i>Similkameen</i>	<i>0.9939</i>	<i>114,000</i>

**Numbers tagged**

The 2012 brood Okanogan summer Chinook from the Similkameen facility were 99.4% CWT and adipose fin-clipped (Table 10.3). Table 10.4 summarizes the number of hatchery summer Chinook that have been PIT-tagged and released into the Okanogan River basin. No fish from the 2012 brood year were PIT tagged.

**Table 10.4.** Summary of PIT-tagging activities for Okanogan hatchery summer Chinook, brood years 2008-2011.

Brood year	Release year	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2008	2010	5,700 (high density)	1,169	0	4,531
		5,700 (low density)	1,407	0	4,293
2009	2011	5,100	11	0	5,089
2010	2012	0	0	0	0
2011	2013	5,100	64	0	5,036

**Fish size and condition at release**

Size at release of the Similkameen population was 73.3% and 56.8% of the fork length and weight targets, respectively. The CV for fork length exceeded the target by 18.9% (Table 10.5). There was no Bonaparte program for the 2014 release year.

**Table 10.5.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of Okanogan summer Chinook smolts released from the hatchery, brood years 1989-2012. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1989	1991	-	-	41.3	11
1990	1992	143	9.5	37.8	12
1991	1993	125	15.5	22.4	20
1992	1994	120	15.4	20.7	22
1993	1995	132	-	23.2	20
1994	1996	136	16.0	29.6	15
1995	1997	137	8.2	32.8	14
1996	1998	127	12.8	26.2	17
1997	1999	144	9.9	36.0	13
1998	2000	148	5.9	41.0	11
1999	2001	141	15.7	35.4	13
2000	2002	121	13.4	20.4	22
2001	2003	132	8.2	25.7	18
2002	2004	119	13.4	20.8	22
2003	2005	133	10.6	28.9	16
2004	2006	132	9.9	29.8	15
2005	2007	132	9.6	25.9	18
2006	2008	120	12.3	20.9	22
2007	2009	124	12.6	21.9	21
2008	2010	140	12.3	35.1	13
2009	2011	132	11.6	24.7	18
2010	2012	125	10.1	23.2	20
2011	2013	132	9.5	27.9	16
2012	2014	129	7.3	25.8	18
<b>Average</b>		<b>131</b>	<b>11.4</b>	<b>28.2</b>	<b>17</b>
<b>Targets</b>		<b>176</b>	<b>9.0</b>	<b>45.4</b>	<b>10</b>

### Survival Estimates

Overall survival of Okanogan summer Chinook from green (unfertilized) egg to release was above the standard set for the program (Table 10.6). Low survival can be attributed to high mortality after ponding through release because of external fungus. Currently, it is unknown if gamete viability is sex biased or is uniform between sexes and more influenced by between-year environmental variations.

**Table 10.6.** Hatchery life-stage survival rates (%) for Okanogan summer Chinook, brood years 1989-2012. Survival standards or targets are provided in the last row of the table.

Brood year	Rearing facility	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
		Female	Male							
1989 <sup>a</sup>	Similkameen	89.8	99.5	89.9	96.7	99.7	99.4	73.3	57.4	48.7
1990 <sup>a</sup>	Similkameen	93.9	99.0	84.9	97.1	81.2	80.6	97.7	98.6	77.6
1991 <sup>a</sup>	Similkameen	93.1	95.5	88.2	97.1	99.4	99.1	98.4	97.1	76.8
1992 <sup>a</sup>	Similkameen	96.9	99.0	87.0	98.0	99.9	99.9	91.7	92.6	75.2
1993 <sup>a</sup>	Similkameen	82.2	99.4	85.4	97.6	99.8	99.5	92.0	90.2	73.5
1994	Similkameen	96.1	90.0	86.6	100.0	98.1	97.4	73.1	89.8	60.1
1995	Similkameen	91.9	96.2	98.2	84.1	96.5	96.2	92.7	98.2	79.7
1996	Similkameen	95.4	98.1	83.2	100.0	97.7	96.9	86.5	92.5	75.6
1997	Similkameen	91.9	94.6	86.1	98.4	98.7	98.3	98.8	99.4	98.0
1998	Similkameen	84.0	96.2	54.1	98.0	99.4	98.9	96.6	99.6	50.2
1999	Similkameen	98.8	98.7	92.9	96.9	98.0	97.6	96.9	99.0	86.9
2000	Similkameen	90.5	96.9	89.2	98.5	98.2	98.0	93.6	97.2	82.5
2001	Similkameen	96.2	92.3	89.1	97.6	99.7	99.5	7.4	11.9	6.4
2002	Similkameen	97.1	98.1	89.8	98.0	99.7	99.5	51.6	52.2	54.1
2003	Similkameen	96.7	97.5	86.8	97.6	99.3	98.5	98.0	98.8	81.5
2004	Similkameen	93.6	98.2	84.0	97.6	99.6	99.3	97.8	98.8	80.2
	Bonaparte	93.6	98.2	84.0	97.6	99.6	99.3	97.9	98.9	80.3
2005	Similkameen	97.0	89.6	88.0	99.5	99.5	99.0	93.5	94.6	81.8
	Bonaparte	97.0	89.6	88.0	99.5	99.5	99.0	0.0	0.0	0.0
2006	Similkameen	92.9	89.5	86.3	98.3	99.6	99.3	94.1	95.5	79.8
2007	Similkameen	92.6	99.6	80.8	99.1	99.5	99.1	97.0	98.1	77.7
	Bonaparte	92.6	99.6	80.8	99.1	99.5	99.1	95.6	96.7	76.6
2008	Similkameen	97.9	99.6	91.2	96.8	99.7	99.3	89.8	90.5	79.3
	Bonaparte	97.9	99.6	91.2	96.8	99.7	99.3	86.9	87.8	76.7
2009 <sup>b</sup>	Similkameen	93.6	93.5	91.0	98.2	99.7	99.5	97.8	98.6	87.4
	Bonaparte	93.6	93.5	91.0	98.2	99.7	99.5	74.8	75.3	66.8
2010	Similkameen	96.5	100.0	91.2	99.9	97.4	97.1	93.3	96.3	85.0
2011	Similkameen	100.0	90.2	95.9	98.3	99.8	99.1	97.8	98.8	92.2
2012	Similkameen	100.0	100.0	85.1	98.6	99.7	99.3	70.6	71.2	59.3
Mean	Similkameen	94.1	96.3	86.9	97.6	98.3	97.9	86.7	88.2	72.9
	Bonaparte	94.9	96.1	87.0	98.2	99.6	99.2	71.0	71.7	60.1
Standard		90.0	85.0	92.0	98.0	97.0	93.0	90.0	95.0	81.0

<sup>a</sup> Survival rates were calculated from the aggregate population collected at Wells Fish Hatchery volunteer channel and left- and right-ladder traps at Wells Dam.

<sup>b</sup> Survival rates were calculated from aggregate collections at Wells east fish ladder for the Methow and Okanogan/Similkameen programs. About 59% of the total fish collected were used to estimate survival rates.

### 10.3 Disease Monitoring

Rearing of the 2012 brood Okanogan summer Chinook was similar to previous years with fish being held on well water before being transferred for final acclimation on the Similkameen. The Similkameen group was transferred in late October. Fish acclimating at the Similkameen facility experienced mortality because of cold water disease and external fungus from October through December. No additional disease-related problems were noted before the fish were released.

Results of adult broodstock bacterial kidney disease (BKD) monitoring for Methow/Okanogan summer Chinook are shown in Table 9.12 in Section 9.3.

### 10.4 Spawning Surveys

Surveys for Okanogan/Similkameen summer Chinook redds were conducted from late September to mid-November in the Okanogan and Similkameen rivers. Total redd counts (not peak counts) were conducted in the rivers.

#### Redd Counts

During the survey period 1989 through 2014, the number of summer Chinook redds in the Okanogan River basin averaged 1,994 and ranged from 110 to 6,025 (Table 10.7).

**Table 10.7.** Total number of redds counted in the Okanogan River basin, 1989-2014. The Colville Tribes provided data for survey years 2013 to present.

Survey year	Number of summer Chinook redds		
	Okanogan River	Similkameen River	Total count
1989	151	370	521
1990	99	147	246
1991	64	91	155
1992	53	57	110
1993	162	288	450
1994	375*	777	1,152
1995	267*	616	883
1996	116	419	535
1997	158	486	644
1998	88	276	364
1999	369	1,275	1,644
2000	549	993	1,542
2001	1,108	1,540	2,648
2002	2,667	3,358	6,025
2003	1,035	378	1,413
2004	1,327	1,660	2,987
2005	1,611	1,423	3,034
2006	2,592	1,666	4,258
2007	1,301	707	2,008
2008	1,146	1,000	2,146



Survey year	Number of summer Chinook redds		
	Okanogan River	Similkameen River	Total count
2009	1,672	1,298	2,970
2010	1,011	1,107	2,118
2011	1,714	1,409	3,123
2012	1,613	1,066	2,679
2013	2,267	1,280	3,547
2014	2,231	2,022	4,253
<i>Average</i>	<i>990</i>	<i>989</i>	<i>1,979</i>

\* Reach-expanded aerial counts.

### Spawning Escapement

Spawning escapement for Okanogan/Similkameen summer Chinook was calculated as the total number of redds times the fish per redd ratio estimated from fish sampled at Wells Dam. During the survey period 1989 through 2014, the summer Chinook spawning escapement within the Okanogan River basin averaged 5,396 and ranged from 473 to 13,857 (Table 10.8).

**Table 10.8.** Spawning escapements for summer Chinook in the Okanogan and Similkameen rivers for return years 1989-2014. The Colville Tribes provided data for return years 2013 to present.

Return year	Fish/Redd	Spawning escapement		
		Okanogan	Similkameen	Total
1989*	3.30	498	1,221	1,719
1990*	3.40	337	500	837
1991*	3.70	237	337	574
1992*	4.30	228	245	473
1993*	3.30	535	950	1,485
1994*	3.50	1,313	2,720	4,033
1995*	3.40	908	2,094	3,002
1996*	3.40	394	1,425	1,819
1997*	3.40	537	1,652	2,189
1998	3.00	264	828	1,092
1999	2.20	812	2,805	3,617
2000	2.40	1,318	2,383	3,701
2001	4.10	4,543	6,314	10,857
2002	2.30	6,134	7,723	13,857
2003	2.42	2,505	915	3,420
2004	2.25	2,986	3,735	6,721
2005	2.93	4,720	4,169	8,889
2006	2.02	5,236	3,365	8,601
2007	2.20	2,862	1,555	4,417
2008	3.25	3,725	3,250	6,975

Return year	Fish/Redd	Spawning escapement		
		Okanogan	Similkameen	Total
2009	2.54	4,247	3,297	7,544
2010	2.81	2,841	3,111	5,952
2011	3.10	5,313	4,368	9,681
2012	3.07	4,952	3,273	8,225
2013	2.14	4,851	2,739	7,591
2014	2.86	6,381	5,783	12,164
<i>Average</i>	<i>2.97</i>	<i>2,641</i>	<i>2,721</i>	<i>5,363</i>

\* Spawning escapement was calculated using the "Modified Meekin Method" (i.e., 3.1 x jack multiplier).

## 10.5 Carcass Surveys

Surveys for summer Chinook carcasses were conducted during late September to mid-November in the Okanogan and Similkameen rivers.

### Number sampled

During the survey period 1993 through 2012, the number of summer Chinook carcasses sampled in the Okanogan River basin averaged 1,205 and ranged from 115 to 2,460 (Table 10.9). In all years, most were sampled in the upper Okanogan River and lower Similkameen River (Table 10.9).

**Table 10.9.** Numbers of summer Chinook carcasses sampled within each survey reach in the Okanogan River basin, 1993-2012. Reach codes are described in Table 2.11. The Colville Tribes provided data for survey years 2013 to present.

Survey year	Number of summer Chinook carcasses								
	Okanogan						Similkameen		Total
	O-1	O-2	O-3	O-4	O-5	O-6	S-1	S-2	
1993 <sup>a</sup>	0	2	3	0	23	13	73	1	115
1994 <sup>b</sup>	0	4	4	0	27	5	318	60	418
1995	0	0	2	0	30	0	239	15	286
1996	0	0	0	2	5	2	226	0	235
1997	0	0	2	0	9	3	225	1	240
1998	0	1	8	1	7	7	340	4	368
1999	0	0	3	2	23	53	766	48	895
2000	0	2	20	15	47	16	727	41	868
2001	0	26	75	10	127	112	1,141	105	1,596
2002	10	32	83	35	204	572	1,265	259	2,460
2003 <sup>c</sup>	0	0	28	0	17	243	596	381	1,265
2004	0	4	31	24	146	283	1,392	298	2,178
2005	0	8	93	37	371	434	731	276	1,950
2006	4	3	31	16	120	291	508	106	1,079

Survey year	Number of summer Chinook carcasses								
	Okanogan						Similkameen		Total
	O-1	O-2	O-3	O-4	O-5	O-6	S-1	S-2	
2007	2	0	55	1	453	519	658	29	1,717
2008	4	10	40	36	248	665	859	157	2,019
2009	2	7	31	32	348	500	703	150	1,773
2010	3	10	30	42	241	352	627	148	1,453
2011	0	0	55	14	361	478	753	114	1,775
2012	1	0	56	15	256	537	495	54	1,414
2013 <sup>d</sup>	0	0	39		52	432	387		910
2014	1	1	79	54	275	783	770	489	2,452
<b>Average</b>	<b>1</b>	<b>5</b>	<b>35</b>	<b>16</b>	<b>154</b>	<b>286</b>	<b>639</b>	<b>130</b>	<b>1,248</b>

<sup>a</sup> 25 additional carcasses were sampled on the Similkameen and 46 on the Okanogan without any reach designation.

<sup>b</sup> One additional carcasses was sampled on the Similkameen without any reach designation.

<sup>c</sup> 793 carcasses were sampled on the Similkameen before initiation of spawning (pre-spawn mortality) and an additional 40 carcasses were sampled on the Okanogan. The cause of the high mortality (*Ichthyophthirius multifiliis* and *Flavobacterium columnarae*) was exacerbated by high river temperatures.

<sup>d</sup> In 2013, the Colville Tribes combined survey reaches O-3 and O-4, and S-1 and S-2.

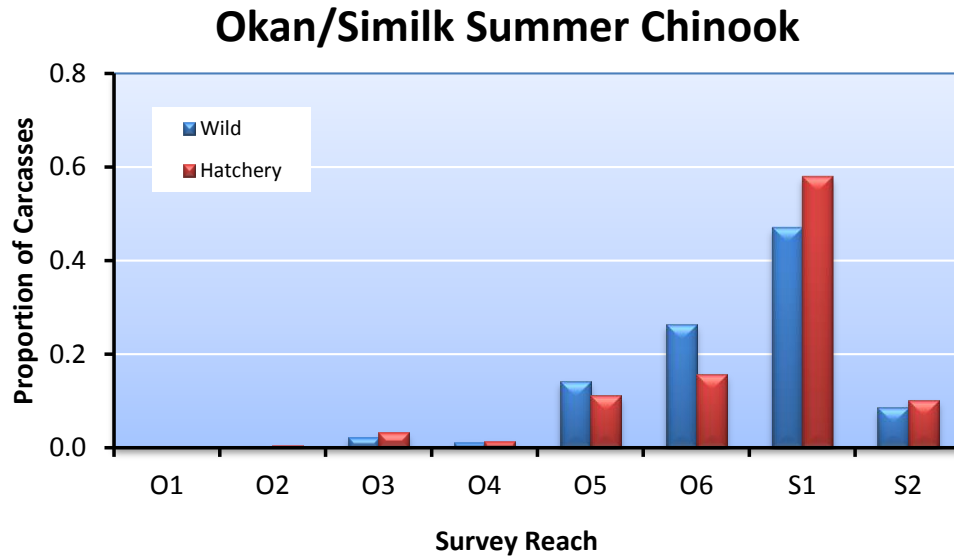
### Carcass Distribution and Origin

Based on the available data (1991-2012), most fish, regardless of origin, were found in Reach 1 on the Similkameen River (Driscoll Channel to Oroville Bridge) (Table 10.10). However, a slightly larger percentage of hatchery fish were found in reaches on the Similkameen River than were wild fish (Figure 10.1). In contrast, a larger percentage of wild fish were found in reaches on the Okanogan River.

**Table 10.10.** Numbers of wild and hatchery summer Chinook carcasses sampled within different reaches in the Okanogan River basin, 1993-2012.

Survey year	Origin	Survey reach								Total
		O-1	O-2	O-3	O-4	O-5	O-6	S-1	S-2	
1993	Wild	0	0	3	0	13	4	48	1	69
	Hatchery	0	2	0	0	10	9	25	0	46
1994	Wild	0	0	1	0	7	1	113	22	144
	Hatchery	0	4	3	0	20	4	205	38	274
1995	Wild	0	0	1	0	10	0	66	4	81
	Hatchery	0	0	1	0	20	0	173	11	205
1996	Wild	0	0	0	1	3	1	53	0	58
	Hatchery	0	0	0	1	2	1	173	0	177
1997	Wild	0	0	1	0	0	3	83	0	87
	Hatchery	0	0	1	0	9	0	142	1	153
1998	Wild	0	1	3	1	6	5	162	4	182
	Hatchery	0	0	5	0	1	2	178	0	186
1999	Wild	0	0	0	0	9	23	293	9	334
	Hatchery	0	0	3	2	14	30	473	39	561

Survey year	Origin	Survey reach								Total
		O-1	O-2	O-3	O-4	O-5	O-6	S-1	S-2	
2000	Wild	0	0	8	8	24	11	189	4	244
	Hatchery	0	2	12	7	23	5	538	37	624
2001	Wild	0	10	23	5	67	42	390	54	591
	Hatchery	0	16	52	5	60	70	751	51	1,005
2002	Wild	6	14	20	10	81	212	340	72	755
	Hatchery	4	18	63	25	123	360	925	187	1,705
2003	Wild	0	0	13	0	12	152	231	124	532
	Hatchery	0	0	15	0	5	91	365	257	733
2004	Wild	0	2	19	19	108	225	1,125	260	1,758
	Hatchery	0	2	12	5	38	58	267	38	420
2005	Wild	0	5	51	21	256	364	531	176	1,404
	Hatchery	0	3	42	16	115	70	200	100	546
2006	Wild	2	2	22	10	105	247	370	73	831
	Hatchery	2	1	9	6	15	44	138	33	248
2007	Wild	1	0	30	1	284	322	405	20	1,063
	Hatchery	1	0	25	0	169	197	253	9	654
2008	Wild	2	1	14	11	107	324	347	41	847
	Hatchery	2	9	26	25	141	341	512	116	1,172
2009	Wild	2	3	13	14	189	347	330	75	973
	Hatchery	0	4	18	18	159	153	373	75	800
2010	Wild	1	5	19	18	154	180	329	69	775
	Hatchery	2	5	11	24	87	172	296	79	676
2011	Wild	0	0	21	4	201	362	216	19	823
	Hatchery	0	0	34	10	160	116	537	95	952
2012	Wild	0	0	18	9	133	427	206	23	816
	Hatchery	1	0	38	6	123	110	288	31	597
<i>Average</i>	<i>Wild</i>	<i>1</i>	<i>2</i>	<i>14</i>	<i>7</i>	<i>88</i>	<i>163</i>	<i>291</i>	<i>53</i>	<i>618</i>
	<i>Hatchery</i>	<i>1</i>	<i>3</i>	<i>19</i>	<i>8</i>	<i>65</i>	<i>92</i>	<i>341</i>	<i>60</i>	<i>587</i>



**Figure 10.1.** Distribution of wild and hatchery produced carcasses in different reaches in the Okanogan River basin, 1993-2012. Reach codes are described in Table 2.11.

### 10.6 Life History Monitoring

Life history characteristics of Okanogan/Similkameen summer Chinook were assessed by examining carcasses on spawning grounds and fish collected or examined at broodstock collection sites, and by reviewing tagging data and fisheries statistics.

#### Migration Timing

Migration timing for Okanogan/Similkameen summer Chinook is described in Section 9.6.

#### Age at Maturity

Because hatchery summer Chinook are released after one year of rearing and natural-origin summer Chinook migrate primarily as age-0 fish, total ages will differ between hatchery and natural-origin Chinook (see Hillman et al. 2011). Therefore, in this section, we evaluated age at maturity by comparing differences in salt (ocean) ages between the two groups.

Most of the wild and hatchery summer Chinook sampled during the period 1993-2012 in the Okanogan River basin were salt age-3 fish (Table 10.11; Figure 10.2). A higher percentage of salt age-4 wild Chinook returned to the basin than did salt age-4 hatchery Chinook. In contrast, a higher proportion of salt age-1 and 2 hatchery fish returned than did salt age-1 and 2 wild fish. Thus, a higher percentage of wild fish returned at an older age than did hatchery fish.

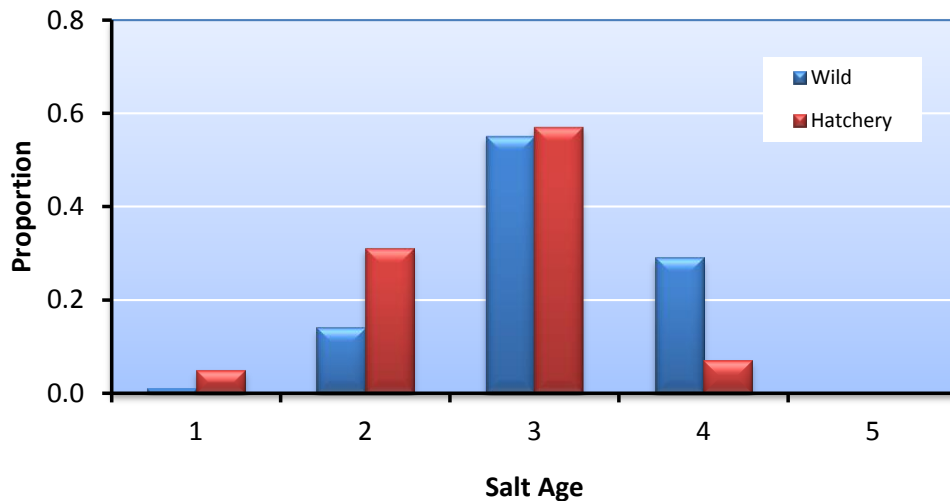
**Table 10.11.** Proportions of wild and hatchery summer Chinook of different salt (ocean) ages sampled on spawning grounds in the Okanogan River basin, 1993-2012.

Sample year	Origin	Salt age					Sample size
		1	2	3	4	5	
1993	Wild	0.00	0.21	0.70	0.10	0.00	63
	Hatchery	0.00	0.98	0.02	0.00	0.00	44
1994	Wild	0.02	0.13	0.54	0.31	0.00	134
	Hatchery	0.02	0.09	0.89	0.00	0.00	290
1995	Wild	0.00	0.19	0.59	0.22	0.00	68
	Hatchery	0.01	0.15	0.36	0.49	0.00	200
1996	Wild	0.03	0.28	0.61	0.08	0.00	36
	Hatchery	0.02	0.22	0.56	0.20	0.01	174
1997	Wild	0.04	0.27	0.53	0.15	0.00	73
	Hatchery	0.00	0.02	0.87	0.11	0.00	148
1998	Wild	0.02	0.35	0.52	0.11	0.00	151
	Hatchery	0.05	0.50	0.23	0.22	0.00	185
1999	Wild	0.00	0.20	0.64	0.16	0.00	268
	Hatchery	0.00	0.12	0.85	0.02	0.00	552
2000	Wild	0.03	0.15	0.62	0.20	0.00	216
	Hatchery	0.12	0.02	0.76	0.10	0.00	545
2001	Wild	0.02	0.18	0.76	0.04	0.00	531
	Hatchery	0.05	0.88	0.02	0.05	0.00	1,005
2002	Wild	0.02	0.15	0.62	0.21	0.00	692
	Hatchery	0.01	0.19	0.80	0.01	0.00	1,681
2003	Wild	0.03	0.18	0.63	0.17	0.00	477
	Hatchery	0.03	0.06	0.79	0.12	0.00	653
2004	Wild	0.01	0.17	0.26	0.55	0.00	1,528
	Hatchery	0.01	0.32	0.45	0.23	0.00	382
2005	Wild	0.00	0.12	0.79	0.08	0.01	1,281
	Hatchery	0.02	0.06	0.77	0.15	0.00	530
2006	Wild	0.00	0.02	0.53	0.45	0.00	830
	Hatchery	0.05	0.18	0.24	0.53	0.00	139
2007	Wild	0.02	0.07	0.12	0.78	0.02	1,061
	Hatchery	0.22	0.30	0.42	0.05	0.01	559
2008	Wild	0.01	0.32	0.63	0.04	0.01	846
	Hatchery	0.02	0.60	0.36	0.02	0.00	1,108
2009	Wild	0.01	0.03	0.81	0.15	0.00	926
	Hatchery	0.05	0.05	0.86	0.03	0.00	783
2010	Wild	0.00	0.16	0.45	0.39	0.00	708
	Hatchery	0.02	0.65	0.27	0.06	0.00	619

Sample year	Origin	Salt age					Sample size
		1	2	3	4	5	
2011	Wild	0.01	0.07	0.82	0.10	0.00	787
	Hatchery <sup>a</sup>	0.16	0.08	0.76	0.00	0.00	873
2012	Wild	0.02	0.23	0.41	0.34	0.00	750
	Hatchery	0.05	0.55	0.35	0.05	0.00	532
Average	Wild	0.01	0.14	0.55	0.29	0.00	571
	Hatchery	0.05	0.31	0.57	0.07	0.00	550

<sup>a</sup> There was one salt age-6 hatchery fish that was not included in this table.

### Okan/Similk Summer Chinook



**Figure 10.2.** Proportions of wild and hatchery summer Chinook of different salt (ocean) ages sampled at broodstock collection sites and on spawning grounds in the Okanogan River basin for the combined years 1993-2012.

#### Size at Maturity

For the period 1993 through 2012, on average, hatchery summer Chinook were about 2 cm smaller than wild summer Chinook sampled in the Okanogan River basin (Table 10.12). This is likely because a higher percentage of wild fish returned as salt age-4 fish than did hatchery fish.

**Table 10.12.** Mean lengths (POH; cm) and variability statistics for wild and hatchery summer Chinook sampled in the Okanogan River basin, 1993-2012; SD = 1 standard deviation.

Sample year	Origin	Sample size	Summer Chinook length (POH; cm)			
			Mean	SD	Minimum	Maximum
1993 <sup>a</sup>	Wild	69	73	7	52	90
	Hatchery	59	62	6	47	75
1994	Wild	136	71	7	40	86
	Hatchery	268	69	8	30	84

Sample year	Origin	Sample size	Summer Chinook length (POH; cm)			
			Mean	SD	Minimum	Maximum
1995	Wild	81	75	6	54	87
	Hatchery	201	73	8	39	87
1996	Wild	22	68	14	22	85
	Hatchery	26	75	8	60	88
1997	Wild	87	70	7	44	84
	Hatchery	148	74	6	48	88
1998	Wild	182	70	8	45	94
	Hatchery	186	65	12	30	87
1999	Wild	333	73	7	56	91
	Hatchery	559	71	7	23	84
2000	Wild	241	70	10	32	86
	Hatchery	624	69	12	24	92
2001	Wild	578	67	9	26	86
	Hatchery	997	61	8	32	90
2002	Wild	755	69	9	28	91
	Hatchery	1705	70	8	33	87
2003	Wild	532	68	9	30	93
	Hatchery	733	69	10	26	90
2004	Wild	1756	71	10	33	94
	Hatchery	417	66	9	41	92
2005	Wild	1403	66	7	41	99
	Hatchery	546	68	8	31	85
2006	Wild	831	72	6	31	91
	Hatchery	248	71	9	33	87
2007	Wild	1063	75	9	27	99
	Hatchery	654	64	13	30	87
2008	Wild	847	65	9	29	86
	Hatchery	1172	65	8	32	89
2009	Wild	973	70	7	28	89
	Hatchery	799	70	9	35	86
2010	Wild	775	71	9	43	90
	Hatchery	676	64	10	22	87
2011	Wild	823	68	7	29	89
	Hatchery	952	66	11	26	86
2012	Wild	816	67	10	27	93
	Hatchery	597	63	9	23	86
<b>Pooled</b>	<b>Wild</b>	<b>12,303</b>	<b>70</b>	<b>8</b>	<b>22</b>	<b>99</b>
	<b>Hatchery</b>	<b>11,567</b>	<b>68</b>	<b>9</b>	<b>22</b>	<b>92</b>



<sup>a</sup> This year includes sizes reported in the annual report. The data contained in the WDFW database do not include all these data.

### Contribution to Fisheries

Most of the harvest on hatchery-origin Okanogan/Similkameen summer Chinook occurred in the Ocean (Table 10.13). Ocean harvest has made up 37-100% of all hatchery-origin Okanogan/Similkameen summer Chinook harvested. Brood years 1997, 1998, 2000, 2004, 2006, and 2008 provided the largest harvests, while brood years 1993 and 1996 provided the lowest.

**Table 10.13.** Estimated number and percent (in parentheses) of hatchery-origin Okanogan/Similkameen summer Chinook captured in different fisheries, brood years 1989-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
1989	2,371 (80)	553 (19)	0 (0)	42 (1)	2,966
1990	355 (89)	34 (8)	0 (0)	12 (3)	401
1991	220 (86)	37 (14)	0 (0)	0 (0)	257
1992	422 (91)	28 (6)	2 (0)	10 (2)	462
1993	24 (80)	6 (20)	0 (0)	0 (0)	30
1994	374 (92)	23 (6)	2 (0)	7 (2)	406
1995	652 (93)	9 (1)	12 (2)	25 (4)	696
1996	6 (100)	0 (0)	0 (0)	0 (0)	6
1997	6,520 (92)	136 (2)	36 (1)	416 (6)	7,108
1998	4,363 (89)	251 (5)	45 (1)	219 (4)	4,898
1999	1,353 (68)	224 (11)	31 (2)	384 (19)	1,992
2000	3,141 (69)	533 (12)	222 (5)	665 (15)	4,561
2001	184 (58)	81 (25)	31 (10)	23 (7)	319
2002	702 (56)	200 (16)	90 (7)	258 (21)	1,250
2003	697 (37)	568 (31)	130 (7)	466 (25)	1,861
2004	3,091 (38)	2,162 (27)	694 (9)	2,165 (27)	8,112
2005	468 (46)	306 (30)	79 (8)	167 (16)	1,020
2006	3,164 (38)	3,352 (40)	469 (6)	1,419 (17)	8,404
2007	1,551 (45)	951 (27)	67 (2)	910 (26)	3,482
2008	4,527 (43)	1,950 (18)	214 (2)	3,948 (37)	10,639
<i>Average</i>	<i>1,710 (58)</i>	<i>570 (19)</i>	<i>106 (4)</i>	<i>557 (19)</i>	<i>2,944</i>

### Straying

Stray rates were determined by examining CWTs recovered on spawning grounds within and outside the Okanogan River basin. Targets for strays based on return year (recovery year) and brood year should be less than 5%.

Few hatchery-origin Okanogan summer Chinook have strayed into basins outside the Okanogan (Table 10.14). Although hatchery-origin Okanogan summer Chinook have strayed into other

spawning areas, they usually made up less than 5% of the spawning escapement within those areas. The Chelan tailrace has received the largest number of Okanogan strays.

**Table 10.14.** Number and percent of spawning escapements within other non-target basins that consisted of hatchery-origin Okanogan summer Chinook, return years 1994-2011. For example, for return year 2002, 1% of the summer Chinook spawning escapement in the Entiat Basin consisted of hatchery-origin Okanogan summer Chinook. Percent strays should be less than 5%.

Return year	Wenatchee		Methow		Chelan		Entiat		Hanford Reach	
	Number	%	Number	%	Number	%	Number	%	Number	%
1994	0	0.0	0	0.0	-	-	-	-	-	-
1995	0	0.0	0	0.0	-	-	-	-	-	-
1996	0	0.0	0	0.0	-	-	-	-	-	-
1997	0	0.0	0	0.0	-	-	-	-	-	-
1998	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1999	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2000	0	0.0	6	0.5	30	4.5	0	0.0	3	0.0
2001	12	0.1	0	0.0	10	1.0	0	0.0	0	0.0
2002	0	0.0	3	0.1	4	0.7	5	1.0	0	0.0
2003	0	0.0	8	0.2	22	5.3	14	2.0	0	0.0
2004	0	0.0	0	0.0	5	1.2	0	0.0	0	0.0
2005	5	0.1	27	1.1	36	6.9	7	1.9	8	0.0
2006	0	0.0	5	0.2	4	1.0	7	1.2	0	0.0
2007	0	0.0	3	0.2	4	2.1	0	0.0	0	0.0
2008	0	0.0	9	0.5	46	9.3	4	1.3	0	0.0
2009	15	0.2	3	0.2	11	1.8	18	7.2	0	0.0
2010	6	0.1	0	0.0	33	3.0	0	0.0	0	0.0
2011	0	0.0	0	0.0	46	3.6	0	0.0	0	0.0
<b>Average</b>	<b>2</b>	<b>0.0</b>	<b>4</b>	<b>0.2</b>	<b>18</b>	<b>2.9</b>	<b>4</b>	<b>1.0</b>	<b>1</b>	<b>0.0</b>

On average, about 1% of the returns have strayed into non-target spawning areas, falling within the acceptable level of less than 5% (Table 10.15). Depending on brood year, percent strays into non-target spawning areas have ranged from 0-4.4%. Few (<1% on average) have strayed into non-target hatchery programs.

**Table 10.15.** Number and percent of hatchery-origin Okanogan summer Chinook that homed to target spawning areas and the target hatchery, and number and percent that strayed to non-target spawning areas and non-target hatchery programs, by brood years 1989-2008. Percent stays should be less than 5%.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1989	3,132	69.7	1328	29.6	2	0.0	31	0.7
1990	729	71.4	291	28.5	0	0.0	1	0.1

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1991	1,125	71.3	453	28.7	0	0.0	0	0.0
1992	1,264	68.5	572	31.0	8	0.4	1	0.1
1993	54	62.1	32	36.8	0	0.0	1	1.1
1994	924	80.8	203	17.7	16	1.4	1	0.1
1995	1,883	85.4	271	12.3	50	2.3	0	0.0
1996	27	100.0	0	0.0	0	0.0	0	0.0
1997	11,659	97.1	309	2.6	34	0.3	3	0.0
1998	2,784	95.4	102	3.5	31	1.1	2	0.1
1999	828	96.7	18	2.1	10	1.2	0	0.0
2000	2,091	93.6	29	1.3	99	4.4	15	0.7
2001	105	98.1	2	1.9	0	0.0	0	0.0
2002	702	96.2	17	2.3	11	1.5	0	0.0
2003	1,580	96.2	47	2.9	16	1.0	0	0.0
2004	4,947	94.4	206	3.9	85	1.6	2	0.0
2005	606	93.2	22	3.4	22	3.4	0	0.0
2006	5,220	97.6	60	1.1	68	1.3	0	0.0
2007	1,396	97.9	20	1.4	10	0.7	0	0.0
2008	3,582	98.3	35	1.0	23	0.6	4	0.1
<b>Average</b>	<b>2,232</b>	<b>88.2</b>	<b>201</b>	<b>10.6</b>	<b>24</b>	<b>1.1</b>	<b>3</b>	<b>0.2</b>

\* Homing to the target hatchery includes Okanogan/Similkameen hatchery summer Chinook that are captured and included as broodstock in the Okanogan/Similkameen Hatchery program. These hatchery fish were typically collected at Wells Dam.

## Genetics

Genetic studies were conducted to investigate relationships among temporally replicated collections of summer Chinook from the Wenatchee River, Methow River, and Okanogan River in the upper Columbia River basin (Kassler et al. 2011; the entire report is appended as Appendix M). Samples from the Eastbank Hatchery – Wenatchee stock, Eastbank Hatchery – Methow/Okanogan (MEOK) stock, and Wells Hatchery were also included in the analysis. Samples of natural and hatchery-origin summer Chinook were analyzed and compared to determine if the supplementation program has affected the genetic structure of these populations. The study also calculated the effective number of breeders for collection locations of natural and hatchery-origin summer Chinook from 1993 and 2008.

In general, population differentiation was not observed among the temporally replicated collection locations. A single collection from the Okanogan River (1993) was the only collection showing statistically significant differences. The effective number of breeders was not statistically different from the early collection in 1993 in comparison to the late collection in 2008. Overall, these analyses revealed a lack of differentiation among the temporal replicates from the same locations and among the collection from different locations, suggesting the

populations have been homogenized or that there has been substantial gene flow among populations. Additional comparisons among summer-run and fall-run Chinook populations in the upper Columbia River were conducted to determine if there was any differentiation between Chinook with different run timing. These analyses revealed pairwise  $F_{ST}$  values that were less than 0.01 for the collections of summer Chinook to collections of fall Chinook from Hanford Reach, lower Yakima River, Priest Rapids, and Umatilla. Collections of fall Chinook from Crab Creek, Lyons Ferry Hatchery, Marion Drain, and Snake River had pairwise  $F_{ST}$  values that were higher in comparison to the collections of summer Chinook. The consensus clustering analysis did not provide good statistical support to the groupings, but did show relationships among collections based on geographic proximity. Overall the summer and fall run Chinook that have historically been spawned together were not differentiated while fall Chinook from greater geographic distances were differentiated.

### Proportionate Natural Influence

Another method for assessing the genetic risk of a supplementation program is to determine the influence of the hatchery and natural environments on the adaptation of the composite population. This is estimated by the proportion of natural-origin fish in the hatchery broodstock (pNOB) and the proportion of hatchery-origin fish in the natural spawning escapement (pHOS). The ratio  $pNOB/(pHOS+pNOB)$  is the approximate Proportionate Natural Influence (PNI) Index. The larger the ratio (PNI), the greater the strength of selection in the natural environment relative to that of the hatchery environment. In order for the natural environment to dominate selection, PNI should be greater than 0.50, and important integrated populations should have a PNI of at least 0.67 (HSRG/WDFW/NWIFC 2004).

For brood years 1993-2003, the approximate PNI Index was less than 0.67 (Table 10.16). However, since brood year 2003, the approximate PNI Index has generally been greater than 0.67, save 2008 and 2011.

**Table 10.16.** Approximate Proportionate Natural Influence (PNI) Index of the Okanogan/Similkameen summer Chinook supplementation program for brood years 1989-2012. PNI was calculated as the proportion of naturally produced Chinook in the hatchery broodstock (pNOB) divided by the proportion of hatchery Chinook on the spawning grounds (pHOS) plus pNOB. NOS = number of natural-origin Chinook on the spawning grounds; HOS = number of hatchery-origin Chinook on the spawning grounds; NOB = number of natural-origin Chinook collected for broodstock; and HOB = number of hatchery-origin Chinook included in hatchery broodstock.

Brood year	Spawners			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
1989	1,719	0	0.00	1,297	312	0.81	1.00
1990	837	0	0.00	828	206	0.80	1.00
1991	574	0	0.00	924	314	0.75	1.00
1992	473	0	0.00	297	406	0.42	1.00
1993	915	570	0.38	681	388	0.64	0.63
1994	1,323	2,710	0.67	341	244	0.58	0.46
1995	979	2,023	0.67	173	240	0.42	0.39
1996	568	1,251	0.69	287	155	0.65	0.49
1997	862	1,327	0.61	197	265	0.43	0.41

Brood year	Spawners			Broodstock			PNI
	NOS	HOS	pHOS	NOB	HOB	pNOB	
1998	600	492	0.45	153	211	0.42	0.48
1999	1,274	2,343	0.65	224	289	0.44	0.40
2000	1,174	2,527	0.68	164	337	0.33	0.33
2001	4,306	6,551	0.60	12	345	0.03	0.05
2002	4,346	9,511	0.69	247	241	0.51	0.43
2003	1,933	1,487	0.43	381	101	0.79	0.65
2004	5,309	1,412	0.21	506	16	0.97	0.82
2005	6,441	2,448	0.28	391	9	0.98	0.78
2006	5,507	3,094	0.36	500	10	0.98	0.73
2007	2,983	1,434	0.32	456	17	0.96	0.75
2008	2,998	3,977	0.57	359	86	0.81	0.59
2009	4,204	3,340	0.44	503	4	0.99	0.69
2010	3,189	2,763	0.46	484	8	0.98	0.68
2011	4,642	5,039	0.52	467	26	0.95	0.65
2012	4,494	3,731	0.45	79	2	0.98	0.69
<b>Average</b>	<b>2,569</b>	<b>2,418</b>	<b>0.42</b>	<b>415</b>	<b>176</b>	<b>0.69</b>	<b>0.63</b>

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel times (arithmetic mean days) of hatchery summer Chinook from the Similkameen River release site to McNary Dam, and smolt to adult ratios (SARs) from release to detection at Bonneville Dam (Table 10.17).<sup>14</sup> Over the three brood years for which PIT-tagged hatchery fish were released, survival rates from the Similkameen River to McNary Dam ranged from 0.432 to 0.720; SARs from release to detection at Bonneville Dam ranged from 0.016 to 0.030. Average travel time from the Similkameen River to McNary Dam ranged from 41 to 44 days. Although there is only one year in which low densities were compared to high densities (brood year 2008), there was little difference in survival rates and travel times between the two groups (Table 10.17).

**Table 10.17.** Total number of Okanogan hatchery summer Chinook released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2008-2011. Standard errors are shown in parentheses. NA = not available (i.e., not all the fish from the release groups have returned to the Columbia River).

Brood year	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2008	4,531 (high density)	0.445 (0.061)	44.0 (10.2)	0.028 (0.002)
	4,293 (low density)	0.432 (0.050)	41.4 (9.7)	0.030 (0.003)
2009	5,089	0.720 (0.102)	41.5 (10.1)	0.016 (0.002)

<sup>14</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

Brood year	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam (d)	SAR to Bonneville Dam (%)
2010	0	--	--	--
2011	5,036	0.682 (0.064)	41.9 (12.3)	NA

### Natural and Hatchery Replacement Rates

Natural replacement rates (NRR) were calculated as the ratio of natural-origin recruits (NOR) to the parent spawning population (spawning escapement). Natural-origin recruits are naturally produced (wild) fish that survive to contribute to harvest (directly or indirectly), to broodstock, and to spawning grounds. We do not account for fish that died in route to the spawning grounds (migration mortality) or died just before spawning (pre-spawn mortality) (see Appendix B in Hillman et al. 2012). We calculated NORs with and without harvest. NORs without harvest include all returning fish that either returned to the basin or were collected as wild broodstock. NORs with harvest include all fish harvested and are based on brood year harvest rates from the hatchery program. For brood years 1989-2007, NRR for summer Chinook in the Okanogan averaged 1.05 (range, 0.16-3.82) if harvested fish were not include in the estimate and 2.35 (range, 0.32-10.26) if harvested fish were included in the estimate (Table 10.18). NRRs for more recent brood years will be calculated as soon as all tag recoveries and sampling rates have been loaded into the database.

Hatchery replacement rates (HRR) are the hatchery adult-to-adult returns and were calculated as the ratio of hatchery-origin recruits (HOR) to the parent broodstock collected. These rates should be greater than the NRRs and greater than or equal to 5.30 (the calculated target value in Hillman et al. 2013). HRRs exceeded NRRs in 16 of the 19 years of data, regardless if harvest was or was not included in the estimate (Table 10.18). Hatchery replacement rates for Okanogan summer Chinook have exceeded the estimated target value of 5.30 in 8 or 12 of the 18 years of data depending on if harvest was or was not included in the estimate.

**Table 10.18.** Broodstock collected, spawning escapements, natural and hatchery-origin recruits (NOR and HOR), and natural and hatchery replacement rates (NRR and HRR; with and without harvest) for wild summer Chinook in the Okanogan River basin, brood years 1989-2007.

Brood year	Broodstock Collected	Spawning Escapement	Harvest not included				Harvest included			
			HOR	NOR	HRR	NRR	HOR	NOR	HRR	NRR
1989	304	1,719	4,493	2,146	14.78	1.25	7,459	3,577	24.54	2.08
1990	288	837	1,021	1,477	3.55	1.76	1,422	2,063	4.94	2.46
1991	364	574	1,578	629	4.34	1.10	1,835	728	5.04	1.27
1992	304	473	1,845	752	6.07	1.59	2,307	942	7.59	1.99
1993	328	1,485	87	1,003	0.27	0.68	117	1,348	0.36	0.91
1994	302	4,033	1,144	2,168	3.79	0.54	1,550	2,946	5.13	0.73
1995	385	3,002	2,204	959	5.72	0.32	2,902	1,267	7.54	0.42
1996	330	1,819	27	466	0.08	0.26	33	574	0.10	0.32
1997	313	2,189	12,005	4,363	38.35	1.99	19,113	6,959	61.06	3.18
1998	352	1,092	2,919	4,166	8.29	3.82	7,817	11,199	22.21	10.26
1999	333	3,617	856	6,641	2.57	1.84	2,848	22,211	8.55	6.14

Brood year	Broodstock Collected	Spawning Escapement	Harvest not included				Harvest included			
			HOR	NOR	HRR	NRR	HOR	NOR	HRR	NRR
2000	334	3,701	2,234	1,716	6.69	0.46	6,795	5,232	20.34	1.41
2001	335	10,857	107	8,946	0.32	0.82	426	35,784	1.27	3.30
2002	333	13,857	730	6,061	2.19	0.44	1,980	16,470	5.95	1.19
2003	337	3,420	1,643	562	4.88	0.16	3,504	1,201	10.40	0.35
2004	335	6,721	5,240	3,112	15.64	0.46	13,352	7,959	39.86	1.18
2005	338	8,889	650	6,173	1.92	0.69	1,670	15,951	4.94	1.79
2006	355	8,601	5,348	2,422	15.06	0.28	13,752	6,242	38.74	0.73
2007	314	4,417	1,426	6,334	4.54	1.43	4,908	21,841	15.63	4.94
<b>Average</b>	<b>331</b>	<b>4,279</b>	<b>2,398</b>	<b>3,163</b>	<b>7.32</b>	<b>1.05</b>	<b>4,936</b>	<b>8,658</b>	<b>14.96</b>	<b>2.35</b>

### Smolt-to-Adult Survivals

Smolt-to-adult survival ratios (SARs) were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery smolts released. Here, SARs were based on CWT returns. For the available brood years, SARs have ranged from 0.00006 to 0.03249 for hatchery summer Chinook in the Okanogan River basin (Table 10.19).

**Table 10.19.** Smolt-to-adult ratios (SARs) for Okanogan/Similkameen summer Chinook, brood years 1989-2008.

Brood year	Number of tagged smolts released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1989	202,125	4,293	0.02124
1990	367,207	972	0.00265
1991	360,380	975	0.00271
1992	537,190	2,282	0.00425
1993	379,139	117	0.00031
1994	217,818	1,528	0.00702
1995	574,197	2,851	0.00497
1996	487,776	32	0.00007
1997	572,531	18,599	0.03249
1998	287,948	7,706	0.02676
1999	610,868	2,776	0.00454
2000	528,639	6,767	0.01280
2001	26,315	424	0.01611
2002	245,997	1,975	0.00803
2003	574,908	3,489	0.00607
2004	676,222	12,896	0.01907
2005	273,512	1,660	0.00607
2006	597,276	13,633	0.02283

Brood year	Number of tagged smolts released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
2007	610,379	4,890	0.00801
2008	516,533	14,225	0.02754
<i>Average</i>	<i>432,348</i>	<i>5,105</i>	<i>0.01168</i>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

## 10.7 ESA/HCP Compliance

### Broodstock Collection

Because summer Chinook adults collected at Wells Dam are used for both the Methow and Okanogan supplementation programs, please refer to Section 9.7 for information on ESA compliance during broodstock collection. Direct and/or indirect take of ESA-listed species during broodstock collection for the Okanogan summer Chinook outside of Wells Dam is covered by permits held by the Colville Tribes.

### Hatchery Rearing and Release

Activities associated with the spawning, rearing, and release of Okanogan summer Chinook that could result in either direct or incidental take of listed species is covered under ESA permits held by the Colville Tribes.

### Hatchery Effluent Monitoring

Per ESA Permits 1196, 1347, 1395, 18118, 18120, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January 2014 through 31 December 2014. NPDES monitoring and reporting for PUD Hatchery Programs during 2014 are provided in Appendix F. NPDES reporting for Okanogan summer Chinook only covers the Similkameen acclimation facility and only during the time fish are present.



## SECTION 11: CHELAN FALLS SUMMER CHINOOK

Although the Chelan Falls summer Chinook program (formerly the Turtle Rock program) is an augmentation program, the production of 200,000 fish is No Net Impact (NNI) compensation for passage mortalities associated with Rocky Reach Dam. In addition, the conversion of the subyearling program to a 400,000 yearling program is compensation for lost spawning habitat as a result of the construction of Rocky Reach Dam. In 2011, as part of the periodic recalculation of NNI for Rocky Reach Dam, the previous 200,000 NNI program was reduced to 176,000 fish. This reduced the combined Chelan Falls summer Chinook production from 600,000 to 576,000 beginning with the 2012 brood.

Before 2012, broodstock were collected at Wells Dam and consisted of volunteers to the Wells Fish Hatchery. Summer Chinook were spawned at Wells Fish Hatchery and fertilized eggs were then transferred to Eastbank Fish Hatchery for hatching and rearing. In 2012, adults were collected at Wells Fish Hatchery and then transferred to Eastbank Fish Hatchery for spawning, hatching, and rearing. Beginning in 2013, broodstock have been collected from Eastbank Hatchery Outfall.

The original program consisted of both subyearling (normal and accelerated groups) and yearling releases. Subyearlings were transferred to Turtle Rock Fish Hatchery for acclimation in May. These fish were released in June after about 30 days of acclimation on Columbia River water. The goal of this program was to release 1,620,000 subyearling summer Chinook (810,000 normal and 810,000 accelerated subyearlings) into the Columbia River at 40 fish per pound. Targets for fork length and weight were 112 mm (CV = 9.0) and 11.4 g, respectively. Over 50% of both subyearling groups were marked with CWTs. In 2010, the subyearling program was converted to a 400,000 yearling program.

The goal of the yearling program was to release 200,000 summer Chinook smolts into the Columbia River from Turtle Rock Fish Hatchery at 10 fish per pound. Targets for fork length and weight were 176 mm (CV = 9.0) and 45.4 g, respectively. Beginning with the 2006 brood year, yearling summer Chinook were acclimated at both Turtle Rock Fish Hatchery and the Chelan River net pens. With the conversion of the subyearling program to a yearling program and the reduction of the NNI component to 176,000, the current goal is to release 576,000 yearling summer Chinook smolts (176,000 from the NNI program plus 400,000 from the converted subyearling program). Beginning in 2012, the 576,000 yearlings are acclimated overwinter at facilities at Chelan Falls Hatchery on Chelan River water. In 2012, the Turtle Rock program officially became the Chelan Falls summer Chinook program.

Over 90% of yearling summer Chinook have been marked with CWTs and all are ad-clipped. In addition, juvenile summer Chinook were PIT tagged within each of the circular reuse and standard raceways.

### 11.1 Broodstock Sampling

Before 2013, broodstock for the program were collected as part of the Wells summer Chinook volunteer program. Refer to Snow et al. (2012) for information related to adults collected for these programs. Beginning in 2013, broodstock for the Chelan Falls program are collected from the Eastbank Hatchery Outfall.

## 11.2 Hatchery Rearing

### Rearing History

#### *Number of eggs taken*

Based on the unfertilized egg-to-release standard of 81%, a total of 688,995 eggs were needed to meet the program goal of 576,000 smolts for brood years 2012 and 2013. An evaluation of the program in 2014 concluded that 696,493 eggs were needed to attain the 576,000 smolts. From 2012-2014, the egg take goal was reached in 2013.

#### *Disease*

Significant health concerns were encountered during rearing of Chelan Falls summer Chinook in 2014 (BY 2012). Specifically, after transfer from Eastbank Fish Hatchery to the Chelan Falls acclimation facility in November, there was an increase in mortality. Diagnosis showed initial transfer trauma, followed by fungus, bacterial cold water disease, and fusobacteria. January to February showed an increase in mortality of emaciated, smaller fish. No treatment was prescribed.

#### *Number of acclimation days*

Rearing of the 2012-brood Chelan Falls summer Chinook was similar to previous years with fish being held on well water. This was the third year that the whole program was transferred to the Chelan Falls Acclimation Facility for final overwinter acclimation. Transfer occurred on 4-14 November 2013. Fish were force released on 15 April 2014 after 153-163 days of acclimation on Chelan River water.

### Release Information

#### *Numbers released*

The subyearling Turtle Rock summer Chinook program was discontinued in 2010; however, releases of subyearling Chinook in past years are shown in Tables 11.1 and 11.2. Production from the subyearling programs was converted to the yearling program.

The 2012 yearling summer Chinook program achieved 94.4% of the 600,000 target goal with about 566,188 fish being released from the Chelan River Acclimation Ponds (Table 11.3). Releases of 2013 yearling Chinook will be reported in the 2015 report.

**Table 11.1.** Numbers of Turtle Rock summer Chinook subyearlings released from the hatchery, brood years 1995-2009. The release target for Turtle Rock summer Chinook subyearlings was 810,000 fish.

Brood year	Release year	CWT mark rate	Number of subyearlings released
1995	1996	0.1873	1,074,600
1996	1997	0.9653	385,215
1997	1998	0.9780	508,060
1998	1999	0.6453	301,777
1999	2000	0.9748	369,026
2000	2001	0.3678	604,892
2001	2002	0.9871	214,059
2002	2003	0.3070	656,399

Brood year	Release year	CWT mark rate	Number of subyearlings released
2003	2004	0.4138	491,480
2004	2005	0.4591	411,707
2005	2006	0.4337	490,074
2006	2007	0.3388	538,392
2007	2008	0.4385	439,806
2008	2009	0.6355	309,003
2009	2010	NA	713,130
<i>Average</i>		<i>0.6111</i>	<i>500,508</i>

**Table 11.2.** Numbers of Turtle Rock summer Chinook accelerated subyearlings released from the hatchery, brood years 1995-2008. The release target for Turtle Rock summer Chinook accelerated subyearlings was 810,000 fish.

Brood year	Release year	CWT mark rate	Number of subyearlings released
1995	1996	0.9834	169,000
1996	1997	0.4163	477,300
1997	1998	0.3767	521,480
1998	1999	0.6033	307,571
1999	2000	0.9556	347,946
2000	2001	0.4331	449,329
2001	2002	0.4086	480,584
2002	2003	0.5492	364,461
2003	2004	0.6414	289,696
2004	2005	0.5471	364,453
2005	2006	0.9783	457,340
2006	2007	0.5510	342,273
2007	2008	0.4745	392,024
2008	2009	0.5295	372,320
<i>Average</i>		<i>0.6034</i>	<i>381,127</i>

**Table 11.3.** Numbers of Turtle Rock summer Chinook yearling smolts released from the hatchery, brood years 1995-2012. The release target for Turtle Rock summer Chinook was 200,000 smolts for the period before brood year 2010. The current release target is 600,000 smolts.

Brood year	Release year	Acclimation facility	CWT mark rate	Number of smolts released
1995	1997	Turtle Rock	0.9688	150,000
1996	1998	Turtle Rock	0.9582	202,727
1997	1999	Turtle Rock	0.9800	202,989
1998	2000	Turtle Rock	0.9337	217,797

Brood year	Release year	Acclimation facility	CWT mark rate	Number of smolts released
1999	2001	Turtle Rock	0.9824	285,707
2000	2002	Turtle Rock	0.9941	279,969
2001	2003	Turtle Rock	0.9824	203,279
2002	2004	Turtle Rock	0.9799	195,851
2003	2005	Turtle Rock	0.9258	215,366
2004	2006	Turtle Rock	0.9578	206,734
2005	2007	Chelan	0.9810	204,644
2006	2008	Chelan	0.9752	99,271
		Turtle Rock	0.9752	43,943
2007	2009	Chelan Falls	0.9426	112,604
		Turtle Rock	0.9426	61,003
2008	2010	Chelan Falls	0.9818	200,999
		Turtle Rock	0.9818	252,762
2009	2011	Chelan Falls <sup>a</sup>	-	190,449
		Turtle Rock	0.9721	250,667
<b>Average (1995-2009)</b>		<b>Chelan Falls</b>	<b>0.9665</b>	<b>137,625</b>
		<b>Turtle Rock</b>	<b>0.9745</b>	<b>233,429</b>
2010	2012	Chelan Falls	0.9702	563,824
2011	2013	Chelan Falls	0.9859	582,460
2012	2014	Chelan Falls	0.9879	566,188
<b>Average (2010-present)</b>		<b>Chelan Falls</b>	<b>0.9813</b>	<b>570,824</b>

<sup>a</sup>No CWT mark rate was provided because of the early release of this group.

### **Numbers tagged**

Brood year 2012 yearling Chinook were 98.0% CWT and adipose fin-clipped.

In 2014, a total of 10,000 summer Chinook from the 2012 brood were PIT tagged at the Chelan River Hatchery during 10-19 March. Fish were tagged in four groups of 2,500 per group. Two groups made up a “small-size fish” group that averaged 123-129 mm and 21-25 g at time of tagging, and the other two made up a “big-size fish” group that averaged 133-138 mm and 25-29 g at time of tagging. The two size groups were developed to identify techniques that maximize performance of hatchery-origin summer yearling Chinook salmon. This is part of the NOAA Fisheries size-target study. Fish were not fed during tagging or for two days before and after tagging. A total of 9,943 PIT-tagged summer Chinook were released into the Chelan River in April 2014. A total of 57 fish died and no fish shed their tags during the period between tagging and release.

Table 11.4 summarizes the number of yearling summer Chinook that have been PIT-tagged and released from the Turtle Rock/Chelan Falls Program.

**Table 11.4.** Summary of PIT-tagging activities for Turtle Rock/Chelan Falls yearling summer Chinook, brood years 2007-2012.

Brood year	Release year	Raceway/Program	Number of fish tagged	Number of tagged fish that died	Number of tags shed	Number of tagged fish released
2007	2009	Circular Reuse	10,104	128	1	9,975
		Standard	10,102	162	3	9,937
2008	2010	Circular Reuse	11,102	15	0	11,087
		Standard	11,100	18	2	11,080
2009	2011	Turtle Rock	5,051	106	0	4,945
		Chelan Net Pens	5,050	2	0	5,048
2010	2012	Chelan Falls	4,200	10	0	4,190
2011	2013	Chelan Falls	4,101	26	0	4,075
2012	2014	Chelan Falls (Small Fish)	5,000	17	0	4,983
		Chelan Falls (Big Fish)	5,000	40	0	4,960

*Fish size and condition at release*

Although the subyearling summer Chinook program was discontinued, sizes of subyearlings released from Turtle Rock Hatchery before 2010 are shown in Tables 11.5 and 11.6.

**Table 11.5.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of Turtle Rock summer Chinook subyearlings released from the hatchery, brood years 1995-2009. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1995	1996	102	6.3	12.6	36
1996	1997	87	8.0	7.4	62
1997	1998	98	6.2	10.2	45
1998	1999	96	6.3	10.7	43
1999	2000	90	9.0	9.8	46
2000	2001	100	7.1	11.3	40
2001	2002	104	7.2	13.4	34
2002	2003	97	7.3	11.8	39
2003	2004	101	8.0	12.0	43
2004	2005	100	7.8	11.4	40
2005	2006	100	6.5	12.5	36
2006	2007	95	7.2	9.5	48
2007	2008	79	7.4	5.6	81
2008	2009	86	7.9	7.9	57
2009 <sup>a</sup>	2010	89	7.1	7.0	65

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
<i>Average</i>		<i>95</i>	<i>7.3</i>	<i>10.2</i>	<i>48</i>
<i>Targets</i>		<i>112</i>	<i>9.0</i>	<i>11.4</i>	<i>40</i>

<sup>a</sup> Pre-release growth sample was conducted using pond mortalities.

**Table 11.6.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of Turtle Rock summer Chinook accelerated subyearlings released from the hatchery, brood years 1995-2008. Size targets are provided in the last row of the table.

Brood year	Release year	Fork length (mm)		Mean weight	
		Mean	CV	Grams (g)	Fish/pound
1995	1996	129	7.1	27.3	17
1996	1997	107	6.5	15.6	29
1997	1998	117	6.0	18.9	24
1998	1999	119	8.0	18.9	24
1999	2000	114	6.7	19.0	24
2000	2001	111	7.0	16.8	27
2001	2002	117	8.4	19.5	23
2002	2003	116	11.3	21.2	21
2003	2004	113	14.9	17.0	30
2004	2005	117	11.3	20.1	23
2005	2006	119	9.1	22.2	21
2006	2007	118	8.3	19.1	24
2007	2008	95	7.7	10.0	45
2008 <sup>a</sup>	2009	97	8.6	10.6	43
<i>Average</i>		<i>114</i>	<i>8.6</i>	<i>18.3</i>	<i>27</i>
<i>Targets</i>		<i>112</i>	<i>9.0</i>	<i>11.4</i>	<i>40</i>

<sup>a</sup> The 2008 brood year was the last year of the accelerated subyearling program.

Size at release of the brood year 2012 yearling summer Chinook was 80.1% and 54.0% of the fork length and weight targets, respectively, for the Chelan Falls group. This group exceeded the target CV for length (Table 11.7).

**Table 11.7.** Mean lengths (FL, mm), weight (g and fish/pound), and coefficient of variation (CV) of Turtle Rock/Chelan summer Chinook yearling releases, brood years 1995-2012. Size targets are provided in the last row of the table.

Brood year	Release year	Acclimation facility	Fork length (mm)		Mean weight	
			Mean	CV	Grams (g)	Fish/pound
1995	1997	Turtle Rock	-	-	-	-
1996	1998	Turtle Rock	166	14.2	60.9	7
1997	1999	Turtle Rock	198	4.6	91.3	5

Brood year	Release year	Acclimation facility	Fork length (mm)		Mean weight	
			Mean	CV	Grams (g)	Fish/pound
1998	2000	Turtle Rock	161	11.9	53.9	8
1999	2001	Turtle Rock	164	18.6	59.0	8
2000	2002	Turtle Rock	170	15.3	59.0	8
2001	2003	Turtle Rock	154	22.3	48.6	9
2002	2004	Turtle Rock	157	16.7	44.0	12
2003	2005	Turtle Rock	173	13.8	54.7	8
2004	2006	Turtle Rock	176	20.6	45.3	7
2005	2007	Turtle Rock	158	11.0	43.5	10
2006	2008	Chelan Nets	172	14.5	58.4	8
		Turtle Rock	157	25.8	54.1	8
2007	2009	Chelan Nets	153	18.8	45.7	10
		Turtle Rock	167	14.6	49.3	9
2008	2010	Chelan Nets	146	22.9	40.6	11
		Turtle Rock	172	15.9	58.5	8
2009	2011	Chelan Nets	158	15.1	46.6	10
		Turtle Rock	174	17.5	59.3	8
2010	2012	Chelan Falls	132	27.4	33.2	14
2011	2013	Chelan Falls	148	18.6	42.6	11
2012	2014	Chelan Falls	129	17.1	24.5	19
<b>Average</b>			<b>161</b>	<b>17.0</b>	<b>51.1</b>	<b>9</b>
<b>Targets<sup>a</sup></b>			<b>161</b>	<b>9.0</b>	<b>45.4</b>	<b>10</b>

<sup>a</sup> For size-target studies, fish per pound (fpp) targets for brood year 2012 were 10, 13, 18, 22 fpp.

## Survival Estimates

### Normal subyearling releases

Overall survival of the normal subyearling Turtle Rock summer Chinook program from green egg to release was below the standard set for the program (Table 11.8). Lower than expected survival at ponding and post-ponding reduced the overall program performance. This program was discontinued in 2010.

**Table 11.8.** Hatchery life-stage survival rates (%) for Turtle Rock subyearling (zero program) summer Chinook, brood years 2004-2009. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
2004	NA	NA	93.5	74.4	93.9	91.4	90.8	99.7	63.1
2005	NA	NA	94.4	87.9	85	84.8	84.2	99.4	69.8
2006	NA	NA	97.8	87.9	85.0	84.8	84.2	99.4	72.4
2007	NA	NA	92.7	84.9	88.5	86.7	84.8	99.6	66.7

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
2008	NA	NA	78.8	95.0	80.7	79.3	79.9	99.8	59.8
2009	NA	NA	95.0	89.4	89.5	89.2	79.7	89.5	67.7
<i>Average</i>	<i>NA</i>	<i>NA</i>	<i>92.0</i>	<i>86.6</i>	<i>87.1</i>	<i>86.0</i>	<i>83.9</i>	<i>97.9</i>	<i>66.6</i>
<i>Standard</i>	<i>90.0</i>	<i>85.0</i>	<i>92.0</i>	<i>98.0</i>	<i>97.0</i>	<i>93.0</i>	<i>90.0</i>	<i>95.0</i>	<i>81.0</i>

### Accelerated subyearling releases

Overall survival of the accelerated subyearling Turtle Rock summer Chinook program from green egg to release was below the standard set for the program (Table 11.9). Lower than expected survival in post-ponding reduced the overall program performance. This program was discontinued in 2010.

**Table 11.9.** Hatchery life-stage survival rates (%) for Turtle Rock subyearling (accelerated program) summer Chinook, brood years 2004-2009. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Unfertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Unfertilized egg-release
	Female	Male							
2004	NA	NA	92.5	98.3	93.4	92.4	90.0	97.8	81.8
2005	NA	NA	93.8	94.6	83.7	83.4	81.7	98.8	72.5
2006	NA	NA	86.1	94.6	83.7	83.4	81.7	98.8	66.5
2007	NA	NA	93.4	95.4	78.4	77.5	76.3	98.9	67.9
2008 <sup>a</sup>	NA	NA	93.4	95.0	79.8	78.8	78.2	99.3	67.1
<i>Average</i>	<i>NA</i>	<i>NA</i>	<i>91.8</i>	<i>95.6</i>	<i>83.8</i>	<i>83.1</i>	<i>81.6</i>	<i>98.7</i>	<i>71.2</i>
<i>Standard</i>	<i>90.0</i>	<i>85.0</i>	<i>92.0</i>	<i>98.0</i>	<i>97.0</i>	<i>93.0</i>	<i>90.0</i>	<i>95.0</i>	<i>81.0</i>

<sup>a</sup> The 2008 brood year was the last year of the accelerated subyearling program.

### Yearling releases

Overall survival of the yearling Chelan Falls summer Chinook program from green egg to release was above the standard set for the program (Table 11.10). Higher than expected survivals in all life stages contributed to the increased program performance.

**Table 11.10.** Hatchery life-stage survival rates (%) for Turtle Rock/Chelan Falls yearling summer Chinook, brood years 2004-2012. Survival standards or targets are provided in the last row of the table.

Brood year	Collection to spawning		Un-fertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Un-fertilized egg-release
	Female	Male							
2004	NA	NA	92.9	97.7	96.8	96.4	95.5	99.6	86.7
2005	NA	NA	89.1	97.5	98.1	97.8	96.6	99.1	83.9
2006	NA	NA	86.2	78.8	97.6	97.1	95.2	98.7	64.8
2007 (Turtle Rock)	NA	NA	80.3	97.6	98.8	98.2	95.4	99.1	74.8



Brood year	Collection to spawning		Un-fertilized egg-eyed	Eyed egg-ponding	30 d after ponding	100 d after ponding	Ponding to release	Transport to release	Un-fertilized egg-release
	Female	Male							
2007 (Chelan Falls)	NA	NA	80.3	97.6	98.8	98.2	94.9	97.1	74.4
2008 (Turtle Rock)	NA	NA	93.5	98.0	99.4	97.2	95.9	98.8	87.8
2008 (Chelan Falls)	NA	NA	93.5	98.0	97.6	98.7	96.4	99.3	88.2
2009 (Turtle Rock)	NA	NA	90.8	96.8	99.7	99.0	97.2	98.1	85.5
2009 (Chelan Falls)	NA	NA	90.9	96.9	99.8	99.0	96.7	97.7	85.2
2010 (Chelan Falls)	NA	NA	94.8	97.7	99.4	95.2	92.4	97.6	85.5
2011 (Chelan Falls)	NA	NA	90.0	99.4	91.7	98.2	83.4	85.2	74.6
2012 (Chelan Falls)	NA	NA	93.5	98.5	99.8	99.3	95.9	96.7	88.3
<b>Average (Chelan)</b>	<b>NA</b>	<b>NA</b>	<b>90.5</b>	<b>98.0</b>	<b>97.9</b>	<b>98.1</b>	<b>93.3</b>	<b>95.6</b>	<b>82.7</b>
<b>Standard</b>	<b>90.0</b>	<b>85.0</b>	<b>92.0</b>	<b>98.0</b>	<b>97.0</b>	<b>93.0</b>	<b>90.0</b>	<b>95.0</b>	<b>81.0</b>

### 11.3 Spawning Surveys

Surveys for summer Chinook redds in the Chelan River were conducted from late September to mid-November 2014. Total redd counts were conducted in the river (see Appendix N for more details).

#### Redd Counts

A total of 400 summer Chinook redds were counted in the Chelan River in 2014 (Table 11.11). This was higher than the overall average of 286 redds.

**Table 11.11.** Total number of redds counted in the Chelan River, 2000-2014.

Survey year	Total redd count
2000	196
2001	240
2002	253
2003	173
2004	185
2005	179
2006	208
2007	86
2008	153
2009	246
2010	398
2011	413
2012	426
2013	729
2014	400
<b>Average</b>	<b>286</b>

### Redd Distribution

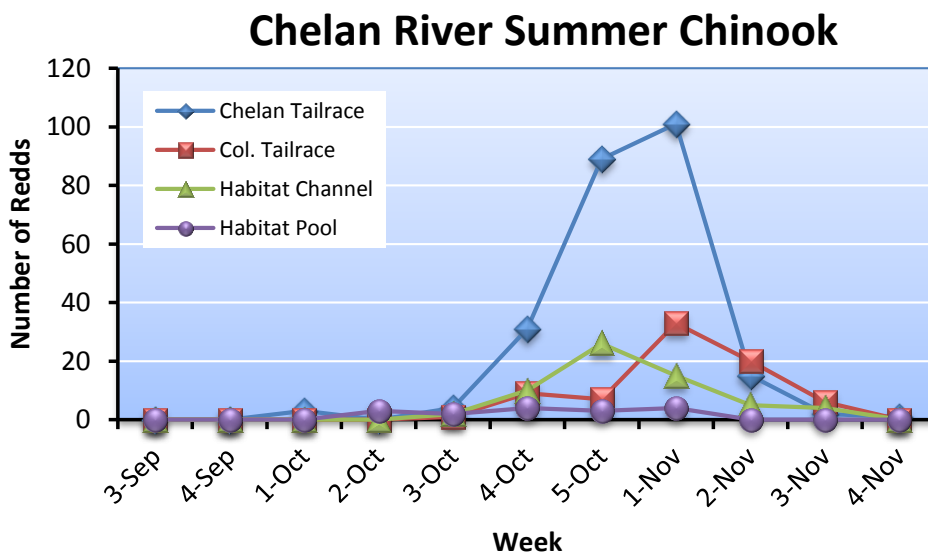
Summer Chinook redds were not evenly distributed among the four sampling areas within the Chelan River. Most redds (62%) were located in the Chelan Tailrace (Table 11.12). Few summer Chinook spawned in the Habitat Pool.

**Table 11.12.** Total number of summer Chinook redds counted in different survey areas within the Chelan River during September through early November, 2014.

Survey area	Total redd count	Percent
Chelan Tailrace	246	61.5
Columbia Tailrace	76	19.0
Habitat Channel	62	15.5
Habitat Pool	16	4.0
<i>Totals</i>	<i>400</i>	<i>100.0</i>

### Spawn Timing

Spawning in 2014 began the last week of September, peaked in late October and early November, and ended late November. Peak spawning occurred in the Chelan Tailrace and Columbia River Tailrace during the first week of November and in the Habitat Channel in late October (Figure 11.1).



**Figure 11.1.** Number of new summer Chinook redds counted during different weeks within different sections of the Chelan River, September through November 2014.

### Spawning Escapement

Spawning escapement for summer Chinook in the Chelan River was calculated as the total number of redds times the fish per redd ratio estimated from fish sampled at Wells Dam. The estimated fish per redd ratio for Methow summer Chinook in 2014 was 2.75. Multiplying this

ratio by the number of redds counted in the Chelan River resulted in a total spawning escapement of 1,100 summer Chinook (Table 11.13).

**Table 11.13.** Spawning escapements for summer Chinook in the Chelan River for return years 2000-2014.

Return year	Fish/Redd	Redds	Total spawning escapement
2000	2.40	196	470
2001	4.10	240	984
2002	2.30	253	582
2003	2.42	173	419
2004	2.25	185	416
2005	2.93	179	524
2006	2.02	208	420
2007	2.20	86	189
2008	3.25	153	497
2009	2.54	246	625
2010	2.81	398	1,118
2011	3.10	413	1,280
2012	3.07	426	1,308
2013	2.31	729	1,684
2014	2.75	400	1,100
<i>Average</i>	<i>2.70</i>	<i>286</i>	<i>775</i>

### 11.4 Carcass Surveys

Surveys for summer Chinook carcasses within the Chelan River were conducted during late September to mid-November 2014 (see Appendix N for more details).

#### Number sampled

A total of 309 summer Chinook carcasses were sampled during September through mid-November in the Chelan River (Table 11.14). This was higher than the overall average of 161 carcasses sampled since 2000.

**Table 11.14.** Numbers of summer Chinook carcasses sampled within each survey area within the Chelan River, 2000-2014; ND = no data.

Survey year	Number of summer Chinook carcasses				
	Chelan Tailrace	Columbia Tailrace	Habitat Channel	Habitat Pool	Total
2000	ND	ND	ND	ND	<b>48</b>
2001	ND	ND	ND	ND	<b>101</b>
2002	ND	ND	ND	ND	<b>145</b>
2003	ND	ND	ND	ND	<b>168</b>
2004	ND	ND	ND	ND	<b>159</b>
2005	ND	ND	ND	ND	<b>103</b>

Survey year	Number of summer Chinook carcasses				
	Chelan Tailrace	Columbia Tailrace	Habitat Channel	Habitat Pool	Total
2006	ND	ND	ND	ND	107
2007	ND	ND	ND	ND	106
2008	ND	ND	ND	ND	132
2009	ND	ND	ND	ND	51
2010	ND	ND	ND	ND	106
2011	ND	ND	ND	ND	201
2012	ND	ND	ND	ND	317
2013	50	120	157	28	355
2014	171	82	50	6	309
<b>Average</b>	<b>111</b>	<b>101</b>	<b>104</b>	<b>17</b>	<b>161</b>

### Carcass Distribution and Origin

Summer Chinook carcasses were not evenly distributed among survey areas within the Chelan River in 2014 (Table 11.14). Most of the carcasses in the Chelan River were found in the Chelan Tailrace.

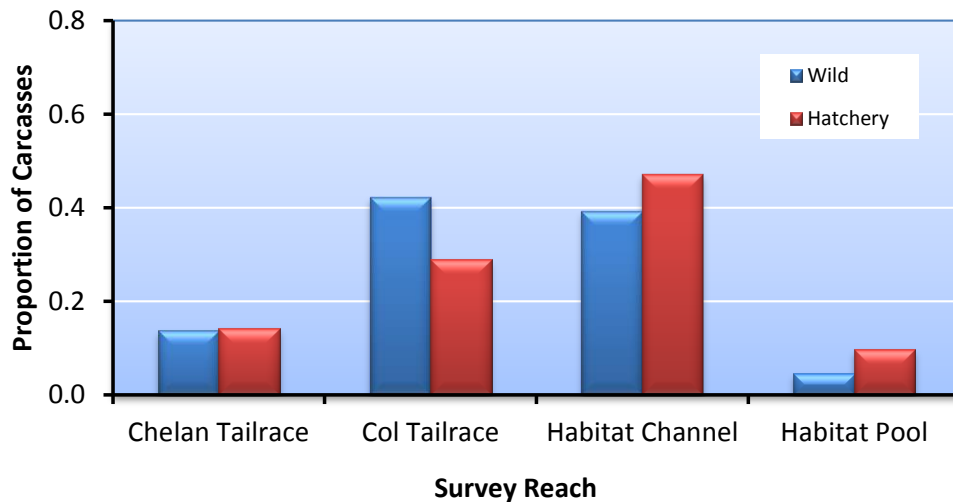
Numbers of wild and hatchery-origin summer Chinook carcasses sampled in 2014 will be available after analysis of CWTs and scales. Based on the available data, hatchery and wild summer Chinook carcasses were not distributed equally among the survey areas within the Chelan River (Table 11.15; Figure 11.2). A larger percentage of hatchery carcasses occurred in the Habitat Channel and Habitat Pool, while a larger percentage of wild summer Chinook carcasses occurred in the Columbia River Tailrace.

**Table 11.15.** Numbers of wild and hatchery summer Chinook carcasses sampled within different survey areas on the Chelan River, 2000-2013; ND = no data.

Survey year	Origin	Survey reach				Total
		Chelan Tailrace	Columbia Tailrace	Habitat Channel	Habitat Pool	
2000	Wild	ND	ND	ND	ND	17
	Hatchery	ND	ND	ND	ND	31
2001	Wild	ND	ND	ND	ND	26
	Hatchery	ND	ND	ND	ND	75
2002	Wild	ND	ND	ND	ND	37
	Hatchery	ND	ND	ND	ND	108
2003	Wild	ND	ND	ND	ND	33
	Hatchery	ND	ND	ND	ND	135
2004	Wild	ND	ND	ND	ND	91
	Hatchery	ND	ND	ND	ND	68
2005	Wild	ND	ND	ND	ND	42
	Hatchery	ND	ND	ND	ND	61
2006	Wild	ND	ND	ND	ND	69
	Hatchery	ND	ND	ND	ND	38

Survey year	Origin	Survey reach				Total
		Chelan Tailrace	Columbia Tailrace	Habitat Channel	Habitat Pool	
2007	Wild	ND	ND	ND	ND	35
	Hatchery	ND	ND	ND	ND	71
2008	Wild	ND	ND	ND	ND	69
	Hatchery	ND	ND	ND	ND	63
2009	Wild	ND	ND	ND	ND	2
	Hatchery	ND	ND	ND	ND	49
2010	Wild	ND	ND	ND	ND	46
	Hatchery	ND	ND	ND	ND	60
2011	Wild	ND	ND	ND	ND	89
	Hatchery	ND	ND	ND	ND	112
2012	Wild	ND	ND	ND	ND	64
	Hatchery	ND	ND	ND	ND	253
2013	Wild	18	55	51	6	130
	Hatchery	23	65	106	22	225
Average	Wild	18	55	51	6	52
	Hatchery	32	65	106	22	96

### Chelan River Summer Chinook



**Figure 11.2.** Distribution of wild and hatchery produced carcasses in different survey areas within the Chelan River, 2014.

#### Sampling Rate

Overall, 28% of the total spawning escapement of summer Chinook in the Chelan River was sampled in 2014 (Table 11.16). Sampling rates among survey reaches varied from 12 to 82%.

**Table 11.16.** Number of redds and carcasses, total spawning escapement, and sampling rates for summer Chinook in the Chelan River, 2014.

Survey reach	Total number of redds	Total number of carcasses	Total spawning escapement	Sampling rate
Chelan Tailrace	246	82	677	12.1
Columbia Tailrace	76	171	209	81.8
Habitat Channel	62	50	171	29.3
Habitat Pool	16	6	44	13.6
<b>Total</b>	<b>400</b>	<b>309</b>	<b>1,100</b>	<b>28.1</b>

### Length Data

Mean lengths (POH, cm) of male and female summer Chinook carcasses sampled during surveys on the Chelan River in 2014 are provided in Table 11.17. The average size of males and females sampled in the Chelan River were 62 cm and 66 cm, respectively.

**Table 11.17.** Mean lengths (postorbital-to-hypural length; cm) and standard deviations (in parentheses) of male and female summer Chinook carcasses sampled in different areas on the Chelan River, 2014.

Stream/watershed	Mean length (cm)	
	Male	Female
Chelan Tailrace	62.2 (6.4)	65.0 (5.2)
Columbia Tailrace	61.8 (9.0)	66.0 (5.0)
Habitat Channel	63.1 (8.7)	65.8 (5.0)
Habitat Pool	68.0 (0.0)	69.0 (7.3)
<b>Total</b>	<b>62.2 (8.6)</b>	<b>65.7 (5.1)</b>

## 11.5 Life History Monitoring

Life history characteristics of Chelan Falls and Turtle Rock summer Chinook were assessed by examining carcasses on spawning grounds and by reviewing tagging data and fisheries statistics.

### Contribution to Fisheries

#### *Normal subyearling releases*

Most of the harvest on Turtle Rock summer Chinook (normal subyearling releases) occurred in the Ocean (10-100% of the fish harvested; Table 11.18). Brood years 1995 and 2006 provided the largest total harvests, while brood year 1997 and 1998 provided the lowest. The subyearling hatchery program was discontinued in 2010.

**Table 11.18.** Estimated number and percent (in parentheses) of Turtle Rock summer Chinook (normal subyearling releases) captured in different fisheries, brood years 1995-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
1995	688 (84)	106 (13)	11 (1)	16 (2)	821
1996	72 (80)	0 (0)	5 (6)	13 (14)	90
1997	10 (100)	0 (0)	0 (0)	0 (0)	10
1998	21 (100)	0 (0)	0 (0)	0 (0)	21
1999	184 (64)	26 (9)	4 (1)	75 (26)	289
2000	36 (55)	8 (12)	8 (12)	14 (21)	66
2001	164 (64)	30 (12)	20 (8)	44 (17)	258
2002	23 (20)	33 (29)	3 (3)	56 (49)	115
2003	9 (10)	55 (61)	2 (2)	24 (27)	90
2004	42 (37)	29 (25)	2 (2)	42 (37)	115
2005	100 (38)	95 (36)	24 (9)	44 (17)	263
2006	305 (41)	288 (38)	53 (7)	104 (14)	750
2007	110 (34)	91 (28)	21 (6)	104 (32)	326
2008	42 (33)	32 (25)	4 (3)	48 (38)	126
<i>Average</i>	<i>129 (54)</i>	<i>57 (21)</i>	<i>11 (4)</i>	<i>42 (21)</i>	<i>239</i>

### *Accelerated subyearling releases*

Most of the harvest on Turtle Rock summer Chinook (accelerated subyearling releases) occurred in ocean fisheries (Table 11.19). Ocean harvest has made up 27% to 100% of all Turtle Rock summer Chinook harvested (no fish from the 2003 brood year were harvested). Brood year 1999 provided the largest total harvest, while brood years 1995, 1997, 2002, and 2003 provided the lowest. This hatchery program was discontinued in 2010.

**Table 11.19.** Estimated number and percent (in parentheses) of Turtle Rock summer Chinook (accelerated subyearling releases) captured in different fisheries, brood years 1995-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
1995	3 (100)	0 (0)	0 (0)	0 (0)	3
1996	77 (89)	5 (6)	5 (6)	0 (0)	87
1997	3 (100)	0 (0)	0 (0)	0 (0)	3
1998	97 (95)	2 (2)	3 (3)	0 (0)	102
1999	1,025 (76)	142 (10)	12 (1)	178 (13)	1,357
2000	117 (100)	0 (0)	0 (0)	0 (0)	117
2001	205 (59)	49 (14)	13 (4)	80 (23)	347
2002	9 (100)	0 (0)	0 (0)	0 (0)	9
2003	0 (0)	0 (0)	0 (0)	0 (0)	0

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
2004	45 (27)	79 (48)	6 (4)	34 (21)	164
2005	65 (59)	12 (11)	26 (24)	7 (6)	110
2006	130 (43)	113 (37)	16 (5)	43 (14)	302
2007	169 (42)	168 (42)	12 (3)	51 (13)	400
2008	20 (54)	2 (5)	4 (11)	11 (30)	37
<i>Average</i>	<i>140 (60)</i>	<i>41 (13)</i>	<i>7 (4)</i>	<i>29 (9)</i>	<i>217</i>

### Yearling releases

Most of the harvest on Turtle Rock/Chelan Falls summer Chinook (yearling releases) occurred in ocean fisheries (Table 11.20). Ocean harvest has made up 39% to 95% of all Turtle Rock summer Chinook harvested. Brood year 2008 provided the largest harvest, while brood years 1995 and 1996 provided the lowest.

**Table 11.20.** Estimated number and percent (in parentheses) of Turtle Rock/Chelan Falls summer Chinook (yearling releases) captured in different fisheries, brood years 1995-2008.

Brood year	Ocean fisheries	Columbia River Fisheries			Total
		Tribal	Commercial (Zones 1-5)	Recreational (sport)	
1995	457 (75)	51 (8)	31 (5)	70 (11)	609
1996	766 (95)	14 (2)	2 (0)	21 (3)	803
1997	2,798 (91)	61 (2)	27 (1)	176 (6)	3,062
1998	4,292 (90)	224 (5)	16 (0)	230 (5)	4,762
1999	1,655 (73)	233 (10)	7 (0)	383 (17)	2,278
2000	1,205 (72)	129 (9)	54 (3)	273 (16)	1,679
2001	1,938 (59)	453 (14)	178 (5)	729 (22)	3,298
2002	1,005 (50)	384 (19)	102 (5)	536 (26)	2,027
2003	753 (46)	449 (27)	70 (4)	378 (23)	1,650
2004	838 (39)	560 (26)	127 (6)	605 (28)	2,130
2005	500 (44)	303 (27)	123 (11)	206 (18)	1,132
2006	1,169 (39)	880 (30)	231 (8)	688 (23)	2,968
2007	753 (50)	367 (24)	66 (4)	323 (21)	1,509
2008	3,543 (56)	971 (15)	148 (2)	1,696 (27)	6,358
<i>Average</i>	<i>1,548 (63)</i>	<i>364 (16)</i>	<i>84 (4)</i>	<i>451 (18)</i>	<i>2,448</i>

## Straying

### Normal subyearling releases

Rates of Turtle Rock summer Chinook (normal subyearling releases) straying into spawning areas in the upper basin have been low. Although Turtle Rock summer Chinook have strayed



into other spawning areas, they made up less than 5% of the spawning escapement within those areas (Table 11.21). The Chelan tailrace has received the largest number of Turtle Rock strays. This hatchery program was discontinued in 2010.

**Table 11.21.** Number (No.) and percent of spawning escapements within other non-target basins that consisted of Turtle Rock summer Chinook (normal subyearling releases), return years 1998-2011. For example, for return year 2003, 0.6% of the summer Chinook spawning escapement in the Okanogan River basin consisted of Turtle Rock summer Chinook. Percent strays should be less than 5%.

Return year	Wenatchee		Methow		Okanogan		Chelan		Entiat		Hanford Reach	
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%
1998	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1999	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2000	8	0.1	3	0.3	13	0.4	63	9.5	0	0.0	0	0.0
2001	0	0.0	5	0.2	13	0.1	0	0.0	0	0.0	0	0.0
2002	0	0.0	0	0.0	13	0.1	0	0.0	0	0.0	0	0.0
2003	7	0.1	7	0.2	19	0.6	6	1.4	0	0.0	0	0.0
2004	5	0.0	4	0.2	13	0.2	6	1.4	0	0.0	0	0.0
2005	5	0.1	0	0.0	5	0.1	0	0.0	2	0.5	0	0.0
2006	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2007	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2008	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2009	0	0.0	16	0.9	0	0.0	2	0.3	9	3.6	0	0.0
2010	0	0.0	26	1.0	0	0.0	0	0.0	14	3.2	0	0.0
2011	0	0.0	14	0.5	0	0.0	34	2.7	0	0.0	0	0.0
<i>Average</i>	<i>2</i>	<i>0.0</i>	<i>5</i>	<i>0.2</i>	<i>5</i>	<i>0.1</i>	<i>8</i>	<i>1.1</i>	<i>2</i>	<i>0.5</i>	<i>0</i>	<i>0.0</i>

On average, about 31% of the brood year returns have strayed into spawning areas in the upper basin (Table 11.22). Depending on brood year, percent strays into spawning areas have ranged from 0-100%. Few (2.5% on average) have strayed into non-target hatchery programs.

**Table 11.22.** Number and percent of Turtle Rock summer Chinook (normal subyearling releases) that homed to the target hatchery and strayed to non-target spawning areas and non-target hatchery programs, by brood years 1995-2008.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1995	-	-	197	74.1	64	24.1	5	1.9
1996	-	-	54	54.5	44	44.4	1	1.0
1997	-	-	2	28.6	5	71.4	0	0.0
1998	-	-	0	0.0	24	100.0	0	0.0
1999	-	-	40	43.5	52	56.5	0	0.0
2000	-	-	5	50.0	5	50.0	0	0.0

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
2001	-	-	56	77.8	16	22.2	0	0.0
2002	-	-	10	100.0	0	0.0	0	0.0
2003	-	-	27	100.0	0	0.0	0	0.0
2004	-	-	71	97.3	2	2.7	0	0.0
2005	-	-	80	92.0	7	8.0	0	0.0
2006	-	-	194	72.1	72	26.8	3	1.1
2007	-	-	113	68.5	34	20.6	18	10.9
2008	-	-	16	80.0	0	0.0	4	20.0
<i>Average</i>	-	-	<i>62</i>	<i>67.0</i>	<i>23</i>	<i>30.5</i>	<i>2</i>	<i>2.5</i>

\* Homing to the target hatchery includes Turtle Rock hatchery fish that were captured and included as broodstock in the Turtle Rock Hatchery program. These hatchery fish were typically collected at Wells Dam and Wells Hatchery.

### *Accelerated subyearling releases*

Rates of Turtle Rock summer Chinook (accelerated subyearling releases) straying into spawning areas in the upper basin have been low. Although Turtle Rock summer Chinook have strayed into other spawning areas, they made up less than 5% of the spawning escapement within those areas (Table 11.23). The Chelan tailrace, Entiat Basin, and Methow River basin have received the largest numbers of Turtle Rock strays. This hatchery program was discontinued in 2010.

**Table 11.23.** Number (No.) and percent of spawning escapements within other non-target basins that consisted of Turtle Rock summer Chinook (accelerated subyearling releases), return years 1998-2011. For example, for return year 2001, 0.2% of the summer Chinook spawning escapement in the Methow River basin consisted of Turtle Rock summer Chinook. Percent strays should be less than 5%.

Return year	Wenatchee		Methow		Okanogan		Chelan		Entiat		Hanford Reach	
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%
1998	3	0.1	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
1999	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2000	7	0.1	0	0.0	0	0.0	24	3.6	0	0.0	0	0.0
2001	0	0.0	12	0.4	31	0.3	0	0.0	0	0.0	0	0.0
2002	0	0.0	5	0.1	0	0.0	0	0.0	0	0.0	0	0.0
2003	0	0.0	45	1.1	0	0.0	22	5.3	13	1.9	16	0.0
2004	0	0.0	7	0.3	0	0.0	14	3.3	0	0.0	18	0.0
2005	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2006	0	0.0	0	0.0	0	0.0	0	0.0	2	0.3	0	0.0
2007	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
2008	0	0.0	7	0.4	0	0.0	27	5.4	0	0.0	0	0.0
2009	19	0.2	0	0.0	0	0.0	2	0.3	0	0.0	0	0.0
2010	0	0.0	19	0.8	0	0.0	0	0.0	10	2.3	0	0.0
2011	17	0.2	10	0.3	10	0.1	0	0.0	4	0.9	0	0.0

Return year	Wenatchee		Methow		Okanogan		Chelan		Entiat		Hanford Reach	
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%
<i>Average</i>	3	0.0	8	0.2	3	0.0	6	1.3	2	0.4	2	0.0

On average, about 28% of the brood year returns have strayed into spawning areas in the upper basin (Table 11.24). Depending on brood year, percent strays into spawning areas have ranged from 0-83%. Few (1% on average) have strayed into non-target hatchery programs.

**Table 11.24.** Number and percent of Turtle Rock summer Chinook (accelerated subyearling releases) that homed to the target hatchery and strayed to non-target spawning areas and non-target hatchery programs, by brood years 1995-2008.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1995	-	-	7	70.0	3	30.0	0	0.0
1996	-	-	33	32.4	69	67.6	0	0.0
1997	-	-	6	100.0	0	0.0	0	0.0
1998	-	-	2	16.7	10	83.3	0	0.0
1999	-	-	138	54.1	117	45.9	0	0.0
2000	-	-	12	40.0	18	60.0	0	0.0
2001	-	-	57	96.6	2	3.4	0	0.0
2002	-	-	0	0.0	0	0.0	0	0.0
2003	-	-	3	100.0	0	0.0	0	0.0
2004	-	-	90	75.6	29	24.4	0	0.0
2005	-	-	64	75.3	19	22.4	2	2.4
2006	-	-	88	88.9	7	7.1	4	4.0
2007	-	-	133	61.9	70	32.6	12	5.6
2008	-	-	21	84.0	2	8.0	2	8.0
<i>Average</i>	-	-	<i>47</i>	<i>64.0</i>	<i>25</i>	<i>27.5</i>	<i>1</i>	<i>1.4</i>

\* Homing to the target hatchery includes Turtle Rock hatchery fish that were captured and included as broodstock in the Turtle Rock Hatchery program. These hatchery fish were typically collected at Wells Dam and Wells Hatchery.

### Yearling releases

Rates of Turtle Rock/Chelan Falls summer Chinook (yearling releases) straying into spawning areas in the upper basin have varied widely depending on spawning area. Most of these fish strayed to spawning areas within the Chelan tailrace, Entiat Basin, and Methow River basin. On average, Turtle Rock summer Chinook have made up 5-15% of the spawning escapement within those basins (Table 11.25). Relatively few, on average, have strayed to spawning areas in the Okanogan River basin, Wenatchee River basin, and the Hanford Reach (i.e., they made up less than 5% of the spawning escapement in these areas).

**Table 11.25.** Number (No.) and percent of spawning escapements within other non-target basins that consisted of Turtle Rock/Chelan Falls summer Chinook (yearling releases), return years 1998-2011. For example, for return year 2003, 4.3% of the summer Chinook spawning escapement in the Methow River basin consisted of Turtle Rock summer Chinook. Percent strays should be less than 5%.

Return year	Wenatchee		Methow		Okanogan		Chelan		Entiat		Hanford Reach	
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%
1998	0	0.0	2	0.3	0	0.0	0	0.0	0	0.0	0	0.0
1999	3	0.1	2	0.2	0	0.0	0	0.0	0	0.0	0	0.0
2000	18	0.3	57	4.8	167	4.5	73	11.0	0	0.0	10	0.0
2001	109	1.0	523	18.9	334	3.1	316	32.1	0	0.0	7	0.0
2002	92	0.6	437	9.4	194	1.4	191	32.8	136	27.1	0	0.0
2003	64	0.5	170	4.3	14	0.4	165	39.4	180	26.0	9	0.0
2004	10	0.1	55	2.5	116	1.7	75	17.9	0	0.0	0	0.0
2005	5	0.1	73	2.9	78	0.9	88	19.8	46	12.5	0	0.0
2006	0	0.0	100	3.7	25	0.3	64	15.2	9	1.6	0	0.0
2007	0	0.0	65	4.8	31	0.7	40	21.2	20	8.2	19	0.1
2008	18	0.3	72	3.7	60	0.9	110	22.1	46	14.4	0	0.0
2009	8	0.1	95	5.4	32	0.4	5	0.8	18	7.2	0	0.0
2010	12	0.2	105	4.2	111	1.9	0	0.0	30	6.9	0	0.0
2011	8	0.1	88	3.0	35	0.4	15	1.2	2	0.4	0	0.0
<b>Average</b>	<b>25</b>	<b>0.2</b>	<b>132</b>	<b>4.9</b>	<b>86</b>	<b>1.2</b>	<b>82</b>	<b>15.2</b>	<b>35</b>	<b>7.5</b>	<b>3</b>	<b>0.0</b>

On average, about 48% of the brood year returns have strayed into spawning areas in the upper basin (Table 11.26). Depending on brood year, percent strays into spawning areas have ranged from 14-86%. Few (<2% on average) have strayed into non-target hatchery programs.

**Table 11.26.** Number and percent of Turtle Rock/Chelan Falls summer Chinook (yearling releases) that homed to the target hatchery and strayed to non-target spawning areas and non-target hatchery programs, by brood years 1995-2008.

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
1995	-	-	180	39.3	278	60.7	0	0.0
1996	-	-	218	27.2	583	72.8	0	0.0
1997	-	-	254	14.2	1,531	85.6	3	0.2
1998	-	-	166	16.1	864	83.8	1	0.1
1999	-	-	181	42.7	243	57.3	0	0.0
2000	-	-	102	29.1	249	70.9	0	0.0
2001	-	-	389	59.8	261	40.2	0	0.0
2002	-	-	303	57.8	220	42.0	1	0.2
2003	-	-	373	62.9	219	36.9	1	0.2

Brood year	Homing				Straying			
	Target stream		Target hatchery*		Non-target streams		Non-target hatcheries	
	Number	%	Number	%	Number	%	Number	%
2004	-	-	287	56.6	219	43.2	1	0.2
2005	149	29.4	202	39.9	144	28.5	11	2.2
2006	429	40.4	376	35.4	220	20.7	36	3.4
2007	123	28.2	218	50.0	63	14.4	32	7.3
2008	873	47.1	658	35.5	255	13.8	66	3.6
<b>Average</b>	<b>394</b>	<b>36.3</b>	<b>279</b>	<b>40.5</b>	<b>382</b>	<b>47.9</b>	<b>11</b>	<b>1.2</b>

\* Homing to the target hatchery includes Turtle Rock/Chelan Falls hatchery fish that were captured and included as broodstock in the Turtle Rock/Chelan Falls Hatchery program. These hatchery fish are typically collected at Wells Dam, Wells Hatchery, and the Eastbank Hatchery Outfall.

### Post-Release Survival and Travel Time

We used PIT-tagged fish to estimate survival rates and travel times (arithmetic mean days) of hatchery summer Chinook from the Turtle Rock/Chelan River release sites to McNary Dam, and smolt to adult ratios (SARs) from release to detection at Bonneville Dam (Table 10.27).<sup>15</sup> Over the six brood years for which PIT-tagged hatchery fish were released, survival rates from the release sites to McNary Dam ranged from 0.552 to 0.722; SARs from release to detection at Bonneville Dam ranged from 0.009 to 0.028. Average travel times from release sites to McNary Dam ranged from 15 to 28 days.

Much of the variation in survival rates and travel time among brood years resulted from releases of different experimental groups (Table 10.27). For example, brood years 2007 and 2008 were each split into two experimental groups (Circular Reuse group and Standard Raceway group). For both brood years, survival from the release site to McNary Dam and SARs appeared to be greater for the Circular Reuse fish than for the Standard Raceway fish. However, the differences between groups were small for brood year 2008. For both brood years, travel time from release to McNary Dam appeared to be longer for the Standard Raceway fish than for the Circular Reuse fish.

Another experiment was conducted with brood year 2012 (Table 10.27). That brood year was split into two different treatment groups (small-size fish and large-size fish). The small-size fish appeared to have a higher survival rate to McNary Dam and faster travel time than did the large-size fish. SARs for these fish will be calculated after all fish have returned to the Columbia River.

<sup>15</sup> It is important to point out that because of fish size differences among rearing tanks or raceways, fish PIT tagged in one tank or raceway may not represent untagged fish rearing in other tanks or raceways.

**Table 10.27.** Total number of Turtle Rock/Chelan Falls yearling summer Chinook released with PIT tags, their survival and travel times (mean days) to McNary Dam, and smolt-to-adult (SAR) ratios for brood years 2007-2012. Standard errors are shown in parentheses. NA = not available (i.e., not all the fish from the release groups have returned to the Columbia River).

Brood year	Raceway/Program	Number of tagged fish released	Survival to McNary Dam	Travel time to McNary Dam	SAR to Bonneville Dam
2007	Circular Reuse	9,975	0.722 (0.036)	22.4 (8.6)	0.017 (0.001)
	Standard	9,546	0.564 (0.037)	28.4 (11.7)	0.009 (0.001)
2008	Circular Reuse	11,082	0.631 (0.040)	26.5 (9.8)	0.028 (0.002)
	Standard	11,070	0.581 (0.038)	27.9 (18.7)	0.025 (0.001)
2009	Turtle Rock	4,945	0.603 (0.061)	15.4 (8.6)	0.018 (0.002)
	Chelan Net Pens	5,048	0.616 (0.059)	19.5 (10.2)	0.012 (0.002)
2010	Chelan Falls	4,186	0.655 (0.050)	22.5 (12.1)	NA
2011*	Chelan Falls	4,075	0.552 (0.054)	27.2 (11.5)	NA
2012	Chelan Falls (Small Fish)	4,983	0.590 (0.049)	25.0 (11.2)	NA
	Chelan Falls (Big Fish)	4,960	0.578 (0.043)	24.4 (10.1)	NA

\* Brood year 2011 experienced high mortality due to fungus, bacterial cold-water disease, bacterial gill disease, and erythrocytic inclusion body syndrome during April 2013.

### Smolt-to-Adult Survivals

Subyearling-to-adult and smolt-to-adult survival ratios (SARs) were calculated as the number of hatchery adult recaptures divided by the number of tagged hatchery subyearling or yearling Chinook released. For these analyses, SARs were based on CWT returns.

#### Normal subyearling releases

For the available brood years, SARs for normal subyearling-released Chinook have ranged from 0.000034 to 0.001886 (Table 11.28). This hatchery program was discontinued in 2010.

**Table 11.28.** Subyearling-to-adult ratios (SARs) for Turtle Rock normal subyearling-released summer Chinook, brood years 1995-2008.

Brood year	Number released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1995	201,230	204	0.001014
1996	371,848	188	0.000506
1997	496,904	17	0.000034
1998	194,723	28	0.000144
1999	197,793	203	0.001026
2000	222,460	28	0.000126
2001	211,306	330	0.001562
2002	200,163	38	0.000190

Brood year	Number released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
2003	203,410	49	0.000241
2004	198,019	91	0.000460
2005	197,135	143	0.000725
2006	188,250	355	0.001886
2007	194,437	216	0.001111
2008	152,993	73	0.000477
<b>Average</b>	<b>230,762</b>	<b>140</b>	<b>0.000679</b>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

### *Accelerated subyearling releases*

For the available brood years, SARs for accelerated subyearling-released Chinook have ranged from 0.000011 to 0.004609 (Table 11.29). This hatchery program was discontinued in 2010.

**Table 11.29.** Subyearling-to-adult ratios (SARs) for Turtle Rock accelerated subyearling-released summer Chinook, brood years 1995-2008.

Brood year	Number released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1995	166,203	13	0.000078
1996	198,720	79	0.000398
1997	196,459	3	0.000015
1998	185,551	69	0.000372
1999	192,665	888	0.004609
2000	194,603	63	0.000324
2001	196,355	167	0.000851
2002	200,165	5	0.000025
2003	185,834	2	0.000011
2004	203,255	156	0.000768
2005	192,045	82	0.000427
2006	186,324	217	0.001165
2007	188,328	299	0.001588
2008	197,136	32	0.000162
<b>Average</b>	<b>191,689</b>	<b>148</b>	<b>0.000771</b>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

### *Yearling releases*

For the available brood years, SARs for yearling-released Chinook have ranged from 0.007212 to 0.028185 (Table 11.30).

**Table 11.30.** Smolt-to-adult ratios (SARs) for Turtle Rock/Chelan Falls yearling-released summer Chinook, brood years 1995-2008.

Brood year	Number released <sup>a</sup>	Estimated adult captures <sup>b</sup>	SAR
1995	145,318	1,048	0.007212
1996	194,251	1,553	0.007995
1997	198,924	4,776	0.024009
1998	215,646	5,772	0.026766
1999	280,683	2,670	0.009513
2000	278,308	2,029	0.007290
2001	199,694	3,905	0.019555
2002	192,234	2,523	0.013125
2003	199,386	2,092	0.010492
2004	202,682	2,605	0.012853
2005	202,329	1,630	0.008056
2006	142,699	4,022	0.028185
2007	161,071	1,841	0.011430
2008	353,450	8,144	0.023041
<b>Average</b>	<b>211,905</b>	<b>3,186</b>	<b>0.014966</b>

<sup>a</sup> Includes all tag codes and CWT released fish (CWT + Ad Clip fish and CWT-only fish).

<sup>b</sup> Includes estimated recoveries (spawning ground, hatcheries, harvest, etc.) and observed recoveries if estimated recoveries were unavailable.

## 11.6 ESA/HCP Compliance

### Broodstock Collection

The 2012 brood Chelan Falls (formerly Turtle Rock) summer Chinook program is supported through adult collections at the volunteer trap at Wells Fish Hatchery and in conjunction with the Wells summer Chinook collections. During 2012, broodstock collections at the volunteer trap were consistent with the 2012 Upper Columbia River Salmon and Steelhead Broodstock Objectives and site-based broodstock collection protocols as required in ESA permit 1347. The 2012 collection target totaled 1,287 summer Chinook (including 318 for the Chelan Falls program).

### Hatchery Rearing and Release

Brood year 2012 releases totaled 566,188 yearling fish. These releases represented 98.3% of the 576,000 Rocky Reach HCP and ESA Section 10 Permit 1347 production for the Chelan Falls yearling summer Chinook production.

### Hatchery Effluent Monitoring

Per ESA Permits 1196, 1347, 1395, 18118, 18119, and 18121, permit holders shall monitor and report hatchery effluents in compliance with applicable National Pollution Discharge Elimination



Systems (NPDES) (EPA 1999) permit limitations. There was one NPDES violation reported at PUD Hatchery facilities during the period 1 January through 31 December 2014. NPDES monitoring and reporting for Chelan PUD Hatchery Programs during 2014 are provided in Appendix F.



## SECTION 12: REFERENCES

- Blankenship, S., J. Von Bargen, K. Warheit, and A. Murdoch. 2007. Assessing the genetic diversity of natural Chiwawa River spring Chinook salmon and evaluating the effectiveness of its supportive hatchery supplementation program. Washington Department of Fish and Wildlife Molecular Genetics Lab, Olympia, WA.
- Environmental Protection Agency (EPA). 1999. National pollutant discharge elimination systems (NPDES) permit program.
- Ford, M., A. Murdoch, and T. Maitland. 2010. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 46273 and 46489, Department of Energy, Bonneville Power Administration, Portland, OR.
- Ford, M., S. Villagecenter, A. Murdoch, and M. Hughes. 2011. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 46273 and 46489, Department of Energy, Bonneville Power Administration, Portland, OR.
- Ford, M., S. Howard, A. Murdoch, and M. Hughes. 2012. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 46273 and 46489, Department of Energy, Bonneville Power Administration, Portland, OR.
- Ford, M., S. Howard, A. Murdoch, and M. Hughes. 2013. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 46273 and 46489, Department of Energy, Bonneville Power Administration, Portland, OR.
- Ford, M., A. Murdoch, and M. Hughes. 2014. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 46273 and 46489, Department of Energy, Bonneville Power Administration, Portland, OR.
- Ford, M., A. Murdoch, and M. Hughes. 2015. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 46273 and 46489, Department of Energy, Bonneville Power Administration, Portland, OR.
- Ford, M., A. Murdoch, and M. Hughes. 2015. Using parentage analysis to estimate rates of straying and homing in Chinook salmon (*Oncorhynchus tshawytscha*). *Molecular Ecology* 24:1109-1121.
- Ford, M., T. Pearsons, and A. Murdoch. 2015. The spawning success of early maturing resident hatchery Chinook salmon in a natural river system. *Transactions of the American Fisheries Society* 144:539-548.
- Hillman, T., J. Mullan, and J. Griffith. 1992. Accuracy of underwater counts of juvenile Chinook salmon, coho salmon, and steelhead. *North American Journal of Fisheries Management* 12:589-603.

- Hillman, T. and M. Miller. 2004. Abundance and total numbers of Chinook salmon and trout in the Chiwawa River Basin, Washington, 2004. BioAnalysts, Inc. Report to Chelan County PUD, Wenatchee, WA.
- Hillman, T., M. Miller, T. Miller, M. Tonseth, M. Hughes, A. Murdoch, J. Miller, and B. Kessee. 2011. Monitoring and evaluation of the Chelan County PUD hatchery programs: 2010 annual report. Report to the HCP Hatchery Committee, Wenatchee, WA.
- Hillman, T., M. Miller, A. Murdoch, T. Miller, J. Murauskas, S. Hays, and J. Miller. 2012. Monitoring and evaluation of the Chelan County PUD hatchery programs: five-year (2006-2010) report. Report to the HCP Hatchery Committee, Wenatchee, WA.
- Hillman, T., T. Kahler, G. Mackey, J. Murauskas, A. Murdoch, K. Murdoch, T. Pearsons, and M. Tonseth. 2013. Updated monitoring and evaluation plan for PUD hatchery programs. Report to the Hatchery Committees, Wenatchee, East Wenatchee, and Ephrata, WA.
- HSRG/WDFW/NWIFC. 2004. Integrated hatchery programs. HSRG/WDFW/NWIFC Technical discussion paper #1, 21 June 2004, Portland, OR.
- Hyatt, K., M. Stockwell, H. Wright, K. Long, J. Tamblyn, and M. Walsh. 2006. Fish and water management tool project assessments: Okanogan adult sockeye salmon (*Oncorhynchus nerka*) abundance and biological traits in 2005. Draft report to JSID-SRE 3-05, Salmon and Freshwater Ecosystems Division, Fisheries and Oceans Canada, Nanaimo, B.C.
- Kassler, W., S. Blankenship, and A. Murdoch. 2011. Genetic structure of upper Columbia River summer Chinook and evaluation of the effects of supplementation programs. Washington Department of Fish and Wildlife Molecular Genetics Lab, Olympia, WA.
- Keller, L. and J. Murauskas. 2012. 2011 Wenatchee River basin summer Chinook and sockeye salmon spawning ground surveys. Appendix H in: Hillman et al. 2012. Monitoring and evaluation of the Chelan County PUD hatchery programs: 2011 annual report. Report to the HCP Hatchery Committee, Wenatchee, WA.
- Lauver, E., T. Pearsons, R. Langshaw, and S. Lowry. 2012. White River spring Chinook salmon captive-brood program 2011 annual summary report. Public Utility District No. 2 of Grant County, Ephrata, WA.
- Mackey, G., T. Pearsons, M. Cooper, K. Murdoch, A. Murdoch, and T. Hillman. 2014. Ecological risk assessment of upper Columbia hatchery programs on non-target taxa of concern. Report produced by the Hatchery Evaluation Technical Team (HETT) for the HCP Wells Hatchery Committee, HCP Rocky Reach Hatchery Committee, HCP Rock Island Hatchery Committee, and the Priest Rapids Hatchery Sub-Committee. Grant County Public Utility District, Ephrata, Washington.
- McElhany, P., M. Ruckelshaus, M. Ford, T. Wainwright, and E. Bjorkstedt. 2000. Viable salmonid populations and the recovery of Evolutionarily Significant Units. NOAA Technical Memorandum.
- Millar, R., S. McKechnie, and C. Jordan. 2012. Simple estimators of salmonid escapement and its variance using a new area-under-the-curve method. *Canadian Journal of Fisheries and Aquatic Sciences* 69:1002-1015.

- Miller, T. 2008. 2007 Chiwawa and Wenatchee River smolt estimates. Technical memorandum from Todd Miller, WDFW to the HCP Hatchery Committee, 13 February 2008, Wenatchee, WA.
- Miller, T. and M. Tonseth. 2008. The integrated status and effectiveness monitoring program: expansion of smolt trapping and steelhead spawning survey. Annual report to the U.S. Department of Energy, Bonneville Power Administration, Division of Fish and Wildlife, Portland, OR.
- Murdoch, A. and C. Peven. 2005. Conceptual approach to monitoring and evaluating the Chelan County Public Utility District Hatchery Program. Final Report for the Chelan PUD Habitat Conservations Plan's Hatchery Committees, Wenatchee, WA.
- Murdoch, A., T. Pearsons, T. Maitland, M. Ford, and K. Williamsons. 2009. Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 00032138, Department of Energy, Bonneville Power Administration, Portland, OR.
- NMFS (National Marine Fisheries Service). 2003. Section 10(a)(1)(b) Permit for takes of endangered/threatened species. Incidental Take Permit 1347 for the artificial propagation of unlisted salmon. Portland, OR.
- NMFS (National Marine Fisheries Service). 2004. Biological Opinion and Magnuson-Stevens Fishery Conservation and Management Act Consultation Interim Protection Plan for Operation of the Priest Rapids Hydroelectric Project FERC Project No. 2114 Columbia River, Grant and Kittitas Counties, Washington Action Agency: Federal Energy Regulatory Commission Consultation Conducted by: NOAA Fisheries Northwest Region Hydropower Division NOAA Fisheries Log Number: 1999/01878. May 3, 2004.
- Pearsons, T., A. Murdoch, G. Mackey, K. Murdoch, T. Hillman, M. Cooper, and J. Miller. 2012. Ecological risk assessment of multiple hatchery programs in the upper Columbia watershed using Delphi and modeling approaches. *Environmental Biology of Fishes* 94:87-100. DOI 10.1007/s10641-011-9884-1.
- Seamons, T., S. Young, C. Bowman, K. Warheit, and A. Murdoch. 2012. Examining the genetic structure of Wenatchee River basin steelhead and evaluating the effects of the supplementation program. Washington Department of Fish and Wildlife Molecular Genetics Lab, Olympia, WA.
- Snow, C., C. Frady, A. Repp, A. Murdoch, M. Small, and C. Dean. 2013. Monitoring and evaluation of Wells and Methow Hatchery Programs: 2012 annual report. Washington Department of Fish and Wildlife. Prepared for Douglas County Public Utility District and the Wells HCP Hatchery Committee, East Wenatchee, WA.
- Tonseth, M. and T. Maitland. 2011. White River spring Chinook salmon captive broodstock program, 2010 annual activity report. Washington Department of Fish and Wildlife, Wenatchee, WA.
- Truscott, K. 2005. Memo to Habitat Conservation Plan (HCP) Hatchery Committee (HC). Brood year 2005-2013 Upper Columbia steelhead stocking allotments for releases in the Wenatchee River basin. February 28, 2005 memo from K. Truscott, Washington Department of Fish and Wildlife, Wenatchee, WA.

Truscott, B., A. Murdoch, J. Cram and K. See. 2015. Upper Columbia spring Chinook salmon and steelhead juvenile and adult abundance, productivity, and spatial scale monitoring. Project # 2010-034-00. Bonneville Power Administration, Portland OR.

<https://pisces.bpa.gov/release/documents/DocumentViewer.aspx?doc=P142786>

Washington Department of Fish and Wildlife (WDFW). 2006. Memo to Habitat Conservation Plan (HCP) Hatchery Committee (HC). 2006 Upper Columbia River salmon and steelhead broodstock objectives and site-based broodstock collection protocols. Memo from K. Truscott, Washington Department of Fish and Wildlife, Wenatchee, WA.

## SECTION 13: APPENDICES

- Appendix A:** Abundance and Total Numbers of Chinook Salmon and Trout in the Chiwawa River Basin, Washington, 2014.
- Appendix B:** Fish Trapping at the Chiwawa and Wenatchee Smolt Traps during 2014.
- Appendix C:** Summary of CSS PIT-Tagging Activities in the Wenatchee River Basin, 2014.
- Appendix D:** Wenatchee Steelhead Spawning Escapement Estimates, 2014.
- Appendix E:** Examining the Genetic Structure of Wenatchee River Basin Steelhead and Evaluating the Effects of the Supplementation Program.
- Appendix F:** NPDES Hatchery Effluent Monitoring, 2014.
- Appendix G:** Steelhead Stock Assessment at Priest Rapids Dam, 2014.
- Appendix H:** Wenatchee Sockeye Salmon Spawning Escapement, 2014.
- Appendix I:** Genetic Diversity of Wenatchee Sockeye Salmon.
- Appendix J:** Genetic Diversity of Natural Chiwawa River Spring Chinook Salmon.
- Appendix K:** Fish Trapping at the Nason Creek Smolt Trap during 2014.
- Appendix L:** Fish Trapping at the White River Smolt Trap during 2014.
- Appendix M:** Genetic Diversity of Upper Columbia Summer Chinook Salmon.
- Appendix N:** Summer Chinook Spawning Ground Surveys in the Methow and Chelan Rivers, 2014.





# Appendix A

**Abundance and Total Numbers of Chinook Salmon and Trout in the  
Chiwawa River basin, Washington, 2014**





January 25, 2015

TO: HCP Hatchery Committee

FROM: Tracy Hillman

**Subject: Abundance and Total Numbers of Chinook Salmon and Trout in the Chiwawa River basin, Washington, 2014**

The Chelan County Public Utility District (PUD) hatchery program is operated through a habitat conservation program (HCP) that was incorporated into the PUD's license in 2004. The HCP directed the signatories to develop a monitoring and evaluation plan within one year of the effective date. This resulted in the development of the Conceptual Approach to Monitoring and Evaluating the Chelan County Public Utility District Hatchery Programs (Murdoch and Pevan 2005). In 2013, the Hatchery Committees updated the hatchery monitoring and evaluation plan (Hillman et al. 2013). This study will help the Hatchery Committees determine if it is meeting Objective 2 in the updated monitoring and evaluation plan.

***Objective 2:** Determine if the proportion of hatchery fish on the spawning grounds affects the freshwater productivity of supplemented stocks.*

We estimated densities and total numbers of age-0 spring Chinook salmon *Oncorhynchus tshawytscha*, trout *Oncorhynchus* sp., and char *Salvelinus* sp. in the Chiwawa River basin, Washington, in August 2014. This was the 22<sup>nd</sup> year of an ongoing study to assess the freshwater productivity (juveniles/redd) of Chinook salmon in the Chiwawa River basin. We used landscape classification to stratify streams in the basin that supported juvenile Chinook salmon (Hillman and Miller 2004). Classification "explained" most of the variability in fish numbers caused by geology, land type, valley bottom type, stream state condition, and habitat type. We identified ten reaches on the lower 31 miles (50 km) of the Chiwawa River and one reach in each of Phelps, Rock, Chikamin, Big Meadow, Alder, Brush, Clear, Y, and Unnamed<sup>1</sup> creeks (Figure 1). Each reach consisted of several combinations of state-type and habitat-type strata. We used classification to find reference areas for reaches in the Chiwawa River. We matched Reach 3 and Reach 8 of the Chiwawa River with a moderately-confined section of Nason Creek (RM 0.62-1.70) and an unconfined area of the Little Wenatchee River (RM 4.39-8.55), respectively

---

<sup>1</sup>Unnamed tributary that drains the eastside of Chiwawa Ridge. Its confluence with the Chiwawa River is about 1 mile (1.6 km) downstream from the mouth of Phelps Creek.

(Hillman and Miller 2004). Because of the supplementation program in Nason Creek, the use of Nason Creek as a reference for the Chiwawa River is no longer valid. However, as directed by the Hatchery Committee, we continue to sample sites in Nason Creek. Following methods described in Hillman and Miller (2004), we used underwater observations to estimate numbers of fish in 179 randomly selected sites.

During sampling in August 2014, discharge in the Chiwawa River averaged 250 cubic feet per second (cfs) and ranged from 165-363 cfs (Figure 2). Stream temperatures during the study period ranged from 10.0 to 18.0°C. Fish species observed in the Chiwawa River basin and reference areas during the 1992-2014 survey period<sup>2</sup> included: spring Chinook salmon, coho salmon *O. kisutch*, sockeye salmon *O. nerka* (in the Little Wenatchee River reference area), steelhead/rainbow trout *O. mykiss* (hatchery rainbow were present only in 1992 and 1993), cutthroat trout *O. clarki lewisi*, bull trout *S. confluentus*, brook trout *S. fontinalis*, mountain whitefish *Prosopium williamsoni*, dace *Rhinichthys* sp., suckers *Catostomus* sp., and sculpin *Cottus* sp. The age-0 spring Chinook that we observed in the Chiwawa River basin during the 2014 survey were produced from 714 redds counted in the fall of 2013 (Hillman et al. 2014). Assuming a mean fecundity of 4,716 eggs per female Chinook (from females collected for broodstock), and that no female produced more than one redd (Murdoch et al. 2009), we estimated that the Chiwawa River basin was seeded with 3,367,224 eggs in 2013 (Appendix A).

In 2014, riffles made up the largest fraction of habitat types in reaches of the Chiwawa River basin (54% of the total stream surface area) (Table 1). Pools (22%), glides (7%), and multiple channels (17%) constituted the remaining 46% of the stream surface area. We consistently found woody debris associated with multiple-channel habitat.

### **Chinook Salmon Abundance**

Chinook salmon were the most abundant salmonid in the Chiwawa River basin. We estimated, based on surface area, that age-0 Chinook salmon numbered 121,240 ( $\pm 11\%$  of the estimated total) in the Chiwawa River basin in August 2014 (Table 2). Extrapolating based on volume of habitat types, age-0 Chinook numbered 122,706 ( $\pm 11\%$ ) in the Chiwawa River basin. About 6% of the juvenile Chinook were in tributaries to the Chiwawa River. During the 1992-2014 surveys, numbers of age-0 Chinook ranged from 5,815 to 149,563 in the Chiwawa River basin (Figure 3; Appendix A and B). Most of the difference in juvenile numbers among years resulted from different seeding (stock) levels (Figure 4). Numbers of Chinook redds in the Chiwawa River basin during 1992-2014 ranged from 13 to 1,078, resulting in seeding levels of 66,248 to 4,984,672 eggs (Appendix A).

As in most years, age-0 Chinook in 2014 were distributed contagiously among reaches in the Chiwawa River (Table 2). In the Chiwawa River, densities of age-0 Chinook were highest in the upper reaches (Reaches 7-10). The highest densities in the Chiwawa River basin were in tributaries to the Chiwawa River (Table 2). Age-0 Chinook were most abundant in multiple channels and least abundant in glides and riffles. We found the majority of the Chinook

---

<sup>2</sup> The study period 1992-2014 includes only 22 years of sampling because there was no sampling in 2000.

associated with woody debris in multiple channels (multiple channel use index = 2.76)<sup>3</sup>. These sites (multiple channels) made up 17% of the total surface area of the Chiwawa River basin, but they provided habitat for 52% of all the age-0 Chinook in the basin in 2014 (Appendix C). In contrast, riffles made up 54% of the total surface area, but provided habitat for only 11% of all age-0 Chinook in the Chiwawa River basin (riffle use index = 0.25). Pools made up 22% of the total surface area and provided habitat for 37% of all age-0 Chinook in the basin (pool use index = 1.60). Few Chinook used glides that lacked woody debris (glide use index = 0.27).

As noted earlier, we assumed that the Chiwawa River was seeded with 3,367,224 Chinook eggs (714 redds times 4,716 eggs/female) in fall, 2013, and that at least 121,240 of those survived to August 2014. This means that the egg-to-parr survival was at least 3.6% (95% confidence bound 3.2-4.0%). During 1992-2014, egg-to-parr survival averaged 8.2% (range 2.7-19.1%) in the Chiwawa River basin (Appendix A). This survival rate comports with those from other streams. For example, Mullan et al. (1992) estimated an egg-to-parr survival rate of 9.8% for spring Chinook salmon in Icicle Creek, a tributary of the Wenatchee River. Using a Beverton and Holt model, Hubble (1993) estimated that egg-to-parr survival of Chinook in the Chewuck River, a tributary to the Methow River, ranged between 13% and 32%, depending on percent seeding level in the basin. Kiefer and Forster (1991) estimated a mean egg-to-parr survival rate of 5.5% (range 5.1-6.7%) for naturally-spawning spring Chinook salmon in the entire upper Salmon River. They also noted that egg-to-parr survival of natural spawners and adult outplants in the headwater streams of the upper Salmon River averaged 24.4% (range 16.1-32.0%). Petrosky (1990) reported an egg-to-parr survival range of 1.2-29.0% for Chinook in the upper Salmon River, Idaho. Konopacky et al. (1986) estimated egg-to-parr survival of Chinook in Bear Valley Creek, Idaho, as 8.1-9.4%. Work by Richards and Cerner (1987) in Bear Valley Creek indicated an egg-to-parr survival of 2.1%.

Mean densities of age-0 Chinook salmon in two reaches of the Chiwawa River were generally less than those in corresponding reference areas (Figure 5). Within both the Chiwawa River and its reference areas, pools and multiple channels consistently had the highest densities of age-0 Chinook.

We estimated a total of 939 ( $\pm 28\%$  of the estimated total) age-1+ Chinook salmon in the Chiwawa River basin in August 2014 (Table 3). This was the second highest estimate since the initiation of the study. In August 1992-2014, numbers of age-1+ Chinook ranged from 5 to 967 in the Chiwawa River basin (Figure 3; Appendix B). These fish occurred throughout the Chiwawa River. We found relatively few age-1+ Chinook in tributaries; although, numbers in Phelps and Rock creeks were higher in 2014 than in past years. Age-1+ Chinook were most abundant in multiple channels and pools.

---

<sup>3</sup>The habitat use index was calculated as follows: Multiple channel use =  $(\text{parr}_{mc}/\text{parr}_t) / (\text{area}_{mc}/\text{area}_t)$ , where  $\text{parr}_{mc}$  = the number of parr counted in multiple channel habitat,  $\text{parr}_t$  = the total number of parr counted within all habitat types,  $\text{area}_{mc}$  = the area of multiple channel habitat within the sampling frame, and  $\text{area}_t$  = the total area of the sampling frame. A multiple channel use index value of 1 would indicate that parr were uniformly distributed among habitat types and exhibited no preference for multiple habitat types. Values of the use index greater than 1 indicate use of multiple channels to a greater extent than the average, while scores between 0 and 1 indicate below-average use of multiple channel habitat.

## Juvenile Chinook Salmon Productivity (Fish/Redd)

Freshwater productivity of juvenile Chinook salmon was estimated as the number of parr (age-0 Chinook) per redd in the Chiwawa River basin. Theoretically, the relationship between number of parr and redds can be explained mathematically provided the relationship between the two parameters goes through the origin, increases monotonically at low spawning levels, and shows some level of density dependence at high spawning levels. We identified four alternative hypotheses that may explain the relationship between spawning level (redds) and numbers of age-0 Chinook:

1. The first hypothesis assumed that the number of juveniles increases constantly toward an asymptote as the number of redds increases. After the asymptote is reached, the number of juveniles neither increases nor decreases. The asymptote represents the maximum number of juveniles the system can support (i.e., carrying capacity for the system). This hypothesis was modeled with a Beverton-Holt curve that took the form:

$$J = \frac{(\alpha R)}{(\beta + R)}$$

where  $J$  is the number of juvenile (age-0) Chinook,  $R$  is the number of redds,  $\alpha$  is the maximum number of juveniles produced, and  $\beta$  is the number of redds needed to produce (on average) juveniles equal to one-half the maximum number of juveniles.

2. The second hypothesis, like the first, assumed that the number of juveniles increases toward an asymptote (carrying capacity) as the number of redds increases. After the carrying capacity is reached, the number of juveniles neither increases nor decreases. The carrying capacity represents the maximum number of juveniles the system can support. This hypothesis was modeled with a smooth hockey stick function that took the form:

$$J = J_{\infty} \left( 1 - e^{-\left(\frac{\alpha}{J_{\infty}}\right)R} \right)$$

where  $J$  and  $R$  are as above,  $\alpha$  is the slope at the origin of the spawner-recruitment curve, and  $J_{\infty}$  is the carrying capacity of juveniles.

3. The third hypothesis assumed that the number of juveniles increases to a maximum and then declines as the number of redds increases. In this case, mortality rate of juveniles (or eggs) is proportional to the initial number of redds. Higher mortality rate is associated with density-dependent growth coupled with size-dependent predation. This hypothesis was modeled with a Ricker curve that took the form:

$$J = \alpha R e^{-\beta R}$$

where  $J$  and  $R$  are as above,  $\alpha$  is the number of juveniles per redd at low spawning levels, and  $\beta$  describes how quickly the juveniles per redd drop as the number of redds increases.

4. The fourth hypothesis, like the first, assumed that the number of juveniles increases constantly, but unlike the first, the number of juveniles does not reach an asymptote. Rather, the number of juveniles increases indefinitely, but at a slowing rate of increase. This hypothesis was modeled with both a Cushing curve and a Gamma function. The

Cushing curve took the form:

$$J = \alpha R^\gamma$$

where  $J$  and  $R$  are as above,  $\alpha$  is the number of juveniles per redd at low spawning levels, and  $\gamma$  describes the level of density dependence at high spawning levels. The Gamma function is a three-parameter model that has the form:

$$J = \alpha R^\gamma e^{-\beta R}.$$

This is an un-normalized gamma function that is similar to the Cushing curve when  $\beta = 0$ .

We used Akaike's Information Criterion for small sample size ( $AIC_c$ ) to determine which model(s) best explained the productivity of juvenile Chinook in the Chiwawa River basin.  $AIC_c$  was estimated as:

$$AIC_c = -2\log(\mathcal{L}(\theta|data)) + 2K + \left(\frac{2K(K+1)}{n-K-1}\right)$$

where  $\log(\mathcal{L}(\theta|data))$  is the maximum likelihood estimate,  $K$  is the number of estimable parameters (structural parameters plus the residual variance parameter), and  $n$  is the sample size (Burnham and Anderson 2002). We used least-squares methods to estimate  $\log(\mathcal{L}(\theta|data))$ , which was calculated as  $\log(\sigma^2)$ , where  $\sigma^2$  = residual sum of squares divided by the sample size ( $\sigma^2 = RSS/n$ ).  $AIC_c$  assesses model fit in relation to model complexity (number of parameters). The model with the smallest  $AIC_c$  value represents the "best approximating" model within the model set. Remaining models were ranked relative to the best model using  $AIC_c$  difference scores ( $\Delta AIC_c$ ), Akaike weights ( $w_i$ ), and evidence ratios. Models with  $\Delta AIC_c$  values less than 2 indicate that there is substantial support for these models as being the best-fitting models within the set (Burnham and Anderson 2002). Models with values greater than 2 have less support. Akaike weights are probabilities estimating the strength of the evidence supporting a particular model as being the best model within the model set. Models with small  $w_i$  values are less plausible as competing models (Burnham and Anderson 2002). If no single model could be specified as the best model, a "best subset" of competing models was identified using (1)  $AIC_c$  differences to indicate the level of empirical support each model had as being the best model, (2) evidence ratios based on Akaike weights to indicate the relative probability that any model is the best model, and (3) coefficients of determination ( $R^2$ ) assessing the explanatory power of each model.

The use of  $AIC_c$  indicated that the Beverton-Holt model best approximated the information in the juveniles/redd data (Table 4; Figure 6). The estimated structural parameters for this model were:

$$Juveniles = \frac{(148,016 \times Redds)}{(183 + Redds)}$$

where the bootstrap estimated standard errors for the two parameters were 18,073 and 55, respectively. The adjusted  $R^2 = 0.83$ . The second-best model was the smooth hockey stick model, which was 1.49  $AIC_c$  units from the best model (Table 4; Figure 6). The estimated parameters for this model were:

$$LN(\text{Juveniles}) = 11.6 + LN\left(1 - e^{-\left(\frac{725.2}{113,027}\right)\text{Redds}}\right)$$

where the bootstrap estimated standard errors of the two parameters were 0.1 and 134, respectively, and the  $R^2 = 0.82$ . The  $AIC_c$  difference scores, Akaike weights, and evidence ratios indicated that there was substantial support for both the Beverton-Holt and smooth hockey stick models (Table 4). There was less support for the remaining models (Ricker, Gamma<sup>4</sup>, and Cushing), which were  $> 2 AIC_c$  units from the best models. This was further supported by the fact that, relative to the best models, the remaining models had evidence ratios greater than 10.

Although the Beverton-Holt, smooth hockey stick, and Ricker models have different biological assumptions, they all indicated a density-dependent relationship between spawning levels (redds) and juvenile Chinook production. This was not only evident in the best approximating models, but there was also a significant negative relationship between juveniles per redd and numbers of redds in the Chiwawa River basin (Figure 7). Although data at high seeding levels are lacking, the Beverton-Holt model would limit the capacity of juvenile Chinook to less than about 183,000 parr in the basin (bootstrap upper 95% CI of  $\alpha$  in the Beverton-Holt model). This equates to about 1,443 Chinook parr per hectare. In contrast, the smooth hockey stick model, which fit the data as well as the Beverton-Holt model, would limit the carrying capacity for juvenile Chinook to about 141,000 parr (bootstrap upper 95% CI of  $J_\infty$  in the smooth hockey stick model). This equates to about 1,112 Chinook parr per hectare. As a comparison, Thorson et al. (2013) estimated the carrying capacity for 15 populations of juvenile Chinook in the Snake River metapopulation as 5,000 juveniles per hectare. However, those authors noted that the estimate could be biased because of imperfect detectability and estimates of spawning numbers.

### **Steelhead/Rainbow Abundance**

Based on stream surface area, we estimated a total of 16,083 ( $\pm 10\%$  of the estimated total) age-0 steelhead/rainbow ( $< 4$  in) in reaches of the Chiwawa River basin in August 2014 (Table 5). During the 1992-2014 survey period, numbers of age-0 steelhead/rainbow ranged from 1,410 to 45,727 in the Chiwawa River basin (Figure 8; Appendix B). In 1992-2014, numbers of age-0 steelhead/rainbow varied among reaches, but were typically highest in the lower reaches of the Chiwawa River. In all years they most often used riffle and multiple channel habitats in the Chiwawa River, although we also found them associated with woody debris in pool and glide habitat. In tributaries they were generally most abundant in small pools. Those that we observed in riffles selected stations in quiet water behind small and large boulders or occupied stations in quiet water along the stream margin. In pool and multiple-channel habitats, we found age-0 steelhead/rainbow using the same kinds of habitat as age-0 Chinook salmon.

We estimated that 5,084 ( $\pm 12\%$  of the estimated total) age-1+ steelhead/rainbow (4-8 in) lived in reaches of the Chiwawa River basin in August 2014 (Table 6). During the survey period 1992-2014, numbers of age-1+ steelhead/rainbow ranged from 2,533 to 22,130 (Figure 8; Appendix B). In most years we found these fish in nearly all reaches, but they were typically most numerous in lower reaches of the Chiwawa River. We observed age-1+ steelhead/rainbow

---

<sup>4</sup>The  $\gamma$  parameter in the Gamma model was greater than 0, which means that this model is nearly identical to the Ricker model.



mostly in pool, riffle, and multiple-channel habitats. Those that we observed in pools were usually in deeper water than age-0 steelhead/rainbow and Chinook. Like age-0 steelhead/rainbow, age-1+ steelhead/rainbow selected stations in quiet water behind boulders in riffles, but we generally did not find the two age groups together. Age-1+ steelhead/rainbow appeared to use deeper and faster water than did age-0 steelhead/rainbow.

We estimated that steelhead/rainbow larger than 8 inches numbered 87 ( $\pm 20\%$  of the estimated total) in the Chiwawa River basin in August 2014 (Table 7). During the period 1992-2014, steelhead/rainbow numbers ranged from 8 to 1,869 (Appendix B). Steelhead/rainbow larger than 8 inches were most abundant in the lower Chiwawa River; however, in 1992 and 1993, they were most abundant near campgrounds in Reaches 8, 9, and 10 (these were mostly hatchery fish planted near the campgrounds). We found very few in tributaries. Most of the steelhead/rainbow larger than 8 inches used deep pools ( $>5$  feet), and occupied stations near the bottom at the upstream end of pools.

### **Bull Trout Abundance**

We estimated, based on surface area that at least 259 ( $\pm 25\%$  of the estimated total) juvenile (2-8 in) bull trout lived in reaches of the Chiwawa River basin in August 2014 (Table 8). We found most of these fish in the upper-most reaches of the Chiwawa River and in Rock and Phelps creeks. During 1992-2014, numbers of juvenile bull trout ranged from 79 to 505 (Figure 9; Appendix B). These estimates and those for adult bull trout are incomplete because we did not sample the entire range of bull trout in all tributaries. We did not extend our surveys into the headwaters of the Chiwawa River because there were no juvenile Chinook there. Areas beyond the distribution of juvenile Chinook salmon are known to support bull trout, steelhead/rainbow, and cutthroat trout (USFS 1993). In addition, our estimates of bull trout abundance were based on daytime snorkel surveys, which may underestimate the actual abundance of bull trout.<sup>5</sup> Several studies (e.g., Goetz 1994; Thurow and Schill 1996; Hillman and Chapman 1996; Bonar et al. 1997) have found bull trout population estimates based on nighttime snorkeling to be in some cases more accurate than daytime snorkeling, especially for juvenile bull trout. Our estimates of adult bull trout numbers may be more accurate than those for juveniles.

In all years we found most juvenile bull trout in the upstream reaches of the Chiwawa River. Of the reaches we surveyed, they were most numerous in Reaches 7-10 on the Chiwawa River. We found the majority of these fish in multiple channels, pools, and riffles, and few in glides. They consistently occupied stations close to the stream bottom over rubble and small boulder substrate or near woody debris. This is similar to the observation of Pratt (1984) in the upper Flathead River Basin in Montana. She found that juvenile bull trout lay close to instream cover and that they tended to conceal themselves. As a result, she found it difficult to accurately estimate their numbers. Although this implies that we underestimated numbers of juvenile bull trout in the Chiwawa River, the relative distribution of juvenile bull trout is valid if we assume that we saw the same fraction of juveniles in all reaches (i.e., detection probability was the same across

---

<sup>5</sup> Because there are no estimates for probability of detecting bull trout with daytime underwater observation methods in the Chiwawa River basin, we could not adjust bull trout numbers based on detectability. Therefore, the numbers reported in this report likely underestimate the “true” number of bull trout in the survey area.

survey sites).

We estimated a total of 875 ( $\pm 12\%$  of the estimated total) adult ( $>8$  in) bull trout in reaches of the Chiwawa River basin in August 2014 (Table 9). In previous years, numbers ranged from 76 to 900 (Figure 9; Appendix B). As with juvenile bull trout, we found most of the adult bull trout upstream from Reach 6; although they were found in all reaches on the Chiwawa River. We found few adult bull trout in tributaries of the Chiwawa River. Adult bull trout primarily used pools and multiple channel habitat, although most of the smaller adults ( $<10$  in) used riffles.

### **Abundance of Other Salmonids**

In August 2014, we estimated that at least 70 brook trout, an exotic species closely related to the bull trout, occurred in the Chiwawa River, Chikamin Creek, Big Meadow Creek, Minnow Creek, and in the Little Wenatchee River survey areas. Brook trout occurred in the lower seven reaches on the Chiwawa River. In both the Chiwawa and Little Wenatchee rivers, brook trout usually used multiple channels. Few appeared to be bull trout/brook trout hybrids. In Chikamin, Minnow, and Big Meadow creeks, brook trout were most abundant in pools. Brook trout lengths ranged from 2-12 inches.

At least 765 westslope cutthroat trout occurred in the Chiwawa River, Rock Creek, Phelps Creek, and Little Wenatchee River survey areas in August 2014. These fish most often occurred in pools and multiple channel habitats. They ranged in size from 2-22 inches. Juvenile coho salmon were observed in Nason Creek and the Chiwawa River.

We observed both juvenile and adult mountain whitefish in the Chiwawa River, Phelps Creek, Rock Creek, Nason Creek, and the Little Wenatchee River survey areas. In sum, at least 7,258 adult and 1,360 juvenile whitefish lived in these streams in August 2014. We found few whitefish in most tributaries to the Chiwawa River.

### **Conclusion**

This was the 22<sup>nd</sup> year of a study to monitor trends in juvenile spring Chinook production in the Chiwawa River basin. As shown in Figure 3, numbers of juvenile Chinook salmon in the Chiwawa River basin have fluctuated widely over the 22-year period. Numbers of juveniles in 2001, 2002, and 2009-2014 were some of the highest recorded, while numbers in the mid-1990s were some of the lowest. Interestingly, the highest spawning escapements (highest redd numbers) resulted in the lowest egg-parr survival rates (Appendix A). This is supported by the fact that the best approximating models clearly demonstrated a density-dependent relationship between seeding levels and juvenile production. Indeed, there was a significant negative relationship between parr per redd and numbers of redds in the Chiwawa River basin. This is an important observation because some of the hypotheses in the revised monitoring and evaluation plan (Hillman et al. 2013) are only valid when the supplemented population is below its carrying capacity.

The proportion of hatchery-origin spawners (pHOS) within the Chiwawa River basin during the survey period has ranged from 0 to 100%. Thus, some of the variation in juvenile productivity may be related to pHOS. Although there appeared to be a negative relationship between juvenile productivity (parr/redd) and pHOS, the correlation was not significant (Figure 10). In addition,

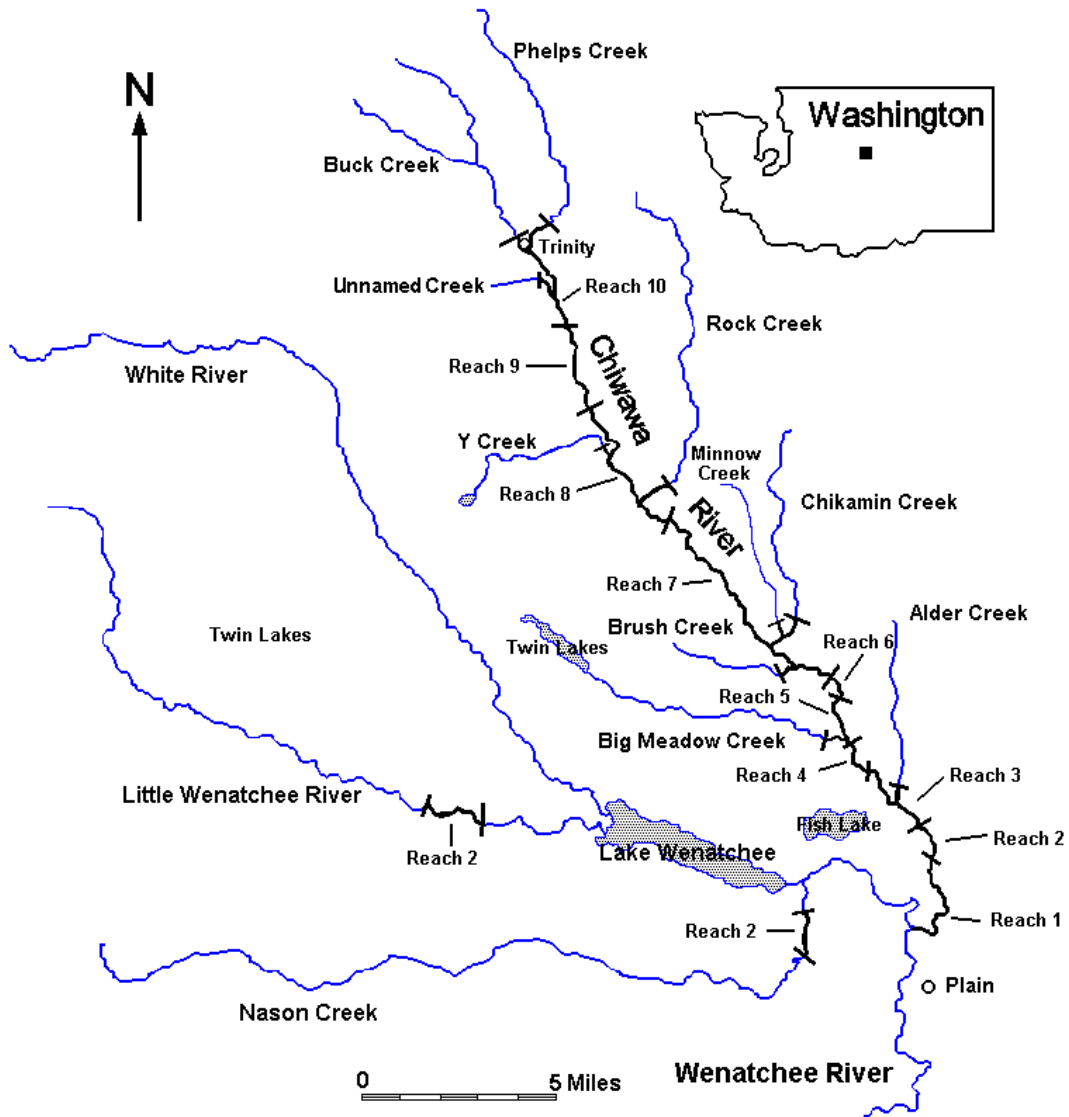
there was no relationship between juvenile productivity and pHOS after the effects of spawning escapement were removed from the analysis (Figure 10). This suggests that spawning escapement has a larger effect on juvenile productivity than does the presence of hatchery spawners.

The presence of density dependence in the early life stages of spring Chinook is not surprising. Rarely does density dependence appear in numbers of adult spring Chinook or on their spawning grounds. The Chiwawa River basin appears to have plenty of spawning habitat, as indicated by the large numbers of spawners and redds widely distributed throughout the basin during high spawning escapements. However, those large spawning escapements did not translate into large numbers of juveniles or smolts. Thus, density-dependent regulation appears to occur sometime during the early life stages of the fish, likely at the fry stage. It is possible that physical habitat (space) during higher flows when fry are emerging may limit juvenile Chinook production in the basin. Low nutrient levels and its effects on food webs may also be a limiting factor in the basin. If spawning escapements remain relatively high, marine-derived nutrients should increase in the basin, resulting in more food for juvenile Chinook salmon.

## References

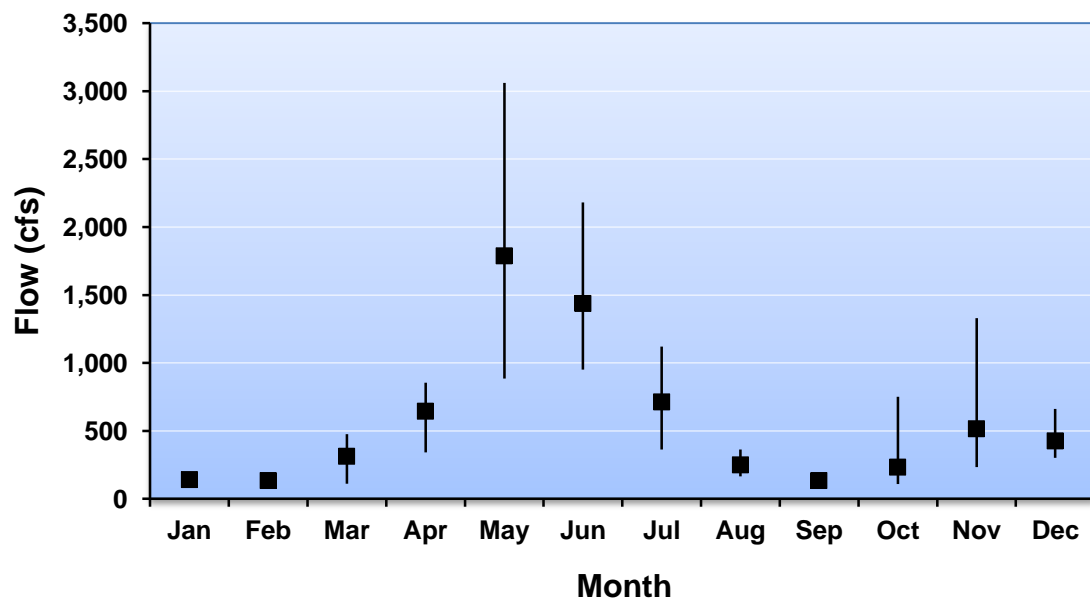
- Bonar, S. A., M. Divens, and B. Bolding. 1997. Methods for sampling the distribution and abundance of bull trout and Dolly Varden. Washington Department of Fish and Wildlife, Research Report No. RAD97-05. Olympia, WA.
- Burnham, K. P. and D. R. Anderson. 2002. Model selection and multimodel inference: a practical information-theoretic approach. Second edition. Springer, New York, N.Y.
- Goetz, F. A. 1994. Distribution and juvenile ecology of bull trout (*Salvelinus confluentus*) in the Cascade Mountains. Master's thesis. Oregon State University, Corvallis.
- Hillman, T. W. and D. W. Chapman. 1996. Comparison of underwater methods and electrofishing for estimating fish populations in the upper Blackfoot River Basin. Report to Seven-Up Pete Joint Venture, Lincoln, MT.
- Hillman, T. W. and M. D. Miller. 2004. Abundance and total numbers of Chinook salmon and trout in the Chiwawa River basin, Washington, 2004. Report to Chelan Public Utility District, Wenatchee, WA. BioAnalysts, Inc., Boise, ID.
- Hillman, T., M. Miller, T. Miller, M. Tonseth, M. Hughes, A. Murdoch, L. Keller, and J. Murauskas. 2013. Monitoring and evaluation of the Chelan County PUD Hatchery programs: 2012 annual report. Report to the HCP Hatchery Committee, Wenatchee, WA.
- Hillman, T., T. Kahler, G. Mackey, J. Murauskas, A. Murdoch, K. Murdoch, T. Pearsons, and M. Tonseth. 2013. Updated monitoring and evaluation plan for PUD hatchery programs. Report to the Hatchery Committees, Wenatchee, East Wenatchee, and Ephrata, WA.
- Hillman, T., T. Kahler, G. Mackey, J. Murauskas, A. Murdoch, K. Murdoch, T. Pearsons, and M. Tonseth. 2013. Monitoring and evaluation plan for PUD hatchery programs; 2013 update. Report to the HCP and PRCC Hatchery Committees, Wenatchee, WA.
- Hubble, J. 1993. Methow valley spring Chinook supplementation project. Yakima Indian Nation. Annual report to Douglas County Public Utility District, East Wenatchee, WA.
- Kiefer, R. and K. Forster. 1991. Idaho habitat and natural production monitoring. Idaho Department of Fish and Game, Annual Report 1989, Project No. 83-7, Contract No. DE-BI79-84BP13381.
- Konopacky, R. C., P. J. Cerner, and E. C. Bowles. 1986. Natural propagation and habitat improvement, Idaho: Salmon River habitat enhancement. Subproject I, Bear Valley Creek: inventory, 1984 and 1985. Shoshone-Bannock Tribes, Fort Hall, ID. Report to U.S. Department of Energy, Bonneville Power Administration, Division of Fish and Wildlife, Project No. 83-359, Contract No. DE-A179-84BP14383, Portland, OR.
- Mullan, J. W., K. R. Williams, G. Rhodus, T. W. Hillman, and J. D. McIntyre. 1992. Production and habitat of salmonids in mid-Columbia River tributary streams. U.S. Fish and Wildlife Service, Monograph I. 489 p.
- Murdoch, A. and C. Peven. 2005. Conceptual approach to monitoring and evaluating the Chelan County Public Utility District hatchery programs. Chelan County Public Utility District

- and the Washington Department of Fish and Wildlife, Wenatchee, WA.
- Murdoch, A., T. Pearsons, and T. Maitland. 2009. The number of redds constructed per female spring Chinook salmon in the Wenatchee River Basin. *North American Journal of Fisheries Management* 29:441-446.
- Petrosky, C. E. 1990. Estimating spring Chinook parr and smolt abundance in wild and natural production areas. Pages 57-61 *in*: D. L. Park, editor. Status and future of spring Chinook salmon in the Columbia River basin--conservation and enhancement. Spring Chinook salmon workshop, U.S. Dept. Comm. NOAA Tech. Mem. NMFS F/NWD-187.
- Pratt, K. L. 1984. Habitat use and species interactions of juvenile cutthroat *Salmo clarki lewisi* and bull trout *Salvelinus confluentus* in the upper Flathead River Basin. Master's thesis. University of Idaho, Moscow, ID. 95 p.
- Quinn, T. P. 2004. The behavior and ecology of Pacific salmon and trout. University of Washington Press, Seattle, WA.
- Richards, C. and P. J. Cernera. 1987. Salmon River habitat enhancement, annual report, 1986. Shoshone-Bannock Tribes, Fort Hall, ID. Report to Bonneville Power Administration, Project No. 83-359, Contract No. DE-A179-84BP14383, Portland, OR.
- Thorson, J. T., M. D. Scheuerell, E. R. Buhle, and T. Copeland. 2013. Spatial variation buffers temporal fluctuations in early juvenile survival for an endangered Pacific salmon. *Journal of Animal Ecology* 2013:1-11.
- Thurow, R. F. and D. J. Schill. 1996. Comparison of day snorkeling, night snorkeling, and electrofishing to estimate bull trout abundance and size structure in a second-order Idaho stream. *North American Journal of Fisheries Management* 16:314-323.
- USFS (United States Forest Service). 1993. Upper Chiwawa River stream survey report. Wenatchee National Forest, Wenatchee, WA.

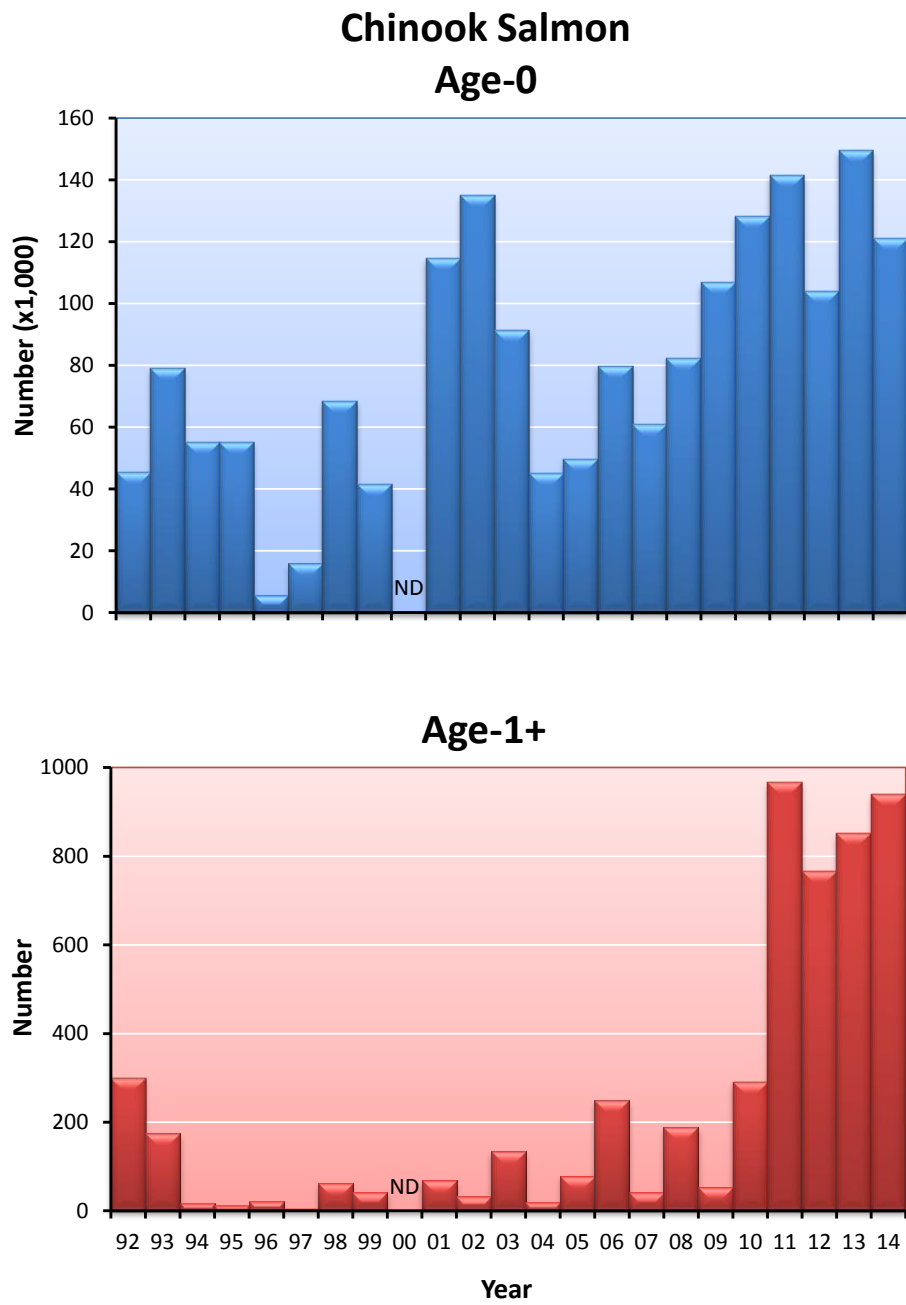


**Figure 1.** Location of study reaches on the Chiwawa River, and Chikamin, Rock, Big Meadow, Unnamed, Alder, Brush and Phelps creeks, Chelan County, Washington. Reach 2 on Nason Creek and Reach 2 on the Little Wenatchee River were matched with Reaches 3 and 8 on the Chiwawa River, respectively.

## Chiwawa River 2014



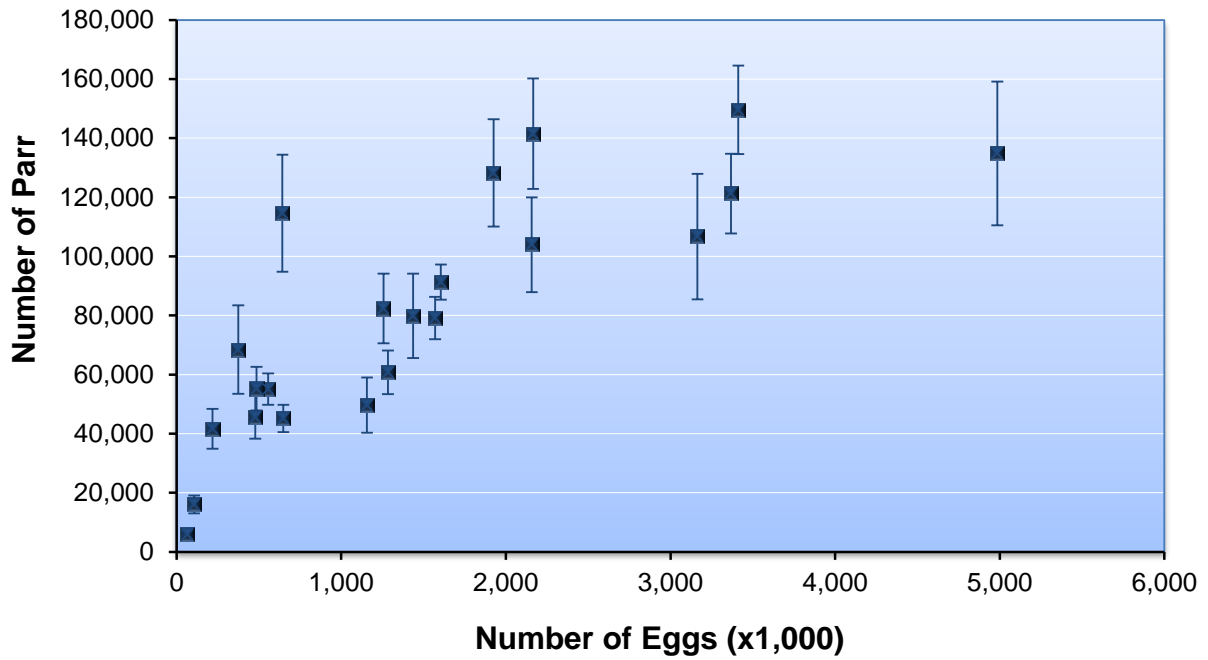
**Figure 2.** Mean, minimum, and maximum monthly flows in the Chiwawa River for 2014.



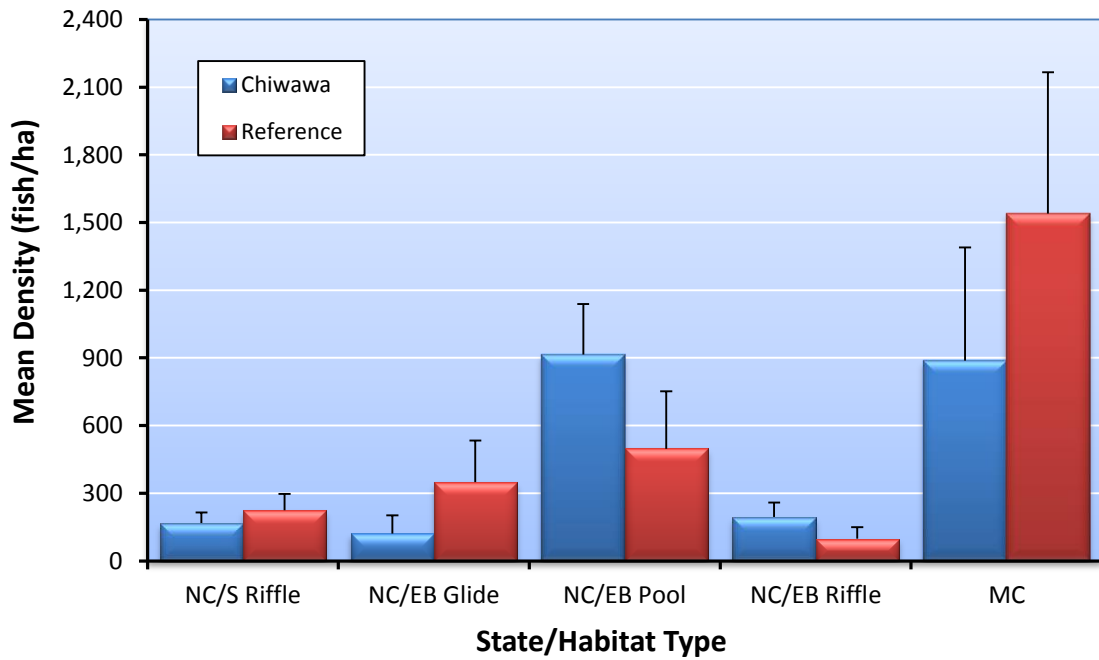
**Figure 3.** Numbers of age-0 and age-1+ Chinook salmon within the Chiwawa River basin in August 1992-2014; ND = no data.



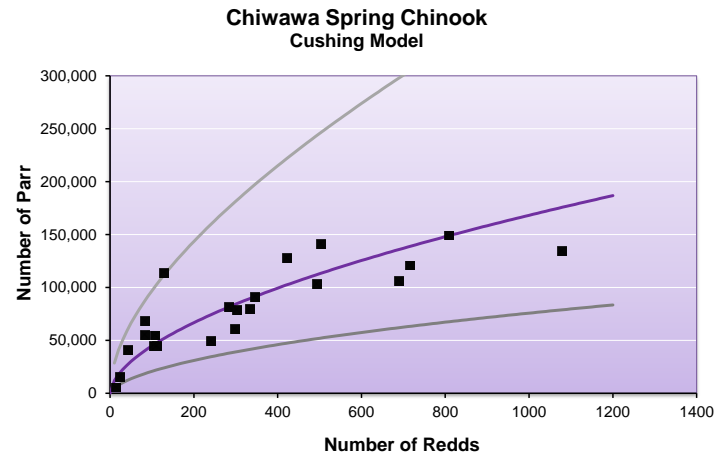
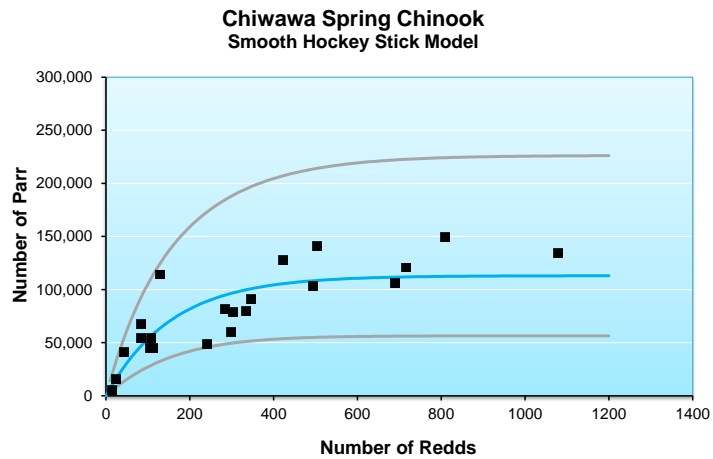
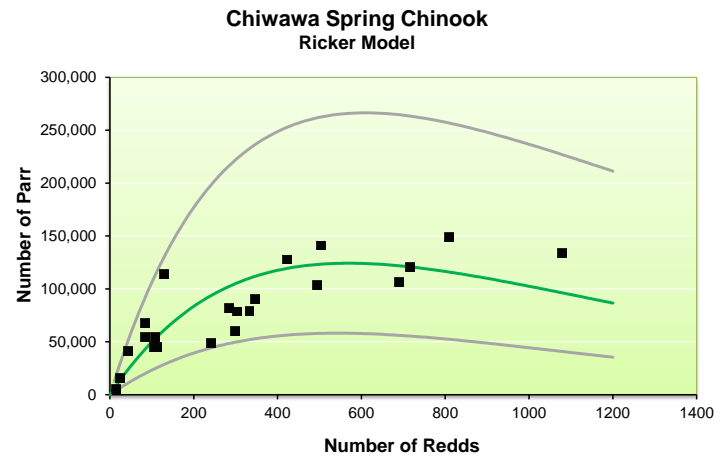
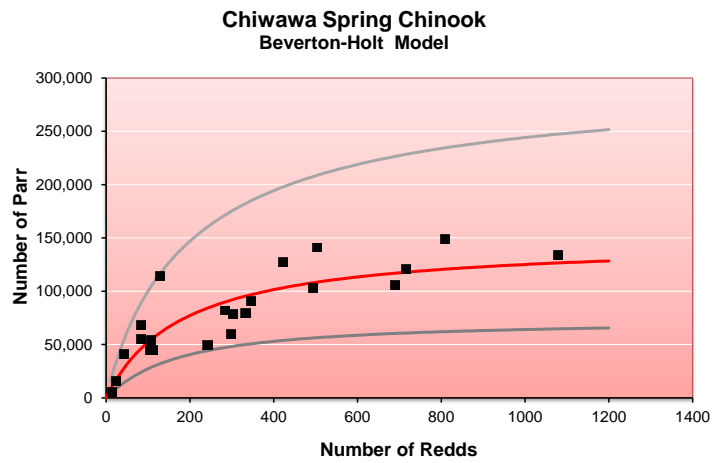
## Chiwawa Spring Chinook



**Figure 4.** Relationship between total numbers of age-0 Chinook salmon (based on fish/ha) and numbers of eggs in the Chiwawa River basin. Vertical bars indicate 95% confidence bounds.

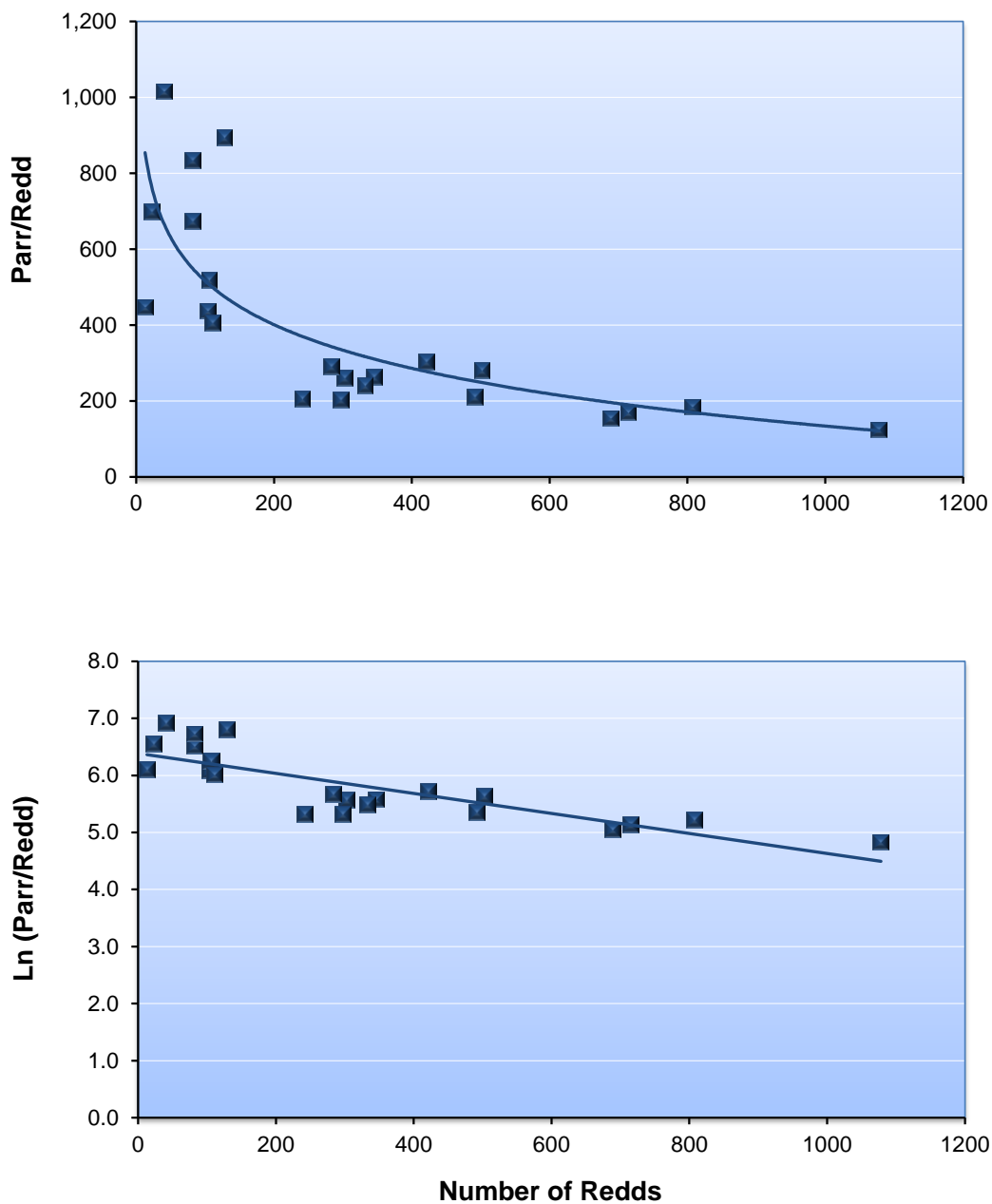


**Figure 5.** Comparison of the means (95% CI) of age-0 Chinook salmon densities (fish/ha) within state/habitat types in Reaches 3 and 8 of the Chiwawa River and their matched reference areas on Nason Creek and the Little Wenatchee River. There was no sampling in 2000 and no sampling in reference areas in 1992.

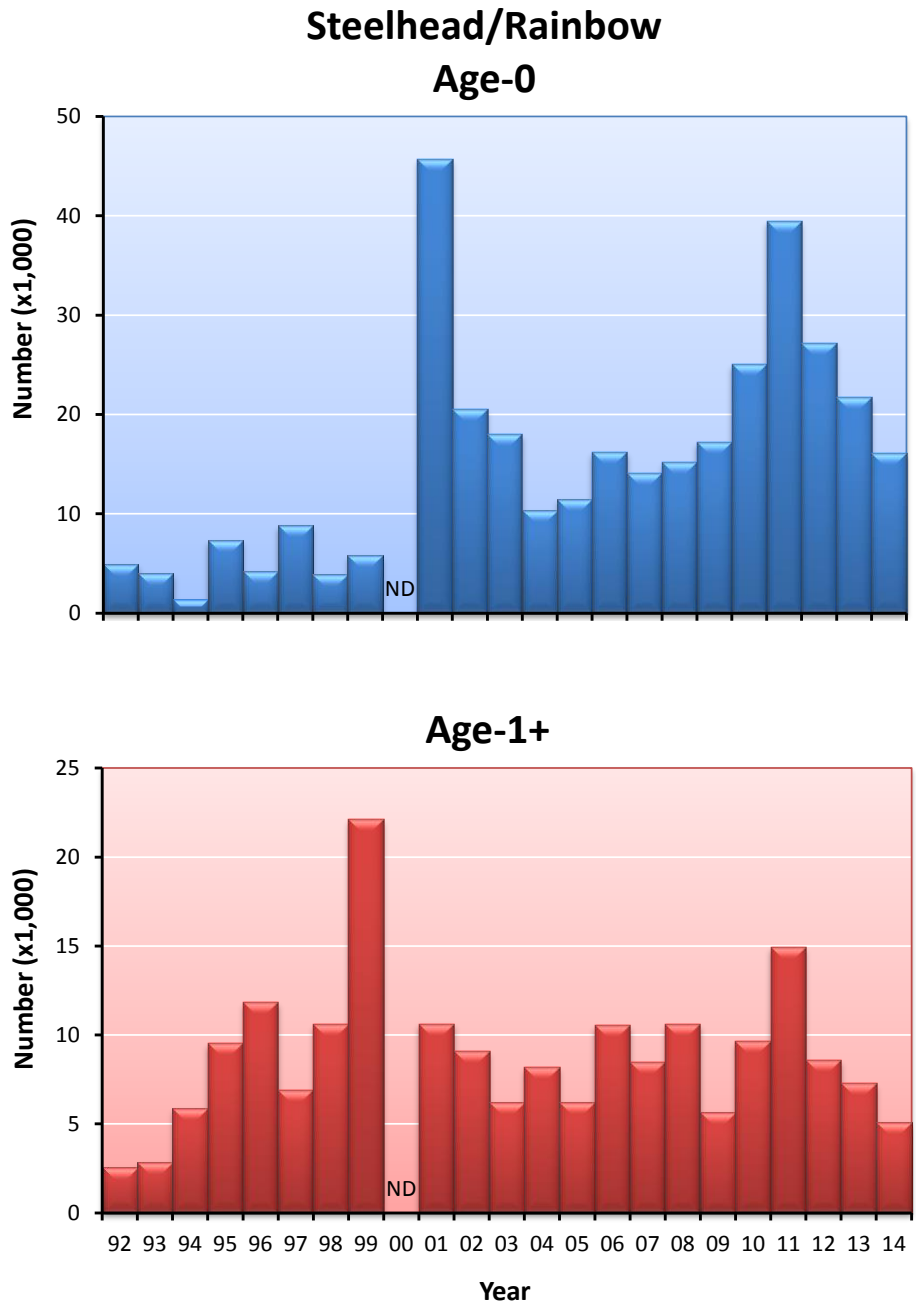


**Figure 6.** Relationship between numbers of juvenile (age-0) Chinook and redds in the Chiwawa River basin, 1992-2014 (no sampling occurred in 2000). Figures show the fit of the Beverton-Holt model, smooth hockey stick, Ricker model, and the Cushing model to the data. Gray lines indicate the upper and lower 95% C.B.

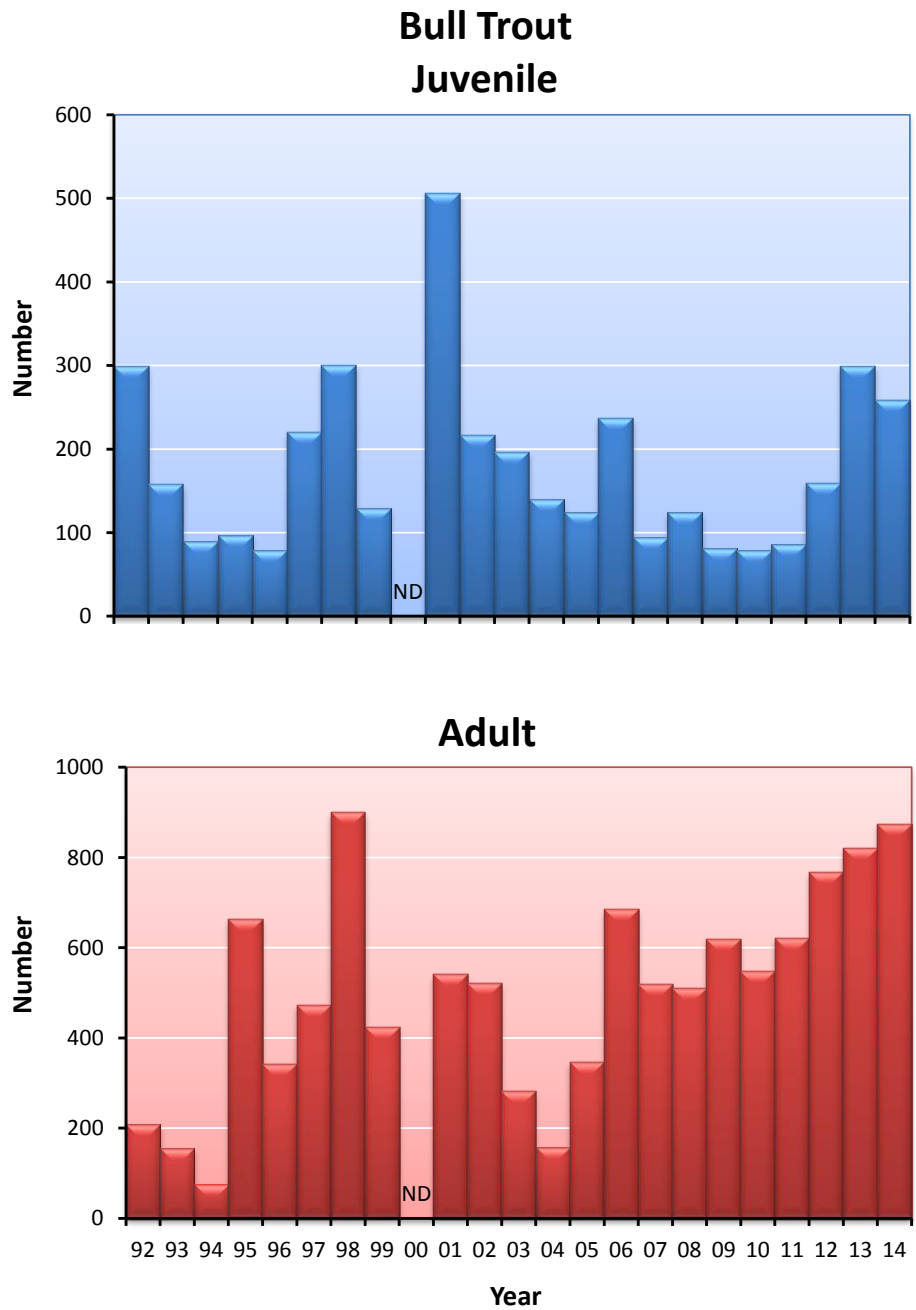
### Chiwawa Spring Chinook



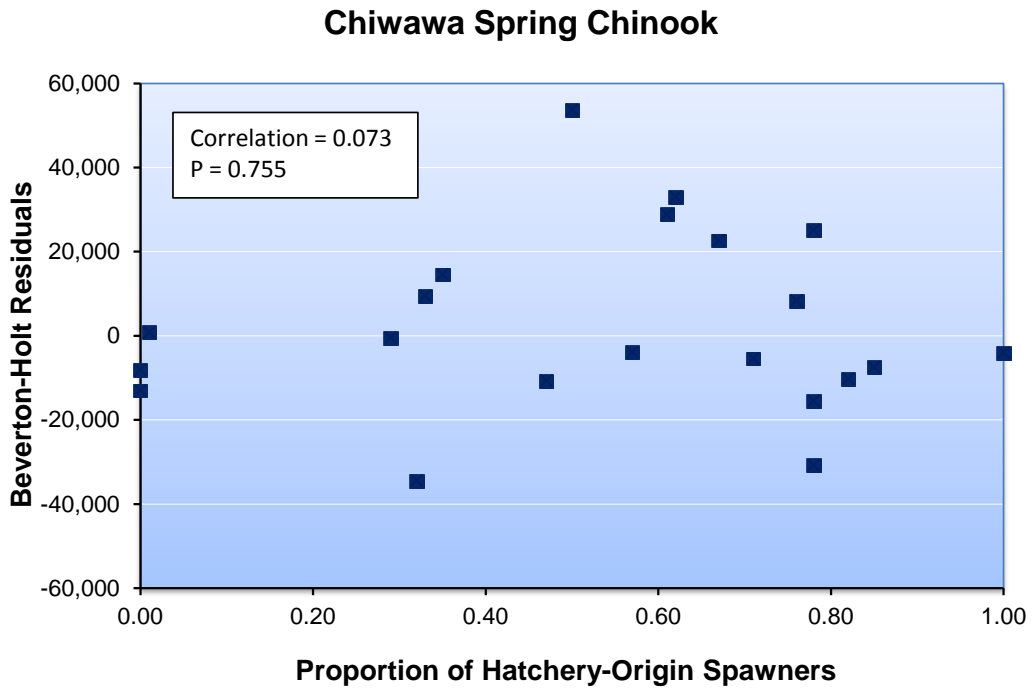
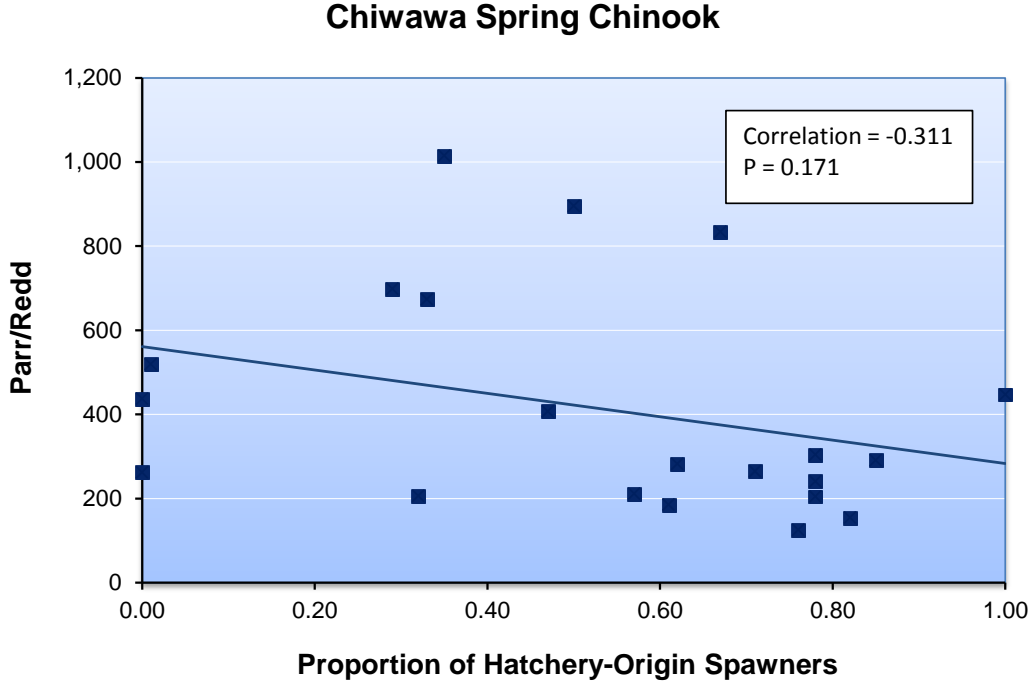
**Figure 7.** Relationship between parr/redd and numbers of redds (top figure) and natural log parr/redd and numbers of redds (bottom figure) in the Chiwawa River basin, 1992-2014. No sampling was conducted in 2000. Estimates for 1992-2014 included the Chiwawa River and its tributaries; the 1992 estimate included only the Chiwawa River. The linear relationship  $\text{LN}(P/R) = 6.15 - 0.002(\text{Redds})$  was significant with  $P = 0.0000$ ;  $R^2 = 0.684$ .



**Figure 8.** Numbers of age-0 (<4 in) and age-1+ (4-8 in) steelhead/rainbow within the Chiwawa River basin in August 1992-2014; ND = no data.



**Figure 9.** Numbers of juvenile (2-8 inches) and adult (>8 inches) bull trout within the Chiwawa River basin in August 1992-2014; ND = no data.



**Figure 10.** Relationship between juvenile productivity (parr/redd) and the proportion of hatchery-origin spawners (pHOS) (top figure) and the relationship between the residuals from the Beverton-Holt stock/recruitment relationship and pHOS (bottom figure).

**Table 1.** Description, location (river mile), and area (hectares) of land-class strata (reaches) used by age-0 Chinook salmon in the Chiwawa River basin, 2014. Reaches were classified according to geologic district, landtype association, valley-bottom type, stream state-type, and habitat type within the Cascade Ecoregion; MCV = moderately confined valley, CC = confined canyon, UCV = unconfined valley, NC = natural channel, EB = eroded banks, S = straight, G = glide, P = pool, R = riffle, and MC = multiple channel. See Hillman and Miller (2004) for definitions of stream state codes.

Reach	RM	Gradient	Geologic district	Landtype association	Valley bottom type	Stream state type	Habitat type	Area (ha)	
								Total	Sample
<b>Chiwawa River</b>									
1	0.00-3.77	0.007	Glacial Drift over Chumstick Formation	Glacial Valley	MCV Alluvial	NC/EB	G	0.60	0.60
						NC/EB	P	1.33	0.92
						NC/EB	R	18.22	1.80
2	3.77-5.51	0.010	Glacial Drift over Chumstick Formation	Glacial Canyon	CC Fluvial	NC/EB	G	0.26	0.26
						NC/EB	P	0.75	0.28
						NC/EB	R	7.20	0.64
3	5.51-7.88	0.009	Glacial Drift over Chumstick Formation	Glacial Valley	MCV Alluvial	NC/S	R	5.96	0.76
						NC/EB	G	0.15	0.15
						NC/EB	R	4.59	0.57
						MC	MC	0.28	0.28
4	7.88-8.90	0.007	Glacial Drift over Chumstick Formation	Glacial Canyon	CC Fluvial	NC/EB	P	0.36	0.27
						NC/EB	R	2.65	0.42
						MC	MC	0.44	0.44
5	8.90-10.83	0.011	Glacial Drift over Chumstick Formation	Glacial Valley	MCV Alluvial	NC/EB	P	0.13	0.13
						NC/EB	R	9.87	1.02
6	10.83-11.80	0.008	Glacial Drift over Chumstick Formation	Glacial Canyon	CC Fluvial	NC/EB	P	0.33	0.33
						NC/EB	R	3.71	0.94
						MC	MC	0.34	0.34
7	11.80-20.03	0.001	Glacial Drift over Chumstick Formation	Glacial Valley	UCV Alluvial	NC	G	2.27	0.58
						NC	P	6.22	0.47
						NC	R	1.05	0.24
						NC/EB	G	2.62	1.41
						NC/EB	P	6.49	1.71
						NC/EB	R	4.55	0.54
8	20.03-25.42	0.003	Glacial Drift over Swakane Gneiss	Glacial Valley	UCV Alluvial	MC	MC	5.05	2.20
						NC/EB	G	2.50	1.14
						NC/EB	P	6.93	1.59
						NC/EB	R	6.04	0.96
						EB	P	0.22	0.22
9	25.42-28.81	0.007	Glacial Drift over Swakane Gneiss	Glacial Valley	MCV Alluvial	EB	R	0.40	0.40
						MC	MC	6.99	3.12
						NC	P	4.55	0.64
						NC	R	2.28	0.58
						MC	MC	2.33	0.82
10	28.81-31.11	0.011	Pre-upper Jurassic Gneiss	Glacial Valley	MCV Alluvial	NC	P	0.43	0.37
						NC	R	1.58	0.86
						MC	MC	5.20	0.38



**Table 1.** Concluded.

Reach	RM	Gradient	Geologic district	Landtype association	Valley bottom type	Stream state type	Habitat type	Area (ha)	
								Total	Sampled
<b>Phelps Creek</b>									
1	0.00-0.35	0.043	Pre-upper Jurassic Gneiss	Glacial Valley	MCV Alluvial	NC	R	0.00	0.00
						NC	MC	0.24	0.24
<b>Chikamin Creek<sup>1</sup></b>									
1	0.00-0.94	0.013	Glacial Drift over Chumstick Formation	Glacial Valley	UCV Alluvial	NC	G	0.01	0.01
						NC	P	0.18	0.05
						NC	R	0.30	0.06
						MC	MC	0.10	0.10
<b>Rock Creek</b>									
1	0.00-0.73	0.020	Glacial Drift over Swakane Gneiss	Glacial Valley	UCV Alluvial	NC	P	0.17	0.05
						NC	R	0.38	0.05
						MC	MC	0.15	0.15
<b>Unnamed Creek</b>									
1	0.00-0.05		Pre-upper Jurassic Gneiss	Glacial Valley	MCV Alluvial	NC	P	0.01	0.01
						NC	R	0.00	0.00
<b>Big Meadow Creek</b>									
1	0.00-0.35	0.025	Glacial Drift over Chumstick Formation	Glacial Valley	MCV Alluvial	NC	G	0.01	0.01
						NC	P	0.13	0.03
						NC	R	0.12	0.02
						NC	MC	0.09	0.09
<b>Alder Creek</b>									
1	0.00-0.01		Glacial Drift over Chumstick Formation	Glacial Valley	MCV Alluvial	NC	P	0.002	0.002
						NC	R	0.006	0.006
<b>Brush Creek</b>									
1	0.00-0.01		Glacial Drift over Chumstick Formation	Glacial Valley	UCV Alluvial	NC	P	0.002	0.002
						NC	R	0.003	0.003
<b>Clear Creek</b>									
1	0.00-0.05		Glacial Drift over Chumstick Formation	Glacial Valley	UCV Alluvial	NC	P	0.002	0.002
						NC	R	0.003	0.003
<b>Y Creek</b>									
1	0.00-0.05		Glacial Drift over Swakane Gneiss	Glacial Valley	UCV Alluvial	NC	P	0.000	0.000
						NC	R	0.000	0.000

<sup>1</sup> Includes the lower 0.2 miles of Minnow Creek.

**Table 2.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of age-0 Chinook salmon in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	252.3	0.067	5,084	±2,423	0.48	4,719	±2,371	0.50
2	147.1	0.036	1,208	±191	0.16	1,142	±214	0.19
3	144.4	0.037	1,586	±25	0.02	1,696	±30	0.02
4	793.0	0.169	2,736	±150	0.05	2,812	±114	0.04
5	207.9	0.058	2,079	±74	0.04	2,191	±118	0.05
6	448.9	0.109	1,966	±77	0.04	1,829	±129	0.07
7	1,126.2	0.179	31,814	±6,122	0.19	31,841	±6,886	0.22
8	1,214.3	0.196	28,026	±9,921	0.35	26,239	±10,325	0.39
9	1,372.9	0.201	12,576	±1,667	0.13	11,998	±3,603	0.30
10	3,716.2	1.081	26,794	±6,114	0.23	30,755	±4,530	0.15
<b>Phelps Creek</b>								
1	1,504.2	0.669	361	±0	0.00	361	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	3,994.9	2.076	2,384	±316	0.13	2,701	±332	0.12
<b>Rock Creek</b>								
1	4,462.9	1.861	3,124	±347	0.11	3,153	±436	0.14
<b>Unnamed Creek</b>								
1	2,000.0	0.571	28	±0	0.00	28	±0	0.00
<b>Big Meadow Creek</b>								
1	3,950.9	1.307	1,367	±705	0.52	1,134	±660	0.58
<b>Alder Creek</b>								
1	1,375.0	1.410	11	±0	0.00	11	±0	0.00
<b>Brush Creek</b>								
1	5,600.0	6.364	28	±0	0.00	28	±0	0.00
<b>Clear Creek</b>								
1	11,333.3	9.189	68	±0	0.00	68	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>956.3</b>	<b>0.197</b>	<b>121,240</b>	<b>±13,518</b>	<b>0.11</b>	<b>122,706</b>	<b>±13,928</b>	<b>0.11</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.

**Table 3.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of age-1+ Chinook salmon in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	1.4	0.000	29	±6	0.21	28	±36	1.29
2	2.3	0.001	19	±12	0.63	16	±22	1.38
3	2.5	0.001	27	±3	0.11	28	±3	0.11
4	7.2	0.002	25	±6	0.24	25	±7	0.28
5	0.3	0.000	3	±0	0.00	4	±0	0.00
6	3.0	0.001	13	±0	0.00	13	±0	0.00
7	15.0	0.002	425	±180	0.42	427	±288	0.67
8	8.5	0.001	197	±165	0.84	188	±195	1.04
9	13.8	0.002	126	±95	0.75	119	±114	0.96
10	0.4	0.000	3	±3	1.00	3	±4	1.33
<b>Phelps Creek</b>								
1	116.7	0.052	28	±0	0.00	28	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	6.7	0.003	4	±0	0.00	4	±0	0.00
<b>Rock Creek</b>								
1	54.3	0.023	38	±38	1.00	39	±37	0.95
<b>Unnamed Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Big Meadow Creek</b>								
1	5.8	0.002	2	±0	0.00	2	±0	0.00
<b>Alder Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Brush Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Clear Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>7.4</b>	<b>0.002</b>	<b>939</b>	<b>±265</b>	<b>0.28</b>	<b>924</b>	<b>±371</b>	<b>0.40</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.

**Table 4.** Summary of the six productivity models of juvenile (age-0) Chinook salmon in the Chiwawa River basin. Models are shown, including the number of parameters ( $K$ ),  $AIC_c$  values,  $AIC_c$  difference scores ( $\Delta_i$ ), the likelihood of the model given the data ( $\ell(g_i|x)$ ), Akaike weights ( $w_i$ ), and adjusted  $R^2$  values. The sample size ( $n$ ) for all models was 22. Models describe the relationship between juvenile Chinook numbers (dependent variable) and redd numbers (independent variable).

Model	$K^a$	$AIC_c$	$\Delta_i$	$\ell(g_i x)$	$w_i$	$Adj R^2$
Beverton-Holt	3	-115.505	0.000	1.000	0.638	0.833
Smooth Hockey Stick	3	-114.014	1.491	0.474	0.302	0.822
Gamma <sup>b</sup>	4	-108.886	6.619	0.037	0.023	0.794
Ricker	3	-108.853	6.652	0.036	0.023	0.775
Cushing	3	-107.835	7.671	0.022	0.014	0.765

<sup>a</sup>  $K$  is the number of structural parameters in the model plus 1 for  $\sigma^2$ .

<sup>b</sup> The  $\gamma$  parameter in the Gamma model was greater than 0, which means that this model is nearly identical to the Ricker model.

**Table 5.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of age-0 (<4 in) steelhead/rainbow in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	118.5	0.030	2,388	±114	0.05	2,140	±163	0.08
2	49.0	0.012	402	±73	0.18	387	±85	0.22
3	184.9	0.049	2,030	±132	0.07	2,224	±146	0.07
4	281.2	0.066	970	±258	0.27	1,101	±273	0.25
5	172.8	0.049	1,728	±100	0.06	1,825	±114	0.06
6	201.8	0.045	884	±85	0.10	759	±108	0.14
7	134.0	0.022	3,785	±1,406	0.37	3,844	±1,604	0.42
8	3.0	0.001	69	±111	1.61	67	±120	1.79
9	0.0	0.000	0	±0	0.00	0	±0	0.00
10	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Phelps Creek</b>								
1	87.5	0.039	21	±0	0.00	21	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	2,679.7	1.379	1,617	±424	0.26	1,813	±436	0.24
<b>Rock Creek</b>								
1	1,484.3	0.610	1,039	±136	0.13	1,033	±145	0.14
<b>Unnamed Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Big Meadow Creek</b>								
1	2,904.6	0.986	1,005	±369	0.37	855	±329	0.38
<b>Alder Creek</b>								
1	4,000.0	4.103	32	±0	0.00	32	±0	0.00
<b>Brush Creek</b>								
1	11,200.0	12.727	56	±0	0.00	56	±0	0.00
<b>Clear Creek</b>								
1	9,500.0	7.703	57	±0	0.00	57	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>126.9</b>	<b>0.026</b>	<b>16,083</b>	<b>±1,563</b>	<b>0.10</b>	<b>16,214</b>	<b>±1,749</b>	<b>0.11</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.

**Table 6.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of age-1+ (4-8 in) steelhead/rainbow in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	48.4	0.012	975	±78	0.08	879	±99	0.11
2	8.2	0.002	67	±17	0.25	57	±20	0.35
3	51.8	0.014	569	±47	0.08	628	±56	0.09
4	100.3	0.024	346	±166	0.48	395	±182	0.46
5	58.1	0.016	581	±37	0.06	614	±56	0.09
6	134.7	0.030	590	±189	0.32	502	±177	0.35
7	49.7	0.008	1,404	±512	0.36	1,424	±551	0.39
8	1.6	0.000	36	±57	1.58	27	±62	2.30
9	0.0	0.000	0	±0	0.00	0	±0	0.00
10	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Phelps Creek</b>								
1	54.2	0.024	13	±0	0.00	13	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	162.7	0.075	96	±27	0.00	97	±32	0.00
<b>Rock Creek</b>								
1	301.4	0.126	211	±100	0.47	214	±92	0.43
<b>Unnamed Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Big Meadow Creek</b>								
1	476.9	0.173	165	±129	0.78	150	±86	0.57
<b>Alder Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Brush Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Clear Creek</b>								
1	5,166.7	4.189	31	±0	0.00	31	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>40.1</b>	<b>0.008</b>	<b>5,084</b>	<b>±605</b>	<b>0.12</b>	<b>5,031</b>	<b>±637</b>	<b>0.13</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.

**Table 7.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of steelhead/rainbow larger than 8 inches in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	0.5	0.000	10	±0	0.00	7	±12	1.71
2	1.0	0.000	8	±7	0.88	6	±12	2.00
3	0.2	0.000	2	±0	0.00	2	±0	0.00
4	0.9	0.000	3	±3	1.00	3	±5	1.67
5	4.1	0.001	41	±7	0.17	41	±8	0.20
6	0.0	0.000	0	±0	0.00	0	±0	0.00
7	0.8	0.000	23	±13	0.57	18	±17	0.94
8	0.0	0.000	0	±0	0.00	0	±0	0.00
9	0.0	0.000	0	±0	0.00	0	±0	0.00
10	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Phelps Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Rock Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Unnamed Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Big Meadow Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Alder Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Brush Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Clear Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>0.7</b>	<b>0.000</b>	<b>87</b>	<b>±17</b>	<b>0.20</b>	<b>77</b>	<b>±25</b>	<b>0.32</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.

**Table 8.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of juvenile bull trout (2-8 in) in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
2	0.0	0.000	0	±0	0.00	0	±0	0.00
3	0.0	0.000	0	±0	0.00	0	±0	0.00
4	0.0	0.000	0	±0	0.00	0	±0	0.00
5	0.0	0.000	0	±0	0.00	0	±0	0.00
6	0.0	0.000	0	±0	0.00	0	±0	0.00
7	2.8	0.000	79	±58	0.73	71	±76	1.07
8	0.0	0.000	0	±0	0.00	0	±0	0.00
9	5.6	0.001	51	±28	0.55	54	±42	0.78
10	15.1	0.004	109	±11	0.10	122	±26	0.21
<b>Phelps Creek</b>								
1	29.2	0.013	7	±0	0.00	7	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Rock Creek</b>								
1	18.6	0.008	13	±13	1.00	14	±12	0.86
<b>Unnamed Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Big Meadow Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Alder Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Brush Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Clear Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>2.0</b>	<b>0.000</b>	<b>259</b>	<b>±66</b>	<b>0.25</b>	<b>268</b>	<b>±91</b>	<b>0.34</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.



**Table 9.** Estimated mean densities (fish/hectare and fish/m<sup>3</sup>), total numbers, 95% confidence bounds on total numbers, and error of the estimated total number of adult bull trout (>8 in) in reaches in the Chiwawa River basin, Washington, August 2014.

Reach	Mean density		Surface area (ha)			Volume (m <sup>3</sup> )		
	Fish/ha	Fish/m <sup>3</sup>	Total No.	95% C.B.	± Error	Total No.	95% C.B.	± Error
<b>Chiwawa River</b>								
1	1.3	0.000	26	±10	0.38	28	±29	1.04
2	3.5	0.001	29	±9	0.31	25	±31	1.24
3	2.6	0.001	29	±2	0.07	32	±4	0.13
4	5.5	0.001	19	±6	0.32	18	±15	0.83
5	0.4	0.000	4	±0	0.00	4	±0	0.00
6	2.1	0.001	9	±0	0.00	8	±0	0.00
7	6.2	0.001	176	±63	0.36	178	±126	0.71
8	8.0	0.001	184	±71	0.39	174	±151	0.87
9	19.1	0.003	175	±37	0.21	173	±72	0.42
10	29.8	0.009	215	±26	0.12	250	±29	0.12
<b>Phelps Creek</b>								
1	16.7	0.007	4	±0	0.00	4	±0	0.00
<b>Chikamin Creek<sup>1</sup></b>								
1	3.4	0.002	2	±0	0.00	2	±0	0.00
<b>Rock Creek</b>								
1	4.3	0.002	3	±5	1.67	4	±4	1.00
<b>Unnamed Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Big Meadow Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Alder Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Brush Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Clear Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Y Creek</b>								
1	0.0	0.000	0	±0	0.00	0	±0	0.00
<b>Grand Total</b>	<b>6.9</b>	<b>0.001</b>	<b>875</b>	<b>±106</b>	<b>0.12</b>	<b>900</b>	<b>±216</b>	<b>0.24</b>

<sup>1</sup> Includes lower 0.2 miles of Minnow Creek.

**APPENDIX A.** Numbers of redds, eggs, age-0 Chinook salmon, parr per redd, and percent egg-to-parr survival in the Chiwawa River basin, brood years 1991-2013; NS = not sampled. Numbers of eggs were calculated as the number of redds times the mean fecundity of females collected for broodstock.

Brood Year	Chinook Salmon			Parr/Redd	Egg-to-parr survival (%)
	Redds	Eggs	Age-0 (parr)		
1991	104	478,400	45,483	437	9.5
1992	302	1,570,098	79,113	262	5.0
1993	106	556,394	55,056	519	9.9
1994	82	485,686	55,240	674	11.4
1995	13	66,248	5,815	447	8.8
1996	23	106,835	16,066	699	15.0
1997	82	374,740	68,415	834	18.3
1998	41	218,325	41,629	1,015	19.1
1999	34	166,090	NS	NS	NS
2000	128	642,944	114,617	895	17.8
2001	1,078	4,984,672	134,874	125	2.7
2002	345	1,605,630	91,278	265	5.7
2003	111	648,684	45,177	407	7.0
2004	241	1,156,559	49,631	206	4.3
2005	332	1,436,564	79,902	241	5.6
2006	297	1,284,228	60,752	205	4.7
2007	283	1,256,803	82,351	291	6.6
2008	689	3,163,888	106,705	155	3.4
2009	421	1,925,233	128,220	305	6.7
2010	502	2,165,628	141,510	282	6.5
2011	492	2,157,420	103,940	211	4.8
2012	808	3,412,184	149,563	185	4.4
2013	714	3,367,224	121,240	170	3.6
<b>Average</b>	<b>314</b>	<b>1,444,803</b>	<b>80,754</b>	<b>247</b>	<b>8.2</b>

**APPENDIX B.** Estimated numbers of salmonids (based on fish/ha) in the Chiwawa River basin, Washington, 1992-2014; NS = not sampled.

Survey year	Chinook salmon		Steelhead/Rainbow			Bull trout		Cutthroat trout
	Age-0	Age-1+	Age-0	Age-1+	>8 in <sup>1</sup>	2-8 in	>8 in	
1992 <sup>2</sup>	45,483	563	4,927	2,533	1,869	299	208	NS
1993	79,113	174	4,004	2,860	768	158	156	NS
1994	55,056	18	1,410	5,856	67	90	76	NS
1995	55,241	13	7,357	9,517	140	97	664	NS
1996	5,815	22	4,245	11,849	78	79	343	NS
1997	16,066	5	8,823	6,905	48	220	472	56
1998	68,415	63	3,921	10,585	78	300	900	93
1999	41,629	41	5,838	22,130	33	130	423	80
2000	NS	NS	NS	NS	NS	NS	NS	NS
2001	114,617	69	45,727	10,623	420	505	542	108
2002	134,874	32	20,521	9,090	181	217	521	111
2003	91,278	134	18,020	6,179	49	196	282	52
2004	45,177	21	10,380	8,190	8	140	157	22
2005	49,631	79	11,463	6,188	48	125	346	23
2006	79,902	388	16,245	10,533	50	238	686	68
2007	60,752	41	14,073	8,448	77	95	520	47
2008	82,351	189	15,230	10,576	144	124	510	109
2009	106,705	54	17,179	5,629	85	82	618	128
2010	128,220	291	25,018	9,616	63	79	547	252
2011	141,510	967	39,446	14,903	65	86	621	240
2012	103,940	767	27,134	8,576	65	159	768	188
2013	149,563	852	21,682	7,253	76	299	820	358
2014	121,240	939	16,083	5,084	87	259	875	761

<sup>1</sup>During 1992-1993, numbers included both hatchery and wild rainbow trout. Thereafter, only wild trout were observed.

<sup>2</sup>Only the Chiwawa River was sampled in 1992. No tributaries were sampled in that year.

**APPENDIX C.** Proportion of total habitat available, fraction of all age-0 Chinook within each habitat type, and densities (fish/ha) and numbers of age-0 Chinook within each habitat type in the Chiwawa River basin, survey years 1992-2014; NS = not sampled.

Habitat	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002
<b>Proportion of total habitat available</b>											
Glide	0.10	0.09	0.10	0.10	0.10	0.09	0.09	0.09	NS	0.07	0.08
Pool	0.19	0.19	0.21	0.18	0.18	0.17	0.16	0.17	NS	0.15	0.16
Riffle	0.61	0.61	0.57	0.59	0.57	0.57	0.58	0.55	NS	0.49	0.48
M. Chan	0.10	0.11	0.12	0.14	0.14	0.17	0.17	0.19	NS	0.29	0.28
<b>Fraction of all age-0 Chinook within habitat types</b>											
Glide	0.07	0.03	0.02	0.01	0.02	0.01	0.01	0.01	NS	0.03	0.01
Pool	0.30	0.28	0.22	0.21	0.30	0.16	0.17	0.14	NS	0.23	0.24
Riffle	0.19	0.16	0.12	0.11	0.43	0.23	0.08	0.11	NS	0.18	0.15
M. Chan	0.45	0.53	0.64	0.67	0.24	0.60	0.74	0.74	NS	0.57	0.60
<b>Densities of age-0 Chinook within habitat types (fish/ha)</b>											
Glide	254	251	93	55	11	12	78	13	NS	351	187
Pool	584	1,049	619	541	82	122	607	257	NS	1,392	1,468
Riffle	116	188	124	91	38	52	79	62	NS	336	300
M. Chan	1,710	3,408	2,985	2,328	84	449	2,620	1,201	NS	1,820	2,069
<b>Number of age-0 Chinook within habitat types</b>											
Glide	2,967	2,458	857	623	137	130	837	157	NS	3,231	1,931
Pool	13,468	21,814	12,131	11,294	1,755	2,553	11,454	5,933	NS	25,890	32,612
Riffle	8,531	12,616	6,698	6,197	2,525	3,699	5,392	4,626	NS	20,629	19,754
M. Chan	20,517	42,225	35,370	36,965	1,396	9,682	50,728	30,912	NS	64,866	80,576

APPENDIX C. Continued.

Habitat	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013
<b>Proportion of total habitat available</b>											
Glide	0.07	0.07	0.08	0.08	0.07	0.09	0.08	0.08	0.08	0.07	0.07
Pool	0.17	0.16	0.16	0.16	0.17	0.23	0.22	0.23	0.18	0.23	0.23
Riffle	0.49	0.50	0.47	0.47	0.47	0.51	0.54	0.53	0.57	0.53	0.53
M. Chan	0.26	0.27	0.29	0.30	0.29	0.17	0.15	0.16	0.17	0.17	0.17
<b>Fraction of all age-0 Chinook within habitat types</b>											
Glide	0.02	0.01	0.01	0.03	0.02	0.03	0.02	0.02	0.04	0.01	0.02
Pool	0.23	0.07	0.19	0.31	0.46	0.40	0.36	0.34	0.34	0.41	0.37
Riffle	0.15	0.14	0.07	0.12	0.12	0.11	0.11	0.11	0.19	0.15	0.13
M. Chan	0.60	0.77	0.73	0.54	0.40	0.45	0.51	0.53	0.43	0.43	0.48
<b>Densities of age-0 Chinook within habitat types (fish/ha)</b>											
Glide	200	58	49	237	113	238	230	286	526	173	321
Pool	951	155	492	1,240	1,211	1,210	1,453	1,436	1,805	1,360	1,890
Riffle	216	101	60	166	118	156	175	200	330	221	281
M. Chan	1,626	1,008	1,057	1,147	603	1,872	2,993	3,293	2,515	2,061	3,190
<b>Number of age-0 Chinook within habitat types</b>											
Glide	1,884	540	442	2,498	1,120	2,668	2,371	3,164	6,122	1,535	2,822
Pool	21,091	3,183	9,626	26,754	28,851	34,314	39,382	44,765	48,846	42,209	55,651
Riffle	13,783	6,501	3,367	10,753	7,809	9,773	11,558	14,446	27,883	15,418	19,619
M. Chan	54,519	34,952	36,196	46,580	25,409	38,275	55,607	69,609	61,944	44,779	73,057

**APPENDIX C.** Concluded.

Habitat	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	Mean
<b>Proportion of total habitat available</b>											
Glide	0.07										<b>0.08</b>
Pool	0.22										<b>0.19</b>
Riffle	0.54										<b>0.53</b>
M. Chan	0.17										<b>0.20</b>
<b>Fraction of all age-0 Chinook within habitat types</b>											
Glide	0.01										<b>0.02</b>
Pool	0.37										<b>0.30</b>
Riffle	0.11										<b>0.14</b>
M. Chan	0.51										<b>0.54</b>
<b>Densities of age-0 Chinook within habitat types (fish/ha)</b>											
Glide	133										<b>175</b>
Pool	1,569										<b>1,035</b>
Riffle	190										<b>165</b>
M. Chan	2,957										<b>1,790</b>
<b>Number of age-0 Chinook within habitat types</b>											
Glide	1,120										<b>1,801</b>
Pool	44,321										<b>24,450</b>
Riffle	13,085										<b>11,121</b>
M. Chan	62,713										<b>44,404</b>

# Appendix B

**Fish Trapping at the Chiwawa and Wenatchee Smolt Traps during  
2014**





**STATE OF WASHINGTON  
DEPARTMENT OF FISH AND WILDLIFE  
FISH PROGRAM -SCIENCE DIVISION  
SUPPLEMENTATION RESEARCH TEAM**

*3515 Chelan HWY, Wenatchee, WA 98801  
Voice (509) 664-3148 FAX (509) 662-6606*

February 4, 2015

To: HCP Hatchery Committee  
From: John Walter, Fish and Wildlife Biologist  
Cc: Distribution List

**Subject: 2014 Wenatchee River Basin Smolt Monitoring**

The primary objective of smolt monitoring in the Wenatchee River basin is to estimate the number of naturally produced migrating smolts at the subbasin and watershed scale (Table 1, Figure 1) for target stocks. The smolt monitoring program also provides valuable data (e.g. freshwater productivity) essential to the monitoring and evaluation of hatchery programs (Hillman et al. 2014)

Table 1. Target stocks and corresponding smolt trapping locations used in 2014.

Stock	Smolt trap location	Smolt trap	
		Number	Diameter (m)
Chiwawa spring Chinook Salmon	Chiwawa	1	2.6
Wenatchee Sockeye Salmon	Lower Wenatchee	2	2.6
Wenatchee spring Chinook Salmon	Lower Wenatchee	2	2.6
Wenatchee summer Chinook	Lower Wenatchee	2	2.6
Wenatchee steelhead	Lower Wenatchee	2	2.6

**Methods**

*Field Operations*

The size of smolt traps operated was determined by water depth and river discharge at each of the locations. The number of smolt traps operated was determined by the expected trap efficiency. Smolt traps were located downstream from all (i.e., Chiwawa spring Chinook Salmon, Wenatchee spring Chinook Salmon, and Wenatchee Sockeye Salmon), or the majority (i.e., Wenatchee summer Chinook Salmon and Wenatchee steelhead) of the spawning areas (Figure 1).

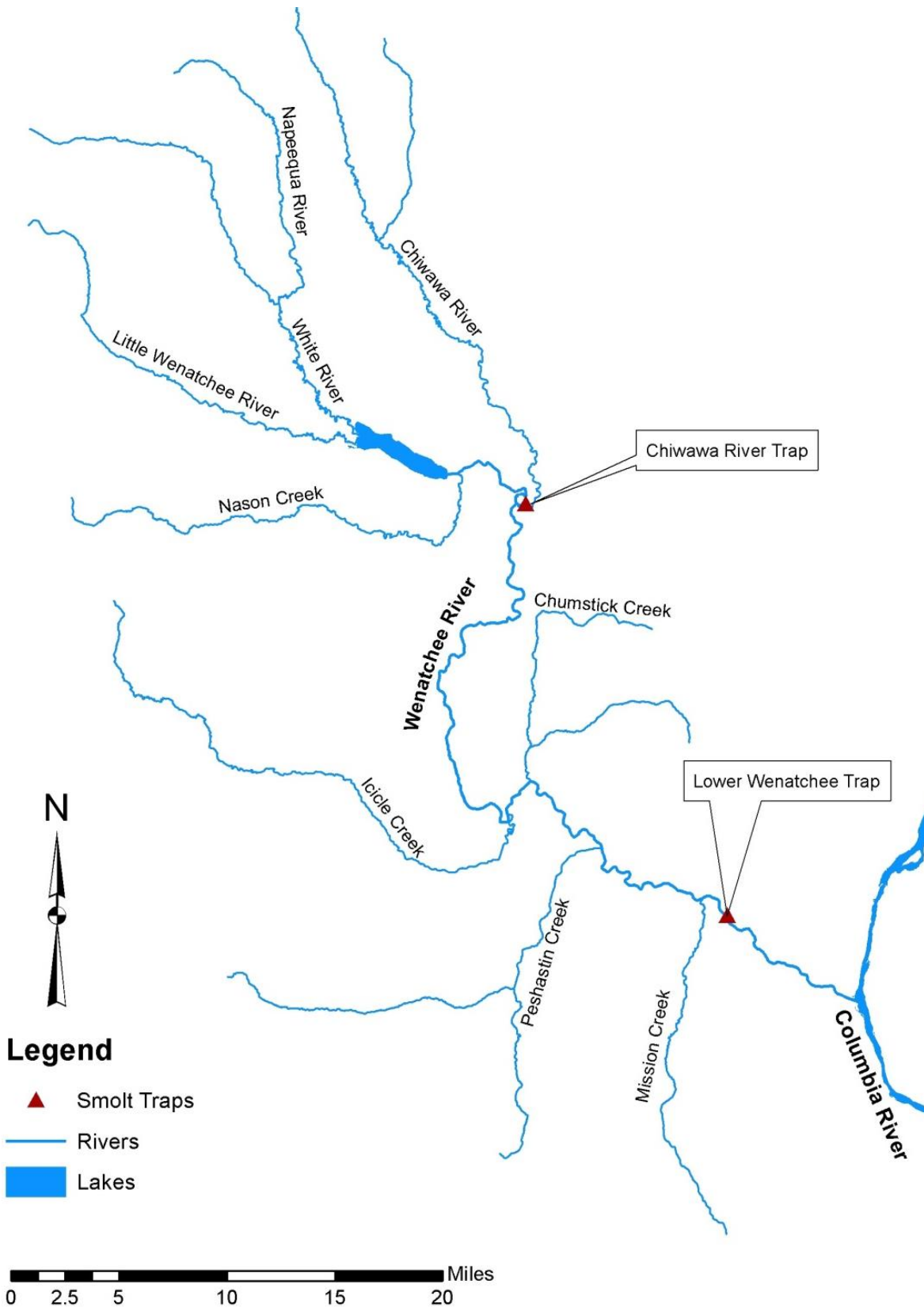


Figure 1. Locations of rotary screw trap sites in the Wenatchee Basin, 2014.

The Chiwawa trap operated nearly continuously during the trapping period. During periods of high water events and heavy river debris the trap was pulled during the day and set in the evening. Conversely, due to heavy recreational use on the lower Wenatchee River, the Wenatchee trap was primarily operated from dawn to dusk. Fish were removed from the traps at a minimum of every morning and anesthetized in a solution of MS-222. Fish were identified to species, then weighed, measured, and enumerated. Target stocks (>60 mm fork length (FL)) were tagged using 12.5 mm FDX PIT tags. All captured fishes were allowed ample recovery time and subsequently released in an area of calm water downstream from the smolt trap. Target species were held in separate live boxes when needed for mark/recapture efficiency trials.

Fork length was measured to the nearest millimeter and weight to the nearest 0.1 g. A Fulton type condition factor ( $WH10^5/FL^3$ ) was calculated for all target species (Anderson et al. 1996). The degree of smoltification (parr, transitional, or smolt) was assessed by visual examination. Juvenile yearling spring Chinook Salmon and steelhead were classified as parr if parr marks were distinct, transitional if parr marks were not distinct, and smolts if parr marks were not visible and the fish exhibited a silvery appearance. Length was used to classify juvenile subyearling Chinook Salmon and steelhead as fry (FL <50 mm) or parr (FL ≥50 mm).

Mark/recapture efficiency trials were conducted throughout the trapping season. The frequency of mark/recapture trials was dependent on the number of fish captured (i.e., no less than 100) and the river discharge. These trials were conducted over the widest range of discharge possible for target stocks at both traps. Fish utilized for mark/recapture trials were marked by clipping the tip of either the upper or lower lobe of the caudal fin or were PIT tagged. Chinook Salmon fry used in mark/recapture trials were dyed using a Bismark Brown solution. Marked fish were distributed evenly on both sides of the river in pools or in calm pockets of water around boulders. Marked fish were released between 1800 h and 2000 h. All recaptures of marked fish typically occurred within 48 h after each trial. Emigration estimates were calculated using estimated daily trap efficiency derived from the regression formula using trap efficiency (dependent variable) and discharge (independent variable). In past years, the Peterson estimator of abundance was used (Seber 1982). A modified Bailey estimator was used for the 2010-2013 brood year spring Chinook Salmon emigration at the Chiwawa Trap (Murdoch et al. 2012).

In order to estimate emigration during the non-trapping period and reduce bias in estimates of abundance, subyearling spring Chinook Salmon parr were remotely captured and tagged during September and October throughout the Chiwawa River basin. The total number of tagged fish (t) divided by the total parr abundance estimate (p) generated using standard snorkeling techniques (Hillman et al. 2013) resulted in an overall tag rate ( $t_i$ ). In order to representatively tag the population throughout all reaches, the number of fish tagged in each reach was based on the reach specific abundance encountered during snorkeling surveys.

### *Peterson Population and Variance Equations*

Trap efficiency was calculated using the following formula:

$$\text{Trap efficiency} = E_i = R / M_i$$

Where  $E_i$  is the trap efficiency during time period  $i$ ;  $M_i$  is the number of marked fish released during time period  $i$ ; and  $R_i$  is the number of marked fish recaptured during time period  $i$ . The number of fish captured was expanded by the estimated daily trap efficiency ( $e$ ) to estimate the daily number of fish migrating past the trap using the following formula:

$$\text{Estimated daily migration} = \hat{N}_i = C_i / \hat{e}_i$$

where  $N_i$  is the estimated number of fish passing the trap during time period  $i$ ;  $C_i$  is the number of unmarked fish captured during time period  $i$ ; and  $e_i$  is the estimated trap efficiency for time period  $i$  based on the regression equation.

The variance for the total daily number of fish migrating past the trap was calculated using the following formulas:

$$\text{Variance of daily migration estimate} = \text{var}[\hat{N}_i] = \hat{N}_i^2 \frac{\text{MSE} \left( 1 + \frac{1}{n} + \frac{(X_i - \bar{X})^2}{(n-1)s_x^2} \right)}{\hat{e}_i^2}$$

where  $X_i$  is the discharge for time period  $i$ , and  $n$  is the sample size. If a relationship between discharge and trap efficiency was not present (i.e.,  $P < 0.05$ ;  $r^2 \sim 0.5$ ), a pooled trap efficiency was used to estimate daily emigration:

$$\text{Pooled trap efficiency} = e_p = \sum R / \sum M$$

The daily emigration estimate was calculated using the formula:

$$\text{Daily emigration estimate} = \hat{N}_i = C_i / e_p$$

The variance for daily emigration estimates using the pooled trap efficiency was calculated using the formula:

$$\text{Variance for daily emigration estimate} = \text{var}[\hat{N}_i] = \hat{N}_i^2 \frac{e_p(1 - e_p) / \sum M}{e_p^2}$$

The total emigration estimate and confidence interval was calculated using the following formulas:

$$\text{Total emigration estimate} = \sum \hat{N}_i$$

$$95\% \text{ confidence interval} = 1.96 \times \sqrt{\sum \text{var}[\hat{N}_i]}$$

## Bailey Population and Variance Equations

Trap efficiency was calculated using the following formula:

$$\text{Trap efficiency} = E_i = R+1 / Mi,$$

$$\text{Estimated daily emigration} = \hat{N}_i = \frac{C_i + 1}{\hat{e}_i}$$

The variance of the total population abundance was calculated as follows:

$$\text{Var}\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i \text{Var}\left(\frac{(C_i + 1)}{\hat{e}_i}\right)}_{\text{Part A}} + \underbrace{\sum_i \sum_j \text{Cov}\left(\frac{(C_i + 1)}{\hat{e}_i}, \frac{(C_j + 1)}{\hat{e}_j}\right)}_{\text{Part B}}$$

Part A is the variance of the daily estimates where  $C_i$  is the number of fish caught in period  $i$ ,  $e_i$  is the estimated trap efficiency for period  $i$ , and  $Cov$  is the between day covariance for days that the same linear model is used (part B). For a more details and derivation of Peterson and Bailey estimation methods see Murdoch et al. (2012).

### Emigration during non-trapping periods

A flow-efficiency regression model was developed for the lower Chiwawa River PIT tag interrogation site (CHL) using the same mark/recapture trials used for estimating efficiency at the smolt trap. This CHL model was used to calculate emigration outside of the trapping period by incorporating the tag rate into the Bailey estimator.

$$\text{Estimated daily emigration} = \left( \hat{N}_i = \frac{C_i + 1}{\hat{e}_i} \right) / t_i$$

$$\text{Where } t_i \text{ is equal to the tag rate} = t_i = \frac{t}{p}$$

## Results

### Emigration during the winter non-trapping period (2012 Brood Year)

WDFW conducted remote tagging and capture of subyearling Chinook Salmon during October 2013. A total of 3,017 PIT tags were applied to subyearling Chinook Salmon. Tags were representatively distributed throughout the basin relative to parr encountered during snorkel surveys in August 2013 conducted by BioAnalyst (Figure 2). During winter non-trapping period, two flow efficiency models for estimating detection probability were used to calculate emigration over the CHL array. Sixteen mark/recapture trials were used to calculate detection efficiency in discharges ranging from 2.7 m<sup>3</sup>/s to 10.9 m<sup>3</sup>/s and eighteen mark/recapture trials were used to calculate detection efficiency during discharges ranging from 8.9 m<sup>3</sup>/s to 27.4 m<sup>3</sup>/s ( $R^2 = 0.75$ ,  $P < 0.01$  and  $R^2 = 0.37$ ,  $P = 0.01$  respectively). Ninety eight detections on the array paired with these models resulted in an estimate (95% C.I.) of 25,305 ( $\pm 7,683$ ) spring Chinook

Salmon (2012 BY) emigrated downstream of the PIT tag array during the non-trapping period. We assumed that all fish that emigrated past the PIT tag array also emigrated past the smolt trap location approximately 150 m downstream.

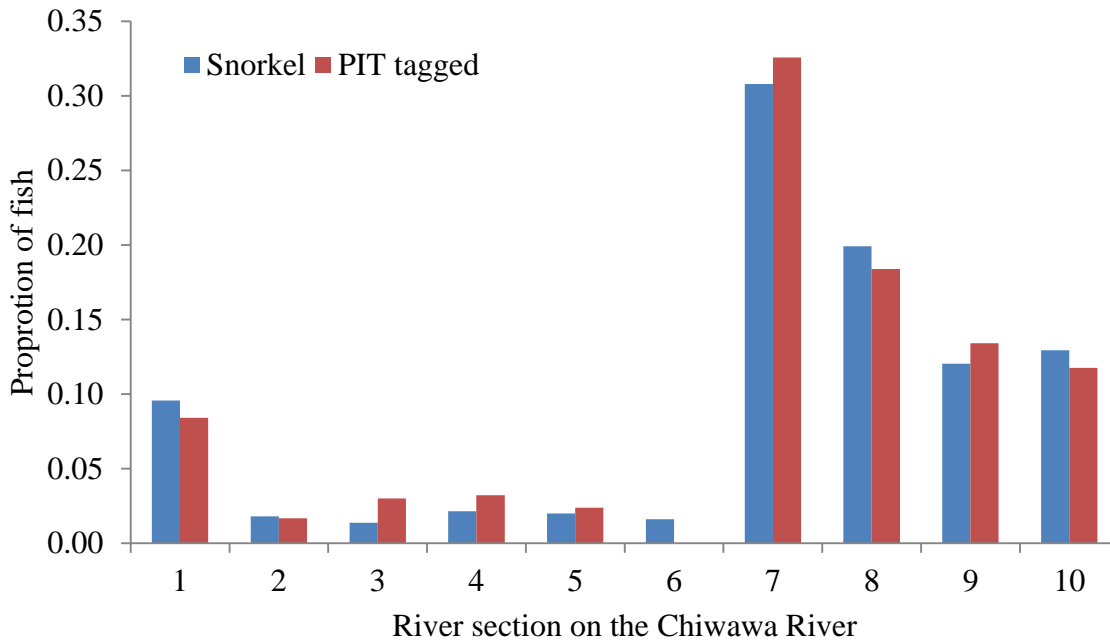


Figure 2. Distribution by reach of spring Chinook Salmon parr encountered during snorkel surveys and PIT tagged parr in the Chiwawa River 2013.

### *Chiwawa River Smolt Trap*

The Chiwawa River smolt trap was located approximately 1 km upstream from the confluence with the Wenatchee River (Figure 1). The smolt trap operated between 18 March and 13 November, when ice and slush prevented further operation of the trap. During that time period the trap was inoperable for 21 days as a result of high river flows, debris, snow/ice, or mechanical failure. During breaks in operation, the estimated number of Chinook Salmon captured was calculated from the mean number of fish captured two days prior and two days after the break in operation. The trap was operated in two positions dependent on river discharge (i.e., lower > 12 m<sup>3</sup>/s and upper < 12 m<sup>3</sup>/s). Daily trap efficiencies were estimated from two regression models (independent variable = discharge) depending on trap position and age class (i.e., upper position subyearling and lower position yearling Chinook Salmon). The subyearling estimate from fall of 2013, non-trapping estimate during the winter of 2013-2014 and the yearling estimate from the spring of 2014 were combined to estimate total abundance of the 2012 brood Chiwawa spring Chinook (Figure 3).

### 2012 Brood Year

Wild yearling spring Chinook Salmon (2012 brood) were primarily captured between 18 March and 25 June (Figure 4). A total of 4,519 yearling Chinook Salmon were captured (Appendix A)

and an estimated 5,108 would have been captured if the trap had operated without interruption. Mortality for the season totaled 28 yearling spring Chinook Salmon (0.62%). Seven mark/recapture efficiency trials were conducted in the lower position with a mean (SD) trap efficiency of 16.53% (0.05). In 2014, mark/recapture trials were conducted at all desired discharge levels and a statistically significant flow-efficiency regression model was obtained for the lower position. The 2014 regression model for the lower position ( $R^2 = 0.68$ ,  $P = 0.02$ ) was used to estimate emigration. The estimated number (95% C.I.) of yearling spring Chinook Salmon that emigrated from the Chiwawa River in 2014 was 34,334 ( $\pm 6,488$ ).

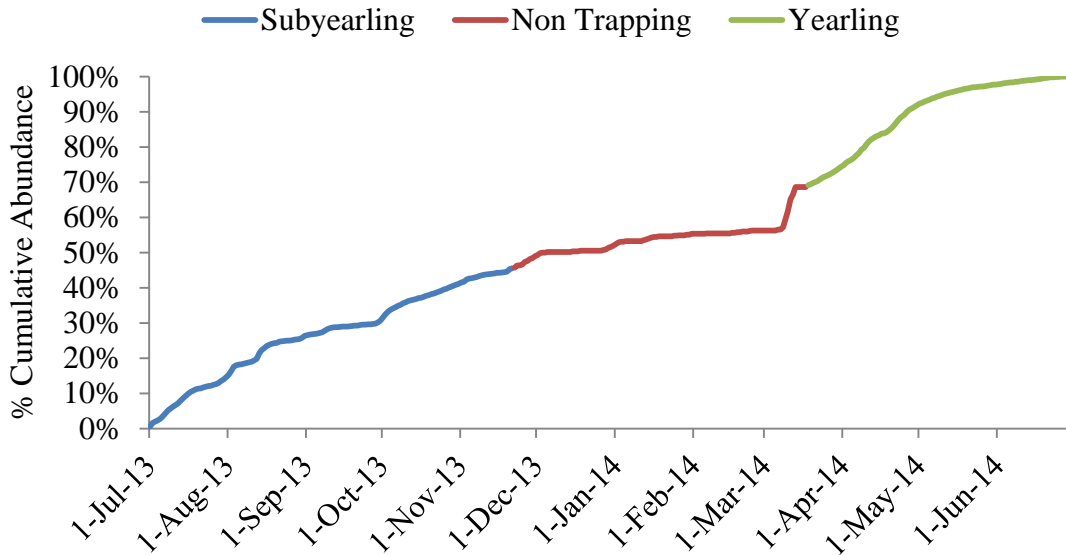


Figure 3. Percent cumulative abundance for the 2012 brood Chiwawa spring Chinook, estimated from the Chiwawa rotary screw trap and lower Chiwawa antenna array.

2013 Brood Year

Wild subyearling spring Chinook Salmon were captured between 18 March and 13 November, with major peaks occurring in July, August and October (Figure 4). A total of 19,528 subyearling parr were captured and an estimated 20,802 subyearling parr would have been captured if the trap had operated without interruption. Mortality for the season totaled 84 subyearling spring Chinook Salmon (0.35%). Thirteen mark/recapture efficiency trials were conducted with a mean (SD) trap efficiency of 25.4% (0.09), which resulted in a significant regression model (i.e., upper trap position;  $R^2 = 0.77$ ,  $P < 0.001$ ). In 2014, the estimated number of subyearling spring Chinook Salmon (excluding fry < 50 mm FL) that moved downstream of the Chiwawa River smolt trap during the sampling period was 73,695 ( $\pm 8,464$ ).

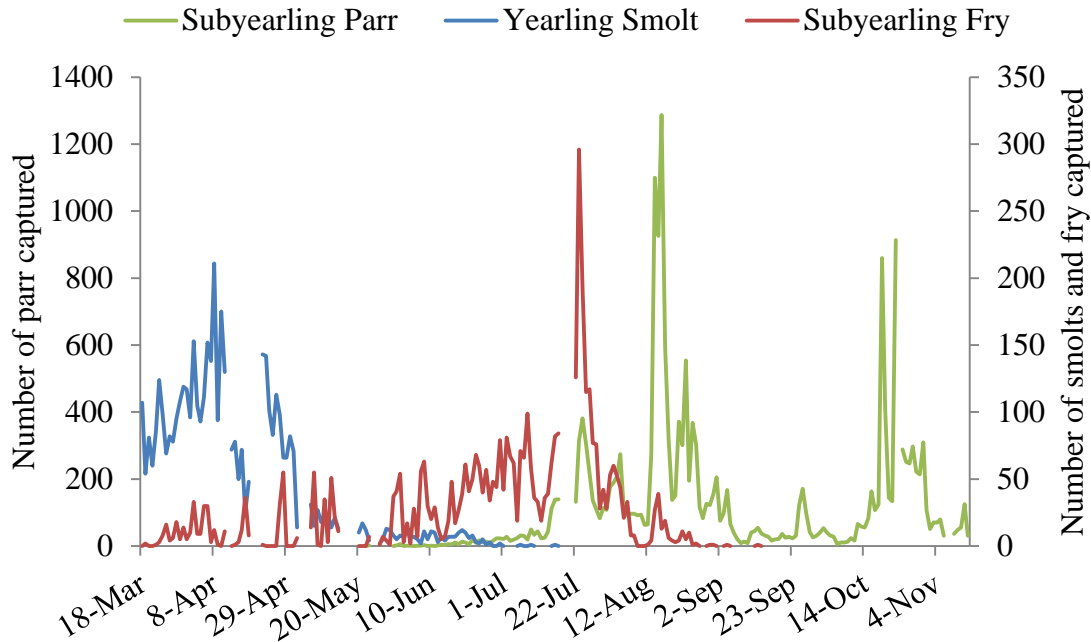


Figure 4. Daily number of spring Chinook Salmon; smolts, parr, and fry captured at the Chiwawa River smolt trap in 2014.

*Emigrant Survival*

The estimated total egg deposition was calculated by multiplying the mean fecundity of the 2012 brood spawners by the total number of redds found during surveys in the Chiwawa River basin in 2012 (Hillman et al. 2013). Egg-to-emigrant survival was calculated by dividing the estimated egg deposition by the total number of subyearling (excluding fry) that emigrated in 2013 and yearling spring Chinook Salmon that emigrated in 2014. The estimated egg-to-emigrant survival for the 2012 brood was 2.9% (Table 2).



Table 2. Estimated egg deposition (# of redds x mean broodstock fecundity) and egg-to-emigrant survival rates for Chiwawa River spring Chinook Salmon.

Brood year	Number of redds	Estimated egg deposition	Estimated number				Egg-to-emigrant survival (%)
			Sub-yearling	Non trapping	Yearling	Total emigrants	
1992	302	1,570,098	25,818		39,723	65,541	4.2
1993	106	556,394	14,036		8,662	22,698	4.1
1994	82	485,686	8,595		16,472	25,067	5.2
1995	13	66,248	2,121		3,830	5,951	9.0
1996	23	106,835	3,708		15,475	19,183	18.0
1997	82	374,740	16,228		28,334	44,562	11.9
1998	41	207,675	2,855		23,068	25,923	11.9
1999	34	166,090	4,988		10,661	15,649	9.4
2000	128	642,944	14,854		40,831	55,685	8.7
2001	1,078	4,836,704	459,784		86,482	546,266	11.0
2002	345	1,605,630	93,331		90,948	184,279	11.5
2003	111	648,684	16,881		16,755	33,637	5.2
2004	241	1,156,559	44,079		72,080	116,158	10.0
2005	333	1,436,564	108,595		69,064	177,659	12.3
2006	297	1,284,228	62,922		45,050	107,972	8.4
2007	283	1,241,521	60,196		25,809	86,006	6.9
2008	689	3,163,199	85,161		35,023	120,184	3.8
2009	421	1,925,233	30,996		30,959	61,955	3.2
2010 <sup>a</sup>	502	2,165,628	53,619		47,511	101,130	4.7
2011 <sup>a</sup>	492	2,157,420	67,982	3,665	37,185	108,832	5.0
2012 <sup>a</sup>	880	3,716,240	49,774	25,305	34,334	109,413	2.9
2013 <sup>a</sup>	714	3,367,224	73,695	--	--	--	--

<sup>a</sup> calculated with Bailey model

### *Length and Weight*

Individual length and weight measurements were recorded from a sample of the daily catch. The mean fork length (SD) of captured yearling and subyearling spring Chinook Salmon (fry excluded) was 89 (7.9) mm and 71 (10.1) mm, respectively (Table 3).

Table 3. Mean fork lengths (mm), weights (g), and body condition factor of spring Chinook Salmon captured in the Chiwawa River smolt trap during 2014.

	Yearling smolts			Subyearling parr		
	Mean	SD	N	Mean	SD	N
Fork length	89	7.9	4,212	71	10.1	12,900
Weight	7.7	2.2	3,956	3.7	1.6	7,122
K factor	1.05	0.08	3,956	1.08	0.10	7,122

### *Non-target Salmonids*

During the trapping period, 49 steelhead smolts and 1,889 steelhead/rainbow parr were captured. Mortality for the season totaled 24 steelhead juveniles (1.24%). The mean fork length (SD) of steelhead parr and smolts captured was 81(31) mm and 137 (37) mm, respectively (Table 4). Bull trout also comprised a large proportion of incidental species captured. During the trapping period, 64 adult (i.e., >300 mm) and 256 juvenile bull trout were captured (Table 5). The total number of steelhead and bull trout that emigrated from the Chiwawa River was not calculated due to the low numbers of fish captured during the sampling period. The monthly totals of all fish captured are presented in Appendix A.

Table 4. Mean fork lengths (mm), weights (g), and body condition factor of steelhead/rainbow parr and steelhead smolts captured in the Chiwawa River smolt trap during 2013.

	Parr			Smolts		
	Mean	SD	<i>N</i>	Mean	SD	<i>N</i>
Fork length	81	31.2	1,693	137	36.6	46
Weight	8.3	13.2	1,583	30.5	19.6	44
K factor	1.04	0.13	1,583	1.02	0.08	44

Table 5. Mean fork lengths (mm), weights (g), and body condition factor of bull trout captured in the Chiwawa River smolt trap during 2013. Weights were not measured on adults.

	Juvenile			Adult		
	Mean	SD	<i>N</i>	Mean	SD	<i>N</i>
Fork length	210	36.55	256	503	79.1	64
Weight	96.7	44.54	254	--	--	--
K factor	0.98	0.23	254	--	--	--

### *Lower Wenatchee River Smolt Trap*

The lower Wenatchee River smolt trap is located at rkm 13.4. Trap operations at this site began 12 February 2014 and lasted until 7 September 2014. The trap did not operate for a total of 12 days due to high flows, heavy debris or major hatchery releases.

Capture for the season totaled 1,700 wild yearling spring Chinook Salmon, 31,290 hatchery yearling Chinook Salmon, and 81,445 wild subyearling summer Chinook Salmon (Figure 5). Steelhead capture for the season totaled 102 wild steelhead parr, 80 wild steelhead smolts, and 494 hatchery steelhead (Figure 6). Wild juvenile Sockeye Salmon capture totaled 7,678 (Figure 7). Additionally, three juvenile bull trout were captured. Mortality of target species for the season totaled 250 (0.31%) subyearling summer Chinook Salmon, 17 (1.00%) wild yearling spring Chinook Salmon, 2 (1.10%) wild juvenile steelhead, 12 (2.43%) hatchery juvenile steelhead, and 128 (1.67%) wild Sockeye Salmon. Monthly totals of all species captured are listed in Appendix B.

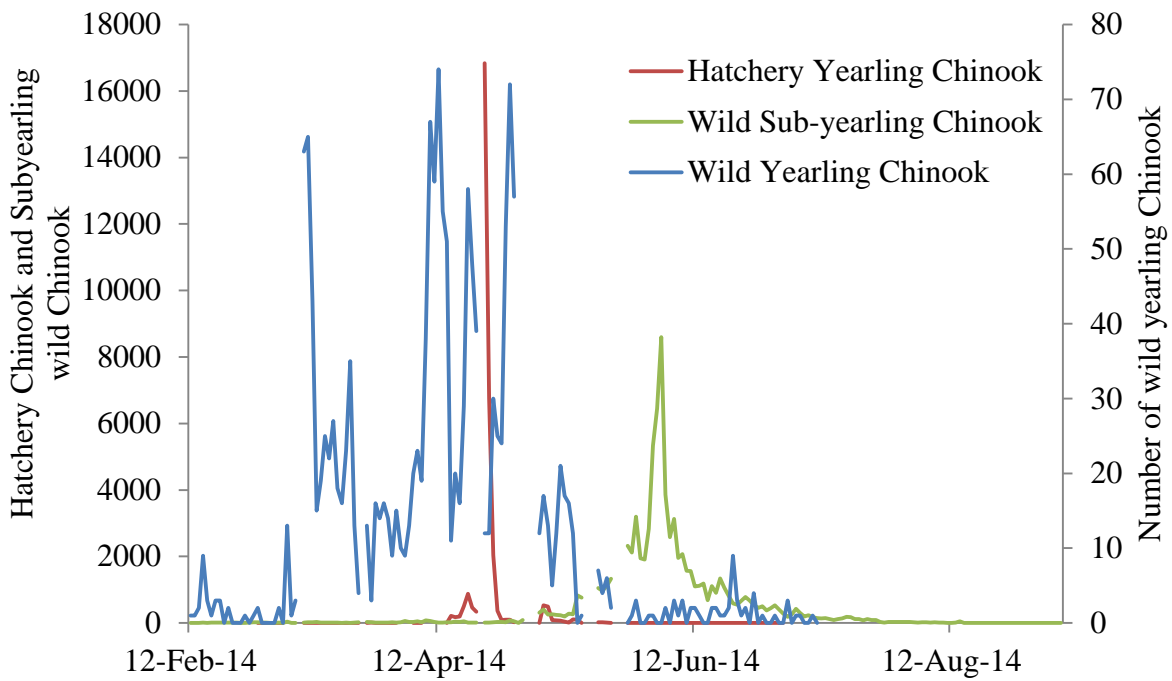


Figure 5. Daily capture of wild yearling Chinook Salmon, hatchery yearling Chinook Salmon and subyearling Chinook Salmon at the lower Wenatchee River smolt trap, 2014.

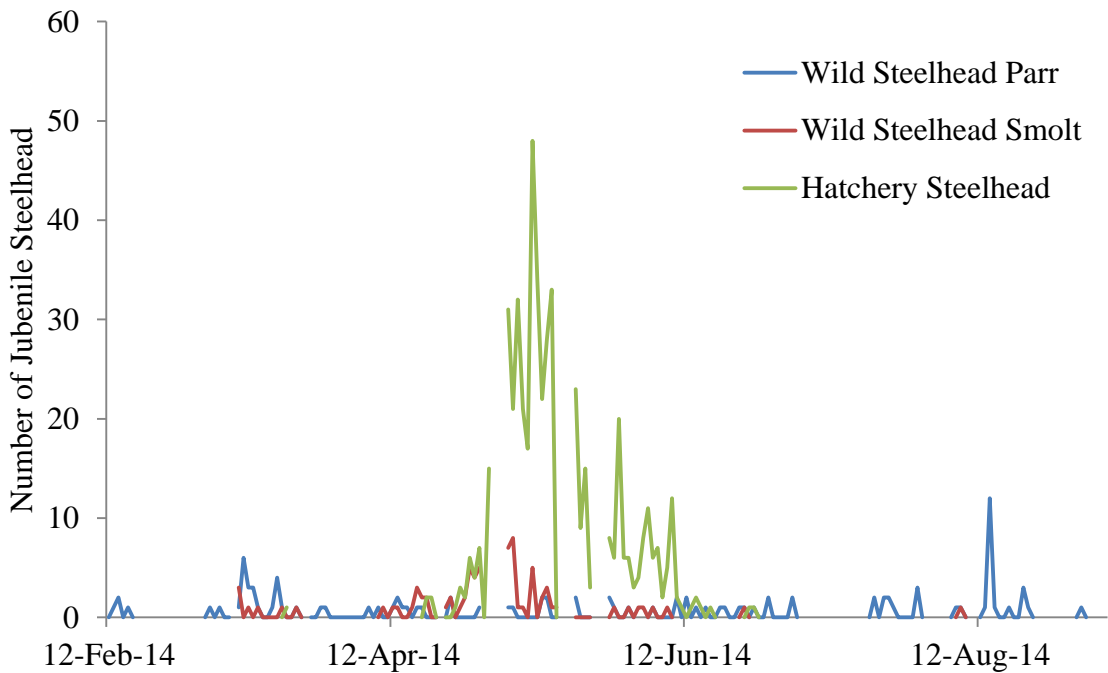


Figure 6. Daily capture of wild and hatchery steelhead at the lower Wenatchee River smolt trap, 2014.

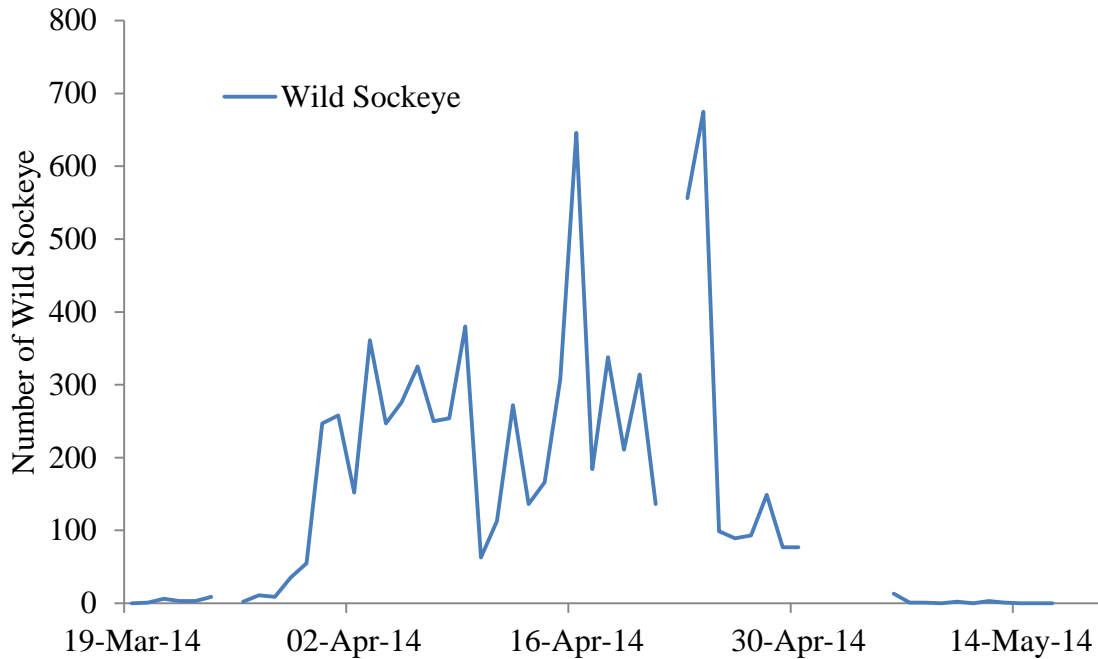


Figure 7. Daily capture of wild Sockeye Salmon at the lower Wenatchee River smolt trap, 2014.

*Wild Yearling Spring Chinook*

Five mark/recapture efficiency trials were carried out using PIT tagged yearling Spring Chinook Salmon and when combined with the previous year's mark/recapture trials a significant discharge efficiency relationship could not be obtained ( $R^2 = 0.01$ ,  $P = 0.77$ ). Therefore, a pooled estimate was calculated at 67,973 ( $\pm 431,135$ ). This resulted in a below average egg to smolt survival of 0.94% (Table 6). Capture of wild yearling Spring Chinook Salmon was primarily in March and April (Figure 5). When statistically valid discharge-efficiency relationships are developed, historical estimates using a pooled efficiency will be recalculated.

Table 6. Estimated egg deposition (# of redds x mean broodstock fecundity) and egg-to-smolt survival rates for Wenatchee Basin spring Chinook Salmon.

Brood year	Number of redds	Estimated egg deposition	Estimated number	
			Total emigrants	Egg-to-smolt survival (%)
2000	350	1,758,050	76,643	4.36
2001	1,876	8,674,624	243,516	2.81
2002	1,139	5,300,906	165,116	3.11
2003	323	1,887,612	70,738	3.75
2004	555	2,663,445	55,619	2.09
2005	829	3,587,083	302,116	8.42
2006	588	2,542,512	85,558	3.37
2007	466	2,069,506	60,219	2.91
2008	1,411	6,479,312	82,137	1.27
2009	--	--	--	--
2010	--	--	--	--
2011	872	3,823,720	89,917	2.35
2012	1,704	7,195,992	67,973	0.94

Table 7. Average length, weight and K factor for wild yearling spring Chinook Salmon sampled at the lower Wenatchee trap

	Mean	SD	N
Fork length	94	9.79	1,695
Weight	8.7	2.78	1,643
K factor	1.04	0.10	1,643

#### *Wild Subyearling Summer Chinook*

Fourteen mark/recapture efficiency trials were carried out using bismark brown subyearling Summer Chinook Salmon during the 2014 trapping season. When combined with five trials from the previous trapping season two discharge efficiency relationships were developed. A high discharge model encompassing flows ranging from 106 m<sup>3</sup>/s to 323 m<sup>3</sup>/s ( $R^2 = 0.58$ ,  $P < 0.001$ ) was used to calculate the majority of the run. This model showed a typical negative relationship where efficiency decreased as river discharge increased. At discharges ranging between 74 m<sup>3</sup>/s and 170 m<sup>3</sup>/s a positive relationship was detected where efficiency increased as discharge increased (Figure 8). However a significant relationship could not be found to model this low discharge range ( $R^2 = 0.33$ ,  $P = 0.13$ ) so a pooled estimate was used for these discharge ranges. The resulting 2013 brood estimate (95% C.I.) for subyearling Summer Chinook Salmon emigrants was 11,936,928 ( $\pm 2,448,536$ ). When expanding this trap estimate by proportion of redds (0.947) found above the trap site (Hillman et al. 2014), the total emigrant estimate was

calculated at 12,605,925 resulting in a 77.99% egg to emigrant survival rate (Table 9). In addition, using the high discharge model that incorporated efficiency trials from 2013 and 2014, the 2012 brood year estimate was recalculated from the previously reported value of 6,286,648 to 9,333,214 (Table 9).

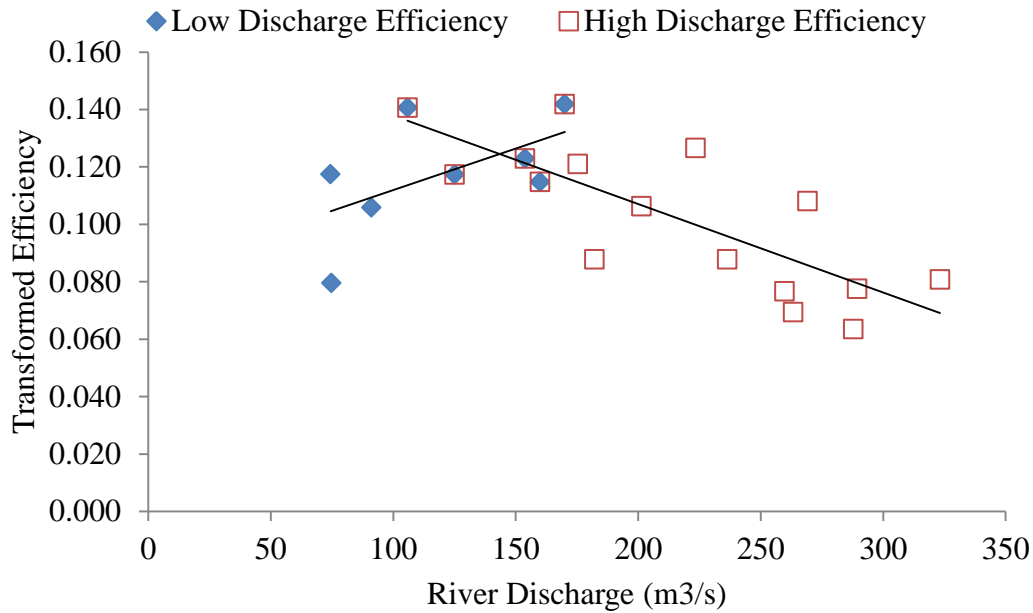


Figure 8. Mark/recapture trials using wild subyearling Summer Chinook Salmon (2013 and 2014 trapping seasons).

Table 8. Fork length, weight and K factor of subyearling Summer Chinook Salmon sampled at the lower Wenatchee smolt trap.

	Fry			Parr			Transition / Smolt		
	Mean	SD	N	Mean	SD	N	Mean	SD	N
Fork length	40	3.15	4,391	58	8.07	729	77	9.10	268
Weight	0.6	0.32	4,170	2.3	1.14	682	5.3	1.85	253
K factor	0.88	0.47	4,170	1.10	0.14	682	1.12	0.11	253

Table 9. Estimated egg deposition (peak total redd expansion x mean broodstock fecundity) and egg-to-emigrant survival rates for Wenatchee Basin summer Chinook Salmon.

Brood year	Peak total redd expansion	Estimated egg deposition	Redds above trap / total redds	Estimated number		
				Trap estimate	Total emigrants	Egg-to-emigrant survival (%)
1999	2,738	13,654,406	0.988	9,572,392	9,685,591	70.93
2000	2,540	13,820,140	0.983	1,299,476	1,322,383	9.57
2001	3,550	18,094,350	0.987	8,229,920	8,340,342	46.09
2002	6,836	37,488,624	0.977	13,167,855	13,475,368	35.95
2003	5,268	28,241,748	0.996	20,336,968	20,426,149	72.33
2004	4,874	26,207,498	0.989	14,764,141	14,935,745	56.99
2005	3,538	17,877,514	0.993	11,612,939	11,695,581	65.42
2006 <sup>b</sup>	8,896	45,663,168	0.979	9,397,044	9,595,512	21.01
2007 <sup>b</sup>	1,970	10,076,550	0.983	4,470,672	4,546,838	45.12
2008 <sup>b</sup>	2,800	14,302,400	0.978	4,309,496	4,405,473	30.80
2009 <sup>b</sup>	3,441	18,206,331	0.983	6,695,977	6,814,805	37.43
2010 <sup>b</sup>	3,261	16,184,343	0.957	--	--	--
2011 <sup>b</sup>	3,078	15,122,214	0.958	--	--	--
2012 <sup>ab</sup>	2,504	12,021,704	0.930	9,333,214	10,034,508	83.47
2013 <sup>b</sup>	3,241	16,162,867	0.947	11,936,928	12,605,925	77.99

<sup>a</sup> trap estimate updated with current model

<sup>b</sup> peak total redd expansion includes redds counted in Icicle Creek

### *Wild Sockeye*

Six mark/recapture efficiency trials were carried out using PIT tagged juvenile Sockeye Salmon. When combined with the one efficiency trial from the 2013 season this yielded a significant discharge efficiency model ( $R^2 = 0.57$ ,  $P < 0.05$ ). Using this model an estimate (95% C.I.) of 1,275,027 ( $\pm 211,615$ ) juvenile sockeye emigrants was calculated for the 2014 trapping season (Table 10).

A total of 5,083 juvenile sockeye were tagged this season at the lower Wenatchee trap. Of those fish tagged 262 fish died and 4,821 live PIT tagged Sockeye were released. Smolt survival (SE) to McNary of those tagged fish was 0.42 (0.05) using a Cormack Jolly Seber estimator.

Table 10. Age structure and estimated number of wild sockeye smolts that emigrated from Lake Wenatchee; 2013-2014.

Run year	Proportion of Wild Smolts			Total Wild Smolts
	Age 1+	Age 2+	Age 3+	
2013	0.933	0.047	0.000	873,096
2014	0.953	0.067	0.000	1,275,027

Table 11. Estimated egg deposition (estimated as mean fecundity times estimated number of females), number of smolts, and survival rates for wild Wenatchee sockeye salmon, 2013-14; NA = Not available.

brood year	# of females	Mean fecundity	Total eggs	# of Wild Smolts				Egg-smolt survival
				Age 1+	Age 2+	Age 3+	Total	
2011	9,722	2,960	28,777,120	814,960	60,382	NA	875,342	3.04
2012	14,753	2,745	40,496,573	1,214,645	NA	NA	1,214,645	3.00

Table 12. Fork length, weight and K factor of wild Sockeye Salmon smolts sampled at the lower Wenatchee smolt trap.

	Mean	SD	N
Fork length	76	8.48	5,623
Weight	3.7	1.54	4,294
K factor	0.81	0.16	4,294

### *Wild Summer Steelhead*

Capture of wild steelhead at the lower site was low totaling only 182 parr and smolts combined. Capture of hatchery steelhead was also low totaling only 494 fish. Due to low capture of both wild and hatchery steelhead, mark/recapture trials using wild steelhead could not be conducted. Using hatchery steelhead as surrogates (wild parents), 473 hatchery steelhead were marked and transported from Chiwawa Ponds and released at Dryden Dam. Two separate trials were conducted resulting in a pooled efficiency of 0.036. The pooled efficiency estimate (95% C.I.) was calculated at 6,149 ( $\pm 32,095$ ) parr and smolt wild emigrant steelhead.

Table 13. Fork length, weight and K factor of wild steelhead sampled at the lower Wenatchee smolt trap.

	Parr			Transitional / Smolt		
	Mean	SD	N	Mean	SD	N
Fork length	90	32.55	97	165	33.79	80
Weight	10.4	13.81	96	46.8	23.13	80
K factor	1.03	0.12	96	0.96	0.15	80



## Discussion

### *Chiwawa River Smolt Trap*

The total production estimate for the 2012 brood year was 109,413 ( $\pm 11,723$ ) and comprises estimates of subyearling emigrants in 2013, emigrants during the non-trapping period and yearling emigrants in 2014. Egg to emigrant survival for this brood year was 2.9%.

The 2013 brood subyearling spring Chinook Salmon trapping ended abruptly on 13 November, 2014 as a result of freezing river conditions and may represent the earliest date trapping has ended on the Chiwawa River. While subyearling Chinook Salmon parr were again remotely captured and tagged upstream of the trap and detections at lower Chiwawa PIT tag array will be used to estimate emigration during the non-trapping period, the fall remote capture and tagging effort was also hindered by large rain and discharge events that made it nearly impossible to safely wade and conduct electrofishing surveys. When river discharge had receded and surveys were continued, the cold water and weather conditions made captures difficult. As a result of the same poor environment conditions that resulted in an early cessation to trapping, the tagging goal of 3000 PIT tagged subyearlings was not met ( $N = 1,024$ ). The lower tag rate and subsequent higher than average river discharge in the Chiwawa River during the non-trapping period will increase the uncertainty in the estimating emigrant abundance in the non-trapping period which will be reported in the 2015 report.

This is the second reporting cycle that an estimate for non-trapping emigration has been reported. The 2012 brood non-trapping period estimate is higher than the previous 2011 brood (25,305 vs. 3,665). This estimate may have been influenced by a lower tag rate for the 2012 brood (0.020 vs. 0.031), but more likely the later than usual installation of the Chiwawa trap (March 18, 2014 vs. February 22, 2013). From March 6 to March 13, 2014 nineteen wild Chinook Salmon tagged the previous fall were detected at the lower Chiwawa array. These detections expanded to 13,506 wild Chinook Salmon leaving the Chiwawa during that time frame which accounts for 53% of the total non-trapping estimate. These data show that maximizing the trapping period, both in the spring and fall, is critically important in reducing bias.

### *Lower Wenatchee River Smolt Trap*

Discharge efficiency models were obtained for two of the four target species at the lower Wenatchee trap during the 2014 trapping season (wild Sockeye Salmon and wild summer Chinook Salmon). While the development of a discharge efficiency model for wild yearling Spring Chinook Salmon continues the pooled estimate should be considered preliminary and is most likely negatively biased as is the pooled estimate for wild steelhead. Conversely, both of the production estimates for wild Sockeye Salmon and wild summer Chinook Salmon have a PSE of 8.5% and 10.5% respectively. While the new trap location has allowed for a greater operational flexibility, it does require the development of new flow-efficiency models. While this can be accomplished relatively quickly with species that are relatively abundant (e.g., summer Chinook and sockeye), it may take several years for those species in low abundance (ESA-listed). Fortunately, given similar operation parameters across time, we will be able to

reexamine past abundance estimates when those models are fully developed. The persistently low number of wild steelhead smolts captured, regardless of trap location, may require the use of hatchery steelhead as surrogates and may be the only option for developing efficiency models to estimate emigration. This approach requires the assumption that hatchery fish behave in a like manner to wild fish. An assumption we will try to test over time when and where possible.

## References

- Anderson, R. O., & Neumann, R. M. (1996). Length, Weight, and Associated Structural Indices. In Murphy B. E. & Willis D. W. (Eds.), *Fisheries Techniques* (2nd ed. pp. 461-480). American Fisheries Society.
- Bailey, N.T.J. 1951. On estimating the size of mobile populations from capture-recapture data. *Biometrika* 38:293-306.
- Hillman, T.W. and M.D. Miller. 2002. Abundance and total numbers of Chinook salmon and trout in the Chiwawa River Basin, Washington, 2001. Report to Chelan county Public Utility District, Wenatchee, Washington.
- Hillman, T., M. Miller, L. Keller, J. Murauskas, T. Miller, M. Tonseth, M. Hughes, A. Murdoch. 2013. Monitoring and evaluation of the Chelan County PUD hatchery programs: 2012 annual report. Report to the HCP Hatchery Committee, Wenatchee, WA.
- Hillman, T., M. Miller, C. Moran, M. Tonseth, M. Hughes, A. Murdoch, L. Keller, C. Willard, B. Ishida, C. Kamphaus, T. Pearsons, and P. Graff. 2014. Monitoring and evaluation of the Chelan and Grant County PUDs hatchery programs: 2013 annual report. Report to the HCP and PRCC Hatchery Committees, Wenatchee, WA.
- Murdoch, A.R., Miller, T.L., Truscott, B.L., Snow, C., Frady, C., Ryding, K., Arterburn, J.E., Hathaway, D. 2012. Upper Columbia Spring Chinook Salmon and Steelhead Juvenile and Adult Abundance, Productivity, and Spatial Structure Monitoring. BPA Project No. 2010-034-00. Bonneville Power Administration, Portland, Oregon.
- Ryding, K.E., 2002. Appendix A - Review: 2000 Skagit River wild 0+ Chinook production evaluation *in* D. Seiler, S. Neuhauser, and L. Kishimoto 2002. 2001 Skagit River wild 0+ Chinook production evaluation, FPA 02-11. Washington Department of Fish and Wildlife, Olympia, WA.
- Seber, G.A.F. 1982. The estimation of animal abundance and related parameters, 2nd edition. Macmillan Co., New York, New York, USA
- Walter, J.B., B.L Truscott, A.R. Murdoch and T.L. Miller. 2011. 2011 Chiwawa and Wenatchee River Smolt Estimates. Report to Chelan county Public Utility District, Wenatchee, Washington.

Appendix A. Monthly total juvenile capture information for the Chiwawa River smolt trap.

Species/Origin	2014										
	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Total
<b>Chinook</b>											
<i>Wild yearling</i>	0	1,278	2,697	360	181	3	0	0	0	0	4,519
<i>Wild subyearling</i>	0	82	295	299	1,288	4,753	9,478	1,635	5,217	708	23,755
<i>Hatchery yearling</i>	0	0	5,289	1	0	0	3	0	0	0	5,293
<b>Steelhead</b>											
<i>Wild</i>											
<i>Smolt</i>	0	1	17	17	12	0	2	0	0	0	49
<i>Parr</i>	0	34	64	334	334	48	495	250	313	96	1,889
<i>Hatchery</i>	0	0	215	62	8	0	5	0	0	0	290
<b>Coho</b>											
<i>Wild yearling</i>	0	2	0	0	0	0	0	0	0	0	0
<i>Wild subyearling</i>	0	0	0	0	5	4	0	0	0	0	12
<i>Hatchery yearling</i>	0	0	1	0	0	0	0	0	0	0	1
<b>Bull trout</b>											
<i>Juvenile</i>	0	4	5	6	19	3	10	100	102	11	260
<i>Adult</i>	0	0	0	0	0	0	0	60	21	0	75
Cutthroat	0	0	0	1	12	5	18	21	2	0	59
Eastern brook	0	0	0	1	2	0	0	2	7	0	12
Whitefish	0	27	5	3	15	469	1,675	747	21	8	2,970
Northern pikeminnow	0	0	0	0	0	0	2	3	0	0	5
Longnose dace	0	18	18	114	710	170	219	964	408	12	2,633
Sucker spp.	0	0	0	0	0	0	2	2	0	0	4
Redside shiner	0	0	0	0	0	1	1	0	0	0	0
Yellow perch	0	0	0	0	0	0	0	0	0	0	0
Sculpin spp.	0	7	4	5	12	30	20	29	24	0	131

Appendix B. Monthly total juvenile capture information for the lower Wenatchee River smolt trap.

Species/Origin	2014										Total
	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	
<b>Chinook</b>											
<i>Wild yearling</i>	29	483	968	169	44	7	0	0	--	--	1,700
<i>Wild subyearling</i>	114	338	860	20,073	54,824	4,975	249	12	--	--	81,445
<i>Hatchery yearling</i>	0	25	29,054	2,196	14	1	0	0	--	--	31,290
<b>Steelhead</b>											
<i>Wild</i>											
<i>Smolt</i>	0	7	31	36	5	0	1	0	--	--	80
<i>Parr</i>	4	25	12	13	14	12	21	1	--	--	102
<i>Hatchery</i>	0	1	27	398	68	0	0	0	--	--	494
<b>Sockeye</b>											
<i>Wild</i>	0	381	7,205	89	2	1	0	0	--	--	7,678
<i>Hatchery</i>	0	0	0	0	0	0	0	0	--	--	0
<b>Coho</b>											
<i>Wild yearling</i>	2	57	72	36	43	3	7	0	--	--	220
<i>Wild subyearling</i>	1	22	19	30	145	162	14	0	--	--	393
<i>Hatchery yearling</i>	0	5	14,131	2,513	258	1	0	0	--	--	16,908
<b>Bull trout</b>											
<i>Juvenile</i>	0	0	1	1	1	0	0	0	--	--	3
<i>Adult</i>	0	0	0	0	0	0	0	0	--	--	0
Cutthroat	0	0	0	1	0	1	1	0	--	--	3
White fish	0	1	1	3	3	7	12	0	--	--	27
Northern pikeminnow	0	1	1	2	14	7	9	3	--	--	37
Longnose dace	12	210	85	34	23	61	104	12	--	--	541
Speckled dace	0	0	0	2	1	31	0	0	--	--	34
Umatilla dace	0	0	0	0	0	0	0	0	--	--	0
Sucker spp.	1	13	6	30	6	36	40	2	--	--	134
Peamouth	0	0	0	1	0	2	6	0	--	--	9
Chiselmouth	0	0	0	1	0	3	60	4	--	--	69
Redside shiner	0	9	0	3	6	40	32	4	--	--	94
Yellow bullhead	0	0	0	0	0	0	0	0	--	--	0
Pacific lamprey	10	171	13	13	56	19	10	0	--	--	292
River lamprey	0	0	0	0	0	0	0	0	--	--	0
Sculpin spp.	3	16	15	12	16	18	42	6	--	--	128
Stickleback (3 spined)	0	0	0	3	2	35	26	0	--	--	66

Appendix C. Yearly total juvenile capture information for the Chiwawa River smolt trap.

Species origin	2013	2012	2011	2010	2009	2008	2007	2006	2005	2004	2003	2002	2001	2000
Chinook														
<i>Wild yearling</i>	3,199	7,626	4,848	6,482	3,765	8,711	4,433	4,974	2,874	4,326	8,012	1,423	2,763	1,791
<i>Wild subyearling</i>	27,621	14,831	20,561	13,344	30,641	12,741	16,286	14,584	10,933	5,257	25,150	53,818	5,188	1,480
<i>Hatchery yearling</i>	15,909	30,751	25,620	22,481	14,097	22,367	17,634	9,796	3,965	7,557	5,893	2,926	0	6
Steelhead														
<i>Wild</i>	2,034	1,921	1,176	1,226	1,957	1,700	1,211	1,789	1,672	2,441	1,662	778	1,091	326
<i>Smolt</i>	85	183	195	210	248	448	152	53	45	280	32	86	63	181
<i>Parr</i>	1,949	1,738	981	1,016	1,709	1,250	1,056	1,736	1,627	2,161	1,630	692	1,028	145
<i>Hatchery</i>	1,539	1,664	8,250	9,921	2,708	2,684	1,964	1,384	2,104	9,678	5,886	2,720	134	45
Coho														
<i>Wild yearling</i>	1	1	3	4	0	0	0	3	4	0	0	0	0	0
<i>Wild subyearling</i>	0	0	4	5	1	13	12	2	0	0	0	0	0	0
<i>Hatchery yearling</i>	10	3	0	3	3	1	0	126	8	0	0	0	2	0
Bull Trout														
<i>Juvenile</i>	310	488	351	499	496	513	250	125	175	238	438	339	264	421
<i>Adult</i>	51	31	7	45	24	33	29	39	41	12	6	8	25	19
Cutthroat	86	60	38	54	66	52	40	56	44	45	28	37	183	22
Eastern brook	13	66	3	0	8	4	3	4	4	2	6	7	25	10
Whitefish	2,108	3,291	990	778	3,340	2,672	2,186	2,267	3,672	3,669	1,212	871	1,825	837
Northern pikeminnow	71	34	20	5	47	7	15	0	0	13	1	3	14	12

## Appendix C. cont.

Species origin	2013	2012	2011	2010	2009	2008	2007	2006	2005	2004	2003	2002	2001	2000
Longnose dace	2257	1,762	1,526	1,393	2,081	2,934	2,349	1,951	3,133	3,162	1,557	604	1,217	1,456
Sucker spp.	6	0	0	0	7	9	1	8	10	5	4	0	6	40
Redside shiner	0	0	0	0	0	0	0	1	2	1	1	0	0	0
Yellow perch	0	0	0	0	0	0	0	0	0	0	0	1	14	0
Sculpin spp.	91	157	129	51	78	143	73	104	23	34	13	58	77	56

Appendix D. Yearly total juvenile capture information for the lower Wenatchee River smolt trap.

Species/Origin	2013	2010	2009	2008	2007	2006	2005	2004	2003	2002	2001
<b>Chinook</b>											
<i>Wild yearling</i>	1,854	1,079	5,346	612	1,906	652	333	1,061	1,619	336	206
<i>Wild subyearling</i>	52,652	50,685	37,568	30,547	86,142	63,580	224,858	225,549	110,528	39,714	70,952
<i>Hatchery yearling</i>	13,979	43,613	6,709	19,440	45,467	35,261	23,709	11,846	20,939	3,421	8,758
<b>Steelhead</b>											
<i>Wild</i>	710	484	264	319	495	151	246	360	413	252	341
<i>Smolt</i>	173	407	216	220	433	105	210	299	343	187	273
<i>Parr</i>	537	77	48	99	62	45	36	61	70	76	68
<i>Hatchery</i>	819	2,735	1,949	2,106	2,697	3,769	2,013	3,465	2,175	2,260	1,711
<b>Sockeye</b>											
<i>Wild</i>	4,520	3,153	1,259	216	6,340	5,204	202	3,224	7,544	5,042	58
<i>Hatchery</i>	72	440	263	207	248	68	79	335	271	281	131
<b>Coho</b>											
<i>Wild yearling</i>	597	188	114	111	292	103	189	58	199	72	0
<i>Wild subyearling</i>	923	2,112	515	1,013	431	1,460	1,846	927	29	1,443	191
<i>Hatchery yearling</i>	12,960	8,013	9,709	4,296	29,305	13,627	11,943	15,455	8,034	12,363	11,265
<i>Bull Trout Juvenile</i>	6	2	0	1	2	1	3	2	0	1	1
<i>Bull Trout Adult</i>	0	0	0	0	0	0	0	0	0	0	0
<i>Cutthroat</i>	0	0	1	1	0	0	0	0	0	0	2
<i>Whitefish</i>	110	48	52	67	23	118	9	34	115	31	78
<i>Northern pikeminnow</i>	39	198	13	57	135	475	90	75	21	93	10
<i>Longnose dace</i>	1,382	643	383	568	1,820	801	659	2,374	488	593	445
<i>Speckled dace</i>	0	0	0	1	0	0	0	5	4	3	7
<i>Umatilla dace</i>	0	0	0	2	0	0	0	2	1	12	36
<i>Sucker spp.</i>	240	390	63	612	339	3,420	203	208	172	169	201
<i>Peamouth</i>	10	62	1	2	1	0	0	0	0	0	0



## Appendix D. cont.

Species/Origin	2013	2010	2009	2008	2007	2006	2005	2004	2003	2002	2001
Chiselmouth	10	1	0	0	1	32	0	7	2	7	1
Redside shiner	423	570	18	69	84	952	166	100	14	47	47
Yellow bullhead	0	1	0	0	0	0	0	0	0	0	0
Pacific lamprey	762	680	1,245	1,431	2,876	1,933	685	650	922	978	1,267
River lamprey	0	0	0	0	0	0	0	0	1	0	18
Sculpin spp.	242	70	123	49	64	118	171	86	71	97	55
Stickleback (3-spined)	196	4	7	4	39	78	51	85	18	48	246



# Appendix C

**Summary of PIT-Tagging Activities in the Wenatchee Basin,  
2014**



**Appendix C.** Numbers of fish captured, PIT tagged, lost, and released in the Wenatchee River basin during February through November, 2014.

Sampling Location	Species and Life Stage	Number held	Number of recaptures	Number tagged	Number died	Shed Tags	Total released	Percent mortality
Chiwawa Trap	Wild Subyearling Chinook	11,803	3,782	11,375	14	3	11,358	0.12
	Wild Yearling Chinook	4,476	75	4,399	16	0	4,383	0.36
	Wild Steelhead/Rainbow	1,288	4	1,195	9	0	1,186	0.70
	Hatchery Steelhead/Rainbow	11	3	3	0	0	3	0.00
	Wild Coho	0	0	0	0	0	0	--
	<b>Total</b>		<b>17,578</b>	<b>3,864</b>	<b>16,972</b>	<b>39</b>	<b>3</b>	<b>16,930</b>
Chiwawa River (Electrofishing)	Wild Subyearling Chinook	1,082	2	1,034	0	2	1,032	0.00
	Wild Yearling Chinook	0	0	0	0	0	0	--
	Wild Steelhead/Rainbow	94	0	23	0	0	23	0.00
	Hatchery Steelhead/Rainbow	0	0	0	0	0	0	--
	Wild Coho	0	0	0	0	0	0	--
	<b>Total</b>		<b>1,176</b>	<b>2</b>	<b>1,057</b>	<b>0</b>	<b>2</b>	<b>1,055</b>
Nason Creek (Electrofishing)	Wild Subyearling Chinook	1,908	28	1,821	4	1	1,816	0.21
	Wild Yearling Chinook	0	0	0	0	0	0	--
	Wild Steelhead/Rainbow	0	0	0	0	0	0	--
	Hatchery Steelhead/Rainbow	0	0	0	0	0	0	--
	Wild Coho	0	0	0	0	0	0	--
	<b>Total</b>		<b>1,908</b>	<b>28</b>	<b>1,821</b>	<b>4</b>	<b>1</b>	<b>1,816</b>
Lower Wenatchee Trap	Wild Subyearling Chinook	36	0	36	0	0	36	0.00
	Wild Yearling Chinook	1,631	107	1,521	15	0	1,506	0.92
	Wild Steelhead/Rainbow	143	5	133	0	0	133	0.00
	Hatchery Steelhead/Rainbow	8	4	4	0	0	4	0.00
	Wild Coho	1	0	1	0	0	1	0.00
	Wild Sockeye	5,096	4	5,083	262	0	4,821	5.14
	<b>Total</b>		<b>6,915</b>	<b>120</b>	<b>6,778</b>	<b>277</b>	<b>0</b>	<b>6,501</b>
<b>Total:</b>	<b>Wild Subyearling Chinook</b>	<b>14,829</b>	<b>3,812</b>	<b>14,266</b>	<b>18</b>	<b>6</b>	<b>14,242</b>	<b>0.12</b>
	<b>Wild Yearling Chinook</b>	<b>6,107</b>	<b>182</b>	<b>5,920</b>	<b>31</b>	<b>0</b>	<b>5,889</b>	<b>0.51</b>
	<b>Wild Steelhead/Rainbow</b>	<b>1,525</b>	<b>9</b>	<b>1,351</b>	<b>9</b>	<b>0</b>	<b>1,342</b>	<b>0.59</b>
	<b>Hatchery Steelhead/Rainbow</b>	<b>19</b>	<b>7</b>	<b>7</b>	<b>0</b>	<b>0</b>	<b>7</b>	<b>0.00</b>
	<b>Wild Coho</b>	<b>1</b>	<b>0</b>	<b>1</b>	<b>0</b>	<b>0</b>	<b>1</b>	<b>0.00</b>
	<b>Wild Sockeye</b>	<b>5,096</b>	<b>4</b>	<b>5,083</b>	<b>262</b>	<b>0</b>	<b>4,821</b>	<b>5.14</b>
<b>Grand Total:</b>		<b>27,577</b>	<b>4,014</b>	<b>26,628</b>	<b>320</b>	<b>6</b>	<b>26,302</b>	<b>1.16</b>



# Appendix D

**Wenatchee Steelhead Spawning Escapement Estimates, 2014**





# Wenatchee Steelhead Spawning Escapement Estimates in 2014

Kevin See

August 15, 2014

## Introduction

Redd counts are an established method to provide an estimate of escapement. In the Wenatchee subbasin, index reaches are surveyed weekly during the steelhead spawning season (Mar 14, 2014 - Jun 02, 2014) and non-index reaches are surveyed once during the peak spawning period. The goal of this work is to:

- Predict observer net error, based on a model developed with data from steelhead redd surveys in the Methow as described Truscott et al. (2015).
- Use estimates of observer net error rates and redd life to estimate total escapement for each index reach, using a Gaussian area under the curve (GAUC) technique described in Millar et al. (2012).
- Estimate escapement to the non-index reaches by matching them with an index reach and assuming some of the same parameters estimated by the GAUC model for the matching index reach.

## Methods

The model for observer net error (observed redd counts / true number of redds) is a model averaging of the two best models that were fit to 43 data points in the Methow. Both models contained covariates of observed redd density (redds / m<sup>2</sup>) and mean thalweg CV as a proxy for channel complexity. One model also contained discharge while the other also contained total redd survey experience as an additional covariate. Predictions were made using each model and the 2014 steelhead data, and were then averaged together using the model weights (based on AICc). From these survey specific estimates of net error, a mean and standard error of net error was calculated for each reach. The standard deviation was calculated by taking the square root of the sum of the squared standard errors for all predictions within a reach.

Redd life data was recorded in 2014, and for each reach the mean and standard deviation of redd life was calculated. One reach, W8, did not have any estimates of redd life. Therefore, we used the mean of redd life, and the mean of the standard deviation of redd life, in other index reaches in the Wenatchee for W8.

Escapement estimates were made for each index reach using the GAUC model described in Millar et al. (2012). Part of that model estimates the day of peak spawning for a reach, and the variation around that peak day. For the non-index reaches, those two parameters were borrowed from an associated index reach (Blue curves in Fig 1). The third parameter in the GAUC model was then estimated based on the single data point from the non-index reach (Fig 2). Net error rates were estimated for the non-index reaches in the same way as the index reaches. Estimates of the mean and standard deviation of redd life were borrowed from the associated index reach. All of these estimates were then inserted into the GAUC model to generate estimates of escapement for the non-index reaches.

## Results

Net error rates were estimated to be well below 1, suggesting the observed number of redds was an underestimate of the true number of redds. The standard error of that error was quite high however, indicating the large amount of uncertainty in redd counts (Table 1). The GAUC model fit the data in the index reaches well (Figure 1), and provided estimates of peak spawning day and the variance around when steelhead spawned in that reach (Table 2). These estimates were applied to the non-index reaches (Figure 2), and then estimates of the number of redds in each reach were calculated (Table 3). The estimated number of redds was always higher than the observed number of redds.

Table 1. Estimated net error and redd life for each index reach in the Wenatchee River in 2014.

Reach	Net Error	Net Error SE	Net Error CV	Redd Life Mean	Redd Life SD	Redd Life CV
W10	0.62	1.13	1.83	27.6	13.2	0.48
W2	0.75	1.28	1.72	18.0	0.0	0.00
W6	0.42	1.46	3.45	34.6	12.0	0.35
W8	0.49	0.92	1.88	26.8	9.1	0.34
W9	0.61	1.45	2.37	27.1	11.2	0.41

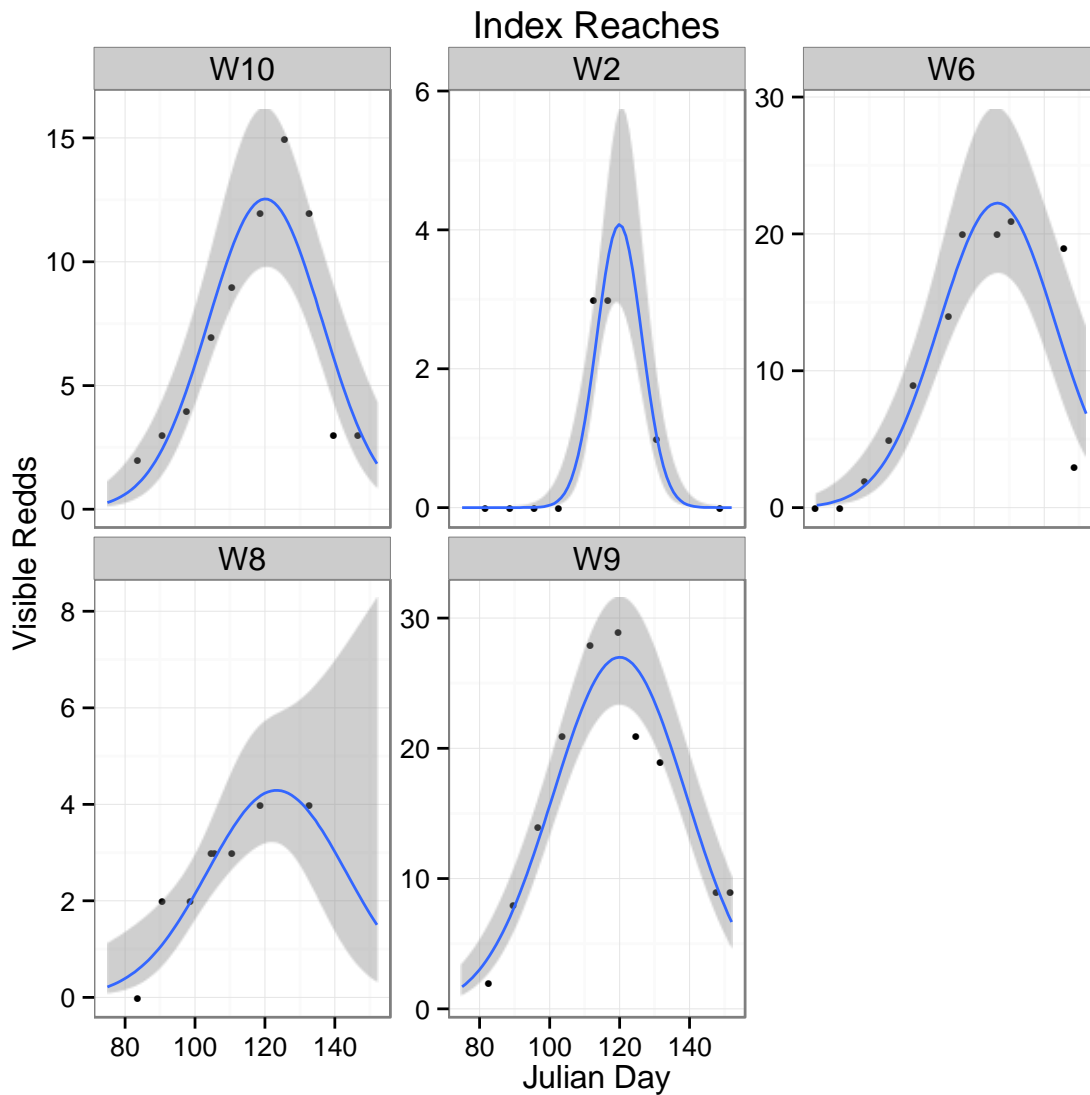


Figure 1. Plots of observed redd counts (black dots) through time for each index reach, and the fitted curve from the GAUC model (blue line) with associated uncertainty (gray).

Table 2. Estimated Julian day of peak spawning for each index reach, and variation around that day in the Wenatchee River in 2014.

Reach	Peak Spawning	Spread
W10	120	265
W2	120	41
W6	127	274
W8	123	392
W9	120	366

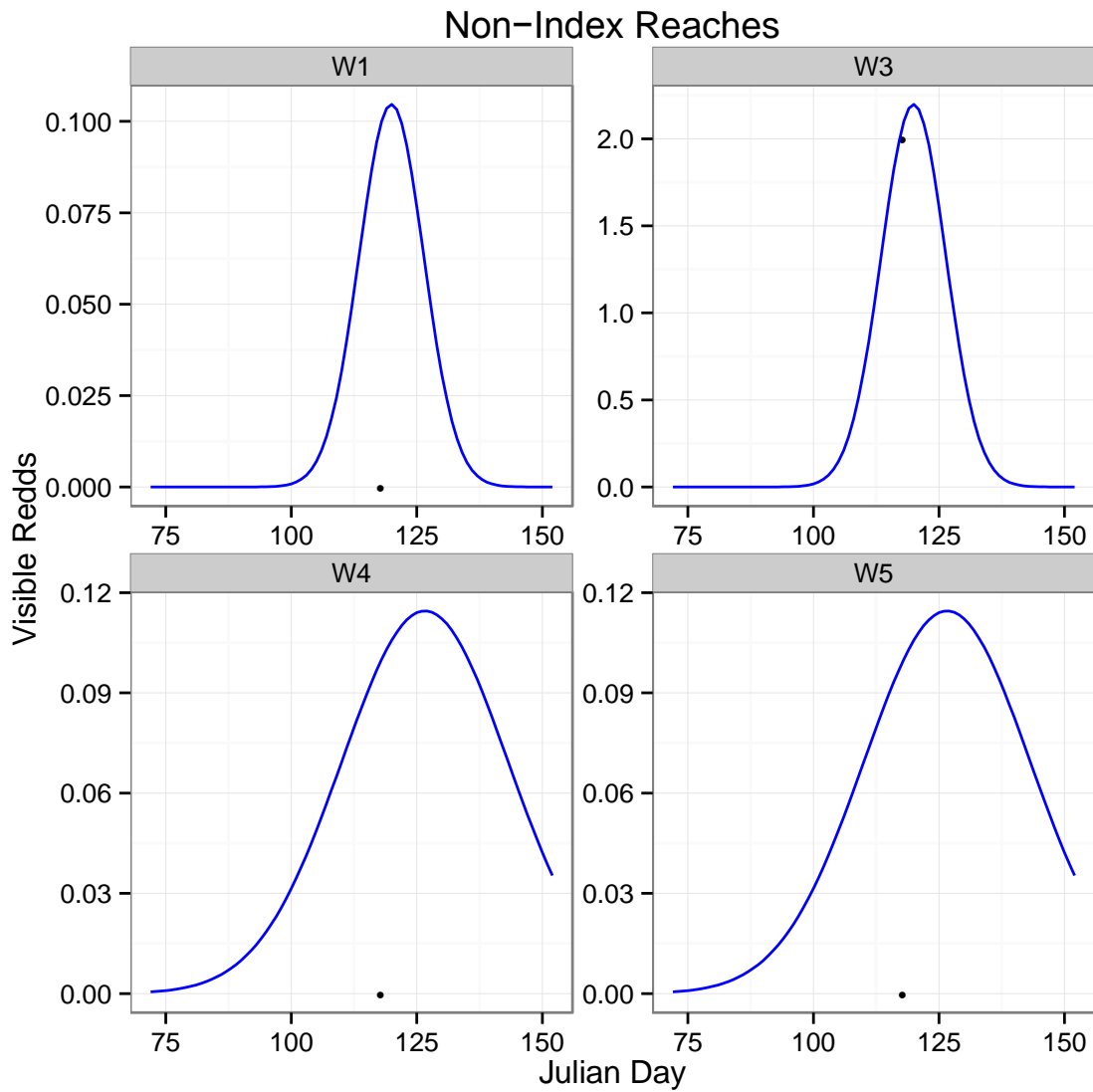


Figure 2. Observed redd counts for non-index reaches in the Wenatchee River in 2014 (black points). The blue curve shows the GAUC estimated spawning curve associated with that reach.

Table 3: Estimated number of redds for each reach in the Wenatchee River in 2014.

Reach	Type	Redds Counted	Net Error	Net Error SE	Estimated redds	
					Number	SE
W10	Index	16	0.62	1.13	29.9	21.73
W2	Index	4	0.75	1.28	4.9	4.69
W6	Index	25	0.42	1.46	63.1	39.14
W8	Index	4	0.49	0.92	16.1	7.50
W9	Index	46	0.61	1.45	77.9	70.23
W1	Non-Index	0	0.81	0.38	0.1	0.04
W3	Non-Index	2	0.88	0.38	2.2	0.79
W4	Non-Index	0	0.85	0.39	0.2	0.07
W5	Non-Index	0	0.85	0.39	0.2	0.07
All		97	0.70	0.17	195	0.43

## References

- Millar, R.B, S. McKechnie, C.E. Jordan, (2012) Simple estimators of salmonid escapement and its variance using a new area-under-the-curve method. *Canadian Journal of Fisheries and Aquatic Sciences* 69 (6) 1002-1015.
- Truscott, B.I., A.R. Murdoch, J.M. Cram and K. See. 2015. Upper Columbia spring Chinook salmon and steelhead juvenile and adult abundance, productivity and spatial scale monitoring, Project # 2010-034-00. Bonneville Power Administration, Portland OR. <https://pisces.bpa.gov/release/documents/DocumentViewer.aspx?doc=P142786>



# Appendix E

## Genetic Diversity of Wenatchee Summer Steelhead





# **Examining the Genetic Structure of Wenatchee Basin Steelhead and Evaluating the Effects of the Supplementation Program**

Developed for

Chelan County PUD

and the

Rock Island Habitat Conservation Plan Hatchery Committee

Developed by

Todd R. Seamons, Sewall Young, Cherril Bowman, and Kenneth I. Warheit

WDFW Molecular Genetics Laboratory

Olympia, WA

and

Andrew R. Murdoch

Supplementation Research Team

Wenatchee, WA

**17 January 2012**

## Table of Contents

Table of Contents .....	2
Executive Summary .....	3
Introduction.....	5
Materials and methods .....	7
Sample collections.....	7
Sample processing.....	8
Evaluation of loci .....	9
Allele frequencies, genetic distances and population differentiation.....	10
Effective spawning population.....	11
Results and Discussion .....	13
Collections and samples received.....	13
Evaluation of loci.....	13
Objective 3.1, 3.2 – Allele frequencies and Genetic distances .....	14
Allele frequencies .....	14
Analysis of Molecular Variance .....	14
Pair-wise $F_{ST}$ estimates .....	14
Principal Components.....	16
Objective 3.3 – Effective spawning population .....	18
Summary .....	19
Acknowledgements.....	20
Literature Cited.....	21
Figures.....	25
Tables.....	38

## **Executive Summary**

In 1997, Wenatchee River summer steelhead, as part of the upper Columbia River evolutionarily significant unit (ESU), were listed as threatened under the Endangered Species Act (ESA). To address concerns about effects of hatchery supplementation, the hatchery program for hatchery produced (HOR) summer steelhead to be planted in the Wenatchee River changed from using mixed ancestry broodstock collected in the Columbia River to using Wenatchee River broodstock collected in the Wenatchee River. Three monitoring and evaluation (M&E) indicators were developed to measure the genetic effects of hatchery production on wild fish populations. To address these indicators, temporal collections of tissue samples from Wenatchee River hatchery-produced (HOR) and natural origin (NOR) adults captured and sampled at Dryden and Tumwater dams and from NOR juveniles from three Wenatchee River tributaries and the Entiat River were surveyed for genetic variation with 132 genetic (SNPs) markers. Peshastin Creek (a Wenatchee River tributary) and the Entiat River served as no-hatchery-outplant controls, meaning they have stopped receiving HOR juvenile outplants. As per the M&E plan, we interrogated these data for the presence or absence of spatial and temporal trends in allele frequencies, genetic distances, and effective population size.

Allele frequencies – Changes to the summer steelhead hatchery supplementation program had no detectable effect on genetic diversity of wild populations. On average, HOR adults had higher minor allele frequencies (MAF) than NOR adults, which may simply reflect the mixed ancestry of HOR adults. Both HOR and NOR adults had MAF similar to juveniles collected in spawning tributaries and in the Entiat River. There was no temporal trend in allele frequencies or observed heterozygosity in adult or juvenile collections and allele frequencies in control populations were no different than those still receiving hatchery outplants. This suggests that the hatchery program has had little effect on allele frequencies since broodstock sources changed in 1998.

Genetic distances – As intended, interbreeding of Wenatchee River HOR and NOR adults reduced the genetic differences between Wells Hatchery HOR adults and Wenatchee River NOR adults observed in the first few years after changing the broodstock collection protocol. Though there were detectable genetic differences between HOR and HOR adults, the magnitude of that

difference declined over time. HOR adults were genetically quite different from NOR adults and juveniles based on pair-wise  $F_{ST}$  and principal components analysis (PCA), most likely because of the much smaller effective population size ( $N_b$ ) in the hatchery population (see below). Pair-wise  $F_{ST}$  estimates and genetic distances between HOR and NOR adults collected the same year declined over time suggesting that the interbreeding of HOR and NOR adults in the hatchery (and presumably in the wild) is slowly homogenizing Wenatchee River summer steelhead. Analyses using brood year (the year fish were hatched, determined using scale-based age estimates) were inconclusive because of limitations of the data.

Effective population size ( $N_b$ ) – Although the effective population size of the Wenatchee River hatchery summer steelhead program was consistently small, it does not appear to have caused a reduction in the effective population size of wild populations. On average, estimates of  $N_b$  were much lower and varied less for HOR adults than for NOR adults and juveniles. Estimates of  $N_b$  for HOR adults declined from the earliest brood years to a stable new low value after broodstock practices were changed in 1997. There was no indication that this had any effect on  $N_b$  in NOR adults and juveniles;  $N_b$  estimates for NOR adults and juveniles were, on average, higher and varied considerably over the time period covered by our dataset (1998 – 2010) and showed no temporal trend.

## Introduction

The National Marine Fisheries Service (NMFS) recognizes 15 Evolutionary Significant Units (ESU) for west coast steelhead (*Oncorhynchus mykiss*). The Upper Columbia ESU, which contains steelhead in the Wenatchee Basin, was listed as endangered under the Endangered Species Act (ESA) in 1997. Included in this listing were the Wells hatchery steelhead (program initiated in the late 1960s) that originated from a mixed group of native steelhead and are considered to be genetically similar to natural spawning populations above Wells Dam. Juvenile steelhead from Wells Fish Hatchery was the primary stock released into the Wenatchee River (Murdoch et al. 2003). The 1998 steelhead status review identified several areas of concern for this ESU including the risk of genetic homogenization due to hatchery practices and the high proportion (65% for the Wenatchee River) of hatchery fish present on the spawning grounds (Good et al. 2005). The Biological Review Team (BRT) further identified the relationship between the resident and anadromous forms of *O. mykiss* and possible changes in the population structure ('genetic heritage of the naturally spawning fish') in the basin as two areas requiring additional study. Furthermore, the West Coast Steelhead BRT (2003) recommended that stocks in the Wenatchee, Entiat, and Methow rivers, within the Upper Columbia ESU, be managed as separate populations.

A review of the presence of resident *O. mykiss* in the Upper Columbia ESU (Good et al. 2005) shows that rainbow trout are relatively abundant in upper Columbia River tributaries currently accessible to steelhead as well as in upriver tributaries unavailable to anadromous access by Chief Joseph and Grand Coulee dams (Kostow 2003). U.S. Fish and Wildlife Service (USFWS) biologists surveyed the abundance of trout and steelhead juveniles in the Wenatchee, Entiat, and Methow river drainages in the mid-1980s and found adult trout (defined as those with fork length > 20 cm) in all basins (Mullan et al. 1992). The results also supported the hypothesis that resident *O. mykiss* are more abundant in tributary or mainstem areas upstream of the areas used by steelhead for rearing. No samples of rainbow trout from the Wenatchee were available for this study.

In addition to the mixed ancestry Wells Hatchery steelhead, Skamania Hatchery (Washougal River steelhead ancestry) steelhead were also released into the Wenatchee River basin for several years in the late 1980s (L. Brown, Washington Dept. of Fish and Wildlife [WDFW], personal communication). In 1996, broodstock for the Wenatchee River steelhead program were collected from Priest Rapids Dam and Dryden (rkm 24.9) and Tumwater (rkm 52.6) dams on the Wenatchee River. Because of the ESA listing, broodstock collection after 1996 was restricted to the Wenatchee River in an effort to develop a localized broodstock (Murdoch et al. 2003). Thus, starting in 1998, all juvenile steelhead released into the Wenatchee River and Wenatchee River tributaries were offspring of only Wenatchee River captured broodstock.

In response to the need for evaluation of the supplementation program, both a monitoring and evaluation plan (Murdoch and Peven 2005) and the associated analytical framework (Hays et al. 2006) were developed for the Habitat Conservation Plans Hatchery Committee through the joint effort of the fishery co-managers (Confederated Tribes of the Colville Reservation [CCT], NMFS, USFWS, WDFW, and Yakama Nation [YN]) and Chelan County, Douglas County, and Grant County Public Utility Districts (PUD). These reports outline 10 objectives to be applied to various species assessing the impacts of hatchery operations mitigating the operation of Rock Island and Rocky Reach Dams. This report pertains to Wenatchee River basin steelhead (*O. mykiss*) and the steelhead supplementation program as addressed by objective 3, specifically the first three evaluation indicators.

**Objective 3:** Determine if genetic diversity, population structure, and effective population size have changed in natural spawning populations as a result of the hatchery program. Additionally, determine if hatchery programs have caused changes in phenotypic characteristics of natural populations.

### **3.1 Allele Frequency**

### **3.2 Genetic Distances Between Populations**

### **3.3 Effective Spawning Population**

To address these evaluation indicators the WDFW Molecular Genetics Lab (MGL) obtained pertinent tissue collections and samples, surveyed genetic variation with SNP markers using our standard laboratory protocols, and calculated the relevant genetic metrics and statistics. We used collections from both the Entiat River and Wenatchee River basins. Both have received hatchery plants from non-local stocks [i.e. Entiat was stocked with both Wenatchee and Wells program juveniles averaging 12K and 18K respectively during 1995-2001, and Wenatchee received on average 177K juveniles from the Wells program during 1995-2001; (Good et al. 2005)], and both have all or some part of the basin designated as natural production “reference” drainage – no hatchery outplanting (i.e., the entire Entiat Basin, and Peshastin Creek in the Wenatchee River basin) (Good et al. 2005).

## **Materials and methods**

### **Sample collections**

To address objectives 3.1 through 3.3, we obtained samples from hatchery (HOR, adipose fin clipped) and natural origin (NOR, adipose fin intact) adult summer steelhead captured at Dryden or Tumwater diversion dams in the summer and fall of 1997 through 2009 (excepting 2004 and 2005; Table 1). All or some fraction of these fish was later used as hatchery broodstock the calendar year following the sampling year. In order to keep things simple we have reported years as the spawning year, i.e., the calendar year the fish were spawned, not the calendar year they were captured.

To address objective 3.2, it was necessary to have samples from natural origin fish from each of the spawning populations in the basin. It is difficult to obtain adult samples from known spawning populations due to the life history and behavior of steelhead, without tributary weirs or some other blocking method of collection. The NOR adult samples used as broodstock collected from Dryden and Tumwater Dams were a mixed collection representing all of the spawning populations located upstream. Therefore to determine population substructure within the basin we obtained collections of juvenile fish from smolt traps located within tributaries representing three major populations in the basin and from the Entiat River (Chiwawa River, Nason Creek, and Peshastin Creek; Table 2). We also obtained two collections of juvenile fish caught in a

smolt trap in the lower Wenatchee River. These, like the NOR adult collections, were a mixed collection presumably representing all populations located upstream. Fin tissue was taken from each fish and preserved in 95% ethanol.

### **Sample processing**

Fin tissue samples were processed for 1468 HOR and NOR adult steelhead broodstock (Table 1) and for 1542 juvenile *O. mykiss* from the Wenatchee and Entiat Rivers (Table 2). Samples were genotyped at 152 single nucleotide polymorphism loci (SNPs, Tables 3, 4). We originally proposed to use microsatellites, but WDFW MGL and other regional genetic laboratories (Columbia River Inter-Tribal Fish Commission [CRITFC], Idaho Fish and Game [IDFG], USFWS) are moving toward using SNPs and they provide the same kinds of information with faster processing. Twenty SNP loci were developed to discriminate among trout species; 14 distinguish *O. mykiss* from coastal cutthroat trout (*O. clarkii clarkii*) and westslope cutthroat (*O. clarkii lewisi*), and 6 distinguish steelhead and coastal cutthroat from westslope cutthroat (Table 4). The remaining 132 SNP loci were developed to be used for population structure, parentage assignment, or other population genetic studies of *O. mykiss* (Table 3). These markers comprised the current standard set of SNP markers used for genetic studies of *O. mykiss* at WDFW MGL.

We used Qiagen DNEasy® kits (Qiagen Inc., Valencia, CA), following the recommended protocol for animal tissues, to extract and isolate DNA from fin tissue. SNP genotypes were obtained through PCR and visualization on Fluidigm EP1 integrated fluidic circuits (chips). Protocols followed Fluidigm's recommendations for TaqMan SNP assays as follows: Samples were pre-amplified by Specific Target Amplification (STA) following Fluidigm's recommended protocol with one modification. The 152 assays were pooled to a concentration of 0.2X and mixed with 2X Qiagen Multiplexing Kit (Qiagen, Inc., Valencia CA), instead of TaqMan PreAmp Master Mix (Applied Biosystems), to a volume of 3.75µl, to which 1.25µl of unquantified sample DNA was added for a total reaction volume of 5µl. Pre-amp PCR was conducted on a MJ Research or Applied Biosystems thermal cycler using the following profile: 95°C for 15 min followed by 14 cycles of 95°C for 15 sec and 60°C for 4 minutes. Post-PCR reactions were diluted with 20µl dH<sub>2</sub>O to a final volume of 25µl.



Specific SNP locus PCRs were conducted on the Fluidigm chips. Assay loading mixture contained 1X Assay Loading Reagent (Fluidigm), 2.5X ROX Reference Dye (Invetrogen) and 10X custom TaqMan Assay (Applied Biosystems); sample loading mixture contains 1X TaqMan Universal PCR Master Mix (Applied Biosystems), 0.05X AmpliTaq Gold DNA polymerase (Applied Biosystems), 1X GT sampling loading reagent (Fluidigm) and 2.1  $\mu$ L template DNA. Four  $\mu$ L assay loading mix and 5  $\mu$ L sample loading mix were pipetted onto the chip and loaded by the IFC loader (Fluidigm). PCR was conducted on a Fluidigm thermal cycler using a two step profile. Initial mix thermal profile was 70°C for 30min, 25°C for 5 min, 52.3° for 10 sec, 50.1°C for 1 min 50sec, 98°C for 5 sec, 96°C for 9 min 55 sec, 96°C for 15 sec, 58.6°C for 8 sec, and 60.1°C for 43 sec. Amplification thermal profile was 40 cycles of 58.6°C for 10 sec, 96°C for 5 sec, 58.6°C for 8 sec and 60.1°C for 43 sec with a final hold at 20°C.

The SNP assays were visualized on the Fluidigm EP1 machine using the BioMark data collection software and analyzed using Fluidigm SNP genotyping analysis software. To ensure all SNP markers were being scored accurately and consistently, all data were scored by two researchers and scores of each researcher were compared. Disputed scores were called missing data (i.e., no genotype).

### **Evaluation of loci**

A two-tailed exact test of Hardy–Weinberg equilibrium (HWE) was performed for each locus in each collection or population using the Markov Chain method implemented in GENEPOP v4.1 (dememorization number 1000, 100 batches, 1000 iterations per batch; Raymond and Rousset 1995; Rousset 2008). Significance of probability values was adjusted for multiple tests using false discovery rate (Verhoeven et al. 2005).  $F_{IS}$ , a measure of the fractional reduction in heterozygosity due to inbreeding in individuals within a subpopulation and an additional indicator of scoring issues, was calculated according to Weir and Cockerham (1984) using GENEPOP v4.1. Allele frequencies were calculated using CONVERT v1.0 (Glaubitz 2004). Expected and observed heterozygosities were calculated using GDA v1.1 (Lewis and Zaykin 2001).

### **Allele frequencies, genetic distances and population differentiation**

To evaluate Q1 of Objective 3.1 and 3.2, we evaluated trends and patterns in allele frequencies, genetic distances and population differentiation. To test for temporal patterns in allele frequencies, we compared sample or spawn year to two diversity metrics, allele frequency and observed heterozygosity, from each adult and juvenile collection. Each SNP locus had only one or two alleles, so we used the minor allele frequency (MAF) of each SNP locus for each adult collection and averaged across loci. We also calculated the average observed heterozygosity ( $H_o$ ) for each SNP locus within each adult and juvenile collection. We examined the presence or absence of a temporal trend in average allele frequency and observed heterozygosity with logistic regression analysis in R (R Development Core Team 2009).

To partition genetic variance into temporal, spatial (juvenile) and origin (adult) fractions, we performed hierarchical analysis of molecular variance (AMOVA) using ARLEQUIN v3.0 (Excoffier et al. 2005) with 1,000 permutations. We performed this analysis separately for juvenile and adult collections. Juveniles were grouped by sampling location (tributary) and adults were grouped by origin (HOR or NOR). To estimate the magnitude of genetic differences among temporal and spatial collections we calculated pairwise  $F_{ST}$  estimates among collections using FSTAT (Goudet 1995) with 1000 permutations. Statistical significance was adjusted using false discovery rate (Verhoeven et al. 2005).

To evaluate the temporal changes in genetic relationships, we compared spawn year to within spawn year pairwise  $F_{ST}$  estimates between NOR and NOR adults using beta regression (Simas and Rocha 2010). We used beta regression because the dependent variable was bound by zero and one but not binomial. Analysis was performed in R (package "betareg", Cribari-Neto and Zeileis 2010), with a loglog link.

We used principal component analyses (PCA) to explore the relationship between the covariation among the SNP loci within each collection and genetic differentiation between HOR and NOR collections, and to determine if the degree of differentiation has changed with time. Since each SNP is represented by only two alleles, only one allele per SNP is necessary to fully describe the covariation among all SNPs. We used MATLAB® scripts (2007a, The Mathworks, Natick, MA)

to calculate the principal components from SNP allele frequencies using only the major allele (1-MAF) for each SNP. We defined the major allele as the allele with the higher mean frequency across all collections, regardless of its status within any individual collection. We conducted three PCA analyses using: (1) all adult samples, aggregated based on origin (HOR versus NOR) and spawn year (i.e., the year the adult fish were used as broodstock) (N = 1437, 22 collections), (2) same as #1, but with the addition of all juvenile samples (N = 2938, 37 collections), and (3) only those adults samples with available age information (Mike Hughes, WDFW, personal communication) aggregated based on origin, and spawn year or brood year (i.e., the year the fish were hatched) (N = 1313, 20 spawn-year or 25 brood-year collections).

Molecular differentiation between HOR and NOR adults within a year was calculated based on principal component scores using Euclidian distances. We calculated pair-wise Euclidian distances between HOR and NOR fish within a spawn year or brood year using the first three principal components, and standardized each distance by subtracting from it the mean Euclidian distance calculated across all pair-wise distances. We used Mahalanobis distances to calculate the variation among HOR and NOR collections (calculated separately), again using the first three principal components. Here, we calculated Mahalanobis distances as the Euclidian distances between each collection and the centroid of all collections (HOR and NOR combined), but the Euclidian distances are scaled based on the dispersion of collections around the centroid (i.e., the variance). Euclidian and Mahalanobis distances were calculated using MATLAB scripts.

### **Effective spawning population**

To evaluate Q1 of Objective 3.3, we estimated  $N_e$  using the single-sample linkage disequilibrium methods implemented in the program LDNE (Waples and Do 2008). This method requires that you input the  $P_{crit}$  value, the minimum frequency at which alleles were included in the analysis, since results can be biased depending on this setting (Waples and Do 2010). SNP markers typically have only one or two alleles; if one of two alleles is excluded based on its frequency in the collection it essentially excludes the locus, reducing the overall dataset. Therefore, we used  $P_{crit}$  values ranging from 0.1 to 0.001 to evaluate whether trends in  $N_e$  changed given which loci were used. Confidence intervals were calculated using a jackknife procedure.

We calculated an estimate of  $N_e$  for all adult and juvenile collections individually. However, the intention of an integrated hatchery program such as the Wenatchee River steelhead hatchery program is that HOR and NOR fish are integrated and progress as a single population through intentional interbreeding in the hatchery and presumed natural interbreeding in the wild. Thus, we also combined annual HOR and NOR collections to calculate an overall  $N_e$  estimate as has been done in other genetic monitoring and evaluation analyses (e.g., Small et al. 2007, [Chinook salmon, *O. tshawytscha*]).

Estimates of  $N_e$  from linkage refer to the generations that produced the sample. To calculate the ratio of effective population size to census size ( $N_e/N$ ), we obtained the number of fish spawned in the hatchery (1993 through 2006, i.e., those that produced the adipose fin clipped adults that returned to spawn in the Wenatchee River 1998 through 2010) and the estimated escapement of fish spawning naturally (HOR and NOR separately) for the same time period. Estimates of census population size in spawning tributaries was obtained by multiplying the fraction of redds counted within tributaries (Chad Herring, WDFW, unpublished data) by the total Wenatchee River census population estimate (Andrew Murdoch, WDFW, unpublished data). To calculate  $N_e/N$ , we performed two analyses. First, for adults, we assumed a five year generation time for natural origin adults and a four year generation time for hatchery origin adults and divided the  $N_e$  estimate by the census population estimate from four or five years earlier. For juveniles, we assumed an age at outmigration of two years and divided the  $N_e$  estimates by the estimate of census population size for the appropriate tributary. Second, we used available adult age data to parse individuals into cohorts originating in brood years (rather than spawn years) and then used LDNE to estimate  $N_e$  from cohort collections. We performed both analyses to make full use of all available data; age data were not available for many adults, and because of variable survival and sampling not all cohorts had sufficient numbers of HOR and NOR adults. According to Luikart et al. (2010), estimates produced using linkage disequilibrium should be interpreted as something between effective population size ( $N_e$ ) and the effective number of breeders ( $N_b$ ). Using cohorts, the estimate produced by LDNE is clearly an estimate of  $N_b$  rather than  $N_e$ . In order to keep things simple, we have referred to all estimates as  $N_b$ .

## Results and Discussion

### *Collections and samples received*

From 1468 samples from HOR and NOR adult steelhead broodstock, 1437 produced sufficient genetic data for further analysis (Table 1). From 1542 samples from NOR juvenile steelhead from Wenatchee River tributaries and the Entiat River, 1501 produced sufficient genetic data for further analysis and were genetically identified as *O. mykiss* (Table 2). Samples genetically identified as *O. clarki* (2 samples from the Chiwawa River, 1 from the Entiat River) or *O. clarki/O. mykiss* hybrids (4 – lower Wenatchee River, 4 – Nason Creek, 4 – Chiwawa River, and 1 – Entiat River) were omitted from further analysis.

### *Evaluation of loci*

Three loci showed deviations from HWE in 10 or more of 37 Wenatchee steelhead collections before correcting for multiple tests (AOmy016, AOmy051, AOmy252, Table A1) indicating possible scoring issues. These loci were omitted from further analysis. Nine of the remaining loci were monomorphic or nearly monomorphic in all collections (average MAF < 0.1, AOmy023, AOmy028, AOmy123, AOmy129, AOmy132, AOmy209, AOmy229, AOmy270, AOmy271, Table A1) contributing little or nothing to analytical power. These loci were also omitted from further analysis. No genetic data was available for collection 10FD due to poor PCR amplification at locus AOmy213 for the entire collection. AOmy213 had a relatively low MAF in most collections so rather than re-processing this collection at this locus or running different sets of loci for different tests, we omitted this locus from further analysis. Only six tests of deviation from HWE were significant after correcting for 4348 tests using false discovery rate. Two of these tests were in loci already omitted. The remaining four tests were spread among the remaining loci, indicating no more loci needed to be omitted from further analysis.

### **Objective 3.1, 3.2 – Allele frequencies and Genetic distances**

#### *Allele frequencies*

Average MAF of SNP loci ranged from 0.00 to 0.60 in HOR adult collections and from 0.00 to 0.61 in NOR adult collections (Table A1). Observed heterozygosity ranged from 0.00 to 0.75 in HOR adult collections and from 0.01 to 0.67 in NOR adult collections. Juvenile collections produced similar ranges of MAF and  $H_o$  (Table A1). Average MAF and  $H_o$  of HOR adult collections appeared to be greater than those of natural origin collections. However, logistic regression analysis indicated there was no significant temporal trend in either diversity statistic (Figure 1). Similarly, there was no consistent temporal trend in MAF or  $H_o$  of juvenile collections (Figure 2). Both the Chiwawa River and Nason Creek, the two tributaries that currently still receive hatchery juvenile outplants, both appeared to have declining allele frequencies, but neither was statistically significant ( $P > 0.90$ ). However, the power to detect significant trends was limited by the small sample sizes ( $n = 3$  sample years).

#### *Analysis of Molecular Variance*

Analysis of molecular variance (AMOVA) of adult collections (i.e., temporal and origin structure) indicated most of the genetic variance was among individuals or among individuals within populations (99.04%). Most of the remaining variance was temporal variation within hatchery and natural origin groups (0.61%) with the remaining variation from origin (0.35%). AMOVA of juvenile collections (i.e., spatial structure) indicated most of the genetic variance was among individuals (98.44%) or among individuals within populations (0.94%). Most of the remaining variance existed among temporal collections within tributary collections (0.37%) with the smallest fraction as among tributary variance (0.24%). Thus, overall, there was more variability among years than among tributaries or origins, but no trend in the temporal variability.

#### *Pair-wise $F_{ST}$ estimates*

HOR adults were genetically different than NOR adults as estimated by  $F_{ST}$  (full pair-wise table in Table A2, all pair-wise  $F_{ST}$  estimates with  $P$ -values  $\leq 0.05$  before correcting for multiple tests

were significantly different from zero after correcting for multiple tests using false discovery rate). On average, HOR adult collections were as different from one another (mean  $F_{ST} = 0.011$ ) as they were from NOR adult collections among years (mean  $F_{ST} = 0.009$ ) or from NOR adult collections within years (mean  $F_{ST} = 0.010$ ). Among year comparisons of NOR adult collections were, on average, nearly an order of magnitude lower (mean = 0.002). These patterns held whether spawn year or brood year (data not shown) was used to group individuals. Over time, within spawn year pair-wise  $F_{ST}$  estimates between HOR and NOR adults declined over time ( $\beta = -0.014$ ,  $P = 0.0185$ ; Figure 3), suggesting that the integration of hatchery and wild fish is slowly genetically homogenizing the groups. That relationship disappeared when adults were grouped by brood year (i.e., comparing fish produced the same year) and all brood years were used ( $\beta = -0.009$ ,  $P = 0.615$ , data not shown). However, when the dataset was restricted to just those brood years when all typical (age at maturation frequency among all years > 0.10) age classes were present in the dataset (HOR = age 3, 4; NOR = age 4, 5, 6; brood years 1996-1998, 2004-2005) a non-significant ( $P = 0.278$ ) negative relationship ( $\beta = -0.12$ ) of  $F_{ST}$  and brood year was apparent. When the data were further restricted to just the years after the hatchery program changed to only collecting broodstock in the Wenatchee River (brood years 1998, 2004-2005), the slope was also negative ( $\beta = -0.09$ ), but the relationship was not statistically significant ( $P = 0.962$ ).

Within tributary among sample year pair-wise comparisons of juvenile collections were, on average, only very slightly smaller than comparisons among tributaries (0.005 vs. 0.006, respectively, Table 5, all pair-wise  $F_{ST}$  estimates with  $P$ -values  $\leq 0.05$  before correcting for multiple tests were significantly different from zero after correcting for multiple tests using false discovery rate). Nason Creek and Peshastin Creek on average showed higher among sample year  $F_{ST}$  estimates (0.010 and 0.007, respectively) than the Chiwawa or Entiat Rivers (0.004 and 0.002, respectively). The pair-wise comparison of the two collections of lower Wenatchee River smolts, presumably a mix of Chiwawa, Nason, Peshastin smolts and smolts from other spawning tributaries, was an order of magnitude smaller ( $F_{ST} = 0.0002$ ), and not significantly different than zero (Table 5). There was no temporal trend in pair-wise comparisons of juvenile collections. However with, at most, four annual collections, detecting any temporal trend was unlikely. We also had no collections from years prior to 1998 (the first year of new hatchery program

broodstock collecting protocols) with which to compare contemporary data, nor could we find any reports or papers containing pre-hatchery-program-change genetic comparisons among Wenatchee River tributary populations, making it impossible to determine whether or not changing the hatchery program has had any effect at all on population structure. However, these data will be useful for future studies.

### *Principal Components*

Each principal component analysis (Figures 4, 5) indicated that the genetic structure among HOR collections differed from that among NOR collections, and that this difference has decreased with time. When adult fish were aggregated based on origin and spawn-year, there was a clear differentiation between HOR and NOR adult collections along PC 1, and a separation among HOR collections, differentiating the early spawn-years (1998 – 2003) from the later spawn-years (2004 – 2010) along PC 2 and PC 3, respectively (Figure 4). The pair-wise genetic distances between HOR and NOR collections from the same spawn year (i.e., the HOR and NOR fish used as broodstock within the same year) decreased from the largest distance in 1998 to small distances in 2009 and 2010, although the smallest distance occurred in 2004 (Figure 4, top right). That is, within hatchery broodstock, the genetic difference between HOR and NOR fish decreased, on average, from 1998 to 2010, and the decrease appeared to be a mutual convergence of NOR fish shifting right along PC 1 and HOR fish shifting downward along PC 2 and PC 3. This increasing similarity in adult fish mirrored that seen in within year pair-wise  $F_{ST}$  estimates between HOR and NOR adults which also declined over time (Figure 3).

Overall, there was considerably more genetic variation among the HOR collections than there was among the NOR collections with average Mahalanobis distances (distance between each collection and the overall centroid [0,0,0]) among the HOR and NOR collections being 4.2 and 1.5, respectively. Since each NOR collection was generally composed of 3-4 brood-years, while HOR collections rarely were composed of more than two brood-years, we attributed the lower year-to-year genetic variability of the NOR broodstock to the greater homogenizing effect of including four or more brood-years compared with only two brood years for the HOR broodstock.



Including the 15 juvenile collections, along with the 22 adult collections, did not materially alter the principal component structure (Figure 6), although the total genetic variation accounted for by the three principal components decreased from 44% using only the adults to 33% when juveniles were included. For the most-part, the juvenile fish appeared intermediate between HOR and NOR fish, but there was greater overlap in principal component scores (and therefore greater genetic similarity) of the juvenile and NOR collections, than of the juvenile and HOR collections. The average Euclidian distance between the juvenile and HOR collections was 0.49, compared to 0.23 between the juvenile and NOR collections, which was no different than 0.23 and 0.22 for the within juvenile and NOR collections, respectively.

By using the available adult age data, we were able to compare the genetic differentiation among the same set of fish when they are aggregated by origin (hatchery versus natural) and brood-year (year fish were hatched) with aggregates based on origin and spawn-year (year adult fish were spawned). A brood-year analysis compares within a year the genetic diversity generated from hatchery broodstock with that naturally produced in the spawning grounds. A spawn-year analysis compares the HOR and NOR genetic diversity that was mixed among cohorts of the parental generations. The same basic pattern of genetic structure that we have seen in spawn-year analyses (Figure 4, Figure 6, and the right side of Figure 5) also occurred in the brood-year analysis (left side of Figure 5). That is, from Figure 5 we saw (1) that HOR and NOR fish were differentiated from each other; (2) there was considerably more genetic variation (temporal variation) among the hatchery-origin collections than there was among the natural-origin collections (for brood-year, Mahalanobis distances = 5.18 and 0.75, respectively; for spawn-year, Mahalanobis distances = 4.25 and 1.25, respectively), and (3) that the genetic distances between HOR and NOR collections were lower in the more recent brood- and spawn-years, than in the earlier brood- and spawn-years (Figure 7;  $R^2 = 0.41$  or 41%,  $P < 0.05$ ). This indicated that the HOR and NOR fish used as broodstock in 2010 were more similar to each other than they were at the inception of the new hatchery program.

The relationship between genetic distance and brood-year was not the same as the relationship between genetic distance and spawn-year. For brood-year, although the slope was negative (i.e.,

trending downward or decreased differentiation with time) and the two most-recent brood years (2005-2006) showed relatively small HOR and NOR adult differentiation, the negative slope was not significantly different from zero and the regression accounted for only 7% of the variation. This was likely the result of insufficient sampling of certain age classes from many brood years (especially from NOR adults) due to two un-processed sample years (2005 and 2006).

### **Objective 3.3 – Effective spawning population**

There was no difference in the temporal trends in estimates of  $N_b$  with  $P_{crit}$  set from 0.1 to 0.001 (Figure 8, data not shown for all collections), so we have reported only results with  $P_{crit} = 0.001$ , i.e., the full genetic dataset. Using either spawn-year or brood year, estimates of NOR adult  $N_b$  were higher and varied more than those of HOR adults (Figures 9, 10), concordant with the PCA analysis. Estimates for HOR adults ranged from 17 to 174 (by spawn year, mean = 65) or from 6 to 130 (by brood year, mean = 39). Estimates for NOR adults ranged from 36 to 982 (by spawn year, mean = 405) or from 59 to 2966 (by brood year, mean = 645). Many  $N_b$  estimates for NOR adults had confidence intervals extending to infinity on the upper bound. This reflected the difficulty in obtaining precise estimates of  $N_b$  for large populations (Waples and Do 2010).

Estimates of  $N_b$  for HOR steelhead dropped by approximately half from 1994, when broodstock were still collected at Wells Hatchery, to 1998, when the program used Wenatchee River trapped adults only, suggesting an effect of changing broodstock collection practices, which began in 1997 (Figures 8, 9). Since 1997, the hatchery population  $N_b$  remained at a relatively stable lower level (Figures 8, 9, and 10). There was no obvious change in  $N_b$  for NOR steelhead since 1993; the  $N_b$  estimate for 1993 was the largest, however the confidence interval overlapped estimates from many other years. The temporal trend in  $N_b$  estimates from combined collections mirrored those of the HOR collections alone, though estimates using combined collections were slightly larger (Figure 11).

As with  $N_b$  estimates, estimates of the ratio of  $N_b/N$  for NOR adults varied more than those of HOR adults (Figures 12, 13). However, using spawn year, i.e., mixtures of cohorts, the average  $N_b/N$  ratio for HOR adults was equal to that of NOR adults (mean  $N_b/N = 0.26$ ), whereas when using brood year, the average  $N_b/N$  ratio for NOR adults was double that of HOR adults (NOR

average = 0.40, HOR average = 0.20). This is likely a consequence of the homogenizing effect of mixed cohorts. Estimates of  $N_b$  for HOR adults using spawn year were close to those estimated using brood year because of the lower diversity in age at maturation, whereas for NOR, grouping by brood year produces different estimates than when grouping by spawn year because of higher diversity in age at maturation. Regardless of which estimate was used, there was no temporal trend in  $N_b/N$  for either NOR or HOR adults.

## Summary

On average, HOR adults had higher minor allele frequencies (MAF) than NOR adults, and both had similar MAF as juveniles collected in spawning tributaries and in the Entiat River. There was no temporal trend in allele frequencies or observed heterozygosity in adult or juvenile collections and allele frequencies in control populations were no different than those still receiving hatchery outplants suggesting that the hatchery program has had little effect on allele frequencies since 1998.

HOR adults were genetically quite different from NOR adults and juveniles based on pair-wise  $F_{ST}$  and principal components analysis (PCA), most likely because of the much smaller effective population size ( $N_b$ ) in the hatchery population. Pair-wise  $F_{ST}$  estimates and genetic distances between HOR and NOR adults collected the same year declined over time suggesting that the interbreeding of HOR and NOR adults in the hatchery (and presumably in the wild) is slowly homogenizing Wenatchee River summer steelhead. Analyses using brood year (the year fish were hatched, determined using scale-based age estimates) were inconclusive because of limitations of the data.

On average, estimates of  $N_b$  were much lower and varied less for HOR adults than for NOR adults and juveniles. Estimates of  $N_b$  for HOR adults declined from the earliest brood years to a stable new low value after broodstock practices were changed in 1997. There was no indication that this had any effect on  $N_b$  in NOR adults and juveniles;  $N_b$  estimates for NOR adults and juveniles were, on average, higher and varied considerably over the time period covered by our dataset (1998 – 2010) and showed no temporal trend. Small  $N_b$  sizes increase the risk of loss of

genetic diversity due to inbreeding and random effects (genetic drift). The  $N_b$  of the hatchery component of the population may be increased by spawning more families, using specific mating designs, and minimizing variance in reproductive success. However, given the apparent lack of effects overall, changes to the hatchery protocol may not be necessary.

Overall, hatchery practices appear to have had little effect on natural origin Wenatchee summer steelhead neutral genetic diversity or  $N_b$ . We cannot accurately assess their effects on population structure at this time. However, it is interesting to note that when juvenile collections are analyzed separately from adult collections, Peshastin Creek, which has received fewer hatchery outplants in the past and is currently a refuge from hatchery outplants, is genetically different than other tributaries and the Entiat River (data not shown). On the other hand, the Entiat River, which is also a refuge from hatchery outplants and is not a tributary of the Wenatchee River, is genetically very similar to Nason Creek and the Chiwawa River, both Wenatchee River tributaries. This suggests, though it does not conclude, that within basin population structure may have existed before summer steelhead hatchery production began in the upper Columbia River and that the population structure was eliminated by hatchery influence long before 1998.

## **Acknowledgements**

We thank Chad Herring, Clint Deason, John Walters and the numerous technicians that sampled these thousands of fish. We thank Sonia Peterson and Sarah Bell for help in the laboratory and thank Maureen Small for help with some analyses. The project was implemented with funding from the Chelan Co. PUD and Washington State general funds.

## Literature Cited

- Aguilar, A., and J. C. Garza. 2008. Isolation of 15 single nucleotide polymorphisms from coastal steelhead, *Oncorhynchus mykiss* (Salmonidae). *Molecular Ecology Resources* 8(3):659-662.
- Campbell, N. R., and S. R. Narum. 2009. Identification and characterization of heat shock response-related single-nucleotide polymorphisms in *O. mykiss* and *O. tshawytscha*. *Molecular Ecology Resources* 9(6):1460-1466.
- Campbell, N. R., K. E. N. Overturf, and S. R. Narum. 2009. Characterization of 22 novel single nucleotide polymorphism markers in steelhead and rainbow trout. *Molecular Ecology Resources* 9(1):318-322.
- Cribari-Neto, F., and A. Zeileis. 2010. Beta regression in R. *Journal of Statistical Software* 34:1-24.
- Excoffier, L., G. Laval, and S. Schneider. 2005. Arlequin (version 3.0): An integrated software package for population genetics data analysis. *Evolutionary Bioinformatics* 1:47-50.
- Finger, A. J., M. R. Stephens, N. W. Clipperton, and B. May. 2009. Six diagnostic single nucleotide polymorphism markers for detecting introgression between cutthroat and rainbow trouts. *Molecular Ecology Resources* 9(3):759-763.
- Glaubitz, J. C. 2004. CONVERT: A user-friendly program to reformat diploid genotypic data for commonly used population genetic software packages. *Molecular Ecology Notes* 4:309-310.
- Good, T. P., R. S. Waples, and P. Adams. 2005. Updated status of federally listed ESUs of West Coast salmon and steelhead. U.S. Dept. of Commerce, NMFS-NWFSC-66.

- Goudet, J. 1995. FSTAT (Version 1.2): a computer program to calculate F-statistics. *Journal of Heredity* 86:485-486.
- Hansen, M. H. H., and coauthors. 2011. Assembling a dual purpose TaqMan-based panel of single-nucleotide polymorphism markers in rainbow trout and steelhead (*Oncorhynchus mykiss*) for association mapping and population genetics analysis. *Molecular Ecology Resources* 11:67-70.
- Hays, S., and coauthors. 2006. Analytical framework for monitoring and evaluating PUD hatchery programs.
- Kostow, K. 2003. The biological implications of nonanadromous *Oncorhynchus mykiss* in Columbia Basin steelhead ESUs. Report to NOAA Fisheries and ODFW, 13 January 2003. (Available from Northwest Fisheries Science Center, 2725 Montlake Blvd. E., Seattle, WA 98112.).
- Lewis, P. O., and D. Zaykin. 2001. Genetic Data Analysis: Computer program for the analysis of allelic data. Free program distributed by the authors over the internet from <http://lewis.eeb.uconn.edu/lewishome/software.html>.
- Luikart, G., N. Ryman, D. Tallmon, M. Schwartz, and F. Allendorf. 2010. Estimation of census and effective population sizes: the increasing usefulness of DNA-based approaches. *Conservation Genetics* 11(2):355-373.
- McGlaflin, M. T., and coauthors. 2010. High-resolution melting analysis for the discovery of novel single-nucleotide polymorphisms in rainbow and cutthroat trout for species identification. *Transactions of the American Fisheries Society* 139(3):676-684.

- Mullan, J. W., K. R. Williams, G. Rhodus, T. W. Hillman, and J. D. McIntyre. 1992. Production and habitat of salmonids in mid-Columbia River tributary streams. U.S. Fish and Wildlife Service, Monograph I, Leavenworth, WA.
- Murdoch, A., T. Miller, T. Maitland, M. Tonseth, and L. Prave. 2003. Annual progress report for Wenatchee summer steelhead, 2001 brood. Washington Dept. of Fish and Wildlife.
- Murdoch, A., and C. Peven. 2005. Conceptual approach for monitoring and evaluating the Chelan County Public Utility District hatchery programs. Chelan County Public Utility District, Wenatchee, WA.
- R Development Core Team. 2009. R: A language and environment for statistical computing
- Raymond, M., and F. Rousset. 1995. An exact test for population differentiation. *Evolution* 49(6):1280-1283.
- Rousset, F. 2008. GENEPOP'007: a complete re-implementation of the GENEPOP software for Windows and Linux. *Molecular Ecology Resources* 8(1):103-106.
- Sánchez, C. C., and coauthors. 2009. Single nucleotide polymorphism discovery in rainbow trout by deep sequencing of a reduced representation library. *BMC Genomics* 10(1):559.
- Simas, A. B., and A. V. Rocha. 2010. Improved estimators for a general class of beta regression models. *Computational Statistics & Data Analysis* 54:348-366.
- Small, M. P., K. I. Warheit, C. Dean, and A. Murdoch. 2007. Genetic monitoring of Methow Spring Chinook. Washington Department of Fish and Wildlife, Olympia, WA.
- Sprowles, A. E., M. R. Stephens, N. W. Clipperton, and B. P. May. 2006. Fishing for SNPs: A targeted locus approach for single nucleotide polymorphism discovery in rainbow trout. *Transactions of the American Fisheries Society* 135(6):1698-1721.

- Verhoeven, K. J. F., K. L. Simonsen, and L. M. McIntyre. 2005. Implementing false discovery rate control: increasing your power. *Oikos* 108(3):643-647.
- Waples, R. S., and C. Do. 2008. LDNE: a program for estimating effective population size from data on linkage disequilibrium. *Molecular Ecology Resources* 8(4):753-756.
- Waples, R. S., and C. Do. 2010. Linkage disequilibrium estimates of contemporary  $N_e$  using highly variable genetic markers: a largely untapped resource for applied conservation and evolution. *Evolutionary Applications* 3(3):244-262.
- Weir, B. S., and C. C. Cockerham. 1984. Estimating F-statistics for the analysis of population structure. *Evolution* 38(6):1358-1370.



## Figures

Figure 1. Observed average minor allele frequencies (MAF) and observed heterozygosities ( $H_o$ ) of 119 SNP loci from 11 annual collections of hatchery-produced (HOR) and natural origin (NOR) adult steelhead from the Wenatchee River. Trend lines are from a logistic regression. Note the X axis does not cross the Y axis at the origin. Neither the slopes nor the intercepts were statistically significant.

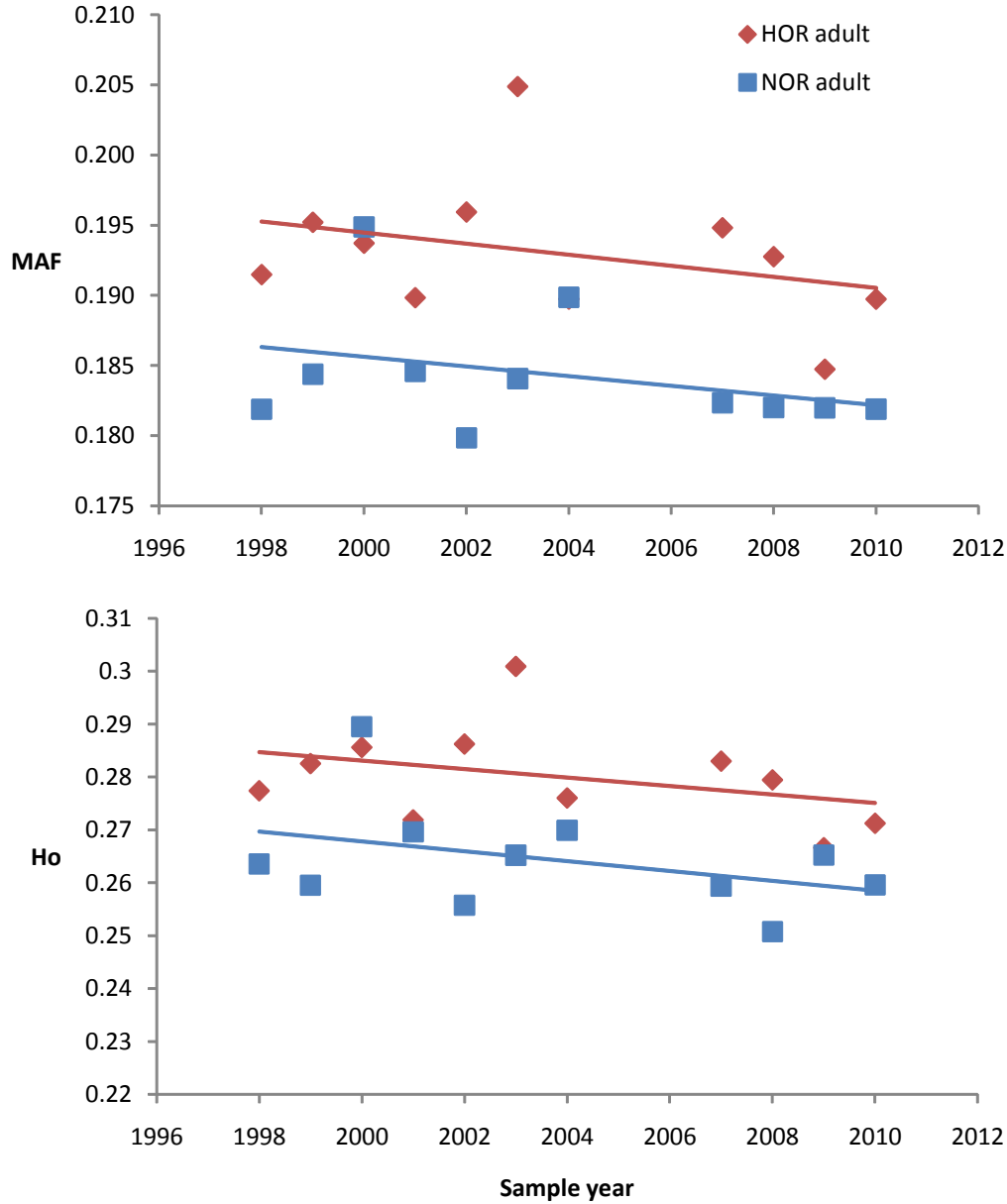


Figure 2. Observed average minor allele frequencies (MAF) and observed heterozygosities (Ho) of 119 SNP loci from 15 collections of natural origin juvenile steelhead from Wenatchee River tributaries, the lower Wenatchee River and the Entiat River. There were no consistent temporal trends in MAF or Ho in these collections.

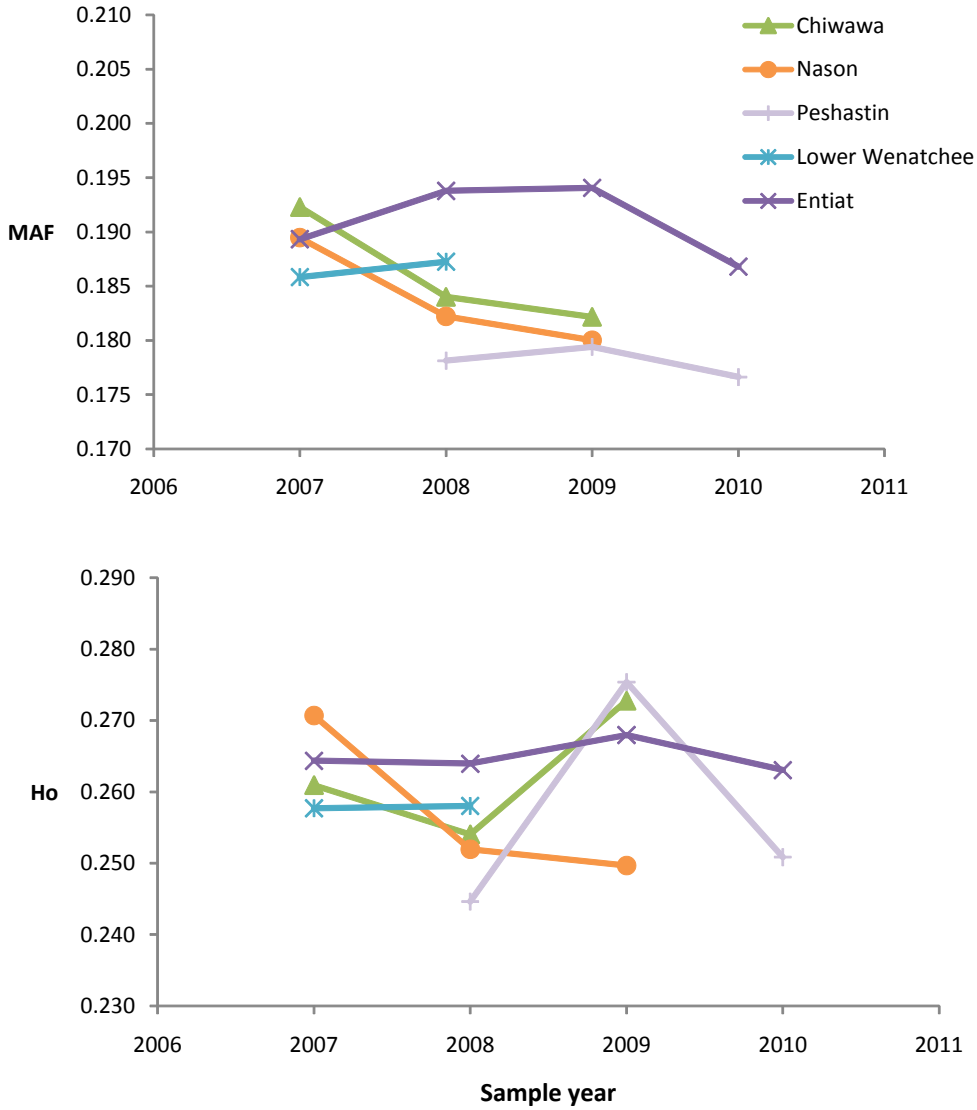


Figure 3. The relationship of time with pairwise  $F_{ST}$  estimates between hatchery-produced (adipose fin clipped) and natural origin (unclipped) adults of the same sample year. The line is the prediction based on beta regression.

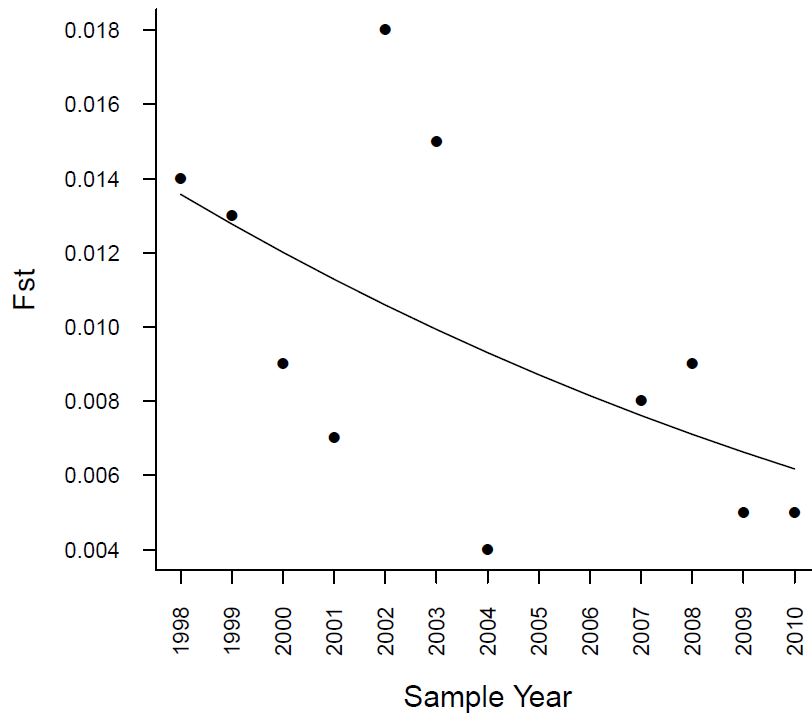


Figure 4. Principal component (PC) 1 versus 2 (top left), PC 1 versus 3 (bottom left), and PC 2 versus 3 (bottom right) based on an analysis using all adults aggregated into origin and spawn-year collections. Natural-origin spawn-years are shown in italicized typeface. The percentage within the label of each axis convey the percent of total genetic variance that is accounted for by that axis. Taken together, the three principal components account for 44% of the total SNP variation. Top right shows pairwise Euclidian distances versus spawn-year, with zero distance equal to average distance across all pairwise distances. Blue line is least-squares fit with  $R^2 = 0.45$ .

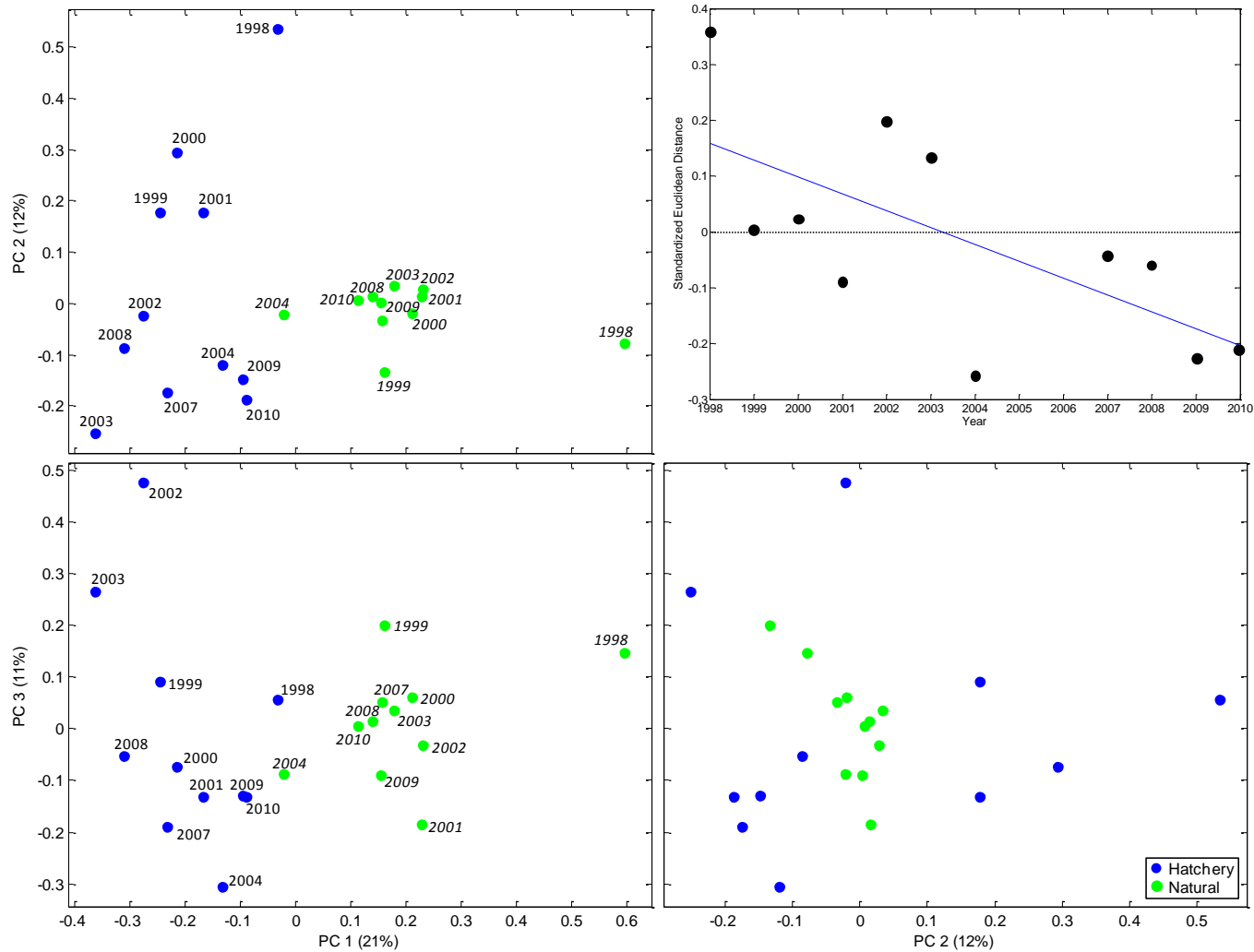


Figure 5. Principal components (PC) 1 versus 2 (top) and 3 (bottom) for adults aggregated into brood-year (BY; left) and spawn-year (SY; right). Spawn-year analysis is the same as in Figure x1, except fewer individuals per collection were included (see methods). Note that for the SY analysis here PC 2 and 3 are similar to PC 3 and 2, respectively, in Figure x1. Only BY1995 (earliest year with paired hatchery-natural data), BY2000 (extreme PC 1 score), and BY2006 (latest year with paired hatchery-natural data) are labeled. Hatchery- and natural-origin individuals from BY1995, BY2000, and BY2006, returned to spawn (spawn-year) in 1999 (hatchery)/1999-2001 (natural), 2003-2004 (hatchery)/2004 and 2007 (natural), and 2009-2010 (hatchery)/2010 (natural), respectively. These years are labeled in the upper right figure. Only 4 year-old BY 2006 natural-origin fish are represented in the SY 2010 collection.

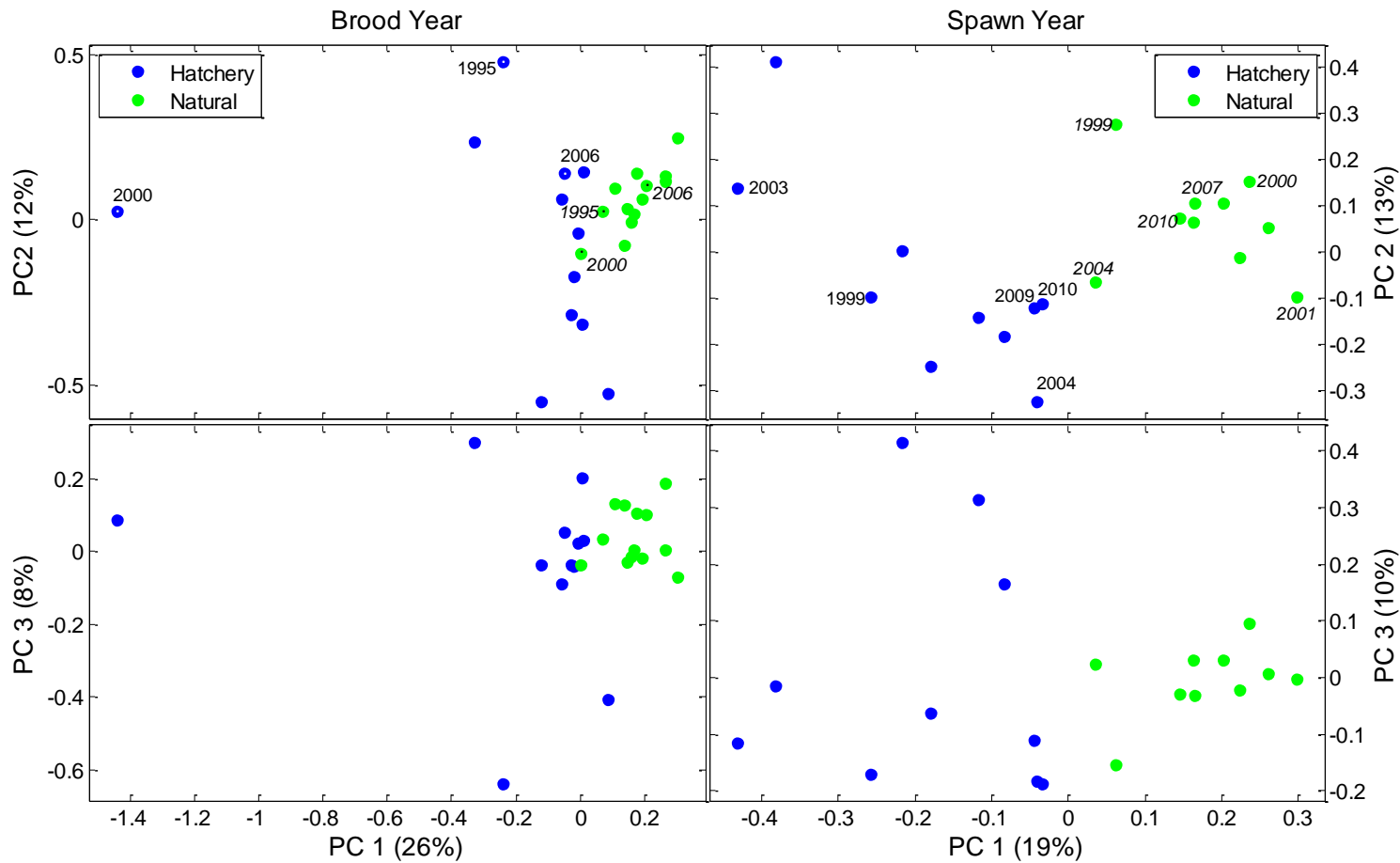


Figure 6. Principal component (PC) 1 versus 2 (top) and PC 1 versus 3 (bottom) based on an analysis using all adult and juvenile fish aggregated into age (juvenile versus adult), origin (hatchery versus adult) and spawn-year collections.

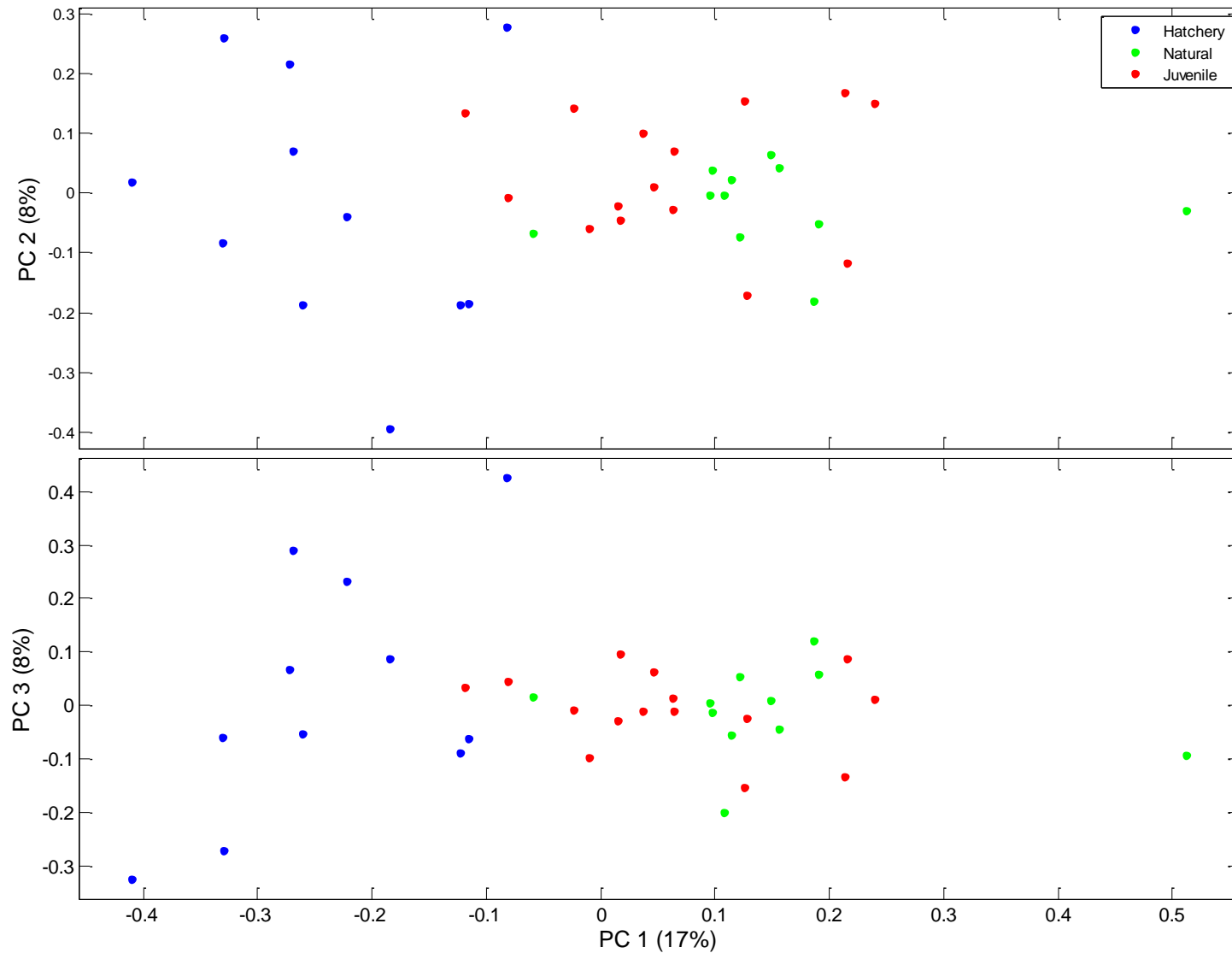


Figure 7. Pairwise Euclidian distances versus brood-year (top) and spawn-year (bottom), with zero distance equal to average distance across all pairwise distances. Blue lines are least-squares fits, which is not significant (slope = 0) for brood-year, but significant (slope > 0) for spawn-year.

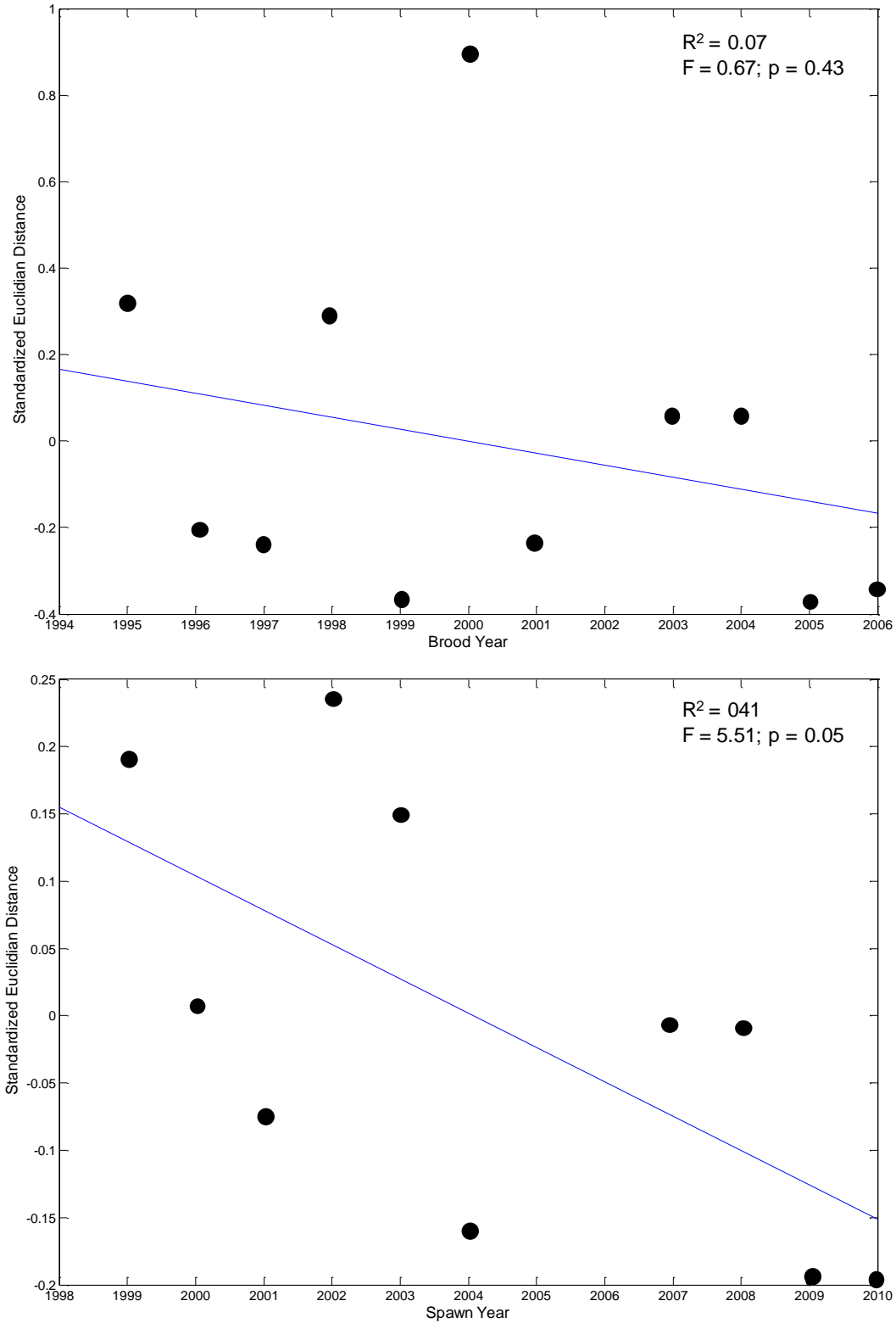


Figure 8. Effective population size estimates ( $N_b$ ) from Wenatchee River adult hatchery-produced steelhead annual collections calculated using single sample methods implemented in the program LDNE (Waples and Do 2008). Each line connects annual estimates of  $N_b$  estimated with a different value of  $P_{crit}$ , the smallest allelic proportion allowed during analysis. With SNP data, omitting an allele omits the locus. Estimates of  $N_b$  changed very little when  $P_{crit}$  varied from 0.1 to 0.001. Setting  $P_{crit} = 0.001$  forced the use of all available loci.

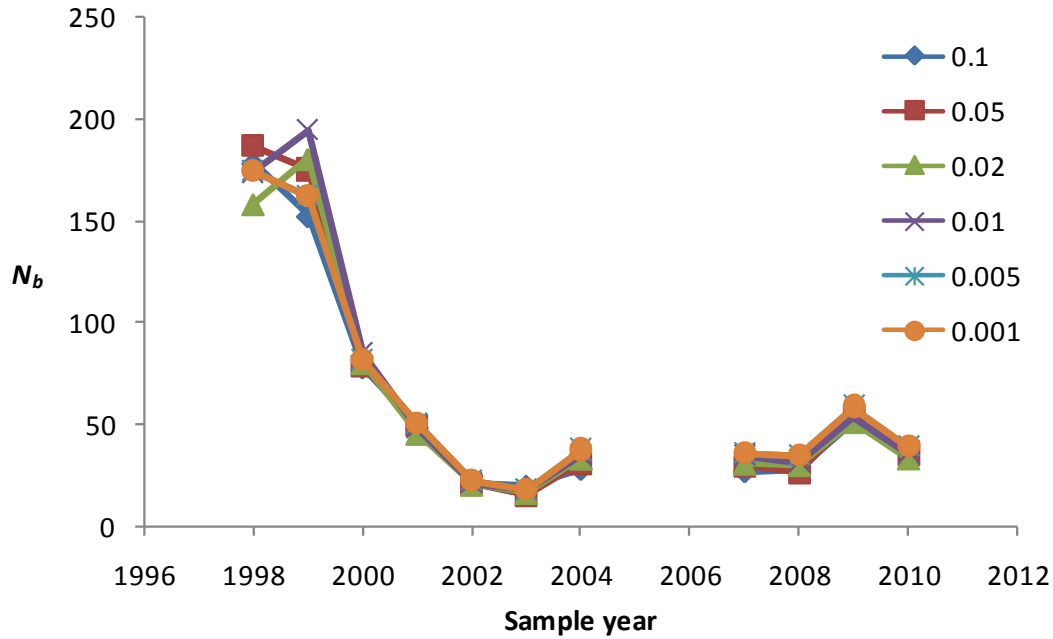




Figure 9. Estimates of Wenatchee River steelhead effective number of breeders ( $N_b$ ) estimated using the single sample methods incorporated in the program LDNE (Waples and Do 2008). Estimates of  $N_b$  refer to parental (and even grandparental) generations.  $N_b$  data were plotted against their estimated parental brood year. We assumed a 5 year generation time for natural origin adults (NOR), a 4 year generation time for hatchery-produced adults (HOR) and an age of smolt outmigration of age 2 for smolt collections from Wenatchee River tributaries (Chiwawa River, Nason Creek, Peshastin Creek), the lower Wenatchee River, and the Entiat River. Bars represent the 95% confidence interval estimated by jackknife procedure. Bars that exceed the upper limit of the Y axis are labeled with the upper bound (Inf. = infinity).

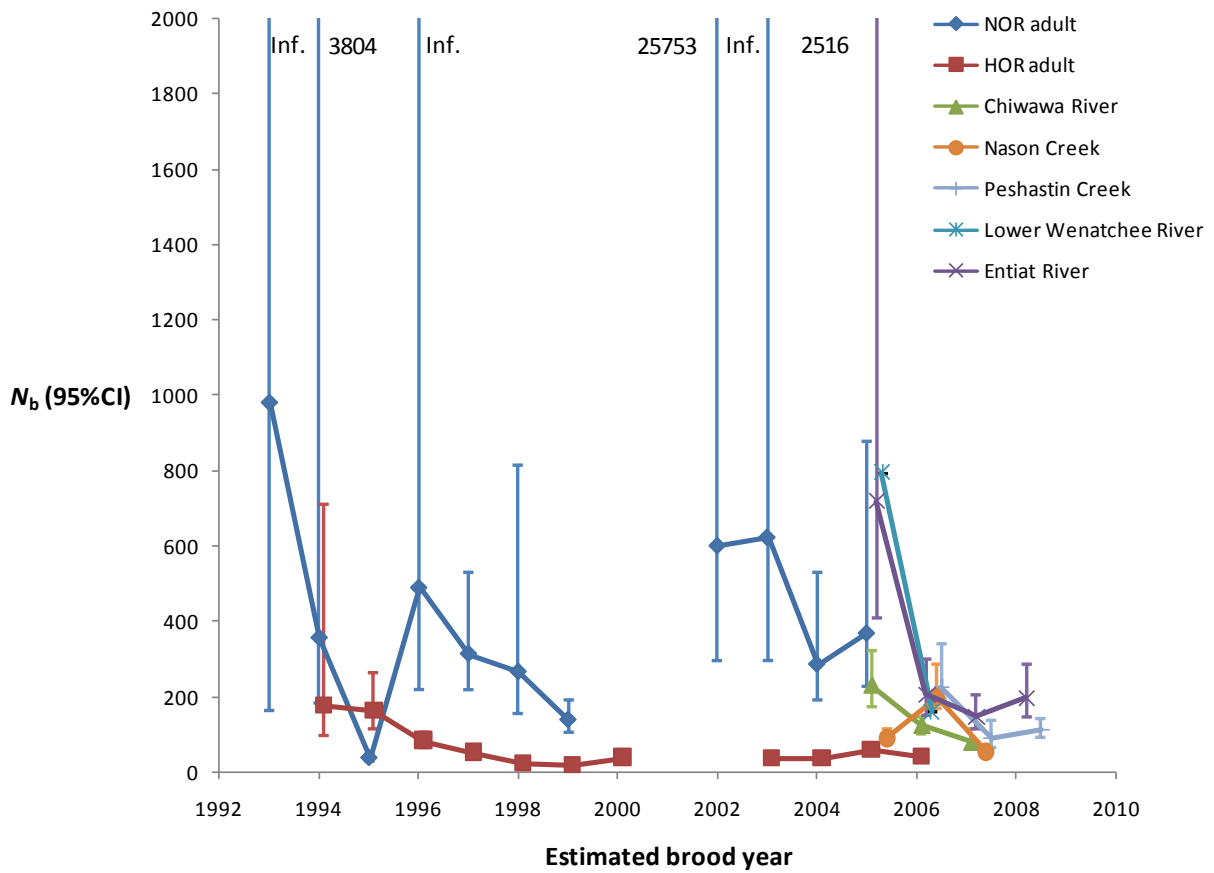


Figure 10. Estimates of  $N_b$  for collections of hatchery-produced (HOR) and natural origin (NOR) Wenatchee River summer steelhead grouped by brood year rather than spawn year. Brood year was estimated using scale-based age data. Error bars that extend past the top of the chart are all bounded by infinity.

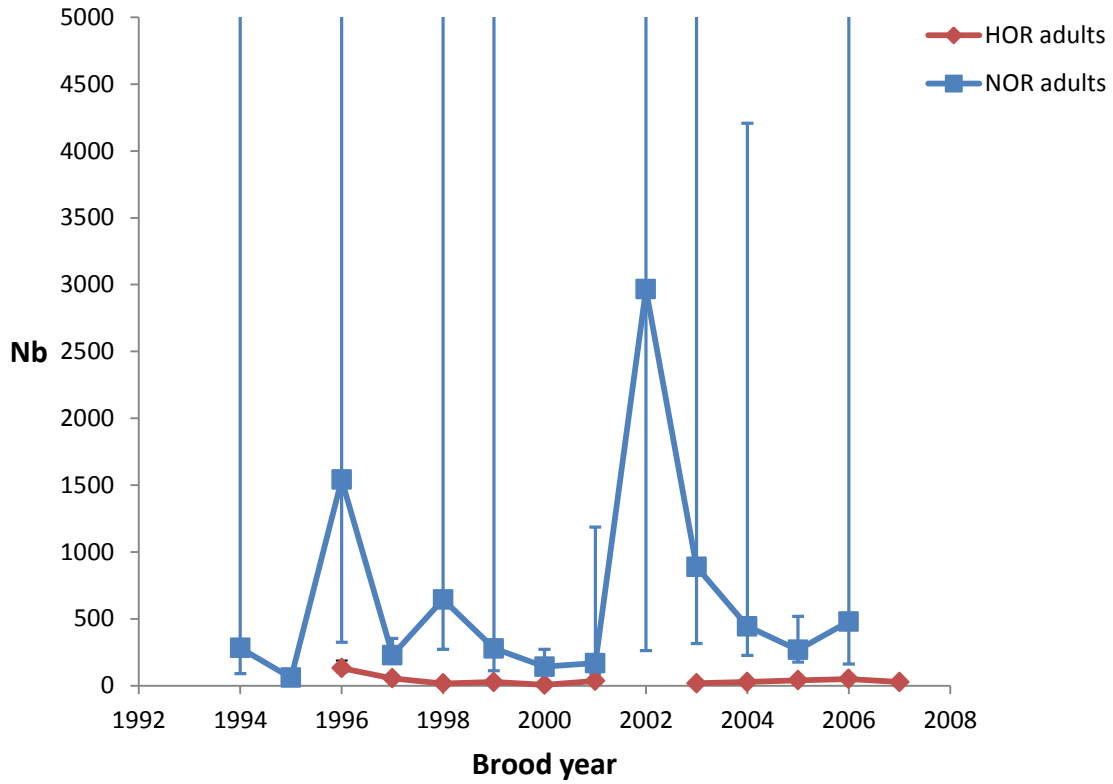


Figure 11. Estimates of  $N_b$  for combined annual adult hatchery-produced (HOR) and natural origin (NOR) steelhead and for HOR adults alone. The temporal patterns are similar, though estimates from combined collections are larger than those from HOR collections alone.

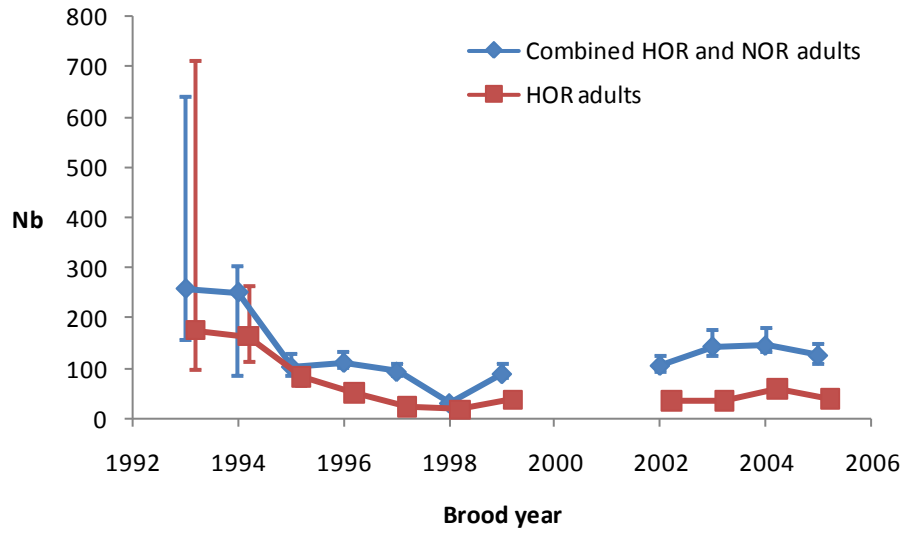


Figure 12.  $N_b/N$  ratios for hatchery-produced (HOR) and natural origin (NOR) adult Wenatchee River summer steelhead grouped by spawn year. The average  $N_b/N$  ratios are not different, though in later years NOR adults appear to have lower  $N_b/N$  ratios.

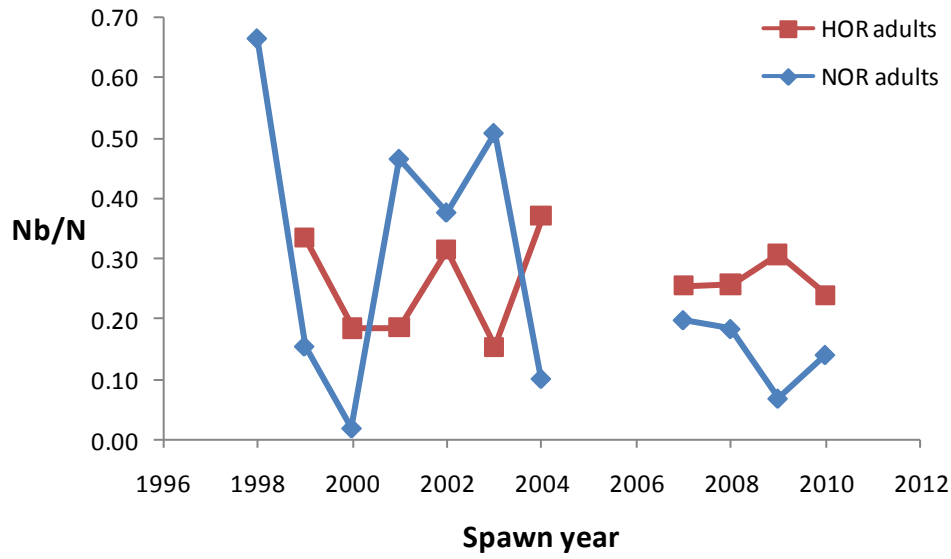
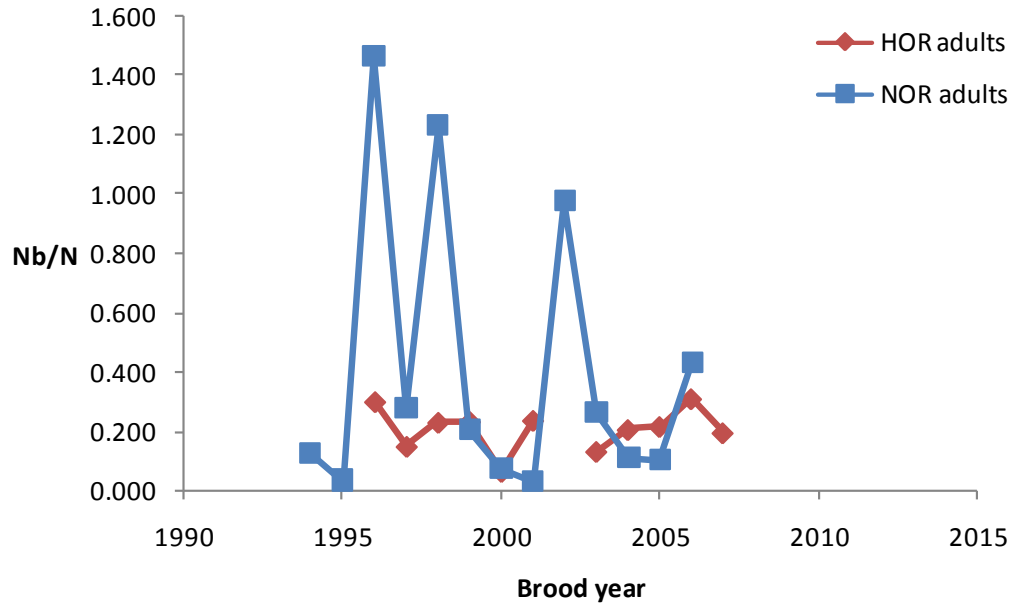


Figure 13.  $N_b/N$  ratios for hatchery-produced (HOR) and natural origin (NOR) adult Wenatchee River summer steelhead collections with individuals grouped in brood years rather than spawn years. Individual brood year was estimated using scale-based age data.



## Tables

Table 1. Samples of adult steelhead collected for Wenatchee Program broodstock and used for genetic monitoring and evaluation.

Origin	Sampling Location	Year spawned	WDFW Collection code	Samples (N)	Unused Samples <sup>a</sup>
Hatchery	Dryden/Tumwater Dams	1998	98AE	32	4
		1999	98LJ	62	2
		2000	99NE	60	5
		2001	00DQ	99	1
		2002	01MS	64	
		2003	02NP	89	
		2004	03KW	61	
		2007	06CW	64	1
		2008	08AG	56	
		2009	09AV	74	
		2010	10FE	76	1
		Total	737	14	
Natural	Dryden/Tumwater Dams	1998	98AF	30	5
		1999	99AA	51	1
		2000	99ND	33	3
		2001	00DP	50	
		2002	01MR	95	
		2003	02NO	50	
		2004	03KV	71	3
		2007	06CX	74	
		2008	08AF	74	1
		2009	09AU	82	2
		2010	10FD	90	2
		Total	700	17	

<sup>a</sup>Samples were not used if they had incomplete ( $\leq 80\%$  or 95 of 119 loci) or duplicate genotypes.

Table 2. Samples of natural origin juvenile steelhead and rainbow trout collected from four Wenatchee basin rivers or creeks and the Entiat River.

Sampling Location	Collection	WDFW Collection	Samples (N)	Unused samples <sup>a</sup>
	Year	Code		
Chiwawa River	2007	07AO	127	5
	2008	08CG	143	1
	2009	09NF	35	2
Entiat River	2007	07AL	134	4
	2008	08CI	82	4
	2009	09NC	74	1
	2010	10OX	82	1
Lower Wenatchee River	2007	07AM	139	5
	2008	08CE	98	2
Nason Creek	2007	07AN	81	4
	2008	08CF	133	6
	2009	09NG	103	2
Peshastin Creek	2008	08CH	142	2
	2009	09NE	34	1
	2010	10OY	94	1
		Total	1501	41

<sup>a</sup>Samples were not used if they were genetically identified as cutthroat trout or cutthroat/rainbow trout hybrids, or if they had incomplete ( $\leq 80\%$  or 95 of 119 loci) or duplicate genotypes.

Table 3. List of 132 general use, diploid single nucleotide polymorphic (SNP) loci genotyped in Wenatchee River basin and Entiat River steelhead.

WDFW Name	Locus Name	Allele 1	Allele 2	Reference
AOmy005	Omy_aspAT-123	T	C	(Campbell et al. 2009)
AOmy014	Omy_e1-147	G	T	(Sprowles et al. 2006)
AOmy015	Omy_gdh-271	C	T	(Campbell et al. 2009)
AOmy016	Omy_GH1P1_2	C	T	(Aguilar and Garza 2008)
AOmy021	Omy_LDHB-2_e5	T	C	(Aguilar and Garza 2008)
AOmy023	Omy_MYC_2	T	C	(Aguilar and Garza 2008)
AOmy027	Omy_nkef-241	C	A	(Campbell et al. 2009)
AOmy028	Omy_nramp-146	G	A	(Campbell et al. 2009)
AOmy047	Omy_u07-79-166	G	T	WDFW - S. Young unpubl.
AOmy051	Omy_121713-115	T	A	(Abadía-Cardoso et al. 2011)
AOmy056	Omy_128693-455	T	C	(Abadía-Cardoso et al. 2011)
AOmy059	Omy_187760-385	A	T	(Abadía-Cardoso et al. 2011)
AOmy061	Omy_96222-125	T	C	(Abadía-Cardoso et al. 2011)
AOmy062	Omy_97077-73	T	A	(Abadía-Cardoso et al. 2011)
AOmy063	Omy_97660-230	C	G	(Abadía-Cardoso et al. 2011)
AOmy065	Omy_97954-618	C	T	(Abadía-Cardoso et al. 2011)
AOmy067	Omy_aromat-280	A	T	WSU - J. DeKoning unpubl.
AOmy068	Omy_arp-630	G	A	(Campbell et al. 2009)
AOmy071	Omy_cd59-206	C	T	WSU - J. DeKoning unpubl.
AOmy073	Omy_colla1-525	C	T	WSU - J. DeKoning unpubl.
AOmy079	Omy_g12-82	T	C	WSU - J. DeKoning unpubl.
AOmy081	Omy_gh-475	C	T	(Campbell et al. 2009)
AOmy082	Omy_gsdf-291	T	C	WSU - J. DeKoning unpubl.
AOmy089	Omy_hsp90BA-193	C	T	(Campbell and Narum 2009)
AOmy094	Omy_inos-97	C	A	WSU - J. DeKoning unpubl.
AOmy095	Omy_mapK3-103	A	T	CRITFC - N. Campbell unpubl.
AOmy096	Omy_mcsf-268	T	C	WSU - J. DeKoning unpubl.
AOmy100	Omy_nach-200	A	T	WSU - J. DeKoning unpubl.



AOmy107	Omy_Ots249-227	C	T	(Campbell et al. 2009)
AOmy108	Omy_oxct-85	A	T	WSU - J. DeKoning unpubl.
AOmy110	Omy_star-206	A	G	WSU - J. DeKoning unpubl.
AOmy111	Omy_stat3-273	G	Deletion	WSU - J. DeKoning unpubl.
AOmy113	Omy_tlr3-377	C	T	WSU - J. DeKoning unpubl.
AOmy117	Omy_u09-52-284	T	G	WDFW - S. Young unpubl.
AOmy118	Omy_u09-53-469	T	C	WDFW - S. Young unpubl.
AOmy120	Omy_u09-54.311	C	T	WDFW - S. Young unpubl.
AOmy123	Omy_u09-55-233	A	G	WDFW - S. Young unpubl.
AOmy125	Omy_u09-56-119	T	C	WDFW - S. Young unpubl.
AOmy129	Omy_BAMBI4.238	T	C	WDFW - S. Young unpubl.
AOmy132	Omy_G3PD_2.246	C	T	WDFW - S. Young unpubl.
AOmy134	Omy_II-1b-028	T	C	WDFW - S. Young unpubl.
AOmy137	Omy_u09-61.043	A	T	WDFW - S. Young unpubl.
AOmy151	Omy_p53-262	T	A	CRITFC - N. Campbell unpubl.
AOmy173	BH2VHSVip10	C	T	Pascal & Hansen unpubl.
AOmy174	OMS00003	T	G	(Sánchez et al. 2009)
AOmy176	OMS00013	A	G	(Sánchez et al. 2009)
AOmy177	OMS00018	T	G	(Sánchez et al. 2009)
AOmy179	OMS00041	G	C	(Sánchez et al. 2009)
AOmy181	OMS00052	T	G	(Sánchez et al. 2009)
AOmy182	OMS00053	T	C	(Sánchez et al. 2009)
AOmy183	OMS00056	T	C	(Sánchez et al. 2009)
AOmy184	OMS00057	T	G	(Sánchez et al. 2009)
AOmy185	OMS00061	T	C	(Sánchez et al. 2009)
AOmy186	OMS00062	T	C	(Sánchez et al. 2009)
AOmy187	OMS00064	T	G	(Sánchez et al. 2009)
AOmy189	OMS00071	A	G	(Sánchez et al. 2009)
AOmy190	OMS00072	A	G	(Sánchez et al. 2009)
AOmy191	OMS00078	T	C	(Sánchez et al. 2009)
AOmy192	OMS00087	A	G	(Sánchez et al. 2009)

AOmy193	OMS00089	A	G	(Sánchez et al. 2009)
AOmy194	OMS00090	T	C	(Sánchez et al. 2009)
AOmy195	OMS00092	A	C	(Sánchez et al. 2009)
AOmy196	OMS00094	T	G	(Sánchez et al. 2009)
AOmy197	OMS00103	A	T	(Sánchez et al. 2009)
AOmy198	OMS00105	T	G	(Sánchez et al. 2009)
AOmy199	OMS00112	A	T	(Sánchez et al. 2009)
AOmy200	OMS00116	T	A	(Sánchez et al. 2009)
AOmy201	OMS00118	T	G	(Sánchez et al. 2009)
AOmy202	OMS00119	A	T	(Sánchez et al. 2009)
AOmy203	OMS00120	A	G	(Sánchez et al. 2009)
AOmy204	OMS00121	T	C	(Sánchez et al. 2009)
AOmy205	OMS00127	T	G	(Sánchez et al. 2009)
AOmy206	OMS00128	T	G	(Sánchez et al. 2009)
AOmy207	OMS00132	A	T	(Sánchez et al. 2009)
AOmy208	OMS00133	A	G	(Sánchez et al. 2009)
AOmy209	OMS00134	A	G	(Sánchez et al. 2009)
AOmy210	OMS00153	T	G	(Sánchez et al. 2009)
AOmy211	OMS00154	A	T	(Sánchez et al. 2009)
AOmy212	OMS00156	A	T	(Sánchez et al. 2009)
AOmy213	OMS00164	T	G	(Sánchez et al. 2009)
AOmy215	OMS00175	T	C	(Sánchez et al. 2009)
AOmy216	OMS00176	T	G	(Sánchez et al. 2009)
AOmy218	OMS00180	T	G	(Sánchez et al. 2009)
AOmy220	Omy_1004	A	T	(Hansen et al. 2011)
AOmy221	Omy_101554-306	T	C	(Abadía-Cardoso et al. 2011)
AOmy222	Omy_101832-195	A	C	(Abadía-Cardoso et al. 2011)
AOmy223	Omy_101993-189	A	T	(Abadía-Cardoso et al. 2011)
AOmy225	Omy_102505-102	A	G	(Abadía-Cardoso et al. 2011)
AOmy226	Omy_102867-443	T	G	(Abadía-Cardoso et al. 2011)
AOmy227	Omy_103705-558	T	C	(Abadía-Cardoso et al. 2011)

AOmy228	Omy_104519-624	T	C	(Abadía-Cardoso et al. 2011)
AOmy229	Omy_104569-114	A	C	(Abadía-Cardoso et al. 2011)
AOmy230	Omy_105075-162	T	G	(Abadía-Cardoso et al. 2011)
AOmy231	Omy_105385-406	T	C	(Abadía-Cardoso et al. 2011)
AOmy232	Omy_105714-265	C	T	(Abadía-Cardoso et al. 2011)
AOmy233	Omy_107031-704	C	T	(Abadía-Cardoso et al. 2011)
AOmy234	Omy_107285-69	C	G	(Abadía-Cardoso et al. 2011)
AOmy235	Omy_107336-170	C	G	(Abadía-Cardoso et al. 2011)
AOmy238	Omy_108007-193	A	G	(Abadía-Cardoso et al. 2011)
AOmy239	Omy_109243-222	A	C	(Abadía-Cardoso et al. 2011)
AOmy240	Omy_109525-403	A	G	(Abadía-Cardoso et al. 2011)
AOmy241	Omy_110064-419	T	G	(Abadía-Cardoso et al. 2011)
AOmy242	Omy_110078-294	A	G	(Abadía-Cardoso et al. 2011)
AOmy243	Omy_110362-585	G	A	(Abadía-Cardoso et al. 2011)
AOmy244	Omy_110689-148	A	C	(Abadía-Cardoso et al. 2011)
AOmy245	Omy_111005-159	C	T	(Abadía-Cardoso et al. 2011)
AOmy246	Omy_111084-526	A	C	(Abadía-Cardoso et al. 2011)
AOmy247	Omy_111383-51	C	T	(Abadía-Cardoso et al. 2011)
AOmy248	Omy_111666-301	T	A	(Abadía-Cardoso et al. 2011)
AOmy249	Omy_112301-202	T	G	(Abadía-Cardoso et al. 2011)
AOmy250	Omy_112820-82	G	A	(Abadía-Cardoso et al. 2011)
AOmy252	Omy_114976-223	T	G	(Abadía-Cardoso et al. 2011)
AOmy253	Omy_116733-349	C	T	(Abadía-Cardoso et al. 2011)
AOmy254	Omy_116938-264	A	G	(Abadía-Cardoso et al. 2011)
AOmy255	Omy_117259-96	T	C	(Abadía-Cardoso et al. 2011)
AOmy256	Omy_117286-374	A	T	(Abadía-Cardoso et al. 2011)
AOmy257	Omy_117370-400	A	G	(Abadía-Cardoso et al. 2011)
AOmy258	Omy_117540-259	T	G	(Abadía-Cardoso et al. 2011)
AOmy260	Omy_117815-81	C	T	(Abadía-Cardoso et al. 2011)
AOmy261	Omy_118175-396	T	A	(Abadía-Cardoso et al. 2011)
AOmy262	Omy_118205-116	A	G	(Abadía-Cardoso et al. 2011)

AOmy263	Omy_118654-91	A	G	(Abadía-Cardoso et al. 2011)
AOmy265	Omy_120255-332	A	T	(Abadía-Cardoso et al. 2011)
AOmy266	Omy_128996-481	T	G	(Abadía-Cardoso et al. 2011)
AOmy267	Omy_129870-756	C	T	(Abadía-Cardoso et al. 2011)
AOmy268	Omy_131460-646	C	T	(Abadía-Cardoso et al. 2011)
AOmy269	Omy_98683-165	A	C	(Abadía-Cardoso et al. 2011)
AOmy270	Omy_cyp17-153	C	T	WSU - J. DeKoning unpubl.
AOmy271	Omy_ftzf1-217	A	T	WSU - J. DeKoning unpubl.
AOmy272	Omy_GHSR-121	T	C	CRITFC - N. Campbell unpubl.
AOmy273	Omy_metA-161	T	G	CRITFC - N. Campbell unpubl.
AOmy274	Omy_UBA3b	A	T	(Hansen et al. 2011)

---

Primer and probe sequences for unpublished loci available by request.

Table 4. List of 20 species identification single nucleotide polymorphic (SNP) loci genotyped in Wenatchee River basin and Entiat River steelhead.

WDFW Name	Locus Name	Expected genotype			Reference
		<i>O. mykiss</i>	<i>O. clarkii clarkii</i>	<i>O. clarkii lewisi</i>	
ASpI001	Ocl_Okerca	T	C	C	(McGlaufflin et al. 2010)
ASpI002	Ocl_Oku202	A	C	C	(McGlaufflin et al. 2010)
ASpI003	Ocl_Oku211	G	T	T	(McGlaufflin et al. 2010)
ASpI004	Ocl_Oku216	C	C	A	(McGlaufflin et al. 2010)
ASpI005	Ocl_Oku217	C	C	A	(McGlaufflin et al. 2010)
ASpI006	Ocl_SsaHM5	A	A	G	(McGlaufflin et al. 2010)
ASpI007	Ocl_u800	T	C	C	(McGlaufflin et al. 2010)
ASpI008	Ocl_u801	A	T	T	(McGlaufflin et al. 2010)
ASpI009	Ocl_u802	C	C	T	(McGlaufflin et al. 2010)
ASpI010	Ocl_u803	C	T	T	(McGlaufflin et al. 2010)
ASpI011	Ocl_u804	G	G	C	(McGlaufflin et al. 2010)
ASpI012	Omy_B9_228	A	A	C	(Finger et al. 2009)
ASpI013	Omy_CTDL1_243	C	A	A	(Finger et al. 2009)
ASpI014	Omy_F5_136	C	G	G	(Finger et al. 2009)
ASpI016	Omy_myclarp404-111	T	G	G	CRITFC - S. Narum - unpubl.
ASpI017	Omy_myclgh1043-156	C	T	T	CRITFC - S. Narum - unpubl.
ASpI018	Omy_Omyclmk436-96	A	C	C	CRITFC - S. Narum - unpubl.
ASpI019	Omy_RAG11_280	T	A	A	(Sprowles et al. 2006)
ASpI020	Omy_URO_302	T	C	C	(Finger et al. 2009)
ASpI021	Omy_BAC-F5.238	C	G	G	WDFW - S. Young unpubl.

Primer and probe sequences for unpublished loci available by request.

Table 5. Pairwise  $F_{ST}$  estimates for collections from Wenatchee River tributaries and the Entiat River (below diagonal) and associated bootstrap estimated  $P$ -values (above diagonal).

Population	Year	Chiwawa River			Nason Creek			Peshastin Creek			Lower Wenatchee River		Entiat River			
		2007	2008	2009	2007	2008	2009	2008	2009	2010	2007	2008	2007	2008	2009	2010
Chiwawa River	2007		<b>0.000</b>	<b>0.003</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.002</b>	<b>0.000</b>	<b>0.001</b>	<b>0.001</b>	<b>0.000</b>	<b>0.001</b>	<b>0.000</b>	<b>0.000</b>
	2008	0.004		<b>0.004</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>
	2009	0.004	0.003		<b>0.000</b>	<b>0.001</b>	0.061	<b>0.000</b>	<b>0.001</b>	<b>0.000</b>	0.086	<b>0.050</b>	<b>0.022</b>	0.108	<b>0.005</b>	<b>0.045</b>
Nason Creek	2007	0.011	0.010	0.007		<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>
	2008	0.007	0.007	0.005	0.009		<b>0.003</b>	<b>0.000</b>	<b>0.002</b>	<b>0.000</b>	0.079	<b>0.000</b>	<b>0.001</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>
	2009	0.007	0.007	0.003	0.014	0.006		<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>
Peshastin Creek	2008	0.010	0.011	0.008	0.013	0.010	0.013		<b>0.001</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>
	2009	0.005	0.005	0.006	0.010	0.007	0.008	0.003		<b>0.002</b>	<b>0.002</b>	<b>0.047</b>	<b>0.028</b>	<b>0.004</b>	<b>0.005</b>	<b>0.001</b>
	2010	0.010	0.011	0.008	0.015	0.008	0.011	0.003	0.003		<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>	<b>0.000</b>
Lower Wenatchee River	2007	0.003	0.003	0.000	0.005	0.008	0.007	0.009	0.010	0.008		0.112	<b>0.020</b>	<b>0.012</b>	<b>0.002</b>	<b>0.017</b>
	2008	0.002	0.005	0.002	0.003	0.004	0.005	0.007	0.009	0.006	0.000		<b>0.049</b>	0.459	<b>0.047</b>	<b>0.002</b>
Entiat River	2007	0.005	0.006	0.002	0.005	0.006	0.005	0.005	0.007	0.006	0.001	0.002		0.451	0.173	<b>0.000</b>
	2008	0.004	0.004	0.000	0.007	0.005	0.007	0.008	0.009	0.011	0.002	0.001	0.000		0.644	<b>0.002</b>
	2009	0.005	0.006	0.002	0.003	-0.001	0.003	0.002	0.003	0.004	0.003	0.002	0.002	0.000		<b>0.028</b>
	2010	0.005	0.006	0.003	0.006	0.004	0.006	0.006	0.006	0.008	0.009	0.002	0.003	0.003	0.003	0.002

$P$ -values in bold were significant at  $\alpha = 0.05$  after correcting for multiple tests using false discovery rate.







# Appendix F

**NPDES Hatchery Effluent Monitoring, 2014**



## NPDES MONITORING FOR WDFW FACILITIES

All WDFW hatcheries monitor their discharge in accordance with the National Pollutant Discharge Elimination System (NPDES) permit. This permit is administered in Washington by the Washington Department of Ecology under agreement with the United States Environmental Protection Agency. The permit was renewed effective 1 August 2010 and will expire 1 August 2015.

Facilities are exempted from sampling during any month that pounds of fish on hand falls below 20,000 lbs and pounds of feed used falls below 5,000 lbs, with the exception of offline settling basin discharges, which are to be monitored once per month when ponds are in use and discharging to receiving waters.

Sampling at permitted facilities includes the following parameters:

<FLOW	Measured in millions of gallons per day (MGD) discharge.
<SS EFF	Average net settleable solids in the hatchery effluent, measured in ml/L.
<TSS COMP	Average net total suspended solids, composite sample (6 x/day) of the hatchery effluent, measured in mg/L.
<TSS MAX	Maximum daily net total suspended solids, composite sample (6 x/day) of the hatchery effluent, measured in mg/L.
<SS PA	Maximum settleable solids discharge from the pollution abatement pond, measured in ml/L.
<SS %	Removal of settleable solids within the pollution abatement pond from inlet to outlet, measured as a percent. No longer required under permit effective 1 June 2000.
<TSS PA	Maximum total suspended solids effluent grab from the pollution abatement pond discharge, measured in mg/L.
<TSS %	Removal of suspended solids within the pollution abatement pond from inlet to outlet, measured as a percent. No longer required under permit effective 1 June 2000.
<SS DD	Settleable solids discharged during drawdown for fish release. One sample per pond drawdown, measured in ml/L.
<TRC	Total residual chlorine discharge after rearing vessel disinfection and after neutralization with sodium thiosulfate. One sample per disinfection, measured in ug/L.

In addition, at Similkameen Hatchery only, the following sampling was conducted at the request of Washington Department of Ecology, but is not required under NPDES permit:

<SS IW	Settleable solids influent grab taken as wastes are pumped into the pollution abatement pond, measured in mg/L. No longer monitored as of January 2008.
<TSS IW	Total suspended solids influent grab as wastes are pumped into the pollution abatement pond, measured in mg/L. No longer monitored as of January 2008.

**Eastbank Hatchery**  
**NPDES Permit Number WAG13-5011**

		<b>FLO W</b>	<b>SS EFF</b>	<b>TSS COMP</b>	<b>TSS MAX</b>	<b>FLOW PA</b>	<b>SS PA</b>	<b>SS %</b>	<b>TSS PA</b>	<b>TSS %</b>	<b>Lbs of Fish</b>	<b>Lbs of Feed</b>
2014	JAN	23.91	0	1	1	7000	0.01		6.6		43982	11617
	FEB	23.91	0	0.9	1	8500	0.01		22.2		56061	10468
	MAR	30.37	0	0	0	10000	0.01		15		33145	3853
	APR	15.51	0	0	0	3000	0.01		13		5220	2440
	MAY	21.32	0	0.2	0.2	7500	0.01		8.2		12718	3928
	JUN	23.26	0	0	0	5000	0.01		6.2		12204	2614
	JUL	31.02	0	0	0	5000	0.01		22.2		10343	3756
	AUG	23.26	0	0.15	0.2	7500	0.01		17.2		16967	6373
	SEP	31.02	0	0.8	0.8	15000	0.01		20.2		24733	10184
	OCT	31.02	0	1.2	1.2	15000	0.01		12.8		24536	12651
	NOV	23.6	0	0.6	0.6	7500	0.01		17.4		18276	3703
	DEC	23.26	0	0.6	0.6	5000	0.01		15.2		18768	4604

**Wells Hatchery**

**NPDES Permit Number WAG13-5009**

		FLO W	SS EFF	TSS COMP	TSS MAX	FLOW PA	SS PA	SS %	TSS PA	TSS %	Lbs of Fish	Lbs of Feed	SS DD	TSS DD
2014	JAN	14.86	0	1.6	1.6	**	**		**		85852	13611		
	FEB	17.4	0.01	0.4	0.4	**	**		**		101614	11870		
	MAR	18.5	0	0.8	0.8	**	**		**		122936	20220		
	APR	15.59	0	-0.5	-0.4	17	0.01		4.2		125516	8546		
	MAY	9.13	0	-1.2	-1.2	**	**		**		27562	3391	0.2	4.6
	JUN	2.36	0.01	1.4	1.4	17	0.1		5.6		3728	2264		
	JUL	3.87	0	0	0	17	0.1		1.8		7687	2251		
	AUG	3.8	0	3	3	17	0.1		10.6		9048	5837		
	SEP	7.42	0.02	0.1	0.1	17	0.5		3.8		23287	8741		
	OCT	8.74	0.01	3.93	6.4	17	0.3		14.6		27500	10688		
	NOV	9.88	0	0	0	**	**		**		41490	9634		
	DEC	13.27	0	-0.6	-0.6	**	**		**		49582	10098		

\*\* PA pond - No discharge this month

**Chiwawa Ponds - Chiwawa River**

**NPDES Permit Number WAG13-5015**

		FLOW	SS EFF	TSS COMP	TSS MAX	Lbs of Fish	Lbs of Feed	SS DD	TSS DD
2014	JAN	4.23	0	-0.2	-0.2	12732	390		
	FEB	3	0	0.6	0.6	13038	378		
	MAR	3.8	0	-0.2	-0.2	13043	1348		
	APR	4.52	0.03	-0.8	-0.8	12460	2364	0.04	1.2
	MAY	No Monitoring				0	0		
	JUN	No Monitoring				0	0		
	JUL	No Monitoring				0	0		
	AUG	No Monitoring				0	0		
	SEP	4.62	0	0	0	6200	44		
	OCT	4.59	0	1	1	6193	748		
	NOV	4.22	0	1.2	1.2	8423	562		
	DEC	3.95	0	1	1	8143	436		

**Chiwawa Ponds - Wenatchee River  
NPDES Permit Number WAG13-5015**

		<b>FLO W</b>	<b>SS EFF</b>	<b>TSS COMP</b>	<b>TSS MAX</b>	<b>Lbs of Fish</b>	<b>Lbs of Feed</b>	<b>SS DD</b>	<b>TSS DD</b>
2014	JAN	4.6	0	-0.4	-0.4	14861	1962		
	FEB	4.73	0	1.6	1.6	15765	852		
	MAR	1.9	0	0	0	16595	2120		
	APR	3.73	0	1.2	1.2	21083	3766		
	MAY	6.01	0.03	0.8	1	16348	88		
	JUN	No Monitoring				0	0		
	JUL	No Monitoring				0	0		
	AUG	No Monitoring				0	0		
	SEP	No Monitoring				0	0		
	OCT	No Monitoring				0	0		
	NOV	7.69	0	-0.6	-0.6	10929	658		
	DEC	4.96	0	0	0	11927	882		

**Carlton Acclimation  
Pond  
NPDES Permit Number WAG13-5013**

2014 Permit became inactive in January. No Monitoring or discharge reports required for inactive permit.

**Methow Hatchery**

**NPDES Permit Number WAG13-5000**

		FLO W	SS EFF	TSS COMP	TSS MAX	FLOW PA	SS PA	SS %	TSS PA	TSS %	Lbs of Fish	Lbs of Feed	SS DD	TSS DD
2014	JAN	10.3	0	0.6	0.6	14400	0.1		2.8		10700	1200		
	FEB	10.3	0	2.5	3.2	14400	0.1		0.4		13000	1400		
	MAR	8.71	0	0	0	14400	0.1		7.6		14600	1450		
	APR	2.16	0	1.8	2.4	14400	0.1		1.8		1400	450		
	MAY	2.16	0	0.8	1	14400	0.1		0.8		1800	550	0.1	1.9
	JUN	2.16	0	0	0	14400	0.1		0		2250	800		
	JUL	3.89	0	0.2	0.2	14400	0.1		0.6		2700	1000		
	AUG	3.89	0	0	0	14400	0.1		0.8		3100	1200		
	SEP	3.89	0	0	0	14400	0.1		2		3500	1300		
	OCT	4.57	0	0	0	14400	0.1		0.2		7100	600		
	NOV	5.26	0	0.4	0.4	14400	0.1		2		7150	600		
	DEC	10.15	0	0.2	0.2	14400	0.1		2.8		4550	715		

**Similkameen Hatchery**

**NPDES Permit Number WAG13-5007**

		FLO W	SS EFF	TSS COMP	TSS MAX	FLOW PA	SS IW	TSS IW	Lbs of Fish	Lbs of Feed	SS DD	TSS DD
2014	JAN	6.62	0	0.8	1				4050	0		
	FEB	6.62	0	0.6	0.6				4040	0		
	MAR	6.62	0	4.4	4.4				4941	968		
	APR	6.62	0	0.6	0.6				2468	1892	0	28.8
	MAY	No Monitoring							0	0		
	JUN	No Monitoring							0	0		
	JUL	No Monitoring							0	0		
	AUG	No Monitoring							0	0		
	SEP	No Monitoring							0	0		
	OCT	No Monitoring							0	0		
	NOV	6.62	0	-1.6	-1.6				9591	836		
	DEC	6.62	0	-0.8	-0.8				9560	0		

**Chelan Hatchery**

**NPDES Permit Number WAG13-5006**

		<b>FLO</b>	<b>SS</b>	<b>TSS</b>	<b>TSS</b>	<b>FLOW</b>					<b>Lbs of</b>	<b>Lbs of</b>
		<b>W</b>	<b>EFF</b>	<b>COMP</b>	<b>MAX</b>	<b>PA</b>	<b>SS PA</b>	<b>SS %</b>	<b>TSS PA</b>	<b>TSS %</b>	<b>Fish</b>	<b>Feed</b>
2014	JAN	7	0.05	-0.4	-0.4	68000	0.05		3.2		20436	4978
	FEB	7	0.05	0	0	68000	0.05		28.2		25460	4301
	MAR	8.5	0.05	1.6	1.8	68000	0.05		14		29005	8773
	APR	6.48	0.04	0.8	0.8	**	**		**		40000	4285
	MAY	6.48	0.05	0.6	0.6	68000	0.05		7.8		4500	2250
	JUN	10.5	0.05	1.2	1.4	68000	0.05		1.6		4891	3673
	JUL	9.5	0.05	-0.6	-0.6	9500000	0.05		2.8		8533	3860
	AUG	9.9	0.05	-0.6	-0.6	9900000	0.05		6		21791	5237
	SEP	10.7	0.05	*5.4	5.4	107000000	0.05		5.2		41823	11213
	OCT	11	0.05	0	0	68000	0.05		3.8		12689	4900
	NOV	5.53	0.05	0	0	68000	0.05		2		1968	2280
	DEC	6	0.05	0.2	0.2	68000	0.05		1.8		4711	2862

\*\*PA pond - No discharge this month

\*One violation reported.

**Chelan Falls Hatchery**

**NPDES Permit Number WAG13-7019**

		<b>FLO</b>	<b>SS</b>	<b>TSS</b>	<b>TSS</b>	<b>FLOW</b>					<b>Lbs of</b>	<b>Lbs of</b>
		<b>W</b>	<b>EFF</b>	<b>COMP</b>	<b>MAX</b>	<b>PA</b>	<b>SS PA</b>	<b>SS %</b>	<b>TSS PA</b>	<b>TSS %</b>	<b>Fish</b>	<b>Feed</b>
2014	JAN	7	0.05	-2.7	-2.6	857	0.05		23.4		28000	3644
	FEB	12.8	0.05	-0.2	-0.2	857	0.05		6.8		31111	5107
	MAR	12.8	0.05	-12.4	-12.4	857	0.05		2.8		35000	9772
	APR	12.8	0.05	-8.4	-8.4	857	0.05		3.2		22512	5728
	MAY	No Monitoring									0	0
	JUN	No Monitoring									0	0
	JUL	No Monitoring									0	0
	AUG	No Monitoring									0	0
	SEP	No Monitoring									0	0
	OCT	No Monitoring									0	0
	NOV	7	0.04	-1	-1	3000	0.05		0.8		24489	1830
	DEC	7	0.04	-0.4	-0.4	3000	0.05		3		27090	5937



**Dryden Acclimation Pond**  
**NPDES Permit Number WAG13-5014**

		<b>FLO</b>	<b>SS</b>	<b>TSS</b>	<b>TSS</b>	<b>Lbs of</b>	<b>Lbs of</b>		
		<b>W</b>	<b>EFF</b>	<b>COMP</b>	<b>MAX</b>	<b>Fish</b>	<b>Feed</b>	<b>SS DD</b>	<b>TSS DD</b>
2014	JAN	No Monitoring				0	0		
	FEB	No Monitoring				0	0		
	MAR	9.43	0	0.2	0.2	40315	836		
	APR	16.16	-0.01	0.7	1.6	48973	5236		
	MAY	No Monitoring				0	0		
	JUN	No Monitoring				0	0		
	JUL	No Monitoring				0	0		
	AUG	No Monitoring				0	0		
	SEP	No Monitoring				0	0		
	OCT	No Monitoring				0	0		
	NOV	No Monitoring				0	0		
	DEC	No Monitoring				0	0		

**Priest Rapids**  
**NPDES Permit Number WAG13-7013**

		FLO W	SS EFF	TSS COMP	TSS MAX	FLOW PA	SS PA	TSS PA	Lbs of Fish	Lbs of Feed	SS DD	TSS DD
2014	JAN	9.76	0	0	0				7000	0		
	FEB	12.3	0	0.8	0.8				14756	592		
	MAR	15.45	0	-0.2	-0.2				9170	4974		
	APR	21.28	0	1	1		0	22.2	27129	12666		
	MAY	40.25	0	2.4	2.4		0	23.2	78430	30166		
	JUN	35.48	0	3.4	3.4		0	23.2	145000	35896	0	2
	JUL	No Monitoring							0	0		
	AUG	No Monitoring							0	0		
	SEP	68.5	0			**	**	**	19490	0		
	OCT	68.5	0			**	**	**	58210	0		
	NOV	68.5	0			**	**	**	39830	0		
	DEC	23.25	0	-0.2	-0.2	**	**	**	8750	0		

New Pollution Abatement pond began discharging in April.

\*\*PA pond - No discharge this month

# Appendix G

**Steelhead Stock Assessment at Priest Rapids Dam, 2012-2013**



# **Priest Rapids Dam 2012-2013 Adult Upper Columbia River Steelhead Run-Cycle Stock Assessment Report**

## **Introduction**

Upper Columbia River (UCR) steelhead stock assessment sampling at Priest Rapids Dam (PRD) is authorized through the Endangered Species Act (ESA) Section 10 Permit 1395 (NMFS 2003). Permit authorizations include interception and biological sampling of up to 10 percent of the UCR steelhead passing PRD to determine upriver population size, estimate hatchery to wild ratios, determine age class contribution and evaluate the need for managing hatchery steelhead consistent with ESA recovery objectives which include fully seeding spawning habitat with naturally produced UCR steelhead supplemented with artificially propagated enhancement steelhead (NMFS 2003).

## **Stock Assessment**

The 2012 steelhead sampling at Priest Rapids Dam began 10 July and concluded 14 November. Sampling consisted of operating the Priest Rapids Off Ladder Trap (OLAFT), located on the left bank Priest Rapids Dam, 8 hours per day, on Tuesdays and Thursdays, for a total of 37 sampling days. Steelhead were trapped, handled and released in accordance with Section 2.1 and 2.2.1 of the National Marine Fisheries Service (NMFS) Biological Opinion for ESA Permit 1395 (NMFS 2003). The cumulative sample rate attained during 2012 totaled 13.5%.

The Washington Department of Fish and Wildlife (WDFW) sampled 2,318 steelhead of the 2012/2012 run-cycle passing PRD, totaling 17,192 steelhead, for an overall sampling rate of 13.5%. Of the 2,716 steelhead sampled, 1,875 (80.9%) were hatchery origin and 443 (19.1%) were wild origin. The estimated 2012-2013 run- cycle total wild steelhead return was 3,284 representing 118.3% of the 1986-2011 average and about 62.9% of the most recent 5-year average (Table 1).

Based on external marks and external and internal tags, 1,875 hatchery origin steelhead were sampled at Priest Rapids Dam during the 2012 return cycle and included, 19.6% Wenatchee hatchery-origin steelhead and 58.3% “above Wells Dam” hatchery origin steelhead <sup>1/</sup> (Table 2), while 12.5% of the hatchery origin steelhead sampled could not be assigned to a specific hatchery program. Ringold FH origin steelhead represented about 5.4% of the sample (Table 2).

1/- Defined as “above Wells Dam” because hatchery origin, adipose-clipped steelhead release into the Methow and Okanogan rivers from the Wells FH and Winthrop NFH have the same marks and are indistinguishable from one another.

**Table 1.** Priest Rapids Dam adult steelhead returns and stock composition, 1974-2012

Run-cycle <sup>1/</sup>	Hatchery	Wild	Wild percent	Total run
1974				2,950
1975				2,560
1976				9,490
1977				9,630
1978				4,510
1979				8,710
1980				8,290
1981				9,110
1982				10,770
1983				32,000
1984				26,200
1985				34,010
1986	20,022	2,342	10.5	22,364
1987	9,955	4,058	29.0	14,013
1988	7,530	2,670	26.2	10,200
1989	8,033	2,685	25.1	10,718
1990	6,252	1,585	20.2	7,837
1991	11,169	2,799	20.0	13,968
1992	12,102	1,618	11.8	13,720
1993	4,538	890	16.4	5,428
1994	5,880	855	12.7	6,735
1995	3,377	993	22.7	4,370
1996	7,757	843	9.8	8,600
1997	8,157	785	8.8	8,942
1998	4,919	928	15.9	5,847
1999	6,903	1,374	16.6	8,277
2000	9,023	2,341	20.6	11,364
2001	24,362	5,715	19.0	30,077
2002	12,884	2,983	18.8	15,867
2003	14,890	2,837	16.0	17,729
2004	15,670	2,985	16.0	18,655
2005	10,352	3,127	23.2	13,479
2006	8,738	1,677	16.1	10,415
2007	12,160	3,097	20.3	15,257
2008	13,528	3,030	18.3	16,558
2009	32,557	7,439	18.6	39,996
2010	18,784	7,647	28.9	26,431
2011	15,910	4,896	23.5	20,806
2012	13,908	3,284	19.1	17,192
<b>1986-2011 average</b>	<b>11,748</b>	<b>2,777</b>	<b>18.7</b>	<b>14,102</b>
<b>2007-2011 average</b>	<b>18,589</b>	<b>5,220</b>	<b>21.9</b>	<b>23,810</b>

<sup>1/</sup> A return cycle is the combined total of steelhead passing PRD from 1 June – 30 November during year (x), plus steelhead passing PRD between 15 April and 31 May on year (x+1).

**Table 2.** Origin classification of steelhead sampled at Priest Rapids Dam, 10 July – 14 November 2012.

Wild Wild			Steelhead origin																		
			Wenatchee					Hatchery				Ringold FH			Unk. Hat.			Total	Total	Total	
Criteria			VIE					Criteria				Criteria			Criteria			Total	Total	Total	
NS	NM	Total	LTGR	RTGR	RTOR	RTPK	RTRD	Total	AD	LTYL	LV	Total	AD	RV	Total	SD	NM	Total	Wild	Hatchery	Total
x	x	443	x					126	x			1,077	x	x	126	x	x	288			1,875
				x				89		x		13									
					x			0			x	4									
						x		152													
							x	0													
<b>Total</b>		<b>443</b>						<b>367</b>				<b>1,094</b>			<b>126</b>			<b>288</b>	<b>443</b>	<b>1,875</b>	<b>2,318</b>
<b>% Hatchery</b>								<b>19.6</b>				<b>58.3</b>			<b>6.7</b>			<b>15.4</b>			<b>100.0</b>
<b>% Total</b>		<b>19.1%</b>						<b>15.8</b>				<b>47.2</b>			<b>5.4</b>			<b>12.5</b>	<b>19.1</b>	<b>80.9</b>	<b>100.0</b>

Reconciliation of salt water age of wild and hatchery steelhead sampled at Priest Rapids Dam during 2012 was accomplished through scale sample analysis. Salt-age analysis of the 2012 UCR steelhead run-cycle provides an estimated hatchery-origin return dominated by 1- salt and 2-salt age composition of 43.2% and 56.2%, respectively (Table 3). Natural origin steelhead salt ages were 35.4% and 61.9% for salt ages 1 and 2, respectively. Three-salt age fish represented only 0.9% of the combined hatchery/wild sample (Table 3).

**Table 3.** Salt-water age composition of 2012 - 2013 return cycle Upper Columbia River steelhead sampled at Priest Rapids Dam, corrected by scale age/origin determination.

Salt-age	Origin					
	Hatchery		Wild		Combined	
	<i>N</i>	%	<i>N</i>	%	<i>N</i>	%
1-salt	801	43.2	155	35.4	956	41.7
2-salt	1,043	56.2	271	61.9	1,314	57.3
3-salt	11	0.6	9	2.1	20	0.9
4-salt	1	0.1	3	0.7	4	0.2
<b>Total</b>	<b>1,856</b>	<b>80.9</b>	<b>438</b>	<b>19.1</b>	<b>2,294</b>	<b>100</b>

Freshwater residency of naturally produced Upper Columbia River steelhead present in the 2012-2013 run cycle were dominated by age-2 freshwater fish (67.5%), and was slightly higher than the 1986-2011 average of 75.2% (Table 4).

**Table 4.** 2012 return year freshwater age of wild Upper Columbia River steelhead sampled at Priest Rapids Dam during steelhead stock assessment activities, compared to July – November 1986-2011 average.

Freshwater age	2012-2013 run cycle		1986-2011 proportion	
	<i>N</i>	%	<i>N</i>	%
1.x	51	13.1	407	8.0
2.x	262	67.5	3,824	75.2
3.x	71	18.3	814	16.0
4.x	3	0.8	36	0.7
5.x	1	0.3	2	<0.1
<b>Total</b>	<b>388</b>		<b>5,083</b>	

Wild and hatchery origin steelhead exhibited similar saltwater growth in the 2012 run-cycle. Wild 1 and 2-salt adults were slightly larger than their hatchery cohorts (Table 5). Age 1-salt hatchery and age 1 and 2-salt wild steelhead observed in the 2012-2013 adult run-cycle return past PRD were comparable in size to the 1986-2011 run-cycle average (Table 5).



**Table 5.** Average fork length of 1-salt and 2-salt, Upper Columbia River steelhead sampled at Priest Rapids Dam during July – November 2012 and the period between 1986-2011.

Salt age	Average fork length (cm)			
	2012-2013 run cycle		1986-2011 run cycle	
	Wild	Hatchery	Wild	Hatchery
x.1	59.2	57.7	60.1	59.1
x.2	72.4	71.2	72.7	71.8



# Appendix H

**Wenatchee Sockeye Salmon Spawning Escapement, 2014**



**PUBLIC UTILITY DISTRICT NUMBER 1 OF CHELAN COUNTY**  
**Natural Resource Division**  
**Fish and Wildlife Department**  
*327 N. Wenatchee Ave., Wenatchee WA 98801 (509) 663-8121*

**March 20, 2015**

To: HCP Hatchery Committee

From: Catherine Willard

**Subject: 2014 Wenatchee Sockeye Mark/Recapture-Based Sockeye Escapement Estimates to Tributaries**

### **Introduction**

In 2014, the Chelan County Public Utility District (District) estimated sockeye escapement to tributaries based on mark-recapture methodology. The purpose of this document is to report the spawning escapement estimates for the Little Wenatchee and White River subbasins. This information is used to track and/or estimate viable salmonid population parameters (VSP): abundance, productivity, spatial structure, and diversity (McElhaney et al. 2000).

### **Methods**

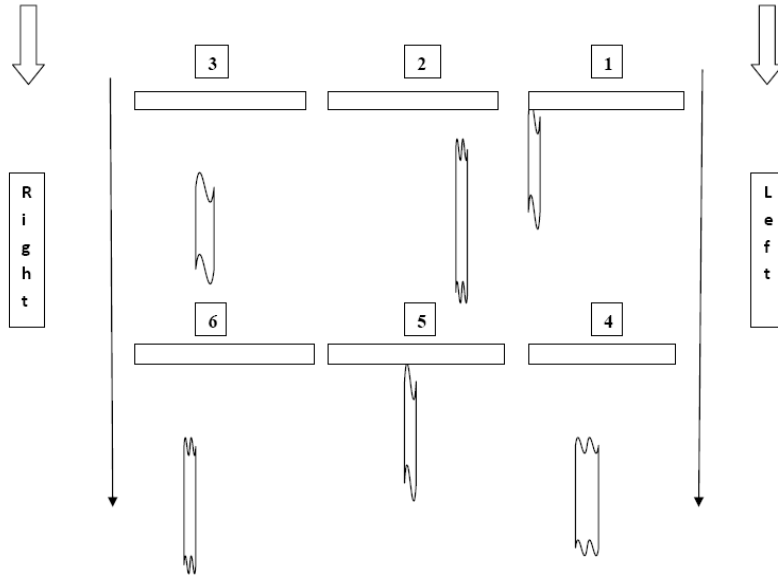
#### ***Mark-Recapture Method:***

Detection efficiencies of the in-stream arrays were calculated for the Little Wenatchee River and White River in 2014. The in-stream arrays include a series of upstream and downstream coils (Figure 1). Combined, these coils represented the upstream and downstream detection arrays, respectively. Overall detection efficiency  $P_{all}$  of the arrays was calculated based on observed detection probabilities of individual arrays:

$$P_{all} = 1 - (1 - P_{array\ 1})(1 - P_{array\ 2})$$

where the probability of missing a fish on both the upstream  $P_{array1}$  and downstream  $P_{array2}$  arrays were combined for an overall efficiency  $P_{all}$  (Connolly et al. 2008).

Adult sockeye salmon were tagged at adult fishways within the Columbia River and at Tumwater Dam. Additionally, adult returns that were PIT tagged as juveniles were used in the analyses. Total passage of adult sockeye salmon through Tumwater Dam was obtained from Columbia River Data Access in Real Time (DART 2014). Resulting tag files were queried in PTAGIS (2014), providing detection histories for each study fish.



**Figure 1.** Schematic of a PIT array configuration.

Resulting data from passage at Tumwater Dam, mark and recapture using PIT tags, and detection efficiency estimates can provide estimation of escapement to spawning tributaries. Assumptions include: (1) the study population is “closed,” i.e., no individuals die or emigrate between the initial mark and subsequent recaptures; (2) tags are not lost and detections are correctly identified; (3) all individuals have the same probability of being detected, and (4) the number of recapture events are proportional to the total population. Lastly, it was assumed that PIT-tagging efforts at Tumwater have negligible influence on fish behavior and tagged individuals behave similarly to untagged individuals. The resulting escapement rate, adjusted for detection efficiency, was then applied to the total population as such:

$$Escapement = \left( \frac{\left( \frac{Obs_{LWN}}{Eff_{LWN}} + \frac{Obs_{WTL}}{Eff_{WTL}} \right)}{PITs_{TUM}} \right) \times Counts_{TUM}$$

where the PIT tag detections (*Obs*) at the Little Wenatchee (*LWN*) and White River (*WTL*) were adjusted for detection efficiency (*Eff*), compared to the number released (*PITs*) at Tumwater Dam (*TUM*), and the resulting proportion was applied to the population observed (*Counts*) passing Tumwater Dam.

## Results

### *Sockeye Salmon Mark-Recapture Method*

Fishway enumeration at Tumwater Dam indicated that 99,898 adult sockeye salmon passed the facility during the 2014 migration, which was a sufficient return to open a recreational fishery in Lake Wenatchee for 2014. PIT tags were implanted in 1,407 of these fish at Tumwater and 322 of these fish were PIT-tagged prior to passing Tumwater; 76 fish were subsequently detected at the Little Wenatchee PIT tag array and 848 fish were subsequently detected at the White River PIT tag array (Table 1). Based on the recapture of PIT-tagged adult sockeye and assigned detection efficiency, total estimated escapement from Tumwater Dam to the Little Wenatchee River was 4,391 adult sockeye and 49,021 adult sockeye to the White River (Table 2).

**Table 1.** Number of adult sockeye salmon PIT-tagged, released, and detected upstream of Tumwater Dam in 2009 through 2014, and mark/recapture based tributary escapement estimates.

Year	Number of PIT-tagged adults detected or tagged at Tumwater <sup>1</sup>	White River <sup>2</sup>		L. Wenatchee River <sup>3</sup>		Chiwawa R.	Nason Creek
		Observed	Estimated	Observed	Estimated	Observed	Observed
2009	1,085	381	939	38	39	37	7
2010	1,164	571	635	67	67	3	1
2011	484	40	N/A <sup>4</sup>	84	0	0	0
2012	1,154	410	435	74	75	0	0
2013	719	152	N/A <sup>4</sup>	55	67	0	0
2014	1,729	848	848	76	76	0	3

<sup>1</sup> Also includes fish detected downstream of release point (fallbacks).

<sup>2</sup> Based on a detection efficiency  $p_{all} = 0.406$  in 2009 (assigned from 2010 data),  $p_{all} = 0.900$  in 2010,  $p_{all} = 0.943$  in 2012 and  $p_{all} = 0.999$  in 2014.

<sup>3</sup> Based on a detection efficiency  $p_{all} = 0.971$  in 2009,  $p_{all} = 1.000$  in 2010,  $p_{all} = 0.987$  in 2012  $p_{all} = 0.818$  in 2013 and  $p_{all} = 1.000$  in 2014.

<sup>4</sup> Technical difficulties with the White River PIT array prevented the calculation of detection efficiency and a mark-recapture based escapement estimate.

**Table 2.** Estimated escapement of adult sockeye salmon to Little Wenatchee and White rivers based on mark-recapture events, in-stream detection efficiency, and adult enumeration at Tumwater Dam, 2009-2014.

Year	Tumwater count	Recreational harvest	Little Wenatchee	White River	Combined	Escapement
2009	16,034	2,229	576	13,876	14,452	0.901
2010	35,821	4,129	2,062	19,542	21,604	0.603
2011 <sup>1</sup>	18,634	0	2,431	14,582	17,013	0.913
2012	66,520	12,107	4,607	23,866	28,473	0.428
2013 <sup>1</sup>	29,015	6,262	2,426	14,294	16,720	0.576
2014	99,898	16,255	4,319	49,021	53,412	0.535
<i>Average</i>	33,205	4,945	2,420	17,232	19,652	0.684

<sup>1</sup> Escapement was calculated using AUC counts for the Little Wenatchee River and a linear regression relationship to the Little Wenatchee River for the White River.



## References

- Hillborn, R. B.G. Bue, and S. Sharr. 1999. Estimating spawning escapements from periodic counts: a comparison of methods. *Can. J. Fish. Aquat. Sci.* 56: 888-896.
- Hyatt, K.D., M.M. Stockwell, H. Wright, K. Long, J. Tamblyn, and M. Walsh. 2006. Fish and Water Management Tool Project Assessments: Okanogan Adult Sockeye Salmon (*Oncorhynchus nerka*) Abundance and Biological Traits in 2005. DRAFT Report to file: JSID-SRe 3-05. Salmon and Freshwater Ecosystems Division, Fisheries and Oceans Canada, Nanaimo, B.C. V9T 6N7.
- McElhany, P., M. H. Ruckelshaus, M. J. Ford, T. C. Wainwright, and E. P. Bjorkstedt. 2000. Viable salmonid populations and the recovery of Evolutionarily Significant Units. NOAA Technical Memorandum.
- Mullan, J. W. 1987. Status and propagation of Chinook salmon in the mid-Columbia River through 1985. *U.S. Fish and Wildlife Serv. Biol. Rep.* 87(3) 111 pp.
- Murdoch, A. and C. Peven. 2005. Conceptual Approach to Monitoring and Evaluating the Chelan County Public Utility District Hatchery Programs. *Prepared for:* Chelan PUD Habitat Conservation Plan's Hatchery Committee. Chelan PUD, Wenatchee, WA.
- Perrin, C.J. and J.R. Irvine. 1990. A review of survey life estimates as they apply to the area-under-the-curve method for estimating the spawning escapement of Pacific salmon. *Canadian Tech. Rep. of Fisheries and Aquatic Sciences* No. 1733. Department of Fisheries and Oceans, Nanaimo, B.C. V9R 5K6.
- Peven, C. M. 1990. The life history of naturally produced steelhead trout from the Mid-Columbia River Basin. MS Thesis, University of Washington, Seattle.



# Appendix I

## **Genetic Diversity of Wenatchee Sockeye Salmon**



**Assessing the Genetic Diversity of Lake Wenatchee Sockeye Salmon  
And Evaluating The Effectiveness Of Its Supportive Hatchery  
Supplementation Program**

Developed for

Chelan County PUD

and the

Habitat Conservation Plan's Hatchery Committee

Developed by

Scott M. Blankenship, Cheryl A. Dean, Jennifer Von Bargaen

WDFW Molecular Genetics Laboratory

Olympia, WA

and

Andrew Murdoch

Supplementation Research Team

Wenatchee, WA

**March 2008**

<b>Executive Summary</b> .....	1
<b>Introduction</b>	
Lake Wenatchee Sockeye Salmon .....	3
Sockeye Artificial Propagation In Lake Wenatchee .....	5
Previous Genetic Analyses .....	6
Study objectives .....	7
<b>Methods</b>	
Tissue collection .....	9
Laboratory Analysis .....	9
Genetic Analysis	
Assessing within collection genetic diversity .....	10
Assessing among-collection genetic differentiation .....	10
Effective population size.....	11
<b>Results/Discussion</b> .....	12
<b>Conclusions</b> .....	13
<b>Acknowledgements</b> .....	14
<b>Literature Cited</b> .....	15
<b>Tables</b> .....	19

## Executive Summary

Nine spawning populations of sockeye (*Oncorhynchus nerka*) salmon have been identified in Washington, including stocks in the Lake Wenatchee basin (SaSI 5800) (Washington Department of Fisheries et al. 1993). Lake Wenatchee sockeye are classified as an Evolutionary Significant Unit (ESU), and consists of sockeye salmon that spawn primarily in tributaries above Lake Wenatchee (the White River, Napeequa River, and Little Wenatchee Rivers). Since 1990, the Wenatchee Sockeye Program has released juveniles into Lake Wenatchee to supplement natural production of sockeye salmon in the basin. The program's broodstock are predominantly natural-origin sockeye adults returning to the Wenatchee River captured at Tumwater Dam (Rkm 52.0), where a net-pen system is used to house both maturing adults and juveniles prior to release into Lake Wenatchee to over-winter.

Previous genetic studies have generally found a lack of concordance between population genetic relationships and their geographic distributions. These studies indicate that the nearest geographic neighbors of sockeye salmon populations are not necessarily the most genetically similar. Specifically for the Columbia River Basin, sockeye from Lake Wenatchee, Okanogan River, and Redfish Lake may be more closely related to a population from outside the Columbia River (depending on marker used) than to each other.

In this study we investigated the temporal and spatial genetic structure of Lake Wenatchee sockeye collections, without regard to sockeye populations outside of the Lake Wenatchee area. Our primary objective here was to determine if the Wenatchee Sockeye Program affected the natural Lake Wenatchee sockeye population. More specifically, we were tasked to determine if the genetic composition of Lake Wenatchee sockeye population had been altered by a supplementation program that was based on the artificial propagation of a small subset of that population. Using microsatellite DNA allele frequencies, we investigated population differentiation between temporally replicated collections of natural-origin Lake Wenatchee sockeye and program broodstock. We analyzed thirteen collections of Lake Wenatchee sockeye (Table 1), eight temporally replicated collections of natural-origin Lake Wenatchee sockeye (N=786) and five temporally replicated collections of Wenatchee Sockeye Program broodstock (N=248). Paired natural – broodstock collections were available from years 2000, 2001, 2004, 2006, and 2007.

### *Conclusions*

We observed that allele frequency distributions were consistent over time, irrespective of collection origin, resulting in small and statistically insignificant measures of genetic differentiation among collections. We interpreted these results to indicate no year-to-year differences in allele frequencies among natural-origin or broodstock collections. Furthermore, there were no observed difference between pre- and post-supplementation collections. Therefore, we accepted our null hypothesis that the allele frequencies of the broodstock collections equaled the allele frequencies of the natural collections, which

equaled the allele frequency of the donor population. Given the small differences in genetic composition among collections, the genetic model for estimating  $N_e$  produced estimates with extremely large variances, preventing the observation of any trend in  $N_e$ .



## Introduction

A report titled “Conceptual Approach to Monitoring and Evaluating the Chelan County Public Utility District Hatchery Programs” was prepared July 2005 by Andrew Murdoch and Chuck Peven for the Chelan PUD Habitat Conservation Plan’s Hatchery Committee. This report outlined 10 objectives to be applied to various species assessing the impact (positive or negative) of hatchery operations mitigating the operation of Rock Island Dam. This current study pertains only to Lake Wenatchee sockeye and objective 3:

*Determine if genetic diversity, population structure, and effective population size have changed in natural spawning populations as a result of the hatchery program. Additionally, determine if hatchery programs have caused changes in phenotypic characteristics of natural populations.*

In order to evaluate cause and effect of hatchery supplementation, WDFW Molecular Genetics Lab surveyed genetic variation of Lake Wenatchee sockeye. The conceptual approach for this project follows that of a parallel study regarding the Wenatchee River spring Chinook supplementation program (Blankenship et al. 2007). We determined the genetic diversity present in the Lake Wenatchee sockeye population by analyzing temporally replicated collections spanning 1989 – 2007, which included collections from before and following the inception of the Wenatchee Sockeye Program. Documenting the genetic composition of the Lake Wenatchee sockeye population is necessary to assess the effect of the hatchery program on the Lake Wenatchee population. In addition, this work provides a genetic baseline for future projects requiring genetic data. See study objectives below for specific details about how this project addresses Murdoch and Peven (2005) objective 3.

### *Lake Wenatchee Sockeye Salmon*

Nine spawning populations of sockeye (*Oncorhynchus nerka*) salmon have been identified in Washington (Washington Department of Fisheries et al. 1993): 1) Baker

River, 2) Ozette Lake, 3) Lake Pleasant, 4) Quinault Lake, and 5) Okanogan River (classified as native stock); 6) Cedar River (classified as non-native stock); 7) Lake Wenatchee, classified as mixed stock); 8) Lake Washington/Lake Sammamish tributaries; and 9) Lake Washington beach spawners (classified as unknown origin). Chapman et al. (1995) listed four additional spawning aggregations of sockeye salmon that appear consistently in Columbia River tributaries: the Methow, Entiat, and Similkameen Rivers; and Icicle Creek in the Wenatchee River drainage.

Located in north central Washington, the Wenatchee River basin drains a portion of the eastern slope of the Cascade Mountains, including high mountainous regions of the Cascade crest. The headwater area of the Wenatchee River is Lake Wenatchee, a typical low productivity oligotrophic or ultra-oligotrophic sockeye salmon nursery lake (Allen and Meekin 1980, Mullan 1986, Chapman et al. 1995). Sockeye salmon bound for Lake Wenatchee enter the Columbia River in April and May and arrive at Lake Wenatchee in late July to early August (Chapman et al. 1995; Washington Department of Fisheries et al. 1993). The run timing of Lake Wenatchee sockeye salmon, classified as an Evolutionary Significant Unit (ESU), appears to have become earlier by 6 - 30 days during the past 70 years (Chapman et al. 1995; Quinn and Adams 1996). Additionally, scale pattern analysis suggests Wenatchee sockeye migrate past Bonneville Dam earlier than the sockeye bound for the Okanogan River (Fryer and Schwartzberg 1994). The Wenatchee population spawns from mid-September through October in the Little Wenatchee, White, and Napeequa Rivers above Lake Wenatchee (Washington Department of Fisheries et al. 1993), peaking in late September (Chapman et al. 1995). Limited beach spawning is believed to occur in Lake Wenatchee (L. Lavoy pers. com.; Mullan 1986), although Gangmark and Fulton (1952) reported two lakeshore seepage areas in Lake Wenatchee that were used by spawning sockeye salmon. Sockeye salmon fry enter Lake Wenatchee between March and May (Dawson et al. 1973), and typically rear in the lake for one year before leaving as smolts (Gustafson et al. 1997; Peven 1987).

Both the physical properties of the habitat and ecological/biological factors of the sockeye populations differ between the Lake Wenatchee ESU and the geographically

proximate Okanogan ESU. For example: 1) Different limnology is encountered by sockeye salmon in Lakes Wenatchee and Osoyoos; 2) Lake Wenatchee sockeye predominantly return at ages four and five (a near absence of 3-year-olds), where a large percentage of 3-year-olds return to the Okanogan population; and 3) the apparent one month separation in juvenile outmigration-timing between Okanogan- and Wenatchee-origin fish (Gustafson et al. 1997 and references therein).

#### *Sockeye Artificial Propagation In Lake Wenatchee*

The construction of Grand Coulee Dam completely blocked fish passage to the upper Columbia River, and 85% of sockeye salmon passing Rock Island Dam between 1935 and 1936 were estimated to be from natural stocks bound for areas up-river to Grand Coulee Dam (Mullan 1986; Washington Department of Fisheries et al. 1938). To compensate for loss of habitat resulting from Grand Coulee Dam, the federal government initiated the Grand Coulee Fish-Maintenance Project (GCFMP) in 1939 to maintain fish runs in the Columbia River above Rock Island Dam. Between 1939 and 1943, all sockeye salmon entering the mid-Columbia River were trapped at Rock Island Dam, and over 32,000 mixed Lake Wenatchee, Okanogan River, and Arrow Lake adult sockeye salmon were released into Lake Wenatchee (Gustafson et al. 1997 Appendix Table D-2). In addition to adult relocation, between 1941 and 1969 over 52.8 million fry descended from original spawners collected at Rock Island and Bonneville Dams, were released into Lake Wenatchee (Gustafson et al. 1997 Appendix Table D-2).

No releases of artificially-reared sockeye salmon occurred in the Wenatchee watershed during the years 1970 to 1989 (Gustafson et al. 1997 Appendix Table D-2). Since 1990, the Wenatchee Sockeye Program has released juveniles into Lake Wenatchee to supplement natural production of sockeye salmon in the basin. Sockeye adults returning to the Wenatchee River are captured at Tumwater Dam (Rkm 52.0) and transferred to Lake Wenatchee net pens until mature. The Wenatchee Sockeye Program goals are 260 adults with an equal sex ratio, <10% hatchery-origin returns (identified by coded wire tags), and the adults removed for broodstock account for <10% of the run size. Fish are spawned at Lake Wenatchee and their gametes are taken to Rock Island Fish Hatchery

Complex (i.e., Eastbank) for fertilization and incubation. Fry are returned to the Lake Wenatchee net -pens after they are large enough to be coded wire tagged, and are housed in the pens until fall (one year after spawning), when they are liberated into the lake to over-winter. For brood years 1991 – 2004 an average of 218,683 (std. dev. = 71,090) pen-reared Lake Wenatchee-origin juvenile sockeye salmon have been released yearly into Lake Wenatchee.

#### *Previous Genetic Studies*

**Protein (allozyme) variation** – Surveying genetic variation at 12 allozyme loci, Utter et al. (1984) reported moderate population structure among 16 sockeye collections from southeast Alaska through the Columbia River Basin, including Okanogan and Wenatchee stocks, with an apparent genetic association between upper Fraser River and Columbia River sockeye salmon. Winans et al. (1996) surveyed variation at 55 allozyme loci for 25 sockeye salmon and two kokanee collections from 21 sites in Washington, Idaho, and British Columbia, and reported the lowest level of allozyme variability of any species of Pacific salmon and a highest level of inter-population differentiation. Furthermore, these authors reported that there was no clear relationship between geographic and genetic differentiation among the populations within their study. Other studies corroborate the results of Winans et al. (1996), finding a lack of discernible geographic patterning for sockeye salmon populations in British Columbia, Alaska, and Kamchatka (Varnavskaya et al. 1994, Wood et al. 1994, Wood 1995). These studies indicate that the nearest geographic neighbors of sockeye salmon populations are not necessarily the most genetically similar, which contrasts with the other Pacific salmon species that exhibit concordance between geographic and genetic differentiation (Utter et al. 1989, Winans et al. 1994, Shaklee et al. 1991). As part of the comprehensive status review of west coast sockeye salmon (Gustafson et al. 1997), NMFS biologists collected new allozyme genetic information for 17 sockeye salmon populations and one kokanee population in Washington and combined these data for analysis with the existing Pacific Northwest sockeye salmon and kokanee data from Winans et al. (1996). Results of the updated study were consistent with Winans et al. (1996), with no clear concordance between geographic and genetic distances. Sockeye salmon from Lake Wenatchee, Redfish Lake,

Ozette Lake, and Lake Pleasant are very distinct from other collections in the study, and Columbia River populations were not necessarily most closely related to each other. Gustafson et al. (1997) also examined between-year variability within a collection location and found low levels of statistical significance among the five Lake Wenatchee collections included in the study (For 10 pair-wise comparisons using sum-G test, five were statistically significant). Lake Wenatchee brood year 1987 accounted for three of the significant comparisons, which were driven by unusually high frequencies of two allozyme alleles (ALAT\*95 and ALAT\*108) (Winans et al. 1996). Nevertheless, Gustafson et al. (1997) conclude that, in general, temporal variation at a locale was considerably less than between-locale variation.

**Nucleic acid variation** - Beacham et al. (1995) reported levels of variation in nuclear DNA of *O. nerka* using minisatellite probes. They analyzed 10 collections, including a sample from Lake Wenatchee. Cluster analysis showed the Lake Wenatchee sample was different from all the other collections, including those from the Columbia River. Using a similar molecular technique, Thorgaard et al. (1995) examined the use of multi-locus DNA fingerprinting (i.e., banding patterns) to discriminate among 14 sockeye salmon and kokanee populations. Dendrograms based on analysis of banding patterns produced different genetic affinity groups depending on the probes used. While none of the five DNA probes showed a close relationship between Lake Wenatchee and Okanogan River sockeye salmon, if information from all probes were combined, *O. nerka* from Redfish Lake, Wenatchee, and Okanogan were separate from kokanee of Oregon and Idaho and a sockeye salmon sample from the mid-Fraser River.

### *Study Objective*

We documented temporal variation in genetic diversity (i.e., heterozygosity and allelic diversity), and investigated population differentiation between temporally replicated collections of natural-origin Lake Wenatchee sockeye and program broodstock, using microsatellite DNA allele frequencies. Temporally replicated collections from the same location can also be used to estimate effective population size ( $N_e$ ). If populations are “ideal”, the census size of a population is equal to the “genetic size” of the population.

Yet, numerous factors lower the “genetic size” below census, such as, non-equal sex ratios, changes in population size, and variance in the numbers of offspring produced from parent pairs.  $N_e$  is thought to be between 0.10 and 0.33 of the estimated census size (Bartley et al. 1992; RS Waples pers. comm.), although numerous observations differ from this general rule.  $N_e$  can be calculated directly from demographic data, or inferred from observed differences in genetic variance over time. Essentially, when calculated from genetic data,  $N_e$  is the estimated size of an “ideal” population that accounts for the genetic diversity changes observed, irrespective of abundance.

We will address the hypotheses associated with Objective 3 in Murdoch and Peven (2005) using the following four specific tasks:

**Task 1** - Document the observed genetic diversity.

**Task 2** - Test for population differentiation among Lake Wenatchee collections and the associated supplementation program.

Task 2 was designed to address two hypotheses listed as part of Objective 3 in Murdoch and Peven (2005):

- Ho: Allele frequency<sub>Hatchery</sub> = Allele frequency<sub>Naturally produced</sub> = Allele frequency<sub>Donor pop.</sub>
- Ho: Genetic distance between subpopulations<sub>Year x</sub> = Genetic distance between subpopulations<sub>Year y</sub>

Murdoch and Peven (2005) proposed these two hypotheses to help evaluate supplementation programs through a “Conceptual Process” (Figure 5 in Murdoch and Peven 2005). There are two components to the first hypothesis, which must be considered separately for Lake Wenatchee sockeye. The first component involves comparisons between natural-origin populations from Lake Wenatchee to determine if there have been changes in allele frequencies through time starting with the donor population. Documenting a change does not necessarily indicate that the supplementation program has directly affected the natural-origin fish, as additional tests would be necessary to support that hypothesis. The intent of the second component is to determine if the hatchery produced populations have the same genetic composition as the naturally produced populations.

**Task 3** - Calculate  $N_e$  using the temporal method for multiple samples from the same location to document trend.

**Task 4** - Compare  $N_e$  estimates with trend in census size for Lake Wenatchee sockeye.

## Methods and Materials

### *Sampling*

Thirteen collections of Lake Wenatchee sockeye were analyzed, eight temporally replicated collections of natural Lake Wenatchee sockeye (N=786) and five temporally replicated collections of Wenatchee Sockeye Program broodstock (N=248) (Table 1). Paired natural – broodstock collections were available from years 2000, 2001, 2004, 2006, and 2007 (Table 1). All collections were made at Tumwater Dam on the Wenatchee River. Note that collections classified as broodstock were predominantly natural-origin sockeye. A majority of the genetic samples were from dried scales. The tissue collections from 2006 and 2007 were fin clips stored immediately in ethanol after collection. DNA was extracted from stored tissue using Nucleospin 96 Tissue following the manufacturer's standard protocol (Macherey-Nagel, Easton, PA, U.S.A.).

### *Laboratory Analysis*

Polymerase chain reaction (PCR) amplification was performed using 17 fluorescently end-labeled microsatellite marker loci, *One* 2 (Scribner et al 1996) *One* 100, 101, 102, 105, 108, 110, 114, and 115 (Olsen et al. 2000), *Omm* 1130, 1135, 1139, 1142, 1070, and 1085 (Rexroad et al. 2001), *Ots* 3M (Banks et al. 1999) and *Ots* 103 (Small et al. 1998). PCR reaction volumes were 10  $\mu$ L, with the reaction variables being 2  $\mu$ L 5x PCR buffer (Promega), 0.6  $\mu$ L  $MgCl_2$  (1.5 mM) (Promega), 0.2  $\mu$ L 10 mM dNTP mix (Promega), and 0.1  $\mu$ L *Go Taq* DNA polymerase (Promega). Loci were amplified as part of multiplexed sets, so primer molarities and annealing temperatures varied. Multiplex one had an annealing temperature of 55°C, and used 0.09 Molar (M) *One* 108, 0.06 M *One* 110, and 0.11 M *One* 100. Multiplex two had an annealing temperature of 53°C, and used 0.08 M *One* 102, 0.1 M *One* 114, and 0.05 M *One* 115. Multiplex three had an annealing temperature of 55°C, and used 0.08 M *One* 105 and 0.07 M *Ots* 103. Multiplex four had

an annealing temperature of 53°C, and used 0.09 M *Omm* 1135 and 0.08 M *Omm* 1139. Multiplex five had an annealing temperature of 60°C, and used 0.2 M *Omm* 1085, 0.09 M *Omm* 1070, and 0.05 M *Ots* 3M. Multiplex six had an annealing temperature of 48°C, and used 0.06 M *One* 2, 0.08 M *Omm* 1142, and 0.08 M *Omm* 1130. *One* 101 was run in isolation with a primer molarity of 0.06. Thermal cycling was conducted on either PTC200 (MJ Research) or GeneAmp 9700 thermal cyclers as follows: 94°C (2 min); 30 cycles of 94°C for 15 sec., 30 sec. annealing, and 72°C for 1 min.; a final 72°C extension and then a 10°C hold. PCR products were visualized by denaturing polyacrylamide gel electrophoresis on an ABI 3730 automated capillary analyzer (Applied Biosystems). Fragment analysis was completed using GeneMapper 3.7 (Applied Biosystems).

#### *Genetic data analysis*

**Assessing within collection genetic diversity** - Heterozygosity measurements were reported using Nei's (1987) unbiased gene diversity formula (i.e., expected heterozygosity) and Hedrick's (1983) formula for observed heterozygosity. Both tests were implemented using the microsatellite toolkit (Park 2001). For each locus and collection FSTAT version 2.9.3.2 (Goudet 1995) was used to assess Hardy-Weinberg equilibrium, where deviations from the neutral expectation of random associations among alleles were calculated using a randomization procedure. Alleles were randomized among individuals within collections (4160 randomizations for this dataset) and the  $F_{IS}$  (Weir and Cockerham 1984) calculated for the randomized datasets were compared to the observed  $F_{IS}$  to obtain an unbiased estimation of the probability that the null hypothesis was true. The 5% nominal level of statistical significance was adjusted for multiple tests (Rice 1989). Genotypic linkage disequilibrium was calculated following Weir (1979) using GENETIX version 4.05 (Belkhir et al. 1996). Statistical significance of linkage disequilibrium results was assessed using a permutation procedure implemented in GENETIX for each locus by locus combination within each collection.

**Assessing among collection genetic differentiation** - The temporal stability of allele frequencies was assessed by the randomization chi-square test implemented in FSTAT version 2.9.3.2 (Goudet 1995). Multi-locus genotypes were randomized between



collections. The G-statistic for observed data was compared to G-statistic distributions from randomized datasets (i.e., null distribution of no differentiation between collections). Population differentiation was also investigated using pairwise estimates of  $F_{ST}$ . Multi-locus estimates of pairwise  $F_{ST}$ , estimated by a “weighted” analysis of variance (Weir and Cockerham, 1984), were calculated using GENETIX version 4.05 (Belkhir et al. 1996).  $F_{ST}$  was used to quantify population structure, the deviation from statistical expectations (i.e., excess homozygosity) due to non-random mating between populations. To determine if the observed  $F_{ST}$  estimate was consistent with statistically expectations of no population structure, a permutation test was implemented in GENETIX (1000 permutations).

**Effective population size ( $N_e$ )** – Estimates of the effective population size were obtained using a multi-collection temporal method (Waples 1990a). The temporal method assumes that cohorts are used, but we did not decompose the collection year samples into their respective cohorts using age data. Therefore,  $N_e$  estimates that pertain to individual year classes of breeders are not valid; however the harmonic mean over all samples will estimate an  $N_e$  that pertains to the time period from which the collections are derived. Comparing samples from years  $i$  and  $j$ , Waples’ (1990a) temporal method estimates the effective number of breeders ( $\hat{N}_{b(i,j)}$ ) according to:

$$\hat{N}_{b(i,j)} = \frac{b}{2(\hat{F} - 1/\tilde{S}_{i,j})}$$

The standardized variance in allele frequency ( $\hat{F}$ ) is calculated according to Pollack (1983). The parameter  $b$  is calculated analytically from age structure information and the number of years between samples (Tajima 1992). The age-at-maturity information required to calculate  $b$  was obtained from ecological data (Hillman et al. 2007). The harmonic mean of sample sizes from years  $i$  and  $j$  is  $\tilde{S}_{i,j}$ . The harmonic mean over all pairwise estimates of  $\hat{N}_{b(i,j)}$  is  $\tilde{N}_b$ . SALMONNb (Waples et al. 2007) was used to calculate  $\tilde{N}_b$ .

## Results and Discussion

In this section we combine our presentation and interpretations of the genetic analyses. Additionally, this section is organized based on the task list presented in the study plan.

**Task 1** - Document the observed genetic diversity.

Substantial genetic diversity was observed over all Lake Wenatchee sockeye collections analyzed (Table 1), with heterozygosity estimates over all loci having a mean of 0.79. Genetic diversity was consistent with expected Hardy-Weinberg random mating genotypic proportions for all collections. The  $F_{IS}$  observed for each collection was not statistically significant given the distribution of  $F_{IS}$  generated using a randomization procedure. Additionally, there were no statistically significant associations observed between alleles across loci (i.e., linkage equilibrium) (data not shown). We concluded from these results that the genetic data from each collection was consistent with statistical expectations for random association of alleles within and between loci. In other words, each collection represents samples from a single gene pool (i.e., populations), and the genetic diversity observed has no detectable technical artifacts or evidence of natural selection.

**Task 2** - Test for differentiation among Lake Wenatchee collections and the associated supplementation program.

We explicitly tested the hypothesis of no significant differentiation within natural-origin or broodstock collections from Lake Wenatchee using a randomization chi-square test. The null hypothesis for these tests was that the allele frequencies from two different populations were drawn from the same underlying distribution. We show the results for the pairwise comparisons among eight temporally replicated natural-origin collections from Lake Wenatchee (28 pairwise tests), and report all tests were non-significant (Table 2A). Similarly, for five temporally replicated broodstock collections, 10 of 10 pairwise tests were non-significant (Table 2B). We also tested if natural-origin and broodstock

collections were differentiated from each other over time, and report that 40 of 40 tests were non-significant (Table 2C). The nominal level of statistical significance ( $\alpha = 0.05$ ) was adjusted for multiple comparisons using strict Bonferroni correction (Rice 1989). Yet, there are perhaps slight differences between paired natural-broodstock collections. Note that the p-values for comparisons regarding 2006 and 2007 paired collections are lower than for comparisons regarding 2000, 2001, and 2004. The small sample sizes for broodstock collections in 2006 and 2007 may not have been random samples from the Lake Wenatchee sockeye population.

Given the consistencies observed for allele frequency distributions over time, metrics of population structure were expected to be small. This was the case, as the estimated  $F_{ST}$  over all thirteen collections was 0.0003. This observed value fell within the distribution of  $F_{ST}$  values expected if there were no population structure present (permutation test p-value 0.12). Analysis of the paired natural-broodstock collections corroborated this result. Pairwise estimates of  $F_{ST}$  were 0.000 for years 2000, 2001, 2004, and 2007, and 0.002 for 2006. All five estimates were non-significant. Essentially, all 13 sockeye collections could be considered samples from the same population. Given these results, it is valid to combine all collections for statistical analysis. Therefore, we did not calculate genetic distances among any collections, as it is inappropriate to estimate distances that are effectively zero.

### *Conclusions*

We interpret these data to indicate that there appears to be no significant year-to-year differences in allele frequencies among natural-origin or broodstock collections, nor are there observed differences between collections pre- and post-supplementation. As a result, we accept the null hypothesis that the allele frequencies of the broodstock collections equal the allele frequencies of the natural collections, which equals the allele frequency of the donor population. Furthermore, the observed genetic variance that can be attributed to among collection differences was negligible.

**Task 3** - Calculate  $N_e$  using the temporal method for multiple samples from the same location to document trend.

The fundamental parameter for inferring  $N_e$  using genetic data is the standardized variance in allele frequency ( $\hat{F}$ ) (Pollack 1983). Methods estimate  $N_e$  from observed changes in  $\hat{F}$  over temporally replicated collections from the same location. Yet, as previously shown, there were no statistically significant differences detected in allele frequencies. The underlying model for estimating  $N_e$  produced estimates with extremely large variances, given small temporal differences in  $\hat{F}$ , which rendered any trend in  $N_e$  unobservable. Table 3 shows  $N_e$  estimates calculated using temporally replicated natural collections.

**Task 4** - Compare  $N_e$  estimates with trend in census size for Lake Wenatchee sockeye.

See Task 3

## **Acknowledgements**

We would like to thank Jeff Fryer (CRITFC) for providing critical collections of natural-origin sockeye from Lake Wenatchee. We would like to thank Norm Switzler for collection curation and Ken Warheit and Denise Hawkins for helpful comments regarding this project. This project was funded by Chelan County PUD and the Washington State General Fund.

## Literature Cited

- Allen RL and Meekin TK (1980) Columbia River sockeye salmon study, 1971-1974. Wash. Dep. Fish. Prog. Rep. 120, 75 p.
- Banks MA, Blouin MS, Baldwin BA, Rashbrook VK, Fitzgerald HA, Blankenship SM, Hedgecock D (1999) Isolation and inheritance of novel microsatellites in chinook salmon (*Oncorhynchus tshawytscha*). *Journal of Heredity*, 90:281-288.
- Bartley D, Bentley B, Brodziak J, Gomulkiewicz R, Mangel M, and Gall GAE (1992) Geographic variation in population genetic structure of chinook salmon from California and Oregon. *Fish. Bull.*, U.S. 90:77-100.
- Blankenship SM, Von Bargaen J, Warheit KI, and Murdoch AR (2007) Assessing the Genetic Diversity of Natural Chiwawa River Spring Chinook Salmon and Evaluating the Effectiveness of its Supportive Hatchery Supplementation Program. WDFW report to Chelan County PUD, March 2007.
- Belkhir K, Borsa P, Chikhi L et al (1996) GENETIX, logiciel sous Windows TM pour la Génétique des populations. Laboratoire Génome, Populations, Interactions, CNRS UMR 5000, Université de Montpellier II, Montpellier (France).
- Chapman D, Peven C, Giorgi A, Hillman T, and Utter F (1995) Status of spring chinook salmon in the mid-Columbia River. Don Chapman Consultants, Inc., 477 p. (Available from Don Chapman Consultants, 3653 Rickenbacker, Ste. 200, Boise, ID 83705.)
- Crawford BA (1979) The origin and history of trout brood stocks of the Washington Department of Game. Wash. State Game Dep., Fish. Res. Rep. 76 p.
- Dawson JJ, Thorne RE, and Traynor JJ (1973) Acoustic surveys of Lake Wenatchee and Lake Osoyoos in 1973. Final Report, Service Contr. 526, to Wash. Dep. Fish., by Fish. Res. Inst., Coll. of Fish., Univ. Washington, Seattle, WA, 18 p.
- Fryer JK and Schwartzberg M (1994) Identification of Columbia Basin sockeye salmon stocks using scale pattern analyses in 1993. Columbia River Inter-Tribal Fish Commission, Tech. Rep. 94-2, 39 p.
- Gangmark HA and Fulton LA (1952) Status of Columbia blueback salmon runs, 1951. U. S. Fish Wildl. Serv. Spec. Sci. Rep. 74, 29 p.
- Goudet J (1995) FSTAT (Version 1.2): A computer program to calculate F-statistics. *Journal of Heredity* 86: 485-486.

- Gustafson RG, Wainwright TC, Winans GA, Waknitz FW, Parker LT, and Waples RS (1997) Status review of sockeye salmon from Washington and Oregon. U.S. Dept. Commer., NOAA Tech. Memo. NMFS-NWFSC-33, 282 p.
- Hedrick PW (1983) Genetics of Populations, Science Books International, Boston
- Hillman T, Miller M, Peven C, Tonseth M, Miller T, Truscott K, and Murdoch A (2007) Monitoring and Evaluation of the Chelan County PUD Hatchery Programs: 2007 Annual Report.
- Knutzen D (1995) Letter to R. Gustafson, NMFS, from D. Knutzen, WDFW, re. Historical kokanee planting records for Lake Wenatchee, Lake Pleasant, Lake Ozette, Lake Shannon, and Baker Lake from 1981-1994, dated 17 July 1995. 1 p. plus attachment. (Available from West Coast Sockeye Salmon Administrative Record, Environmental and Technical Services Division, Natl. Mar. Fish. Serv., 525 N. E. Oregon Street, Portland, OR 97232.)
- Mullan JW (1986) Determinants of sockeye salmon abundance in the Columbia River, 1880's-1982: A review and synthesis. U.S. Fish Wildl. Serv. Biol. Rep. 86(12), 135 p.
- Murdoch AR and Peven C (2005) Conceptual Approach to Monitoring and Evaluating the Chelan County Public Utility District Hatchery Programs, Final Report.
- Nei M (1978) Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics* 89:583-590.
- Olsen JB, Wilson SL, Kretschmer EJ, Jones KC, Seeb JE (2000) Characterization of 14 tetranucleotide microsatellite loci derived from sockeye salmon. *Molecular Ecology* 9, 2185-2187.
- Park SDE (2001) Trypanotolerance in West African Cattle and the Population Genetic Effects of Selection [ Ph.D. thesis], University of Dublin
- Peven CM (1987) Downstream migration timing of two stocks of sockeye salmon on the Mid-Columbia River. *Northwest Sci.* 61(3):186-190.
- Pollak E (1983) A new method for estimating the effective population size from allele frequency changes. *Genetics* 104, 531-548.
- Quinn TP and Adams DJ (1996) Environmental changes affecting the migratory timing of American shad and sockeye salmon. *Ecology* 77:1151-1162.

- Rexroad CE, Coleman RL, Martin AM, Hershberger WK, Killefer J (2001) Thirty-five polymorphic microsatellite markers for rainbow trout (*Oncorhynchus mykiss*). *Animal Genetics*, 32:317-319.
- Rice WR, (1989) Analyzing tables of statistical tests. *Evolution*. 43:223-225.
- Scribner KT, Gust JR, Fields RL (1996) Isolation and characterization of novel salmon microsatellite loci: cross-species amplification and population genetic applications. *Canadian Journal of Fisheries and Aquatic Sciences*. 53, 833-841.
- Shaklee JB, Klaybor DC, Young S, and White BA (1991) Genetic stock structure of odd-year pink salmon, *Oncorhynchus gorbuscha* (Walbaum), from Washington and British Columbia and potential mixed-stock fisheries applications. *J. Fish. Biol.* 39(A):21-34.
- Small MP, Beacham TD, Withler RE, and Nelson RJ (1998) Discriminating coho salmon (*Oncorhynchus kisutch*) populations within the Fraser River, British Columbia. *Molecular Ecology* 7: 141-155.
- Tajima F (1992) Statistical Method for Estimating the Effective Population Size in Pacific Salmon. *J Hered* 83, 309-311.
- Utter F, Aebersold P, Helle J, and Winans G (1984) Genetic characterization of populations in the southeastern range of sockeye salmon. In J. M. Walton and D. B. Houston (editors), *Proceedings of the Olympic wild fish conference*, p. 17-31. Fisheries Technology Program, Peninsula College, Port Angeles, WA.
- Utter F, Milner G, Stahl G, and Teel D (1989) Genetic population structure of chinook salmon, *Oncorhynchus tshawytscha*, in the Pacific Northwest. *Fish. Bull., U.S.* 87:239-264.
- Varnavskaya NV, Wood CC, and Everett RJ (1994) Genetic variation in sockeye salmon (*Oncorhynchus nerka*) populations of Asia and North America. *Can. J. Fish. Aquat. Sci.* 51(Suppl. 1):132-146.
- Waples RS (1990a) Conservation genetics of Pacific salmon. III. Estimating effective population size. *Journal of Heredity* 81:277-289
- Waples RS, Masuda M, Pella J (2007) SALMONNb: a program for computing cohort-specific effective population sizes ( $N_b$ ) in Pacific salmon and other semelparous species using the temporal method. *Molecular Ecology Notes* 7, 21-24.
- Washington Department of Fisheries (WDF), Washington Department of Game (WDG), and United States Bureau of Fisheries (USBF) (1938) A report on the preliminary investigations into the possible methods of preserving the Columbia River salmon and steelhead at the Grand Coulee Dam. Wash. Dep. Fish, Olympia, WA, 120 p.

- Washington Department of Fisheries (WDF), Washington Department of Wildlife (WDW), and Western Washington Treaty Indian Tribes (WWTIT) (1993) 1992 Washington State salmon and steelhead stock inventory (SASSI). Wash. Dep. Fish Wildl., Olympia, WA, 212 p. plus 5 regional volumes.
- Washington Department of Fish and Wildlife (WDFW) (1996) Letter to M. Schiewe, NMFS, from R. Lincoln, Assistant Director, Fish Management Program, Washington Department of Fish and Wildlife, dated 12 July 1996. 3 p. plus appendix. (Available from West Coast Sockeye Salmon Administrative Record, Environmental and Technical Services Division, Natl. Mar. Fish. Serv., 525 N. E. Oregon Street, Portland, OR 97232.)
- Weir BS (1979) Inferences about linkage disequilibrium. *Biometrics* 35:235-254.
- Weir BS, Cockerham CC (1984) Estimating F-Statistics for the Analysis of Population-Structure. *Evolution* 38:1358-1370.
- Winans GA, Aebersold PB, Urawa S, and Varnavskaya NV (1994) Determining continent of origin of chum salmon (*Oncorhynchus keta*) using genetic stock identification techniques: status of allozyme baseline in Asia. *Can. J. Fish. Aquat. Sci.* 51 (Suppl. 1):95-113.
- Winans GA, Aebersold PB, and Waples RS (1996) Allozyme variability of *Oncorhynchus nerka* in the Pacific Northwest, with special consideration to populations of Redfish Lake, Idaho. *Trans. Am. Fish. Soc.* 205:645-663.
- Wood CC, Riddell BE, Rutherford DT, and Withler RE (1994) Biochemical genetic survey of sockeye salmon (*Oncorhynchus nerka*) in Canada. *Can. J. Fish. Aquat. Sci.* 51(Suppl. 1):114-131.
- Wood CC (1995) Life history variation and population structure in sockeye salmon. *In* J. L. Nielsen (editor), *Evolution and the aquatic ecosystem: defining unique units in population conservation*. *Am. Fish. Soc. Symp.* 17:195-216.



**Table 1** Lake Wenatchee sockeye collections analyzed. MNA is the mean number of alleles per locus, Hz is unbiased heterozygosity, Obs Hz is observed heterozygosity, and HW is the p-value of the null hypothesis of random association of alleles (i.e., Hardy – Weinberg equilibrium). For reference, the nominal level of statistical significance at  $\alpha = 0.05$  is 0.0002 after correction for multiple tests.

Year	Collection Code	Tissue Type	Source	N	MNA	Hz	Obs Hz	HW
1989	89 <sup>1</sup>	Scales	Natural	96	14.35	0.792	0.791	0.424
1990	90 <sup>1</sup>	Scales	Natural	96	13.19	0.793	0.779	0.131
2000	00AAE	Scales	Broodstock	96	12.31	0.787	0.776	0.213
2000	00 <sup>1</sup>	Scales	Natural	96	11.76	0.801	0.826	0.868
2001	01AAS	Scales	Broodstock	53	9.47	0.788	0.793	0.392
2001	01 <sup>1</sup>	Scales	Natural	96	14.35	0.786	0.794	0.456
2002	02 <sup>1</sup>	Scales	Natural	96	14.53	0.794	0.777	0.780
2004	04 <sup>1</sup>	Scales	Natural	96	14.65	0.798	0.803	0.704
2004	04AAV	Scales	Broodstock	43	14.35	0.796	0.795	0.051
2006	06CN	Tissue	Broodstock	38	14.59	0.793	0.785	0.688
2006	06CO	Tissue	Natural	96	14.53	0.806	0.803	0.408
2007	07EE	Tissue	Broodstock	18	14.00	0.790	0.790	0.221
2007	07EF	Tissue	Natural	96	14.35	0.789	0.800	0.347

<sup>1</sup> Samples taken from scale cards provided by Jeff Fryer (CRITFC)

**Table 2** Allelic differentiation for Lake Wenatchee sockeye collections. A single analysis tested (pairwise) the allelic differentiation between all thirteen collections; however p-values for G-statistics are partitioned in the table by A) natural-origin, B) broodstock, and C) natural versus broodstock. Underlined values are for paired natural-broodstock collections from the same year. For reference, the nominal level of statistical significance at  $\alpha = 0.05$  is 0.0006 after correction for multiple tests. No significant values were observed.

---

A) Natural-Origin Collections

	89	90	00	01	02	04	06CO	07EF
89		0.257	0.359	0.531	0.331	0.127	0.031	0.263
90			0.953	0.148	0.753	0.903	0.077	0.283
00				0.328	0.527	0.607	0.604	0.400
01					0.209	0.081	0.127	0.093
02						0.085	0.707	0.235
04							0.312	0.577
06CO								0.435
07EF								

---

B) Broodstock Collections

	00AAE	01AAS	04AAV	06CN	07EE
00AAE		0.189	0.090	0.008	0.058
01AAS			0.122	0.020	0.116
04AAV				0.008	0.031
06CN					0.326
07EE					

---

C) Natural vs. Broodstock

	89	90	00	01	02	04	06CO	07EF
00AAE	0.027	0.309	<u>0.572</u>	0.018	0.041	0.012	0.093	0.040
01AAS	0.115	0.471	<u>0.160</u>	<u>0.219</u>	0.519	0.049	0.654	0.133
04AAV	0.136	0.219	0.210	0.423	0.208	<u>0.328</u>	0.037	0.153
06CN	0.029	0.004	0.053	0.007	0.022	<u>0.004</u>	<u>0.019</u>	0.001
07EE	0.099	0.229	0.053	0.015	0.093	0.178	0.090	<u>0.037</u>

---

**Table 3** Estimation of  $N_e$  for temporally replicated natural-original sockeye collections. Above the diagonal are pairwise estimates of  $N_e$ , where negative values mean sampling variance can account for genetic variance observed (i.e., genetic drift unnecessary). Below the diagonal are variances for pairwise estimates of  $N_e$ . Absent variance values (denoted by -) were too large for SalmonNb to display.

---

Collection	89	90	00	01	02	04	06CO	07EF
89		-3936.6	-1414	-2636.3	671.4	1871.1	1066.1	1951.2
90	2.59E+09		-1490.3	3649.1	-31144	-6808.4	817.6	93190.2
00	1.40E+09	4.45E+09		-592.2	-6842.2	-667.1	-1736.9	-1350.1
01	1.21E+09	1.47E+09	2.33E+09		977.1	6160.4	387.8	2531.5
02	1.91E+09	1.33E+09	1.16E+09	2.29E+09		1495.6	-848.5	3213.6
04	2.21E+09	3.62E+09	4.08E+09	1.27E+09	1.14E+09		896.6	2155.3
06CO	1.34E+09	1.39E+09	1.73E+09	-	4.51E+09	1.2E+09		3278.6
07EF	2.15E+09	1.51E+09	1.18E+09	1.68E+09	-	1.36E+09	2.65E+09	

---



# Appendix J

## **Genetic Diversity of Chiwawa River Spring Chinook Salmon**



**Assessing the Genetic Diversity of Natural Chiwawa River Spring  
Chinook Salmon and Evaluating the Effectiveness of its Supportive  
Hatchery Supplementation Program**

Developed for

Chelan County PUD

and the

Habitat Conservation Plan's Hatchery Committee

Developed by

Scott M. Blankenship, Jennifer Von Bargaen, and Kenneth I. Warheit

WDFW Molecular Genetics Laboratory

Olympia, WA

and

Andrew R. Murdoch

Supplementation Research Team

Wenatchee, WA

**March 30, 2007**

# Table of Contents

<b>Executive Summary</b> .....	3
<b>Introduction</b>	
Reasons for evaluation project .....	7
History of artificial propagation .....	8
Previous genetic analyses .....	10
Study objectives .....	12
<b>Methods</b>	
Tissue collection .....	13
Laboratory Analysis .....	13
Genetic Analysis	
Assessing within population genetic diversity .....	14
Within- and among-population genetic differentiation .....	15
Effective population size.....	17
Individual assignment.....	18
<b>Results/Discussion</b> .....	19
<b>Conclusions</b> .....	44
<b>Acknowledgements</b> .....	46
<b>Literature Cited</b> .....	47
<b>Figures</b> .....	52
<b>Tables</b> .....	59



## **Executive Summary**

The main objective of this study was to determine the potential impacts of the Chiwawa River Supplementation Program on natural spring Chinook in the upper Wenatchee system. We did this by investigating population differentiation between temporally replicated Chiwawa River natural and hatchery samples from the Wenatchee River watershed using microsatellite DNA allele frequencies and the statistical assignment of individual fish to specific populations. Additionally, to assess the genetic effect of the hatchery program, we investigated the relationship between census and effective population sizes using collections obtained before and after the supplementation program. In this summary, we briefly describe the salient results contained within this report; however, each “Task” within the Results/Discussion section below contains extended coverage for each topic along with an expanded interpretation of each result.

Overall, we observed substantial genetic diversity within collections, with heterozygosities equal to roughly 80%, over thirteen microsatellite markers. Microsatellite allele frequencies among temporally replicated collections from the same population (i.e., location) were variable, resulting in significant genetic differentiation among these collections. However, these differences are likely the result of salmon life history in this area, as four-year-old Chinook comprise a majority of returns each year. That is, the genetic tests are detecting the differences of contributing parents from each cohort, rather than a hatchery effect.

### *Analysis of Chiwawa River Collections*

To assess the multiple competing hypotheses regarding population differentiation within and among Chiwawa River collections, we found it necessary to organize the Chiwawa genetic data into three data sets: (1) fish origin (hatchery versus natural), (2) spawning location (hatchery broodstock versus in-river (natural) spawners), and (3) four “treatment” groups (1. hatchery-origin hatchery broodstock, 2. hatchery-origin natural spawner, 3. natural-origin natural spawner, and 4. natural-origin hatchery broodstock). We conducted separate analyses using each of the three data sets, with each analysis

touching on some aspect of the components necessary to move through the Conceptual Process outlined by Murdoch and Peven (2005).

**Origin Dataset** – We report that allele frequencies within and between natural- and hatchery-origin collections are significantly different, but there does not appear to be a robust signal indicating that the recent natural-origin collections have diverged greatly from the pre- or early post-supplementation collections. Genetic drift will occur in all populations, but does not appear to be a major factor affecting allele frequencies within the Chiwawa collections.

**Spawning Location Dataset** – There are significant allele frequency differences within and between hatchery broodstock and natural spawner collections. However, in recent years the allele frequency differences between the hatchery broodstock and natural spawner collections have declined. Furthermore, based on linkage disequilibrium, there is a genetic signal that is consistent with increasing homogenization of allele frequencies within hatchery broodstock collections, but a similar homogenization within the natural spawner collection is not apparent. These data suggest that there exists consistent year-to-year variation in allele frequencies among hatchery and natural spawning collections, but there is a trend toward homogenization of the allele frequencies of the natural- and hatchery-origin fish that compose the hatchery broodstock.

**Four Treatment dataset** – Although there are signals of allelic differentiation among Chiwawa River collections, there are no robust signs that these collections are substantially different from each other. We used two different analyses to measure the degree of genetic variation that exists among individuals and collections within the Chiwawa River. First, we conducted a principal component analysis using all Chiwawa samples with complete genotypes (i.e., no missing alleles from any locus). Although the first two principal component axes account for only 10.5% of the total molecular variance, a substantially greater portion of that variance is among individual fish, regardless of their identity, rather than among hatchery and natural collections. The

variances in principal component scores among individuals are 11 and 13 times greater than the variance in scores among collections.

Secondly, using an Analysis of Molecular Variance (AMOVA), we were able to determine how best to group populations, with “best” being defined as that grouping that accounts for the greatest proportion of among group (i.e., population) variance. Furthermore, by partitioning molecular variance into different hierarchical components, we are able to determine what level accounts for the majority of the molecular variance. The AMOVA results clearly show that nearly all molecular variation, no matter how the data are organized, resides within a collection. The percentage of total molecular variance occurring within collections ranged from 99.68% to 99.74%. These results indicate that the significant differences among collections of Chiwawa fish account for less than one percent of the total molecular variance, and these differences cannot be attributed to fish origin or spawning location.

#### *Effective Population Size ( $N_e$ )*

The contemporary estimate of  $N_e$  calculated using genetic data combined for Chiwawa natural-origin spawners (NOS) and hatchery-origin spawners (HOS) Chinook is  $N_e=386.8$ , which is slightly larger than the pre-hatchery  $N_e$  we estimated using demographic data from 1989 – 1992. Additionally, the  $N_e/N$  ratio calculated using 386.8 for  $N_e$  and the arithmetic mean yearly census of NOS and HOS Chinook from 1989 – 2005 for  $N$  is 0.40. These results suggest the  $N_e$  has not declined during the period of Chiwawa Hatchery Supplementation Program operation.

#### *Analysis Of Upper Wenatchee Tributary Collections*

We compared genetic data for spring Chinook collected from the major spawning aggregates of the Wenatchee River. We observed significant differences in allele frequencies among temporally replicated collections within populations, and among populations within the upper Wenatchee. However, these differences account for a very small portion of the overall molecular variance, and these populations overall are very similar to each other. Of all the populations within the Wenatchee River, the White River

appears to be the most distinct. Yet, this distinction is more a matter of detail than of large significance, as the median  $F_{ST}$  between White River collections and all other collections (except the Little Wenatchee collection; see Results/Discussion) is less than 1.5% among population variance. We consider the implications of these results in the Conclusion section that follows the Results/Discussion section. Additionally, there is no evidence that the Chiwawa River Supplementation Program has changed the allele frequencies in the Nason Creek and White River populations, despite the presence of hatchery-origin fish in both these systems.

## Introduction

Murdoch and Peven (2005) outlined 10 objectives to assess the impact (positive or negative) of hatchery operations mitigating the operation of Rock Island Dam. Two objectives relate to monitoring the genetic integrity of populations:

**Objective 3: Determine if genetic diversity, population structure, and effective population size have changed in natural spawning populations as a result of the hatchery program. Additionally, determine if hatchery programs have caused changes in phenotypic characteristics of natural populations.**

**Objective 5: Determine if the stray rate of hatchery fish is below the acceptable levels to maintain genetic variation between stocks.**

This study addresses Objective 3 (above), and documents analyses and results WDFW completed for populations of spring Chinook (*Oncorhynchus tshawytscha*) in the Wenatchee River watershed. This study was not intended to specifically address Objective 5 (above); however, genetic data provide results relevant to Objective 5. The critical component of Objective 3 is to determine if hatchery supplementation has effected change. Furthermore, change in this context means altering census size and/or genetic marker allele frequencies; we did not attempt to measure changes in fitness. Perhaps a more meaningful rewording of Objective 3 is, “Did the hatchery supplementation program succeed at increasing the census size of a target population while leaving genetic integrity intact?” In order to evaluate cause and effect of hatchery supplementation, we surveyed and compared genetic variation in samples collected before and after potential effects from the Chiwawa Hatchery Supplementation Program. Samples were acquired from the primary spawning aggregates in the upper Wenatchee River watershed: Nason Creek, Little Wenatchee River, White River, and Chiwawa River. Hatchery samples were acquired from programs that could potentially affect genetic composition of Wenatchee stocks, the integrated Chiwawa River stock (local stock), Leavenworth National Fish Hatchery spring Chinook (Carson Stock – non local), and Entiat NFH (Carson Stock – non local). Additionally, the genetic markers used were the Genetic Analysis of Pacific Salmonids (GAPS) (Seeb et al. in review) standardized

microsatellites, so all data from the Wenatchee study will be available for inclusion in the GAPS Chinook coastwide microsatellite baseline.

### *History of Artificial Propagation*

Artificial propagation in the upper Columbia River began in 1899 when hatcheries were constructed on the Wenatchee and Methow rivers (Mullan 1987). These initial operations were small, with the Tumwater Hatchery on the Wenatchee River releasing several hundred thousand fry, and the Methow River hatchery producing few Chinook salmon before it was closed in 1913 (Craig and Suomela 1941, Nelson and Bodle 1990). The Leavenworth State Hatchery operated in the Wenatchee River Basin between 1913 and 1931 using eggs from non-native stocks (Willamette River spring-run and lower Columbia Chinook hatchery fall-run). These early attempts at hatchery production were largely unsuccessful for spring-run Chinook (WDF 1934). Between 1931 and 1939, no Chinook salmon hatcheries were in operation above Rock Island Dam (Rkm 730).

In 1938, the last salmon was allowed to pass upstream through the uncompleted Grand Coulee Dam (Rkm 959). To mitigate the loss of habitat, adult Chinook salmon were trapped, under the auspices of the Grand Coulee Fish Maintenance Project (GCFMP), at Rock Island Dam beginning in May 1939, and relocated into three of the remaining accessible tributaries to the upper Columbia River: the Wenatchee, Entiat, and Methow Rivers. GCFMP transfers continued through the autumn of 1943. Spring- and summer/fall-run fish were differentiated at Rock Island Dam based on a 9 July cutoff date for Chinook arrivals at Rock Island Dam (Fish and Hanavan 1948). Spring-run adults collected at Rock Island Dam (pre 9 July fish) were either transported to Nason Creek on the Wenatchee River to spawn naturally (1939-43), or to the newly constructed Leavenworth NFH (1940) for holding and subsequent spawning (1940-43). Eggs were incubated on site or transferred to the Entiat NFH (1941) and Winthrop NFH (1941). In 1944 spring-run adults were allowed to freely pass Rock Island Dam. The GCFMP did not differentiate among late-run stocks (post 9 July fish) passing Rock Island Dam. Late-run offspring reared at the Leavenworth NFH, Entiat NFH, and Winthrop NFHs were an

amalgamation of summer and fall upper Columbia River populations (Fish and Hanavan 1948). Late-run fish were transplanted into the upper and lower Wenatchee, Methow, and Entiat Rivers.

After 1943, the Winthrop NFH continued to use local spring-run Chinook for hatchery production, while the other NFHs largely focused on summer-run Chinook salmon. Renewed emphasis on spring run production in the mid-1970s saw the inclusion of local and non-local eggs (Carson NFH stock, Klickitat River stock, and Cowlitz River stock) to the NFHs. In the early 1980s, imports of non-native eggs were reduced significantly, and thereafter the Leavenworth, Entiat, and Winthrop NFHs have relied on adults returning to their facilities for their egg needs (Chapman et al. 1995). Regarding late-run Chinook, due to the variety of methods employed to collect broodstock at dams, hatcheries, or the result of juvenile introductions into various areas, Chinook populations and runs (i.e., summer and fall) have been mixed considerably in the upper Columbia system over the past five decades (reviewed in Chapman et al. 1994).

Washington Department of Fish and Wildlife (WDFW) operates two facilities producing spring-run Chinook, the Methow Fish Hatchery (MFH) owned by Douglas County PUD that began operation in 1992 and Eastbank Fish Hatchery (EFH) owned by Chelan County PUD that began operation in 1989. Both programs were designed to implement supplementation (supportive breeding) programs for naturally spawning populations on the Methow and Wenatchee Rivers, respectively (Chapman et al. 1995). As part of the Rock Island Mitigation Agreement between Chelan County Public Utility District and the fishery management parties (RISPA 1989), a supplementation (supportive breeding) program was initiated in 1989 on the Chiwawa River to mitigate smolt mortality resulting from the operation of Rock Island Hydroelectric Project. EFH uses broodstock collected at a weir on the Chiwawa River, although in recent years hatchery fish have been collected at Tumwater Dam. Similarly, the MFHC uses returning adults collected at weirs on the Methow River and its tributaries, the Twisp and Chewuch Rivers (Chapman et al. 1995; Bugert 1998). Although low run size and trap efficiency has resulted in most broodstock being collected from the hatchery outfall or in some years Wells Dam,

progeny produced from these programs are reared at and released from satellite sites on the tributaries where the adults were collected. Numerous other facilities have reared spring-run Chinook salmon on an intermittent basis.

*Previous Genetic Studies – Population differentiation*

Waples et al. (1991a) examined 21 polymorphic allozyme loci in samples from 44 populations of Chinook salmon in the Columbia River Basin. These authors reported three major clusters of Columbia River Basin Chinook salmon: 1) Snake River spring- and summer-run Chinook salmon, and mid and upper Columbia River spring-run Chinook salmon, 2) Willamette River spring-run Chinook salmon, 3) mid and upper Columbia River fall- and summer-run Chinook salmon, Snake River fall-run Chinook salmon, and lower Columbia River fall- and spring-run Chinook salmon. Utter et al. (1995) examined allele frequency variability at 36 allozyme loci in samples of 16 upper Columbia River Chinook populations. Utter et al. (1995) indicated that spring-run populations were distinct from summer- and fall-run populations, where the average genetic distance between spring-run and late-run Chinook were about eight times the average of genetic distances between samples within each group. Additionally, allele frequency differences among spring-run populations were considerably greater than that among summer- and fall-run populations in the upper Columbia River. Utter et al. (1995) also reported hatchery populations of spring-run Chinook salmon were genetically distinct from natural spring-run populations, but hatchery populations of fall-run Chinook salmon were not genetically distinct from natural fall-run populations.

As part of an evaluation of the relative reproductive success for the Chiwawa River supplementation program, Murdoch et al. (2006), used eleven microsatellite loci to assess population differentiation among spring Chinook salmon population samples in the upper Wenatchee River. Murdoch et al. (2006) reported a >99% accuracy of correctly identifying spring-run and fall-run Chinook from the Wenatchee River. They also reported slight, but significantly different genetic variation among wild spring populations and between wild and hatchery stocks. Yet, since the spring-run populations



are genetically similar, identifying individuals genetically from the upper tributaries of the Wenatchee River was difficult. This result is exemplified in their individual assignment results, where < 8% of spring-run individuals, hatchery or wild, were correctly assigned using their criterion of an LOD (log of odds) score greater than 2. Murdoch et al. (2006) also reported contemporary natural spring Chinook show heterozygote deficit and low linkage disequilibrium (LD), while contemporary hatchery spring Chinook show heterozygote excess and high LD.

Williamson et al. (submitted) have continued the work of Murdoch et al. (2006) by analyzing Chiwawa River demographic data from 1989 – 2005 to estimate the proportions of recruits that were produced by Chinook with hatchery or wild origin. In an “ideal” population, the genetic size (i.e., effective size or  $N_e$ ) and the census size are equal; however various demographic factors such as unequal sex ratios and variance in reproductive success among individuals reduces the genetic size below the census size. It is generally thought that the genetic size is approximately 10-33% the census size (Bartley et al. 1992; RS Waples pers. comm.), although values have been reported outside this range (Araki et al. 2007; Arden and Kapuscinski 2003; Heath et al. 2002). Despite being difficult to estimate, the effective population size in many respects is a more important parameter to know than census size, because  $N_e$  determines how genetic diversity is distributed within populations and how the forces of evolution (i.e., forces that change genetic diversity over time) will affect the genetic variation present.

Williamson et al. (submitted) used demographic data to 1) investigate the effect of unequal sex ratio on genetic diversity, 2) investigate the effect of variation in reproductive success on genetic diversity, 3) investigate the effect of fluctuations in population size on genetic diversity, and 4) estimate the effective population size, using the inbreeding method (Ryman and Laikre 1991). Most importantly, they use demographic data from 1989 – 2000 to assess the impact of the Chiwawa Hatchery Supplementation Program on the effective population size of natural-origin Chiwawa River spring Chinook. They estimate that the  $N_e$  of naturally spawning Chiwawa Chinook (i.e., both hatchery- and wild-origin fish on the spawning grounds) from 1989 –

1992 was  $N_e = 2683$  and in 1997 – 2000 was  $N_e = 989$ . They compare spawning ground  $N_e$  to estimates calculated from combined broodstock and naturally spawning Chinook demographic data. The combined inbreeding  $N_e$  estimate from 1989 – 1992 was  $N_e = 147$  and in 1997 – 2000 was  $N_e = 490$ . Williamson et al. (submitted) argue that since the combined  $N_e$  estimate is lower than the naturally spawning estimate, the supplementation program has had a negative impact on the Chiwawa River  $N_e$ .

Williamson et al. (submitted) also present genetic data for Chinook recovered on spawning grounds in upper Wenatchee River tributaries in 2004 and 2005. These genetic data are derived from the Murdoch et al. (2006) study. They compare samples collected from Chiwawa River (i.e., hatchery and wild), White River, Nason Creek, and Leavenworth Hatchery. Additionally, they include a 1994 Chiwawa River wild smolt sample for comparison with the 2004 brood year. Williamson et al. (submitted) report statistically significant genetic differentiation among Chiwawa River, White River and Nason Creek. Additionally, they report that the 1994 and 2004 Chiwawa River wild samples are not statistically different, but the 2004 Chiwawa wild and hatchery collections are statistically different.

### *Study Objectives*

This study investigated within and among population genetic diversity to assess the effect of the Chiwawa Hatchery's supplemental program on the natural Chiwawa River spring Chinook population. Differences among temporal population samples, the census size, heterozygosity, and allelic diversity were documented. We investigated population differentiation between the Chiwawa River natural and hatchery samples, and among all temporally replicated samples from the Wenatchee River watershed using microsatellite DNA allele frequencies and the statistical assignment of individual fish to specific populations. To assess the genetic effect of the hatchery program, correlation between census and effective population sizes were investigated using temporally replicated samples obtained before and after the supplementation program operation. To address the hypotheses associated with Objective 3 in Murdock and Peven (2005) we developed

eleven specific “Tasks” (Blankenship and Murdoch 2006), to which we analyzed specific genetic data. We present the results from these analyses specific to each individual Task.

## **Methods and Materials**

### *Tissue collection and DNA extraction*

We analyzed thirty-two population collections of adult spring Chinook salmon (*Oncorhynchus tshawytscha*) obtained from the Wenatchee River between 1989 and 2006 (Table 1). Nine collections of natural Chinook adults from the Chiwawa River (n=501), and nine collections of Chiwawa Hatchery Chinook (n=595) were collected at a weir located in the lower Chiwawa River. The 1993 and 1994 Chiwawa Hatchery samples are smolt samples from the 1991 and 1992 hatchery brood years, respectively. Additional samples were collected from upper Wenatchee River tributaries, White River, Little Wenatchee River, and Nason Creek. Six collections of natural White River Chinook (n=179), one collection from the Little Wenatchee (n=19), and six collections from Nason Creek (n=268) were obtained. Single collections were obtained for Chinook spawning in the mainstem Wenatchee River and Leavenworth National Fish Hatchery. An additional out-of-basin collection from Entiat River was also included in the analysis. Samples collected in 1992 or earlier are scale samples. All other samples were either fin clips or operculum punches, stored immediately in ethanol after collection. DNA was extracted from stored tissue using Nucleospin 96 Tissue following the manufacturer’s standard protocol (Macherey-Nagel, Easton, PA, U.S.A.).

### *Laboratory analysis*

We performed polymerase chain reaction (PCR) amplification on each fish sample using the 13 fluorescently end-labeled microsatellite marker loci standardized as part of the GAPS project (Seeb et al. in review). GAPS genetic loci are: *Ogo2*, *Ogo4* (Olsen et al. 1998); *Oki100* (unpublished); *Omm1080* (Rexroad et al. 2001); *Ots201b* (unpublished); *Ots208b*, *Ots211*, *Ots212*, and *Ots213* (Grieg et al. 2003); *Ots3M*, *Ots9* (Banks et al.

1999); *OtsG474* (Williamson et al. 2002); *Ssa408* (Cairney et al. 2000). PCR reaction volumes were 10  $\mu$ L, and contained 1  $\mu$ L 10x PCR buffer (Promega), 1.0  $\mu$ L MgCl<sub>2</sub> (1.5 mM final) (Promega), 0.2  $\mu$ L 10 mM dNTP mix (Promega), and 0.1 units/mL Taq DNA polymerase (Promega). Loci were amplified as part of multiplexed sets, so primer molarities and annealing temperatures varied. Multiplex one had an annealing temperature of 50°C, and used 0.37 Molar (M) *Oki100*, 0.35 M *Ots201b*, and 0.20 M *Ots208b*, and 0.20 M *Ssa408*. Multiplex two had an annealing temperature of 63°C, and used 0.10 M *Ogo2*, and 0.25 M of a non-GAPS locus (*Ssa 197*). Multiplex three had an annealing temperature of 56°C, and used 0.18 M *Ogo4*, 0.18 M *Ots213*, and 0.16 M *OtsG474*. Multiplex four had an annealing temperature of 53°C, and used 0.26 M *Omm1080*, and 0.12 M *Ots3M*. Multiplex five had an annealing temperature of 60°C, and used 0.30 M *Ots212*, 0.20 M *Ots211*, and 0.10 M *Ots9*. Thermal cycling was conducted on either a PTC200 thermal cycler (MJ Research) or GeneAmp 9700 (Applied Biosystems) as follows: 95°C (2 min); 30 cycles of 95°C for 30 sec., 30 sec. annealing, and 72°C for 30 sec.; a final 72°C extension and then a 10°C hold. PCR products were visualized by electrophoresis on an ABI 3730 automated capillary analyzer (Applied Biosystems). Fragment analysis was completed using GeneMapper 3.7 (Applied Biosystems). Standardization of genetic data to GAPS allele standards was conducted following Seeb et al. (in review).

### *Genetic data analysis*

**Assessing within population genetic diversity** - Heterozygosity measurements are reported using Nei's (1987) unbiased gene diversity formula (i.e., expected heterozygosity) and Hedrick's (1983) formula for observed heterozygosity. Both tests are implemented using the microsatellite toolkit (Park 2001). We used GENEPOP version 3.4 (Raymond and Rousset 1995) to assess Hardy-Weinberg equilibrium (HWE), where deviations from the neutral expectation of random associations among alleles are calculated using a Markov chain method (5000 iterations in this study) to obtain unbiased estimates of Fisher's exact test. Global estimates of  $F_{IS}$  according to Weir and Cockerham (1984) were calculated using GENEPOP version 3.4. Genotypic linkage disequilibrium was calculated following Weir (1979) using GENEPOP version 3.4.

Linkage results for population collections are reported as the proportion of pairwise (locus by locus) tests that are significant ( $\alpha = 0.01$ ). Linkage disequilibrium is considered statistically significant if more than 5% of the pairwise tests based on permutation are significant for a collection.

**Within- and among-population genetic differentiation** – The temporal stability of allele frequencies within populations, and pairwise differences in allele frequencies among populations were assessed using several different procedures. First, we tested for differences in allele frequencies among populations defined in Table 1 using a randomization chi-square test implemented in GENEPOP version 3.4 (Raymond and Rousset 1995). This procedure tests for differences between pairs of populations where alleles are randomized between the populations (i.e., genic test). The null hypothesis for this test is that the allele frequency distributions between two populations are the same. A low p-value should be interpreted as the allele frequency distributions being compared are unlikely to be samples drawn from the same underlying distribution.

Second, to graphically describe allele frequency differences among populations we conducted a nonmetric multidimensional scaling analysis using allele-sharing distance matrices from two different data sets. Pairwise allele-sharing distances are calculated as  $1 - (\text{mean over all loci of the sums of the minima of the relative frequencies of each allele common to a pair of populations})$ . To calculate the allele-sharing distances for each pair of populations we used PowerMarker v3.25 (Liu and Muse 2005). Nonmetric multidimensional scaling is a technique designed to construct an n-dimensional “map” of populations, given a set of pairwise distances between populations (Manly 1986). The output from this analysis is a set of coordinates along n-axes, with the coordinates specific to the number of n-dimensions selected. To simplify our analysis we selected a 2-dimensional analysis to represent the relative positions of each population in a typical bivariate plot. The goodness of fit between the original allele-sharing distances and the pairwise distances between all populations along the 2-dimensional plot is measured by a “stress” statistic. Kruskal (in Rohlf 2002) developed a five-tier guide for evaluating stress levels, ranging from a perfect fit (stress=0) to a poor fit (stress=0.40). We

conducted the nonmetric multidimensional scaling analysis for one data set containing Chiwawa natural- and hatchery-origin collections, and another data set containing Chiwawa broodstock and in-river spawner collections. We used the `mdscale` module in MATLAB R2006b (The Mathworks 2006) to generate the nonmetric multidimensional scaling coordinates.

We examined the geographic and temporal structure of populations in the upper Wenatchee (Chiwawa River, Nason Creek, and White River, only) using a series of analyses of molecular variance (AMOVAs). Here, we defined an AMOVA as an analysis of variance of allele frequencies, as originally designed by Cockerham (1969), but implemented in Arlequin v2.1 (Schneider et al. 2000). These analyses permit populations to be aggregated into groups, and molecular variance is then partitioned into within collections, among collections, but within groups, and among group components. With this approach, we were able to determine how best to group populations, with “best” being defined as that grouping that accounts for the greatest proportion of among group variance. Furthermore, by partitioning molecular variance into three different hierarchical components, we are able to determine what level accounts for the majority of the molecular variance.

Finally, we explored the partitioning of molecular variance between among-individuals and among-populations using a principal component analysis and multi-locus estimates of pairwise  $F_{ST}$ , estimated by a “weighted” analysis of variance (Weir and Cockerham, 1984). Principal component analysis is a data-reduction technique whereby the correlation structure among variables can be used to combine variables into a series of multivariate components, with each original variable receiving a weighted value for each component based on its correlation with that component. Here, we used a program written by Warheit in MATLAB R2006b (The Mathworks 2006) that treats each allele for each locus as a single variable (13 loci = 26 alleles or variables), and these 26 “variables” were arranged into 26 components, with each component accounting for a decreasing amount of molecular variance. Estimates of  $F_{ST}$  were calculated using GENETIX version 4.05 (Belkhir et al.1996). To determine if the  $F_{ST}$  estimates were

statistically different from random (i.e., no structure), 1000 permutations were implemented in GENETIX version 4.05 (Belkhir et al.1996).

**Effective population size ( $N_e$ )** – Estimates of the effective population size were obtained using two methods, a multi-collection temporal method (Waples 1990), and a single-collection method (Waples 2006) using linkage disequilibrium data. The temporal method assumes that cohorts are used, but we did not decompose the collection year samples into their respective cohorts using age data. Therefore,  $N_e$  estimates that pertain to individual year classes of breeders are not valid; however the harmonic mean over all samples will estimate the contemporary  $N_e$ . Comparing samples from years  $i$  and  $j$ , Waples’ (1990) temporal method estimates the effective number of breeders ( $\hat{N}_{b(i,j)}$ ) according to:

$$\hat{N}_{b(i,j)} = \frac{b}{2(\hat{F} - 1/\hat{S}_{i,j})}$$

The standardized variance in allele frequency ( $\hat{F}$ ) is calculated according to Pollack (1983). The parameter  $b$  is calculated analytically from age structure information and the number of years between samples (Tajima 1992). The age-at-maturity information required to calculate  $b$  was obtained from Murdoch et al. (2006) for this analysis. They observed for Chiwawa Hatchery Chinook that 8.6% matured at age 2, 4% at age 3, 87% at age 4, and 0.4% at age 5. For Chiwawa natural Chinook, Murdoch et al. (2006) observed that 1.8% matured at age 3, 81.6% at age 4, and 16.7% at age 5. The harmonic mean of sample sizes from years  $i$  and  $j$  is  $\tilde{S}_{i,j}$ . Over all pairwise comparisons the harmonic mean of all  $\hat{N}_{b(i,j)}$  is  $\tilde{N}_b$ , the contemporary estimate of the effective population size ( $N_e$ ). SALMONNb (Waples et al. 2007) was used to calculate  $\tilde{N}_b$ . As suggested by authors, alleles with a frequency below 0.05 were excluded from the analysis to reduce potential bias.

The method of Waples (2006) uses linkage disequilibrium (i.e., mean squared correlation of allele frequencies at different gene loci) as a means of estimating effective population size ( $N_e$ ) from a single sample. While this method is biased in some cases where  $N_e/N$

ratio is less than 0.1 and the sample size is less than the true  $N_e$ , it has been shown to produce comparable results to the temporal method. Burrows' delta method is used to estimate LD, and a bias corrected estimate of  $N_e$  is calculated after eliminating alleles with frequency less than 0.05. This test was implemented using LD $N_e$  (Do and Waples unpublished). In age-structured species,  $N_e$  estimates based on LD are best interpreted as the effective number of breeders ( $N_b$ ) that produced the sample (Waples 2006).  $N_b$  should be multiplied by the mean generation length (i.e., 4 in this case) to obtain an overall estimate of  $N_e$  based on an  $N_b$  estimate. We analyzed collections categorized by spawning location (i.e., hatchery broodstock or in-river) and did not analyze collections categorized by origin (i.e., hatchery or natural). Waples' (2006) method estimates  $N_e$  from observed LD, therefore the corresponding  $N_e$  estimates for the hatchery collections would be low and the estimates for the natural collections would be high. Yet, since the supplementation program is integrated, and hatchery fish can spawn naturally, we feel it inappropriate to analyze the hatchery and natural samples as if they were separate, which would essentially partition all the LD into the hatchery samples.

Each collection has an  $N_b$  estimate and an associated confidence interval. If the confidence interval includes infinity, it means that sampling error accounts for all the LD observed (i.e., empirical LD is less than expected LD). The usual interpretation is that there is no evidence for any disequilibrium caused by genetic drift in a finite number of parents. Since the LD method estimates the number of breeders that contributed to the sample being analyzed, in order to calculate an  $N_e/N$  ratio, the appropriate census size must be used. The census size used to derive a ratio was the estimate four years prior to the collection analyzed using LD, which assumed a strict four-year-old lifecycle, although the observed proportion of four-year-olds was approximately 85% each year. The census numbers (Table 2) used to calculate the ratios for Chiwawa broodstock and in-river spawners were combined NOS (natural-origin spawners) and HOS (hatchery-origin spawners) census estimates.

**Individual assignment** – A population baseline file was constructed containing all 1704 individual Chinook from 34 population collections (Table 1; Chiwawa origin data set



plus all samples from other populations). All individuals in the baseline had genotypes that included nine or more loci. Individual Chinook were assigned to their most likely population of origin based on the partial Bayesian criteria of Rannala and Mountain (1997), using a “jack-knife” procedure, where each individual to be assigned was removed from the baseline prior to the calculation of population likelihoods. This procedure was implemented in a program written by Warheit in MATLAB R2006b (The Mathworks 2006). Two assignment criteria were used, 1) the population with the largest posterior probability for an individual was the “most-likely” population of origin (i.e., all individuals assigned to a collection), and 2) an assignment was consider valid only if the posterior probability was greater than or equal to 0.9. Please note that while the analysis used 34 population collections to assign Rannala and Mountain likelihoods for each individual, these likelihoods were aggregated based on “population” (i.e., Chiwawa, Nason, White, and so on) and posterior probabilities were calculated for population location, rather than individual collections.

## **Results and Discussion**

In this section we combine our presentation and interpretations of the genetic analyses. Additionally, this section will be organized based on the task list presented in the study plan. Overall conclusions are provided following this section.

### **Task 1: Determine trend in census size for Chiwawa River spring Chinook.**

Census data from 1989 – 2005 are provided in Table 2 for the Chiwawa Hatchery broodstock and spring Chinook present in the Chiwawa River. The demographic data for naturally spawning Chinook are based on redd sampling and carcass surveys, while broodstock data are based on Chiwawa hatchery records. As the supplementation program is integrated by design, we also present the proportion of natural-origin broodstock (pNOB) incorporated into the hatchery, in addition to the number of natural-origin (NOS) and hatchery-origin (HOS) spawners present in Chiwawa River. The

census size fluctuated yearly, and a general reduction in census size was observed in the mid to late 1990's. This trend was apparent in both the broodstock and in the river. The arithmetic mean census size from 1989 – 2005 for the Chiwawa Hatchery (i.e., broodstock) was  $N=87.5$  per year. The arithmetic mean census size from 1989 – 2005 for the Chiwawa River (i.e., NOS and HOS combined) was  $N=961.9$  per year. For collection years when adult Chiwawa hatchery-origin fish would have been absent in the Chiwawa River (1989 – 1992), the arithmetic mean of natural Chiwawa Chinook census size is  $N=962.7$ . We will use this number as the baseline census size to assess if census size has changed. We used two different values for the contemporary census size in the Chiwawa River, NOS only and NOS + HOS. Additionally, we used collection years 2002 – 2005 for the contemporary NOS and HOS estimates, as these are the most recent data and the number of years included for estimation is the same as the pre-hatchery estimate above (i.e., four years). For NOS only, the arithmetic mean census size from 2002 – 2005 was  $N=536.0$ . For total census size (i.e., NOS and HOS combined), the arithmetic mean census size from 2002 – 2005 was  $N=1324.0$ . For the demographic data presented here, the contemporary census size is larger than the census estimate derived from the years prior to hatchery operation.

## **Task 2: Document the observed genetic diversity.**

### *Genetic Diversity Categorized By Origin*

For Chiwawa River collections categorized by origin (Table 1A), substantial genetic diversity was observed, with heterozygosity estimates over all loci, having a mean of 0.80. Genetic diversity was consistent with expected Hardy-Weinberg random mating genotypic proportions for ten of the eighteen collections. Eight of the nine Chiwawa natural collections were consistent with HWE, and two of nine Chiwawa Hatchery collections were consistent with HWE.  $F_{IS}$  is observed to be slight for all Chiwawa population collections, suggesting individuals within collections do not show excessive homozygosity.

The deviations from HWE observed were generally associated with hatchery collections. The two smolt collections (i.e., 1993 and 1994) showed significant deviations from HWE, which may be a function of non-random hatchery practices involving the contributing natural-origin parental broodstocks (i.e., 1991 and 1992 cohort). Deviations from HWE in the remaining hatchery collections may be the result of few individuals being represented in the broodstock (see below).

Additionally, linkage disequilibrium (LD) was also common for Chiwawa hatchery-origin collections and minimal for Chiwawa natural-origin collections. The random association of alleles between loci (i.e., linkage equilibrium) is expected under ideal conditions. LD is observed when particular genotypes are encountered more than expected by chance. Laboratory artifacts (e.g. null alleles) or physical linkage of loci on the same chromosome can cause LD, but the LD we observed was not associated with certain locus combinations, which you would expect if either artifacts or physical linkage were the cause of LD. LD was observed for seven of the nine hatchery-origin collections. As with the deviations from HWE, the high LD in the 1993 and 1994 hatchery-origin collections may be a result of non-random hatchery practices. The substantial LD observed in the hatchery-origin adult collections (collection years 2000, 2001, 2004, and 2006) might be the result of small parental broodstock sizes contributing to those returning adults. During the mid 1990's, the Chiwawa broodstock size was low, with zero individuals collected in 1995 and 1999; so fewer individuals would be contributing to the hatchery adult returns than the natural. This idea is corroborated by the lower LD observed for the 2005 hatchery-origin collection, which had a contributing parental broodstock size in 2001 (i.e., the major contributing parental generation) approximately eight times as large as the previous few collection years (Table 2). LD reappears in the 2006 Chiwawa hatchery-origin collection, which had a contributing parental broodstock size (i.e., for the most-part, the 2002 hatchery brood year) five times lower (Table 2) than that of the 2005 collection.

While seven of nine hatchery-origin collections showed significant LD, only one natural origin collection showed LD, and for this collection, only 10% of the loci-pairs were in

disequilibrium (Table 1). The fact that LD predominated in the hatchery samples, suggests that variance in reproductive success (i.e., overrepresentation of particular parents) is higher in the hatchery-origin than in natural-origin collections.

#### *Genetic Diversity Categorized By Spawning Location*

For upper Wenatchee River collections categorized by spawning location (Table 1B), substantial genetic diversity was observed, with heterozygosity estimates over all loci, having a mean of 0.79 and ranging from a low of 0.69 (1993 White River) to 0.85 (1993 Little Wenatchee). Genetic diversity was consistent with HWE for nineteen of twenty-nine population collections. For the collections that departed from HWE, seven were from the Chiwawa River, one was from Leavenworth Hatchery, one was the Wenatchee mainstem collection of hatchery-origin – naturally spawning fish, and one was from the White River.  $F_{IS}$  is observed to be slight for all population collections except the 1993 White River collection (10% heterozygote deficit) (Table 1B). Collections deviating with HWE generally correlated with collections having high LD. Twelve population collections showed a proportion of pairwise linkage disequilibrium tests (across all loci) greater than 5% (Table 1B), eight of which were Chiwawa collections.

Starting in 1996, spawning location collections are composed of both natural- and hatchery-origin samples. The LD seen in the later spawning location collections may be caused by an admixing effect (i.e., mixing two populations), where random mating has not had the chance to freely associate alleles into genotypes. Interestingly, there appears to be a trend of reducing LD through time within the broodstock collections (Table 1B), which suggests that a “homogenizing” effect is taking place within the Chiwawa River. This observation is discussed more fully in Task 3 below.

### **Task 3: Test for population differentiation among collections within the Chiwawa River and associated supplementation program.**

#### *Introduction*

Task 3 was designed to address two hypotheses listed as part of Objective 3 in Murdoch and Peven (2005):

- Ho: Allele frequency<sub>Hatchery</sub> = Allele frequency<sub>Naturally produced</sub> = Allele frequency<sub>Donor pop.</sub>
- Ho: Genetic distance between subpopulations<sub>Year x</sub> = Genetic distance between subpopulations<sub>Year y</sub>

Murdoch and Peven (2005) proposed these two hypotheses to help evaluate the Chiwawa supplementation program through the “Conceptual Process” (Figure 5 in Murdoch and Peven 2005; repeated here as Figure 1). There are two components to the first hypothesis, which must be considered separately. The first component involves comparisons between natural-origin populations in the Chiwawa to determine if there have been changes in allele frequencies or genetic distances, through time starting with the donor population. Documenting a change does not necessarily indicate that the supplementation program has directly affected the natural origin fish, as additional tests would be necessary to support that hypothesis. The intent of the second component is to determine if the hatchery produced populations have the same genetic composition as the naturally produced populations.

Although on the surface these two components and their associated comparisons may appear simple, from a hypothesis-testing perspective the analyses are complicated by the fact that natural-origin fish may have had hatchery-origin parents, and hatchery-origin fish may have had natural-origin parents. As such, we organized the Chiwawa genetic data into three data sets: (1) fish origin (hatchery versus natural), (2) spawning location (hatchery broodstock versus in-river (natural) spawners), and (3) four “treatment” groups (1. hatchery-origin hatchery broodstock, 2. hatchery-origin natural spawner, 3. natural-origin natural spawner, and 4. natural-origin hatchery broodstock). We conducted separate analyses using each of the three data sets, with each analysis touching on some aspect of the components necessary to move through the Conceptual Process (Figure 1).

### *Hatchery- Versus Natural-Origin*

We address the following questions with the origin data set:

1. Are there changes in allele frequencies and allele sharing distances in the natural-origin collections from pre-supplementation to today?
2. Are there changes in allele frequencies and allele sharing distances in the hatchery-origin collections from early supplementation to today?
3. Are there significant differences in allele frequencies and large allele sharing distances between hatchery- and natural-origin adults from a collection year, and has this pattern changed through time?

**Genic Differentiation Tests** – We explicitly tested the hypothesis of no significant differentiation within natural- or hatchery-origin collections from the Chiwawa River using a randomization chi-square test. We show the results for the pairwise comparisons among natural-origin collections from the Chiwawa River populations in the first block of the second page of Table 3. Ten of the 36 (28%) pairwise comparisons have highly significant allele frequency differences, while only 12 of the 36 comparisons (33%) showed no significant differences. Eight of these 12 comparisons involved the 1996 collection, which included only eight samples and therefore provided little power to differentiate allele frequencies. If we exclude the 1996 collection, only 14% of the pairwise comparisons showed no significant differences, and here all but one of these comparisons involved the 1989 collection. The 1989 collection appeared to be the least differentiated collection in the natural-origin data set in that all pairwise comparisons were either not significant, or only mildly significant at the nominal critical value. No comparisons involving the 1989 collection were significant using a Bonferroni-corrected critical value, and 1989 is the only natural-origin collection in our data set that can be classified as “pre-supplementation.”

We can interpret these results to indicate that although there appears to be significant year-to-year differences in allele frequencies among post-supplementation collections, the allele frequencies between each post-supplementation collection and the 1989 pre-supplementation collection are not greatly different. However, the level of differentiation

does increase from the early post-supplementation years to the more recent years (2001, 2004-2006), although the statistical level of this significance never exceeds the Bonferroni-corrected critical value. Finally, sample sizes were also small for the 1989 collection ( $n = 36$ ) and we cannot eliminate a reduction in power as a contributing factor for the lack of significance for these tests.

As with the hatchery-origin collections, most pairwise comparisons of allele frequencies between hatchery-origin samples were significant (Table 3, first page, upper block). Out of the 36 pairwise comparisons, all but three are significant at some level, and most comparisons are highly significant. Similar to the natural-origin analysis, the non-significant results were limited to comparisons involving the 1996, which included only eight samples.

As a result of this analysis *we reject the hypothesis that there was no significant differentiation among natural- or hatchery-origin collections from the Chiwawa River.* Furthermore, the allele frequencies of the hatchery-origin collections are significantly different from those of natural-origin collections (Table 3, first page, second block). For those fish collected in the same year, allele frequencies are significantly different between hatchery- and natural-origin collections, although in 2005 the level of significance was below the Bonferroni critical value (Table 3). The next step is to examine the pattern of allelic differentiation to discover first if there is a trend among the data, and second, if this trend suggests that the allele frequency differences among Chiwawa River natural-origin fish collections has been affected by the hatchery-origin fish.

**Allele-sharing and Nonmetric Multidimensional Scaling** – We constructed a pairwise allele-sharing distance matrix for all hatchery- and natural-origin collections from the Chiwawa River and subjected this matrix to a nonmetric multidimensional scaling analysis, restricting the analysis to two dimensions (Figure 2). The stress statistic for this analysis is 0.09, a value Kruskal (in Rohlf 2002) listed as a good to excellent fit between the actual allele-sharing distances and the Euclidean (straight-line) distances in the plot.

In other words, Figure 2 is a good visual representation of the allele sharing distance matrix; collections with a high percentage of alleles shared will be closer to each other than collections with a lower percentage of alleles shared.

With the exception of the two outlier years (1996 and 1998) the Chiwawa natural-origin collections form a tight cluster indicating an overall common set of shared alleles among these collections. Even if we ignore the 1996 and 1998 hatchery-origin collections, there appears to be a greater variance in shared alleles among the Chiwawa hatchery-origin collections than the natural-origin collections (Figure 2). In fact, the median percentage of alleles shared among the Chiwawa natural-origin collections is 76% compared with 69% alleles shared among the Chiwawa hatchery-origin collections.

Also, there appears to be a convergence in allele sharing distances (i.e., a decrease in allele frequency differences) between the hatchery- and natural-origin fish from the late 1980s/early 1990s to 2006. The series of red arrows in Figure 2 represent the progression of change in hatchery-origin allele sharing distances from 1996 (first adult hatchery origin fish in our analysis) to 2006 and this progression is decidedly in the direction of the natural-origin cluster. However, the most recent natural-origin collections (2001, 2004-2006) appear to have pulled closer to the hatchery-origin collections, compared with the 1989 natural-origin collection (note the close proximity of the 2000 and 1989 natural-origin collections). Nevertheless, the cluster of natural-origin collections adjacent to the hatchery-origin collections in Figure 2 also includes the 1993 natural-origin collection. Qualitatively, it appears that the initial hatchery-origin and natural-origin collections were more different from each other in terms of the percentage of shared alleles than are the most recent hatchery- and natural-origin collections. This may have been a result of a non-random sample of natural-origin fish that was used as broodstock in the initial years of the supplementation program (see discussion in Task 2 concerning deviations from HWE and linkage disequilibrium).

That being said, we do need to emphasize that Figure 2 is dominated by five outlier collections (two each from the 1996 and 1998 collections, and the 1994 smolt collection).



The 1996 and 1998 collections are characterized by small samples sizes, and the 1994 smolt collection has nearly all pairs of loci in linkage disequilibrium (Table 1). If we eliminate these five outlier groups, both the hatchery- and natural-origin collections form a relatively tight cluster. Excluding the five outliers, the median percentage of shared alleles among all pairwise combinations of Chiwawa hatchery versus Chiwawa natural collections is 76%. This compares with a median pairwise percentage of 79% among only Chiwawa natural-origin collections. That is, there are nearly as many alleles shared between the hatchery-origin and natural-origin collections as there are among the natural-origin collections themselves. There is also a narrowing of differences between natural- and hatchery-origin fish from the same collection years from 1993 (76% shared alleles) through 2006 (83% shared alleles).

If allelic differentiation among collections is a function of genetic drift, we would expect a positive correlation between the number of years between two collections and the allele sharing distance. That is, if genetic drift is the primary cause of allele frequency differences between two collections, the greater the number of years between the two collections the larger the allele-sharing distance. For both the natural- and hatchery-origin collections we examined the relationship between the number of years between a pair of collections and the collections' allele-sharing distance (Figure 3). Although the relationship between time interval and allele distance appears to be a positive function in the natural collections, the slope of the regression line is 0.0017, and is not significantly different from zero. Furthermore, the correlation coefficient ( $r^2$ ) equals 0.1068, which means that the time interval between collections accounts for only 10% of the pairwise differences in allelic distance. The hatchery-origin collections do show a significantly positive slope (0.0037;  $p = 0.0254$ ) and a regression coefficient nearly three times greater than that for the natural-origin collections. However, the correlation coefficient is still relatively small ( $r^2 = 0.3290$ ), indicating that the time interval between collections accounts for one-third of the pairwise differences in allelic distance. The results suggest that if genetic drift is a factor in allelic differentiation between collections, it is only a minor factor, and appears to have affected the hatchery-origin collections more than the natural-origin collections.

If four-year-old fish dominate each collection year, we would expect a closer relationship among collections that are spaced at intervals of four years. The average percentage of alleles shared between two natural-origin collections that are separated by four years or a multiple of four years is 81%, compared with 78% for natural-origin collections separated by years that are not divisible by four. Likewise, for hatchery-origin collections the average percentage of alleles shared is 80% and 75% for collections separated by years divisible and not divisible by four, respectively. Although the percent differences described above are relatively small, they are consistent with the idea that allelic differences between collections are a function of year-to-year variability among different cohorts of four year-old fish.

**Summary** – The allele frequencies within and between natural- and hatchery-origin collections are significantly different, but there does not appear to be a robust signal indicating that the recent natural-origin collections have diverged greatly from the pre- or early post-supplementation collections. Genetic drift will occur in all populations, but does not appear to be a major factor with the Chiwawa collections. We propose that the differences among collections are a function of differences in allele frequencies among cohorts of the four year-old fish that dominate each collection.

#### *Hatchery Broodstock Versus Natural (In-River) Spawners*

We address the following questions with the spawner data set:

1. Are there changes in allele frequencies and allele sharing distances in the natural spawning collections from pre-supplementation to today?
2. Are there changes in allele frequencies and allele sharing distances in the hatchery broodstock collections from early supplementation to today?
3. Are there significant differences in allele frequencies and large allele sharing distances between hatchery and natural spawning adults from a collection year, and has this pattern changed through time?

**Genic Differentiation Tests** – For the most part there are significant differences in allele frequencies among collections for both the hatchery broodstock and natural spawners (Table 4), and these differences are consistent with the origin data set (Table 3). There are four collection years with paired samples (2001, 2004-2006) where we can compare allele frequency differences between the hatchery broodstock and natural spawners, within the same year. The 2001 hatchery broodstock and natural spawner collections have significantly different allele frequencies, but the level of significance decreased from 2001 to 2004, and become non-significant in 2005 and 2006 (Table 4). This indicates that by 2005, the hatchery broodstock and natural spawners collections were effectively sampling from the same population of fish. Additionally, the percentage of alleles shared between the hatchery broodstock and the natural spawners increased from 76% in 2001 to 86% in 2006 (allele sharing distance matrix, not shown). From this analysis, we conclude that although there are year-to-year differences in allele frequencies within the natural and hatchery spawner collections, *there appears to be a convergence of allele frequencies within collection-year, between the natural and hatchery spawner populations.*

**Linkage Disequilibrium** – Linkage disequilibrium is the correlation of alleles between two loci, and can occur for several reasons. If two loci are physically linked on the same chromosome, than alleles from each of these loci should be correlated. However, linkage between two loci can occur as a result of population bottlenecks, small population sizes, and natural selection. If any of these conditions had occurred or were occurring within the Chiwawa River system, we would expect to find substantial linkage disequilibrium in many or perhaps all Chiwawa collections. However, many Chiwawa collections, especially the natural-origin collections, do not show linkage disequilibrium (Table 1), and it would appear that the linkage disequilibrium within certain Chiwawa collections is not a function of the processes listed above. Linkage disequilibrium can also result if the collection is composed of an admixture. That is, if two or more reproductively isolated populations are combined into a single collection, the collection will show linkage disequilibrium. Each broodstock and natural spawning collection is composed of natural- and hatchery-origin fish. If these hatchery- and natural-origin fish are drawn from the

same population, the spawning collections should not show substantial linkage disequilibrium. However, if the hatchery- and natural-origin fish are from different populations (i.e., full hatchery – natural integration has not been achieved), the spawning collections should show substantial linkage disequilibrium.

There are only three Chiwawa spawning collections that are not composed of both hatchery- and natural-origin samples: 1989 (natural-origin, natural spawner), 1993 (natural-origin, hatchery broodstock), and 2001 (natural-origin, natural spawner). Of the 10 spawning collections with both hatchery- and natural-origin fish, seven show significant linkage disequilibrium. Two of the three collections that did not show linkage disequilibrium are the 1996 and 1998 hatchery broodstock collections, which are composed of only seven natural- and six hatchery-origin fish, and two natural- and 19 hatchery-origin fish, respectively. Within the hatchery broodstock collections with linkage disequilibrium, the percent of loci pairs showing linkage decreased from 32% in 2000 to 13% in 2001 and 2004, to only 1% and 5% in 2005 and 2006, respectively (Table 1). If the homogenization of allele frequencies of natural- and hatchery-origin fish was increasing from 2000 to 2006, we would expect a decrease in linkage disequilibrium among the broodstock collections. This is what occurred within the hatchery broodstock collections, but did not occur within the natural spawner collections, where the percent of loci pairs showing linkage was 18% in 2004, 6% in 2005, and 10% in 2006 (Table 1). Furthermore, the 2001 natural spawner collection, with no hatchery-origin component showed linkage disequilibrium with 9% of loci pairs.

There is no correlation between percent of loci pairs showing linkage disequilibrium and percent of broodstock composed of hatchery-origin fish ( $r^2 = 0.0045$ ). Furthermore, the natural spawner and hatchery broodstock collections were each composed of roughly the same average percentage of hatchery-origin fish (57% and 53%, respectively). If the decrease in linkage disequilibrium among the hatchery broodstock collections from 2000 to 2006 was a result of a homogenization of allele frequencies of natural- and hatchery-origin fish in the broodstock, the same degree of homogenization did not occur within the

natural spawner collections. This would occur if natural- and hatchery-origin fish spawning within the river remain segregated, either by habitat or by fish behavior.

**Summary** – As with the origin data set, there are significant allele frequency differences within and between hatchery broodstock and natural spawner collections. However, in recent years the allele frequency differences between the hatchery broodstock and natural spawner collections has declined. Furthermore, based on linkage disequilibrium, there is a genetic signal that is consistent with increasing homogenization of allele frequencies within hatchery broodstock collections, but a similar homogenization within the natural spawner collection is not apparent. These data suggest that there exists consistent year-to-year variation in allele frequencies among hatchery and natural spawning collections, but there is a trend toward homogenization of the allele frequencies of the natural- and hatchery-origin fish that compose the hatchery broodstock.

#### *Four Treatment Groups*

Analyses of genetic differences between hatchery (broodstock) and natural spawner collections is confounded by the fact that each these two groups are composed of fish of natural- and hatchery-origin. To understand the effects of hatchery supplementation on *natural-origin fish that spawn naturally*, we needed to divide the Chiwawa data set into four mutually exclusive groups: (1) hatchery-origin hatchery broodstock, (2) hatchery-origin natural spawner, (3) natural-origin hatchery broodstock, and (4) natural-origin natural spawner, with each group consisting of multiple collection years, for a total of 25 different groups.

**Allele-sharing and Nonmetric Multidimensional Scaling** –As with previous analyses discussed above, we constructed a pairwise allele-sharing distance matrix for all collections from each of these treatment groups and subjected this matrix to a nonmetric multidimensional scaling analysis, restricting the analysis to two dimensions. Figure 4 shows that five outlier groups dominate the allele-sharing distances within this data set. These outlier groups are also present in Figure 2, as discussed above, and Figure 2 and 4 resemble each other because the same fish are included in each analysis. The difference

between Figures 2 and 4 is that in Figure 4 the fish are grouped into collection year and the four treatment groups, rather than collection year and two treatment groups (hatchery-versus natural-origin).

Figure 4 does not provide useful resolution of the groups within the polygon, because the outlier groups dominate the allele sharing distances. We removed the five outlier groups from Figure 4, recalculated the allele sharing distances and subjected this new matrix to a multidimensional scaling analysis (Figure 5). Figure 5 shows separation among the 2001, 2004-2006 collections, but this separation does not necessarily indicate that within-year collections are more similar to each other than any collection is to a collection from another year. For example, the 2006 natural-origin natural spawner and the 2005 natural-origin hatchery broodstock collections share 81% alleles, while the 2006 natural-origin natural spawner and 2006 hatchery-origin hatchery broodstock collections share 75% alleles. There does not appear to be any discernable pattern of change in allele-sharing distance among the collections relevant to pre- or post-supplementation. Although the 1989 pre-supplementation natural-origin collection appears distinct (Figure 5), the 1993 natural-origin hatchery broodstock collection appears quite similar to the 2005 and 2006 natural-origin collections (Figure 5). The 1993 natural-origin hatchery broodstock collection, although not technically pre-supplementation, is composed of fish whose ancestry cannot be traced to any Chiwawa hatchery fish. Therefore, there is no clear pattern of allele sharing change from pre-supplementation to recent collections.

There does appear to be some change in the average percentage of alleles shared within the 2001 to 2006 collections, with an increase from 74% in 2001 and 2004 to 78% and 79% in 2005 and 2006, respectively. The results provided by this analysis are consistent with the results presented in the origin and spawner data sets. That is, there are allele frequency and allele sharing differences among the collections, but analyses do not strongly suggest that these differences are a function of the supplementation program. Furthermore, there is also a weak signal that the hatchery and natural collections within the most recent years are more similar to each other than in the previous years.

**Overall Genetic Variance** – Although there are signals of allelic differentiation among Chiwawa River collections, there are no robust signs that these collections are substantially different from each other. We used two different analyses to measure the degree of genetic variation that exists among individuals and collections within the Chiwawa River. First, we conducted a principal component analysis using all Chiwawa samples with complete genotypes (i.e., no missing alleles from any locus). Although the first two principal component axes account for only 10.5% of the total molecular variance, a substantially greater portion of that variance is among individual fish, regardless of their identity, rather than among hatchery and natural collections (Figure 6). The variances in principal component scores among individuals are 11 and 13 times greater than the variance in scores among collections, along the first and second axes, respectively.

Second, we conducted a series of analyses of molecular variance (AMOVA) to ascertain the percentage of molecular variance that could be attributed to differences among collections. We organized these analyses to test also for differences in the hierarchical structure of the data. That is, we tested for differences among collections using the following framework:

- No organizational structure – all 25 origin-spawner collections considered separately
- Origin-spawner collections organized into 10 collection year groups
- Origin-spawner collections organized into 2 breeding location groups (hatchery versus natural)
- Origin-spawner collections organized into 2 origin groups (hatchery versus natural)
- Origin-spawner collections organized into the 4 origin-spawner groups

It is clear from this analysis that nearly all molecular variation, no matter how the data are organized, resides within a collection (Table 5). The percentage of total molecular variance occurring within collections ranged from 99.68% to 99.74%. The among group variance component was limited to less than 0.26% and in all organizational structures,

except “no structure,” the among group percentage was not significantly greater than zero. Furthermore, none of the organizational structures provided better resolution than “no structure” in terms of accounting for molecular variance within the data set. *These results indicate that if there are significant differences among collections of Chiwawa fish, these differences account for less than one percent of the total molecular variance, and these differences cannot be attributed to fish origin or spawning location.*

#### *Summary and Conclusions*

We reject the null hypothesis that the allele frequencies of the hatchery collections equal the allele frequencies of the natural collections, which equals the allele frequency of the donor population. Furthermore, because the allele-sharing distances are not consistent within and among collections years, we also reject the second stated hypothesis discussed above. However, there is an extremely small amount of genetic variance that can be attributed to among collection differences. The allelic differentiation that does exist among collections does not appear to be a function of fish origin, spawning location, genetic drift, or collection year. Figure 5 and related statistics does suggest that hatchery and natural collections in 2005 and 2006 are more similar to each other than previous years’ collections, and this would be expected in a successful integrated hatchery supplementation program.

Since each of these collection years are generally composed of four-year-old fish, the differentiation among these collections for the most part is differentiation among specific cohorts. The slightly greater percentage of alleles shared among collections that are separated in time by multiples of four years, compared with collections that are not separated in time as such, suggests that cohort differences may be the most important factor accounting for differences in allele frequencies among collections.

#### **Task 4: Develop a model of genetic drift.**

See Task 3



**Task 5: Analyze spring Chinook population samples from the Chiwawa River and Chiwawa Hatchery from multiple generations.**

See Task 3

**Task 6: Analyze among population differences for upper Wenatchee spring Chinook.**

Supplementation of the Chiwawa River spring Chinook population may affect populations within the Wenatchee River watershed other than the Chiwawa River stock. If the stray rate for Chiwawa hatchery-origin fish is greater than that for natural-origin fish, an increase in gene flow from the Chiwawa population into other populations may result. If this gene flow is high enough, Chiwawa River fish may alter the genetic structure of these other populations. Records from field observations indicate that hatchery-origin fish are present in all major spawning aggregates (A.R Murdoch, unpublished data), and these fish are successfully reproducing (Blankenship et al 2006). The intent of this task is to investigate if there have been changes to the genetic structure of the spring Chinook stocks within upper Wenatchee tributaries during the past 15-20 years, and if changes have occurred, are they a function of the Chiwawa River Supplementation Program? Therefore, we ask the following two questions:

1. Are allele frequencies within populations in the upper Wenatchee stable through time? That is, is there significant allelic differentiation among collections within upper Wenatchee populations?
2. Are the recent collections from the upper Wenatchee populations more similar to the Chiwawa population than earlier collections from the same populations?

For this task we analyzed natural spawning collections from the White River (natural-origin), Little Wenatchee River (natural-origin), Nason Creek (natural-origin), and

Wenatchee mainstem (hatchery-origin), and hatchery collections from Leavenworth NFH and Entiat River NFH (Table 1). We also included in the analysis the natural- and hatchery-origin collections from the Chiwawa River. There are no repeated collections from Leavenworth, Entiat, Little Wenatchee, and Wenatchee mainstem (Table 1), so for many of the analyses we have limited our discussion to the Chiwawa River, White River, and Nason Creek collections. Furthermore, genetic structure of the Little Wenatchee collection, which consisted of only 19 samples, was unexpectedly quite different from the other collections. For example, the  $F_{ST}$  statistic measures the percent of total molecular variation that can be attributed to differences between populations. The median  $F_{ST}$  for all pairwise combinations of collections from all populations, except Little Wenatchee (33 populations, 528 individual  $F_{ST}$  statistics) equals 0.010 (1%), with a range of 0.000 to 0.037 (Table 6). The median  $F_{ST}$  for the Little Wenatchee paired with all other collections (33 individual  $F_{ST}$  statistics) equals 0.106 (10.6%), with a range of 0.074 to 0.121. The ten-fold increase in the  $F_{ST}$  statistic indicates that either the Little Wenatchee spring Chinook is unique among the upper Wenatchee River stocks, or this 1993 collection is somehow aberrant. Therefore, we exclude the Little Wenatchee collection from many other analyses.

**Population Differentiation** – Table 3 provides the levels of significance for all pairwise genic differentiation tests. Most between-collection comparisons are highly significant, with no pattern of increasing or decreasing differentiation with time, and no differences when comparisons are made with Chiwawa hatchery- versus Chiwawa natural-origin fish. For example, excluding the outlier 1996 and 1998 Chiwawa hatchery- and natural-origin collections, Nason Creek showed highly significant allele frequency differences between the Chiwawa hatchery- and natural-origin collections at 100% and 86% of the comparisons, respectively. The same comparisons with the White River produced 100% and 93% highly significant allele frequency comparisons, respectively. Allele frequencies between Nason Creek and White River were likewise differentiated from each other.

The collection allele frequencies within the upper Wenatchee system are significantly different, and these differences do not appear to change as a function of time (Table 3). Nason Creek shows greater within-population year-to-year variation in allele frequencies than does the White River, with 47% of the pairwise comparisons showing highly significant differences, compared with only 13% for the White River. However, the 2005 and 2006 collections from the White River appear to be somewhat more differentiated from not only each other, but from the earlier collections from the White River.

Despite the high degree of temporal and spatial structure suggested by the genic differentiation tests, as described above for within-Chiwawa analysis (Task 3), most of the genetic variation within this data set occurs within populations, rather than between populations (Table 6). The  $F_{ST}$  values for most population comparisons are between 0.01 and 0.02, indicating 1% to 2% among-population variance, with the remaining 98% to 99% variance occurring within populations. The White River shows the highest median  $F_{ST}$  among the natural-origin collections, equal to 0.014, compared with 0.009 for both the Nason Creek and Chiwawa natural-origin collections. The median  $F_{ST}$  for the Chiwawa hatchery-origin collections (0.012) was higher than that for the Chiwawa natural-origin collections.

Table 7 summarizes the information from the  $F_{ST}$  analyses, under five different temporal and spatial scenarios. Under all scenarios, over 99% of the molecular variance is within populations. There is significantly greater spatial structure among populations (“Origin”) in 2005 and 2006 than from 1989 to 1996. That is, there appears to be more spatial structure among the Chiwawa hatchery-origin, Chiwawa natural-origin, White River, and Nason Creek now, than in 1989 to 1996, despite the potential homogenizing and cumulative effect of hatchery strays. However, we stress that the amount of molecular variance associated with the among population differences, despite being significantly greater than 0.00%, is limited to only 0.43%.

**Allele-sharing and Nonmetric Multidimensional Scaling** – As in the Chiwawa River data discussed above, we constructed an allele-sharing distance matrix and then subjected

that matrix to a multidimensional scaling analysis (Figure 7). Consistent with all previously discussed multidimensional scaling analyses, the 1996 and 1998 adult, and the 1994 smolt collections are outliers. There is clear separation between the White River collections and all other natural-origin and Chiwawa hatchery-origin collections, indicating that there are more alleles shared among the Nason Creek and Chiwawa collections, than with the White River collections. Furthermore, there is a slight separation between the Chiwawa natural-origin natural spawner collections and Nason Creek collections, suggesting different groups of shared alleles between these populations. There is more variation in the allele-sharing distances among collections involved with the Chiwawa hatchery (origin or broodstock) than any of the natural-origin collections, even if we exclude the 1994, 1996, and 1998 collections. This suggests that there is more year-to-year variation in the composition of hatchery-origin and hatchery broodstock than within natural-origin populations throughout the upper Wenatchee. All Wenatchee mainstem fish are hatchery-origin, and if these fish are from the Chiwawa Supplementation Program (rather than from Leavenworth), it is not unexpected that this collection would be plotted within the Chiwawa polygon (Figure 7).

**Assignment of Individual to Populations** – Finally, we conducted individual assignment tests whereby we assigned each individual fish to a population, based on a procedure developed by Rannala and Mountain (1997) (Table 8 and 9). Individual fish may be correctly assigned to the population from which they were collected, or incorrectly assigned to a different population. Incorrect assignments may occur if the fish is an actual migrant (i.e., source population different from population where collected), or because the genotype for that fish matches more closely with a population different from its source. If there are many individuals from a population incorrectly assigned to populations other than its source population, that original population is either unreal (i.e., an admixture), or there is considerable gene flow between that population and other populations. Furthermore, in assigning individuals to populations, we can either accept the assignment with the highest probability, regardless of how low that probability may be, or we can establish a more stringent criterion, such as to not accept an assignment unless the posterior probability is equal to or greater than 0.90. This value is roughly

equal to having the likelihood of the most-likely population equal to 10 times that of the second most-likely population.

We provide a summary of the assignments in Tables 8 and 9. On average, nearly 50% of the fish are assigned incorrectly if we accept all assignments (Table 8), but the incorrect assignment rate drops to roughly 10% when we accept only those assignments with probabilities greater than 0.90. However, with this more stringent criterion, nearly 64% of the fish go unassigned. These results indicate that the allele frequency distributions for these populations are very similar, and it would be very difficult to assign an individual fish of unknown origin to the correct population. If all fish are assigned, there is a 50% chance, overall, of a correct assignment. If you accept only those assignment with the 0.90 criterion, nearly two-thirds of the fish would be unassigned, but there is a 90% chance of correctly assigning those fish that are indeed assigned.

Of all the populations in the data set, there are fewer errors associated with assigning fish to the White River. If all fish are assigned (Table 8), 72% of those fish assigned to the White River, are actually from the White River (115 fish out of a total of 159 fish assigned to the White River). This compares to a rate of only 52% and 53% for Nason Creek and Chiwawa natural-origin, respectively, and 60% for the Chiwawa hatchery-origin collections. With the 0.90 criterion (Table 9), 89% of the fish assigned to the White River, are actually from the White River, compared with 70% and 65% for Nason Creek and Chiwawa natural origin, respectively, and 81% for the Chiwawa hatchery origin.

When all fish are assigned, most of the incorrectly assigned fish from Nason Creek and White River are assigned to Chiwawa River, at roughly equal frequencies to the hatchery- and natural-origin populations. Incorrectly assigned fish to other populations occur at a slightly higher rate in Nason Creek than in the White River. However, when only those fish meeting the 0.90 criterion are assigned (Table 9), incorrectly assigned fish from Nason Creek are distributed among White and Chiwawa Rivers, as well as Leavenworth NFH, and the Entiat NFH. Mis-assignment to the Chiwawa hatchery-origin was the

highest among the Nason Creek collections, equal to nearly 14%. This contrasts with the White River where mis-assignments do not exceed 7% anywhere, and there is a roughly even distribution of mis-assignments among Nason Creek and Chiwawa River collections.

**Summary and Conclusions** – There is little geographic or temporal structure among populations within the upper Wenatchee systems. Among population molecular variance is limited to 1% or less. The little variance that can be attributed to among populations indicates that the White River is more differentiated from the Chiwawa and Nason populations than these populations are from each other. Furthermore, although we cannot rule out a hatchery effect on the Nason Creek and White River populations, there is no indication there has been any temporal changes in allele frequencies within these populations that can be attributed directly to the Chiwawa River Supplementation Program. In fact, Table 7 weakly suggests that there is more differentiation among these populations now, than there was before or at the early stages of Chiwawa supplementation.

Therefore, returning to our two original questions, there are significant differences in allele frequencies among collections within populations, and among populations within the upper Wenatchee spring Chinook stocks. However, these differences account for a very small portion of the overall molecular variance, and these populations overall are very similar to each other. There is no evidence that the Chiwawa River Supplementation Program has changed the allele frequencies in the Nason Creek and White River populations, despite the presence of hatchery-origin fish in both these systems. Finally, of all the populations within the Wenatchee River, the White River appears to be the most distinct. Yet, this distinction is more a matter of detail than of large significance, as the median  $F_{ST}$  between White River collections and all other collections (except the Little Wenatchee) is less than 1.5% among population variance.

**Task 7: Calculate the inbreeding effective population size using demographic data for each sample year, and document the ratio of census to effective size.**

This analysis was completed by Williamson et al. (submitted).

**Task 8: Calculate LD  $N_b$  using genetic data for each sample year, and document the ratio of census to effective size.**

We report  $N_e$  estimated for the Chiwawa River collections based on the bias correction method of Waples (2006) implemented in LDNe (Do and Waples unpublished).  $N_e$  estimates based on LD are best interpreted as the effective number of breeders ( $N_b$ ) that produced the sample (Waples 2006).

For collections categorized by spawning location (i.e., hatchery broodstock or natural), estimates of  $N_b$  are shown in Table 10. Considering the hatchery broodstock,  $N_b$  estimates range from 30.4 (1996) to 274.3 (2005). To obtain  $N_e/N$  ratios, the  $N_b$  estimate is multiplied by four (i.e., mean generation length) and divided by the total in river (i.e., NOS [natural-origin spawners] plus HOS [hatchery-origin spawners]) census data from four years prior (i.e., major cohort; see Table 2). The observed  $N_e/N$  ratios for the broodstock collections range from 11% to 54% of the census estimate, excluding the 2000 collection which is 106%. A ratio greater than one is possible under special circumstances, and certain artificial mating schemes within hatcheries can inflate  $N_e$  above  $N$ ; yet, it is unknown if this is the case for this collection. While no direct comparisons are possible, the  $N_b$  estimates reported by Williamson et al. (submitted) for Chiwawa broodstock collections from 2000 – 2003 are similar in magnitude to our estimates. For Chiwawa natural spawner collections, the  $N_b$  estimates range from 5.2 (1989) to 231.5 (2005), with observed  $N_e/N$  ratios of 22% - 48% of the census estimate.

**Task 9: Calculate  $N_b$  using the temporal method for multiple samples from the same location.**

Estimates of effective number of breeders ( $N_b$ ) derived from Waples' (1990) temporal method are shown in Tables 11-13. Eight collection years were used for the Chiwawa broodstock collections (Table 11). The harmonic mean of all pairwise estimates of  $N_b$  ( $\tilde{N}_b$ ) was 269.4. This estimate is the contemporary  $N_e$  for Chiwawa broodstock collections. For the five collection years of Chiwawa in-river spawners (Table 12), the estimated  $\tilde{N}_b = 224.2$ . This estimate is the contemporary  $N_e$  for Chiwawa River natural spawner collections. Since the Chiwawa Supplementation Program is integrated by design, we also performed another estimation of  $N_e$  using composite hatchery and natural samples. There are paired samples from 2004-2006. We combined genetic data for hatchery (HOS) and natural (NOS) origin fish from 2004 – 2006 to create a single Chiwawa River natural spawner sample for each year. The three composite samples from 2004 – 2006 were then analyzed using the temporal method (Table 13), resulting in a  $\tilde{N}_b = 386.8$ . This estimate is the contemporary  $N_e$  for Chiwawa River.

Williamson et al. (submitted) estimated  $N_e$  using Waples' (1990) temporal method for Chinook captured in 2004 and 2005, and used age data to decompose brood years into consecutive cohorts from 2000 – 2003. They report for Chiwawa broodstock a  $\tilde{N}_b = 50.4$ . This estimate is not similar to our Chiwawa broodstock estimate. However, if we analyze the hatchery-origin Chinook only, our estimate is  $\tilde{N}_b = 80.1$  for collection years 1989 – 2006 (data not shown). Williamson et al. (submitted) report for Chiwawa naturally spawning Chinook a  $\tilde{N}_b = 242.7$ , which is slightly higher than our estimate for in-river spawners from 1989 – 2006, but lower than our estimate from combined NOS and HOS Chinook from 2004 – 2006 collection years.



**Task 10: Use available data and the Ryman-Laikre and Wang-Ryman models to determine the expected change of  $N_e$  for natural spring Chinook salmon in the Wenatchee River due to hatchery operation.**

$N_e$  is generally thought to be between 0.10 and 0.33 of the estimated census size (Bartley et al. 1992; RS Waples pers. comm.). We used this range to generate an estimate of  $N_e$  for Chiwawa natural spawners prior to hatchery operation. For brood years 1989 – 1992, the arithmetic mean census size was  $N=962.7$  (Table 2), resulting in an estimated  $N_e$  ranging from 96.3 – 317.7. The contemporary estimate of  $N_e$  calculated using genetic data for the Chiwawa in-river spawners is  $N_e=224.2$  (Table 12), falling in the middle of the pre-hatchery range. The  $N_e/N$  ratio calculated using 224.2 and the arithmetic census of NOS Chinook from 1989 – 2005 is 0.42. A more appropriate contemporary  $N_e$  to compare with the pre-hatchery estimate (i.e., 96.3 – 317.7) is the combined NOS and HOS estimate from natural spawners, since the supplementation program is integrated. As discussed above, the contemporary estimate of  $N_e$  calculated using genetic data for Chiwawa NOS and HOS Chinook is  $N_e=386.8$  (Table 13), which is slightly larger than the pre-hatchery range, suggesting the  $N_e$  has not declined during the period of hatchery operation. The  $N_e/N$  ratio calculated using 386.8 and the arithmetic census of NOS and HOS Chinook from 1989 – 2005 is 0.40. These results suggest the Chiwawa Hatchery Supplementation Program has not resulted in a smaller  $N_e$  for the natural spawners from the Chiwawa River.

Williamson et al. (submitted) argued that since their combined (i.e., broodstock and natural)  $N_e$  estimate was lower than the naturally spawning estimate, the supplementation program likely had a negative impact on the Chiwawa River  $N_e$ . We disagree with this interpretation of these data. Since the natural spawning component is mixed hatchery and natural ancestry, the  $N_e$  estimates from natural spawning data are the results that bear on possible hatchery impacts. The census data show the population declined in the mid 1990's and rebounded by 2000 (Table 2). This trend is reflected in the  $N_e$  results, as shown above, and Williamson et al. (submitted) clearly show in their Table 4 the  $N_e$  was lower in 2000 ( $N_e = 989$ ) than it was in 1992 ( $N_e = 2683$ ). Yet, the important comparison

they make in our view was the natural spawning  $N_e$  versus the natural only component  $N_e$  (i.e., hypothetically excluding hatchery program). Williamson et al. (submitted) report the 1989 – 1992  $N_e$  estimated from naturally spawning Chinook (i.e., NOS and HOS integrated) was essentially the same as the natural only component estimate, 2683 and 2776, respectively. This result is not surprising since no HOS fish were present between 1989 – 1992. They also report that the 1997 – 2000  $N_e$  estimated from naturally spawning Chinook (i.e., NOS and HOS integrated) was  $N_e = 989$ , while the natural-origin estimate of  $N_e$  in 1997 – 2000 was  $N_e = 629$ . Since the natural-origin estimate of 629 is lower than 989, the  $N_e$  estimate from all in-river spawners, we argue that their analysis of demographic data show the  $N_e$  estimated from naturally spawning Chinook (i.e., NOS and HOS integrated) is larger only if the hatchery Chinook in the river are ignored.

**Task 11: Use individual assignment methods to determine the power of self-assignment for upper Wenatchee River tributaries.**

See “Assignment of Individual to Populations” in Task 6

## **Conclusions**

Has the Chiwawa Hatchery Supplementation Program succeeded at increasing the census size of the target population while leaving genetic integrity intact? This is an important question, as hatcheries can impact natural populations by reducing overall genetic diversity (Ryman and Laikre 1991), reducing the fitness of the natural populations through relaxation of selection or inadvertent positive selection of traits advantageous in the hatchery (Ford 2002; Lynch and O’Hely 2001), and by reducing the reproductive success of natural populations (McLean et al. 2003). The census data presented here show that the current natural spawning census size is similar to the pre-supplementation census size. Despite large numbers of hatchery-origin fish on the Chiwawa River spawning grounds, the genetic diversity of the natural-origin collections appear unaffected by the supplementation program; heterozygosities are high, and contemporary  $N_e$  is similar (perhaps slightly higher) than pre-supplementation  $N_e$ . We did find

significant year-to-year differences in allele frequencies in both the origin and spawner datasets, but these differences do not appear to be related to fish origin, spawning area, or genetic drift. However, we do suggest that cohort differences may be the most important factor accounting for differences in allele frequencies among collections.

The main objective of this study was to determine the potential impacts of the hatchery program on natural spring Chinook in the upper Wenatchee system. We did this by analyzing temporally replicated collections from the Chiwawa River, and by comparing genetic diversity prior to the presumed effect of the Chiwawa Hatchery Supplementation Program, with contemporary collections. We report that the genetic diversity present in the Chiwawa River is unchanged (allowing for differences among cohorts) from 1989 – 2006, and the contemporary estimate of the effective population size ( $N_e$ ) using genetic data is approximately the same as the  $N_e$  estimate extrapolated from 1989 – 1992 census data (i.e., pre-hatchery collection years). We observed substantial genetic diversity, with heterozygosities ~80% over thirteen microsatellite markers. Yet, temporal variation in allele frequencies was the norm among temporal collections from the same populations (i.e., location). The genetic differentiation of replicated collections from the same population is likely the result of salmon life history in this area, as four-year-old Chinook comprise a majority of returns each year. The genetic tests are detecting the differences of contributing parents for each cohort. An important point related to the temporal variation, is that the hatchery broodstock is composed in part of the natural origin Chinook from the Chiwawa River. When we compared the genetic data (within a collection year) for Chinook brought into the hatchery as broodstock with the Chinook that remained in the river (years 2001, 2004 – 2006), there was a trend of decreasing statistical differences in allele frequencies from 2001 to 2004, and no differences were detected for 2005 and 2006. While the replicated collections may have detectable differences in allele frequencies, those differences reflect actual differences in cohorts, not the result of hatchery operations, and the hatchery broodstock collection method captures the differences in returning Chiwawa River spring adults each year. We conclude from these results that the genetic diversity of natural spring Chiwawa Chinook has been maintained during the Chiwawa Hatchery Supplementation Program.

We observe slight, but statistically significant population differentiation between Chiwawa River, White River, and Nason Creek collections. Murdoch et al (2006) and Williamson et al. (submitted) also observed population differentiation between Chiwawa River, White River, and Nason Creek collections. Yet, 99.3% of the genetic variation observed was within samples, very little variance could be attributed to population differences (i.e., population structure). The AMOVA analysis and poor individual assignment results suggest the occurrence of gene flow among Wenatchee River locations or a very recent divergence of these groups. While Murdoch et al. 2006 did not perform an AMOVA analysis, their  $F_{ST}$  results provide comparable data to our among-population results. Murdoch et al. 2006 report  $F_{ST}$  ranging from 2%-3% for pairwise comparisons between of Chiwawa, White, and Nason River collections. Since  $F_{ST}$  is an estimate of among-sample variance, these results also imply a majority of the genetic variance (i.e., 97%-98%) resides within collections. To provide further context for the magnitude of these variance estimates, we present the among-group data from Murdoch et al. 2006 comparing summer-run and spring-run Chinook from the Wenatchee River. They report that approximately 91% of observed genetic variance is within-collection for comparisons between collections of summer- and spring-run Chinook. Ultimately, the information provided by this and other reports will be incorporated into the management process for Wenatchee River Chinook. However, we would like to emphasize that the application of these genetic data to management is more about the goals related to the distribution of genetic diversity in the future than specific data values reported. If Chinook are collected at Tumwater Dam instead of within the upper Wenatchee River tributaries, a vast majority of the genetic variation present in the basin would be captured, although any differences among tributaries would be mixed. Alternatively, management policies could be crafted to promote and maintain the among-group genetic diversity that genetic studies consistently observe to be non-zero within the Wenatchee River.

We agree with Murdoch et al. (2006) that it appears hatchery Chinook are not contributing to reproduction in proportion to their abundance. Additionally, if the total census size (i.e., NOS and HOS combined) within the Chiwawa River does not continue

to increase, genetic diversity may decline within this system, given the smaller  $N_e$  within the hatchery-origin collections compared with the natural-origin collections.

## **Acknowledgements**

We would like to thank Denise Hawkins, Craig Busack, and Cheryl Dean for helpful comments regarding this project. This project was funded by Chelan County PUD and the Washington State General Fund.

## Literature Cited

- Araki H, Waples RS, Ardren WR, Cooper B, Blouin MS (2007) Effective population size of steelhead trout: influence of variance in reproductive success, hatchery programs, and genetic compensation between life-history forms. *Molecular Ecology*, **16**:953-966.
- Arden WR and Kapuscinski AR (2003) Demographic and genetic estimates of effective population size ( $N_e$ ) reveals genetic compensation in steelhead trout. *Molecular Ecology* 12: 35-49
- Banks MA, Blouin MS, Baldwin BA, Rashbrook VK, Fitzgerald HA, Blankenship SM, Hedgecock D (1999) Isolation and inheritance of novel microsatellites in chinook salmon (*Oncorhynchus tshawytscha*). *Journal of Heredity*, **90**:281-288.
- Banks MA, Rashbrook VK, Calavetta MJ et al (2000) Analysis of microsatellite DNA resolves genetic structure and diversity of chinook salmon (*Oncorhynchus tshawytscha*) in California's Central Valley. *Canadian Journal of Fisheries and Aquatic Sciences* 57:915-927.
- Bartley D, Bentley B, Brodziak J, Gomulkiewicz R, Mangel M, and Gall GAE (1992) Geographic variation in population genetic structure of chinook salmon from California and Oregon. *Fish. Bull., U.S.* 90:77-100.
- Blankenship SM, Von Bargaen J, and Truscott KD (2006) Genetic analysis of White River juveniles retained for captive brood at AquaSeed to assess the hatchery status of contributing parents. Developed for Grant County PUD.
- Blankenship SM and Murdoch AR (2006) Study Plan For Assessing the Genetic Diversity of Natural Chiwawa River Spring Chinook Salmon And Evaluating The Effectiveness Of Its Supportive Hatchery Supplementation Program. Developed for Chelan County PUD and the Habitat Conservation Plan's Hatchery Committee.
- Bugert R (1998) Mechanics of supplementation in the Columbia River. *Fisheries* 23:11-20.
- Cairney M, Taggart JB, Hoyheim B (2000) Characterization of microsatellite and minisatellite loci in Atlantic salmon (*Salmo salar* L.) and cross-species amplification in other salmonids. *Mol Ecol*, **9**:2175-2178.
- Campton DE (1987) Natural hybridisation and introgression in fishes: methods of detection and genetic interpretations. In: Population genetics and fisheries management. (Eds. Ryman, N. and Utter, F.), pp. 161-192. Washington Sea Grant Program, University of Washington Press, Seattle, USA.

- Chapman D, Giorgi A, Hillman T, Deppert D, Erho M, Hays S, Peven C, Suzumoto B, and Klinge R (1994) Status of summer/fall chinook salmon in the mid-Columbia region. Report for Chelan, Douglas, and Grant County PUDs. 412 p. + app. (Available from Don Chapman Consultants, 3653 Rickenbacker, Ste. 200, Boise, ID 83705.)
- Chapman D, Peven C, Giorgi A, Hillman T, and Utter F (1995) Status of spring chinook salmon in the mid-Columbia River. Don Chapman Consultants, Inc., 477 p. (Available from Don Chapman Consultants, 3653 Rickenbacker, Ste. 200, Boise, ID 83705.)
- Cockerham CC (1969) Variance of gene frequencies. *Evolution* 23:72-83.
- Craig JA, and Suomela (1941) Time of appearance of the runs of salmon and steelhead trout native to the Wenatchee, Entiat, Methow, and Okanogan rivers. U.S. Fish Wildl. Serv.
- Fish FF, and Hanavan MG (1948) A report on the Grand Coulee Fish Maintenance. Project 1939-1947. U.S. Fish Wildl. Serv. Spec. Sci. Rep 55.
- Ford MJ (2002) Selection in captivity during supportive breeding may reduce fitness in the wild. *Conservation Biology* 16(3):815-825
- Frankham R, Ballou JD, Briscoe DA (2002). *Introduction to Conservation Genetics*, Cambridge University Press, Cambridge, UK.
- Greig C, Jacobson DP, Banks MA (2003) New tetranucleotide microsatellites for fine-scale discrimination among endangered chinook salmon (*Oncorhynchus tshawytscha*). *Mol Ecol Notes*, **3**:376-379.
- Heath DD, Busch C, Kelly J, and Atagi DY (2002) Temporal change in genetic structure and effective population size in steelhead trout (*Oncorhynchus mykiss*). *Molecular Ecology* 11:197-214
- Hedrick P, Hedgecock D (1994) Effective population size in winter-run chinook salmon. *Conservation Biology*, **8**:890-892.
- Hill WG (1981) Estimation of effective size from data on linkage disequilibrium. *Genetical Research* 38: 209-216.
- Jensen LF, Hansen MM, Carlsson J et al (2005) Spatial and temporal genetic differentiation and effective population size of brown trout (*Salmo trutta*, L.) in small Danish rivers. *Conservation Genetics* 6:615-621.
- Liu, K and Muse SV (2005) PowerMarker: Integrated analysis environment for genetic marker data. *Bioinformatics* 21:2128-2129.

- Lynch M and O'Hely M (2001) Captive breeding and the genetic fitness of natural populations. *Conservation Genetics* 2:363-378
- Manly, BFJ. (1986) *Multivariate Statistical Methods. A Primer*. Chapman and Hall. London.. 159 + x pp.
- McLean JE, Bentzen P, Quinn TP (2003) Differential reproductive success of sympatric, naturally spawning hatchery and wild steelhead trout (*Oncorhynchus mykiss*) through the adult stage. *Can. J. Fish. Aquat. Sci.*, 60:433–440.
- Mullan, JW (1987) Status and propagation of chinook salmon in the mid-Columbia River through 1985. U.S. Fish Wildl. Serv. Biol. Rep. 87:111.
- Murdoch AR and Peven C (2005) *Conceptual Approach to Monitoring and Evaluating the Chelan County Public Utility District Hatchery Programs, Final Report*.
- Murdoch AR, Pearsons TN, Maitland TW, Ford M, and Williamson K (2006) Monitoring the reproductive success of naturally spawning hatchery and natural spring Chinook salmon in the Wenatchee River. BPA Project No. 2003-039-00, Contract No. 00021391. pp. 96
- Nelson WR, and Bodle J (1990) Ninety years of salmon culture at the Little White Salmon National Fish Hatchery. U.S. Fish Wildl. Serv. Biol. Rep. 90:22.
- Palm S, Laikre L, Jorde PE, et al (2003) Effective population size and temporal genetic change in stream resident brown trout (*Salmo trutta, L.*). *Conservation Genetics* 4:249-264.
- Olsen JB, Bentzen P, Seeb JS (1998) Characterization of seven microsatellite loci derived from pink salmon. *Molecular Ecology*, 7:1087-1089
- Rannala B, Mountain JL (1997) Detecting immigration by using multilocus genotypes. *Proceedings of the National Academy of Sciences* 94:9197-9201.
- Rexroad CE, Coleman RL, Martin AM, Hershberger WK, Killefer J (2001) Thirty-five polymorphic microsatellite markers for rainbow trout (*Oncorhynchus mykiss*). *Animal Genetics*, 32:317-319.
- Rohlf, F. J. (2002) *NTSYSpc: Numerical Taxonomy System*, ver. 2.1. Exeter Publishing, Ltd.
- Ryman N, Laikre L (1991) Effects of supportive breeding on the genetically effective population size. *Conservation Biology*, 5:325-329.



- Seeb L, et al. (in review) Development of a Standardized DNA Database for Chinook Salmon. *Fisheries*
- Schneider S, Roessli D, Excoffier L (2000) Arlequin ver 2.000: A software for population genetic data analysis. Genetics and Biometry Laboratory. University of Geneva, Switzerland.
- The Mathworks (2006) MatLab Release R2006b. Massachusetts.
- Utter FM, Chapman DW, and Marshall AR (1995) Genetic population structure and history of chinook salmon of the Upper Columbia River. *Am. Fish. Soc. Symp.* 17:149-165.
- Wang J (2005) Estimation of effective size from data on genetic markers. *Trans. Royal. Phil. Soc. B* 360: 1395-1409.
- Wang J, and Ryman N (2001) Genetic effects of multiple generations of supportive breeding. *Conservation Biology* 15: 1615-1631.
- Wang J, Whitlock MC (2003) Estimating Effective Population Size and Migration Rates From Genetic Samples Over Space and Time. *Genetics* 163:429-446
- Waples RS (1989) A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics*, 121:379-391.
- Waples RS (1990) Conservation genetics of Pacific salmon. III. Estimating effective population size. *J. Hered.* 81: 277-289.
- Waples RS (1991) Genetic interactions between hatchery and wild salmonids: Lessons from the Pacific Northwest. *Can. J. Fish. Aquat. Sci.* 48(Suppl. 1):124-133.
- Waples RS (2005) Genetic estimates of contemporary effective population size: to what time periods do the estimates apply? *Molecular Ecology*, 14:3335-3352
- Waples RS (2006) A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. *Conservation Genetics* 7:167-184
- Washington Department of Fisheries (WDF). 1934. Forty-second and forty-fifth inclusive annual reports of the State Department of Fisheries for the period from April 1, 1931-March 31, 1935, fiscal years of 1931 to 1934 inclusive. *Wash. Dep. Fish.*, pp. 78
- Williamson K, Cordes J, May B (2002) Characterization of microsatellite loci in chinook salmon (*Oncorhynchus tshawytscha*) and cross-species amplification in other salmonids. *Molecular Ecology Notes*, 2:17-19.

Williamson KS, Murdoch AR, and Ford MJ (submitted) Influence of supportive breeding on genetic diversity of hatchery and natural Wenatchee River spring Chinook salmon.

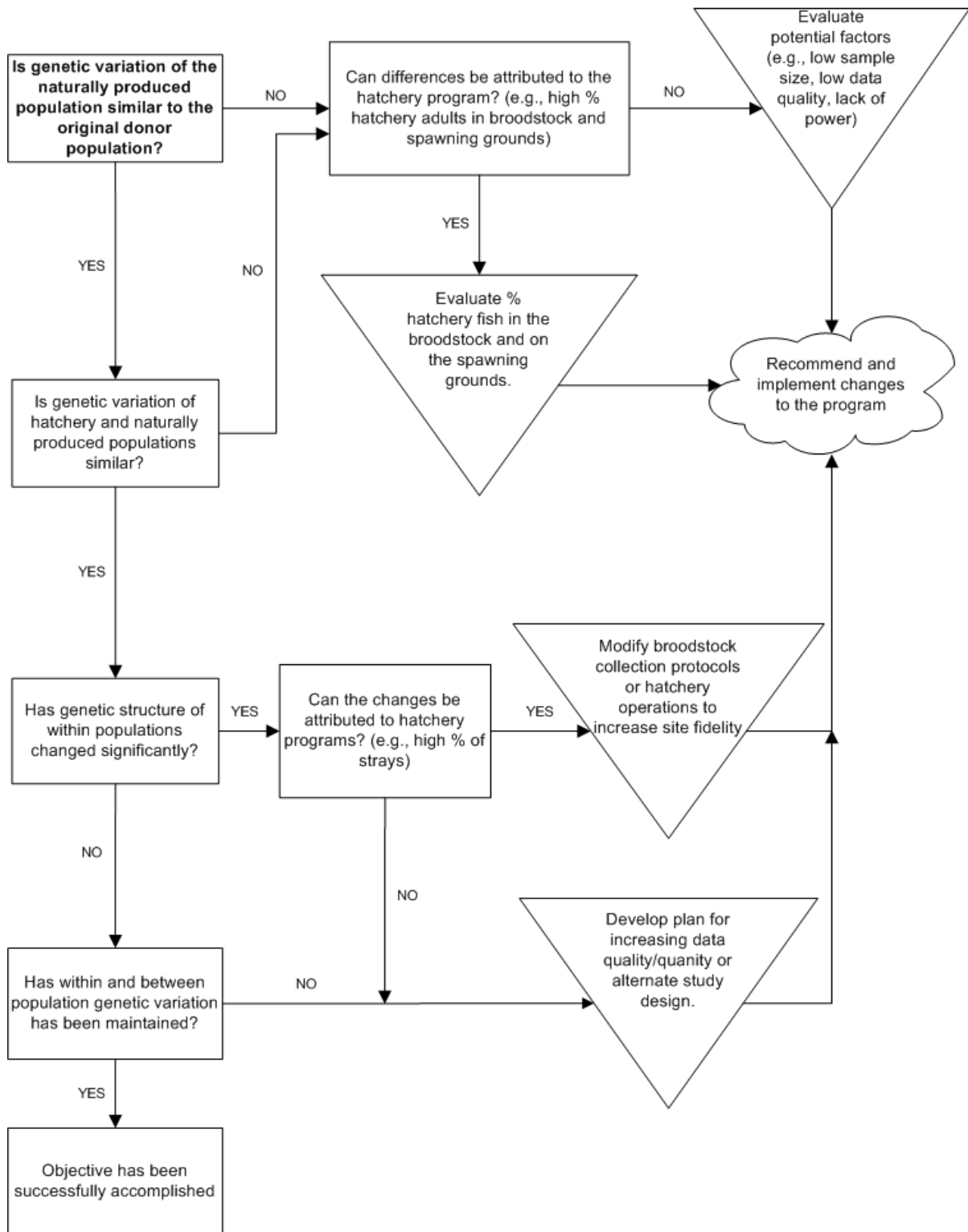


Figure 1. Conceptual process for evaluating potential changes in genetic variation in the Chiwawa naturally produced populations as a result of the supplementation hatchery programs (From Murdoch and Pevan 2005).

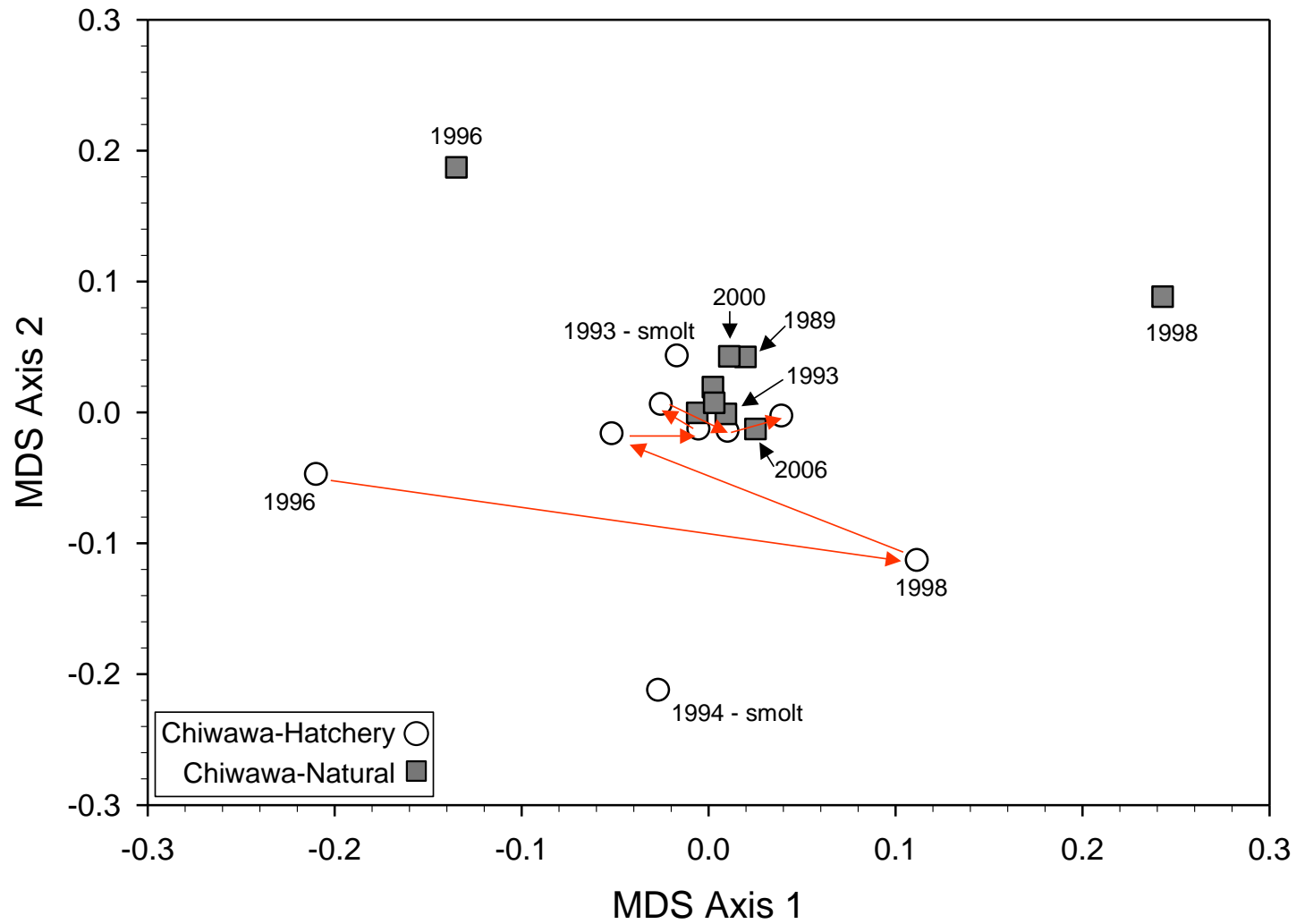


Figure 2. Multidimensional scaling plot from an allele-sharing distance matrix calculated from the Chiwawa data set organized by fish origin (i.e., hatchery versus natural). The red arrows connect consecutive hatchery-origin collections starting with the first adult collection (1996) and ending with the 2006 collection (see Table 1 for collection years).

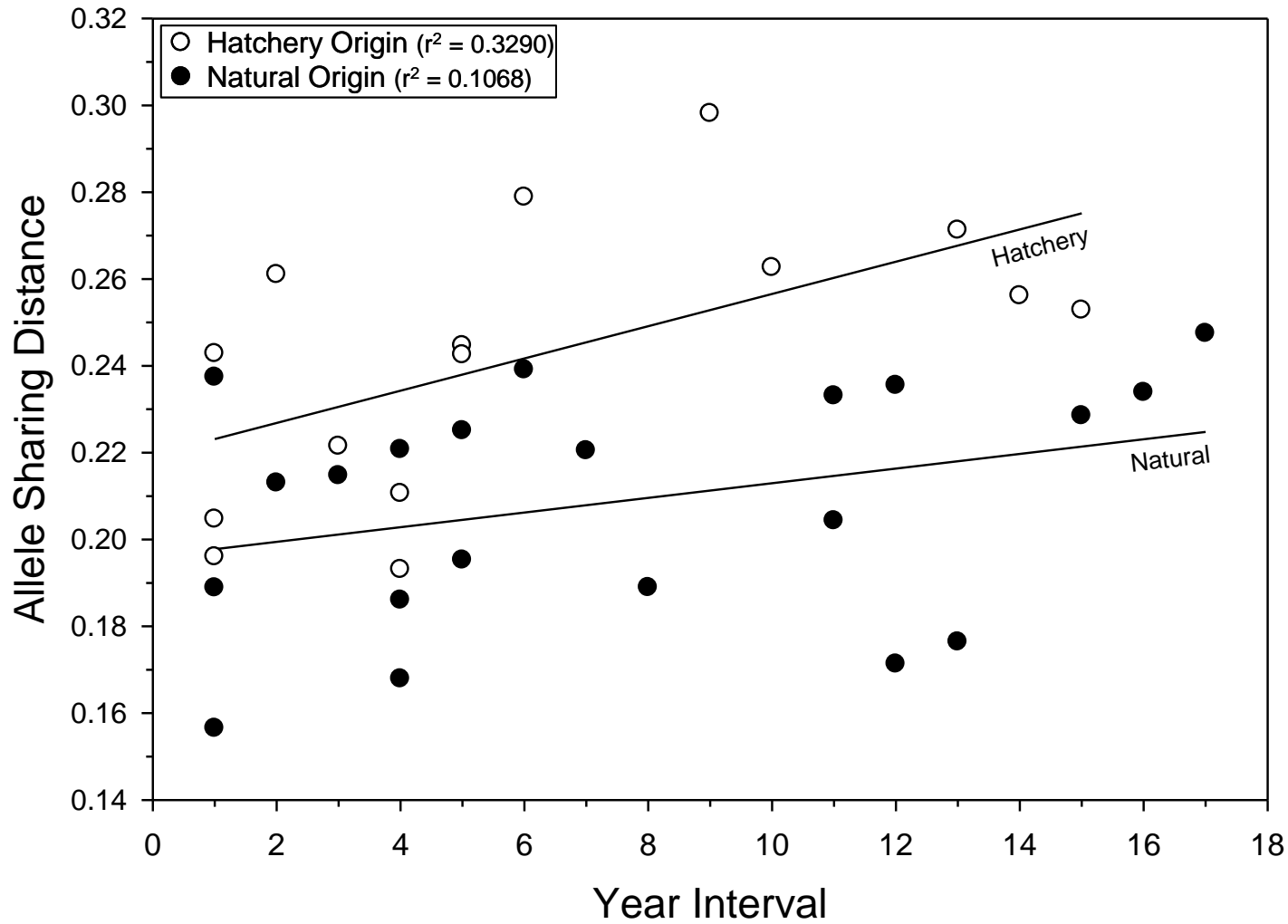


Figure 3. Relationships between the time interval in years and allele sharing distances, with each circle representing the pairwise relationship between two Chiwawa collections. Separate regression lines for the natural- and hatchery-origin collections. The slope for the natural-origin collection is not significantly different from zero ( $p=0.1483$ ), while the slope for hatchery-origin collection is significantly greater than zero ( $p=0.0254$ ) indicating a positive relationship between time interval and allele sharing distance.

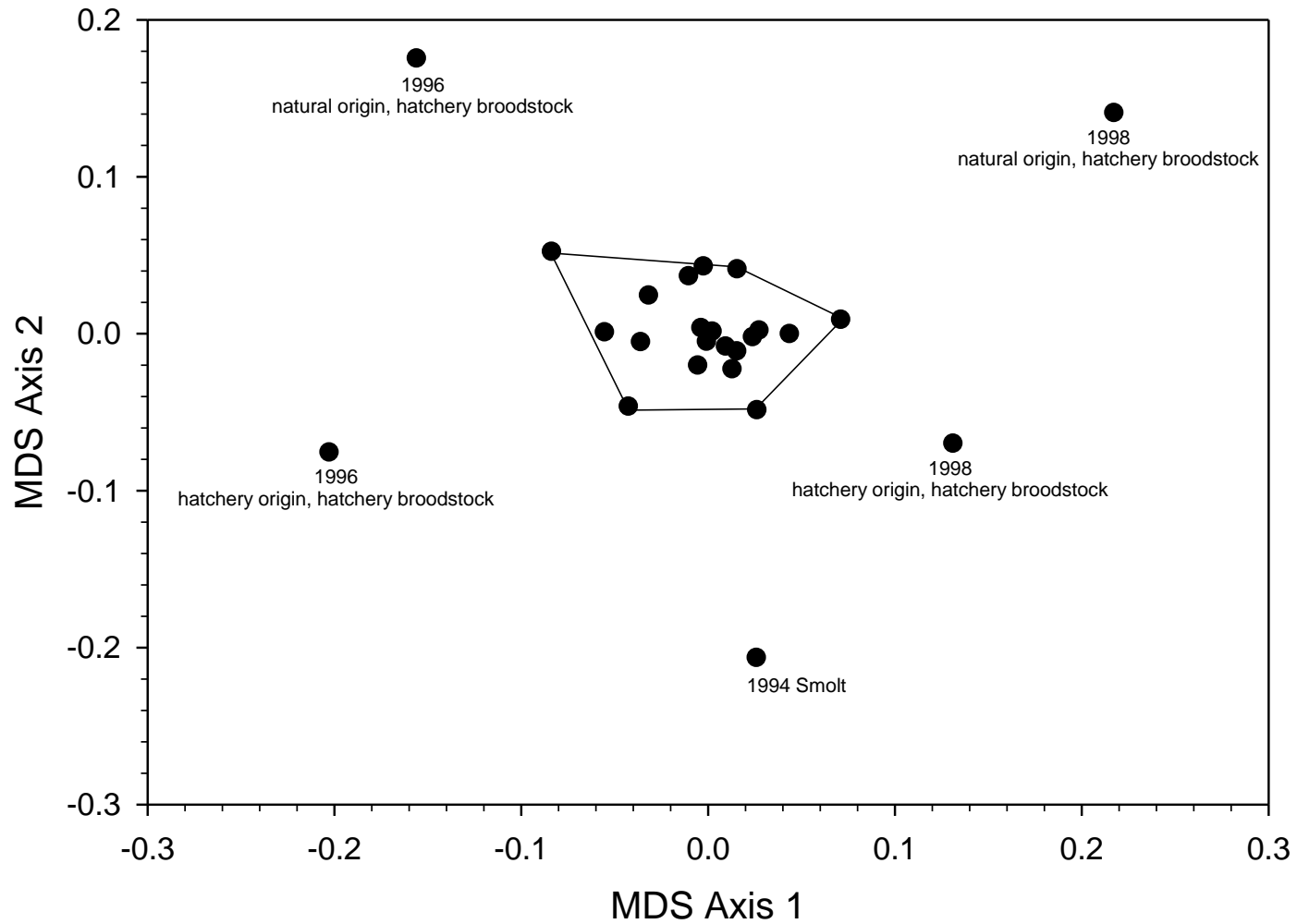


Figure 4. Multidimensional scaling plot from an allele-sharing distance matrix calculated from the Chiwawa data set organized by four treatment groups, as discussed in the text. Each circle represents a single collection within each of the four treatment groups, and the polygon encloses all groups that are not outliers. Each outlier group is specifically labeled.

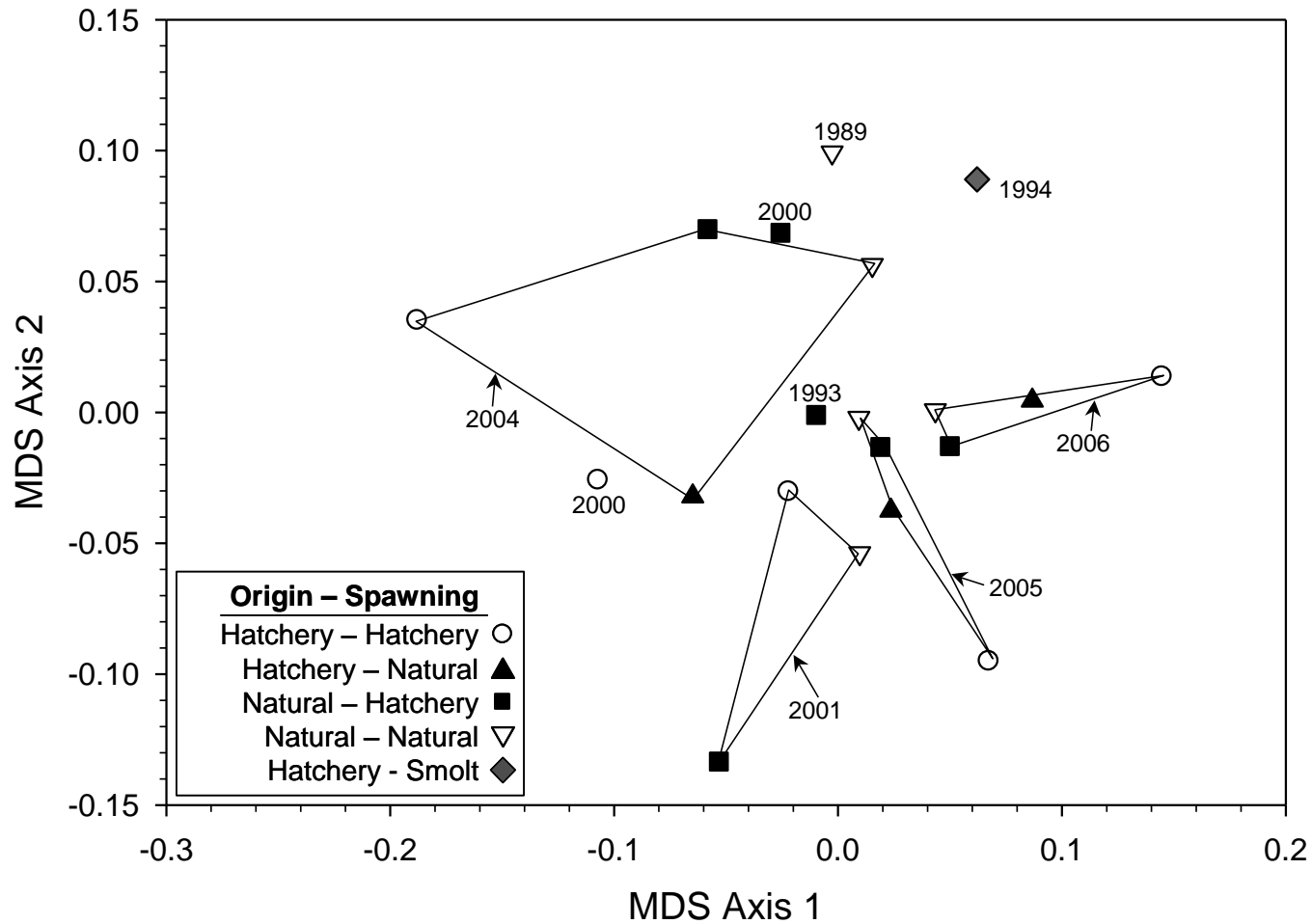


Figure 5. As in Figure 4, but allele-sharing distance matrix recalculated without the five outlier groups shown in Figure 4. Polygons group together treatment groups from the same collection year. Dates associated with symbols also refer to collection year. Collection years 2004-2006 included all four treatment groups, while collection year 2001 did not include a hatchery-origin natural spawner group. Legend is read as follows: Open circles refer to hatchery-origin hatchery spawner group, while filled box refers to natural-origin hatchery spawner group, and so on.

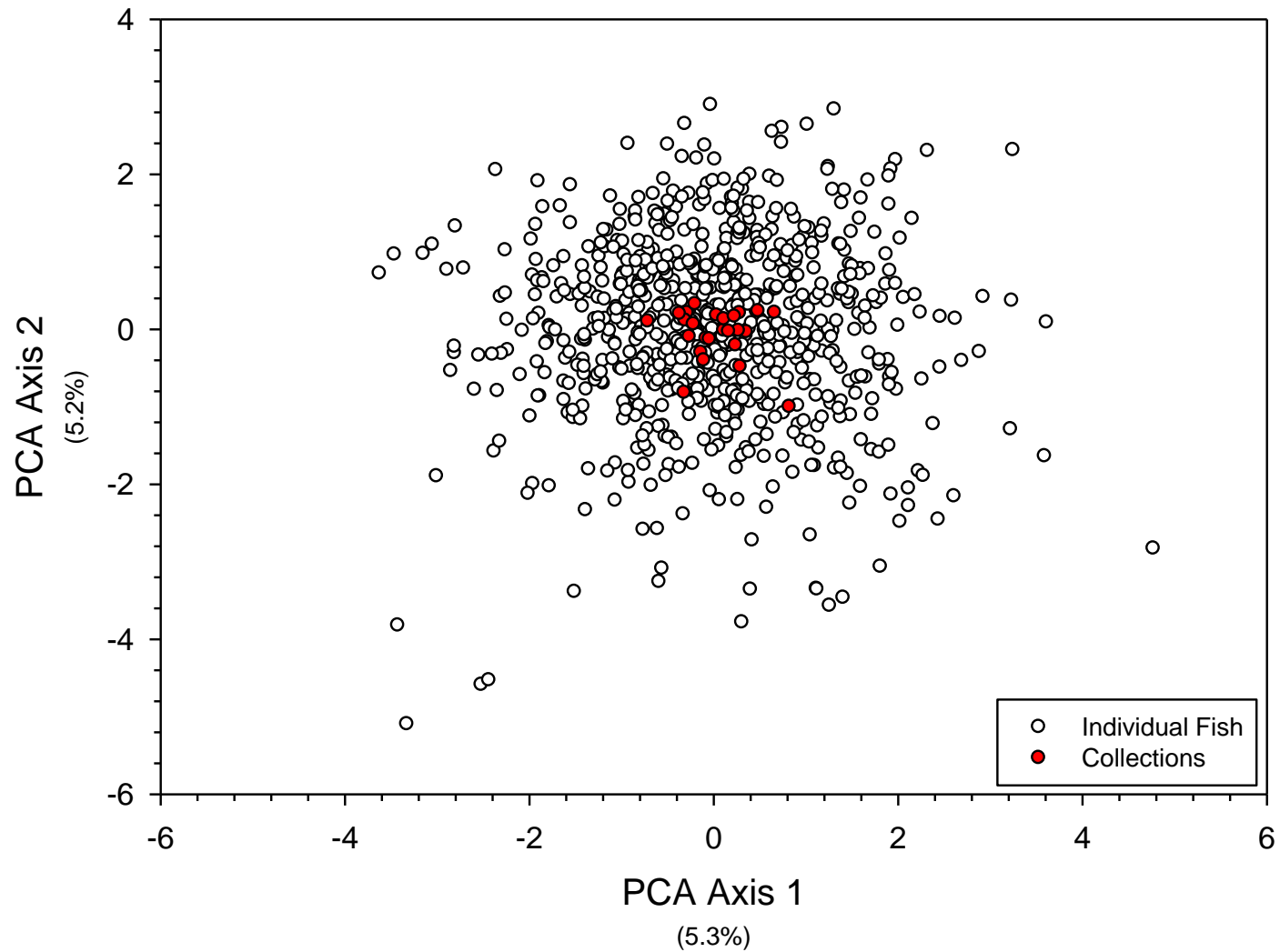


Figure 6. Principal component (PC) analysis of individual fish from the Chiwawa River. Only fish with complete microsatellite genotypes were included in the analysis ( $n = 757$ ). Open circles are the PC scores for individual fish, and the filled circles are the centroids (bivariate means) for each of the 25 groups discussed in the text. PC axes 1 and 2 account for only 10.5% of the total molecular variance.



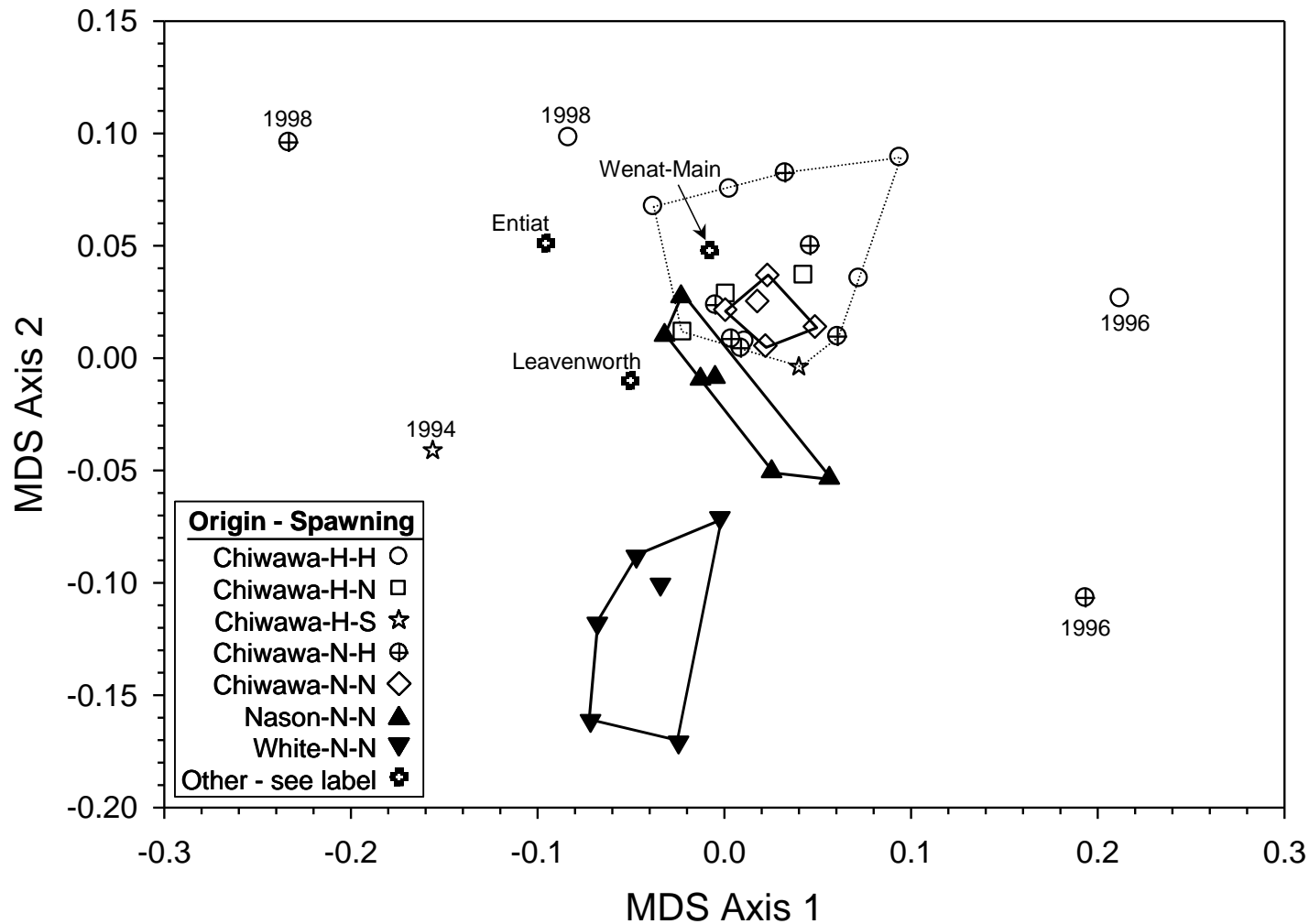


Figure 7. Multidimensional scaling plot from an allele-sharing distance matrix calculated from the Chiwawa origin data set and all other non-Chiwawa collections, except Little Wenatchee River. Legend is read with abbreviations beginning with origin and then spawning location. H=hatchery, N=natural, and S=smolts. Polygons with solid lines enclose the natural-origin natural spawner collections from each population (i.e., river). The polygon with the dotted lines enclose all Chiwawa collections, except for the five outlier collections, as discussed in text.



**Table 1** Summary of within population genetic data. Chiwawa collection data are summarized in A) by origin of the sample (i.e., clipped vs. non-clipped). All collection data are summarized in B) by spawning location (i.e., hatchery broodstock or on spawning grounds). Hz is heterozygosity, HWE is the statistical significance of deviations from Hardy-Weinberg expectations (\* = 0.05, \*\* = 0.01, and \*\*\* = 0.001), LD is the proportion of pairwise locus tests (across all populations) exhibiting linkage disequilibrium (bolded values are statistically significant), and the last column is mean number of alleles per locus.

Collection	Sample size	Gene Diversity	Observed Hz	HWE	F <sub>IS</sub>	LD	Mean # Alleles
A) Origin							
1993 Chiwawa Hatchery	95	0.77	0.79	***	-0.02	<b>0.86</b>	14.00
1994 Chiwawa Hatchery	95	0.76	0.77	***	-0.01	<b>0.91</b>	11.38
1996 Chiwawa Hatchery	8	0.75	0.81	-	-0.01	0.00	8.23
1998 Chiwawa Hatchery	27	0.81	0.82	-	0.00	0.04	12.62
2000 Chiwawa Hatchery	43	0.75	0.78	***	-0.01	<b>0.19</b>	12.46
2001 Chiwawa Hatchery	69	0.77	0.80	***	-0.02	<b>0.14</b>	15.31
2004 Chiwawa Hatchery	72	0.77	0.77	***	0.01	<b>0.45</b>	15.92
2005 Chiwawa Hatchery	91	0.79	0.82	*	-0.03	<b>0.05</b>	16.15
2006 Chiwawa Hatchery	95	0.80	0.84	***	-0.05	<b>0.49</b>	15.85
1989 Chiwawa Natural	36	0.76	0.78	-	0.01	0.00	12.77
1993 Chiwawa Natural	62	0.78	0.81	-	-0.02	0.04	15.85
1996 Chiwawa Natural	8	0.72	0.78	-	-0.02	0.00	7.54
1998 Chiwawa Natural	10	0.78	0.84	-	0.00	0.00	8.23
2000 Chiwawa Natural	39	0.78	0.79	***	0.00	<b>0.10</b>	14.00
2001 Chiwawa Natural	75	0.78	0.80	-	-0.03	0.03	15.31
2004 Chiwawa Natural	85	0.78	0.77	-	0.02	0.01	15.77
2005 Chiwawa Natural	90	0.79	0.79	-	0.01	0.01	16.15
2006 Chiwawa Natural	96	0.80	0.81	-	-0.01	0.01	16.46

**Table 1** Within population genetic data analysis summary continued.

Collection	Sample size	Gene Diversity	Observed Hz	HW	F <sub>IS</sub>	LD	Mean # Alleles
B) Spawning Location							
1993 Chiwawa Broodstock	62	0.78	0.81	-	-0.02	0.00	15.85
1996 Chiwawa Broodstock	16	0.75	0.79	-	-0.02	0.00	10.92
1998 Chiwawa Broodstock	37	0.82	0.83	-	0.00	0.01	14.38
2000 Chiwawa Broodstock	82	0.78	0.78	***	0.00	<b>0.32</b>	15.62
2001 Chiwawa Broodstock	89	0.78	0.80	*	-0.02	<b>0.13</b>	15.77
2004 Chiwawa Broodstock	61	0.77	0.76	*	0.02	<b>0.13</b>	14.92
2005 Chiwawa Broodstock	75	0.79	0.78	*	0.02	0.01	15.85
2006 Chiwawa Broodstock	89	0.80	0.83	-	-0.03	<b>0.05</b>	16.46
1989 Chiwawa River	36	0.76	0.78	-	0.01	0.00	12.77
2001 Chiwawa River	55	0.78	0.80	-	-0.02	<b>0.09</b>	14.00
2004 Chiwawa River	96	0.78	0.78	*	0.01	<b>0.18</b>	17.23
2005 Chiwawa River	106	0.79	0.82	*	-0.02	<b>0.06</b>	16.69
2006 Chiwawa River	102	0.80	0.83	***	-0.03	<b>0.10</b>	16.77
1989 White River	48	0.75	0.75	-	0.01	0.01	12.85
1991 White River	19	0.76	0.76	-	0.03	0.00	10.92
1992 White River	22	0.75	0.79	-	-0.02	0.01	11.00
1993 White River	21	0.75	0.69	*	0.10	0.00	10.15
2005 White River	29	0.75	0.77	-	-0.01	0.03	12.23
2006 White River	40	0.76	0.76	-	0.01	0.04	13.38

**Table 1** Within population genetic data analysis summary continued.

Collection	Sample size	Gene Diversity	Observed Hz	HW	F <sub>IS</sub>	LD	Mean # Alleles
1993 Little Wenatchee R.	19	0.84	0.85	-	0.02	0.00	11.23
1993 Nason Creek	45	0.78	0.80	-	-0.01	0.01	13.77
2000 Nason Creek	51	0.76	0.78	-	-0.02	<b>0.13</b>	13.92
2001 Nason Creek	41	0.79	0.81	-	-0.01	<b>0.08</b>	14.23
2004 Nason Creek	38	0.76	0.76	-	0.02	0.03	13.23
2005 Nason Creek	45	0.78	0.82	-	-0.04	0.03	14.92
2006 Nason Creek	48	0.80	0.82	-	-0.01	0.00	15.77
2001 Wenatchee River	32	0.79	0.80	*	0.00	0.04	12.85
2000 Leavenworth NFH	73	0.80	0.82	*	-0.02	<b>0.15</b>	16.23
1997 Entiat NFH	37	0.81	0.83	-	-0.01	<b>0.06</b>	14.38

**Table 2** Demographic data for Chiwawa Hatchery and Chiwawa natural spring Chinook salmon. BS is census size of hatchery broodstock, pNOB is the proportion of hatchery broodstock of natural origin, NOS is the census size of natural-origin spawners present in Chiwawa River, HOS is the census size of hatchery-origin spawners present in Chiwawa River, Total is NOS and HOS combined, and pNOS is the proportion of spawners present in Chiwawa River of natural origin.

Brood Year	Hatchery		In River			
	BS	pNOB	NOS	HOS	Total	pNOS
1989	28	1	1392	0	1392	1.00
1990	18	1	775	0	775	1.00
1991	32	1	585	0	585	1.00
1992	78	1	1099	0	1099	1.00
1993	94	1	677	491	1168	0.58
1994	11	0.64	190	90	280	0.68
1995	0	0	8	50	58	0.14
1996	18	0.44	131	51	182	0.72
1997	111	0.29	210	179	389	0.54
1998	47	0.28	134	45	178	0.75
1999	0	0	119	13	132	0.90
2000	30	0.3	378	310	688	0.55
2001	371	0.3	1280	2850	4130	0.31
2002	71	0.28	694	919	1613	0.43
2003	94	0.44	380	223	603	0.63
2004	215	0.39	820	788	1608	0.51
2005	270	0.33	250	1222	1472	0.17

**Table 3** Levels of significance for pairwise tests of genic differentiation among all hatchery- and natural-origin collections used in this analysis. HS = highly significant ( $P < 0.000095$ ; the Bonferroni corrected p-value for an alpha = 0.05); \* =  $P < 0.05$  (nominal critical value for most statistical test); - =  $P > 0.05$  (not significant). A significant result between pairs of populations indicates that the allele frequencies between the pair are significantly different. Results are read by comparing the collections along the rows to collections along columns. The top block for each section is a symmetric matrix, as it compares collections within the same group.

		Chiwawa – Hatchery Origin								
		1993	1994	1996	1998	2000	2001	2004	2005	2006
Chiwawa – Hat. Origin	1993		HS	*	HS	HS	HS	HS	HS	HS
	1994	HS		HS	HS	HS	HS	HS	HS	HS
	1996	*	HS		*	-	*	-	-	*
	1998	HS	HS	*		HS	HS	HS	HS	HS
	2000	HS	HS	-	HS		HS	*	HS	HS
	2001	HS	HS	*	HS	HS		HS	*	HS
	2004	HS	HS	-	HS	*	HS		HS	HS
	2005	HS	HS	-	HS	HS	*	HS		HS
	2006	HS	HS	*	HS	HS	HS	HS	HS	
Chiwawa – Natural Origin	1989	HS	HS	-	HS	HS	*	HS	HS	HS
	1993	HS	HS	-	HS	HS	-	HS	*	HS
	1996	*	HS	-	*	-	-	-	-	-
	1998	HS	HS	-	-	HS	*	*	*	-
	2000	HS	HS	-	HS	HS	HS	*	HS	HS
	2001	HS	HS	-	HS	HS	HS	HS	*	HS
	2004	HS	HS	-	HS	HS	HS	HS	HS	HS
	2005	HS	HS	-	HS	HS	*	HS	*	HS
	2006	HS	HS	-	*	HS	HS	HS	HS	HS
Nason	1996	HS	HS	-	HS	HS	HS	HS	HS	HS
	2000	HS	HS	*	HS	HS	HS	HS	HS	HS
	2001	HS	HS	-	HS	HS	HS	HS	HS	HS
	2004	HS	HS	-	HS	HS	HS	HS	HS	HS
	2005	HS	HS	-	HS	HS	HS	HS	HS	HS
	2006	HS	HS	-	*	HS	HS	HS	HS	HS
White	1989	HS	HS	HS	HS	HS	HS	HS	HS	HS
	1991	HS	HS	-	HS	HS	HS	HS	HS	HS
	1992	HS	HS	*	HS	HS	HS	HS	HS	HS
	1993	HS	HS	*	HS	HS	HS	HS	HS	HS
	2005	HS	HS	-	HS	HS	HS	HS	HS	HS
	2006	HS	HS	HS	HS	HS	HS	HS	HS	HS
Other	Wen-M	HS	HS	*	HS	HS	*	*	-	HS
	Leaven	HS	HS	*	HS	HS	HS	HS	HS	HS
	Entiat	HS	HS	*	HS	HS	HS	HS	HS	HS

**Table 3 (con't)**

		<b>Chiwawa – Natural Origin</b>								
		<b>1989</b>	<b>1993</b>	<b>1996</b>	<b>1998</b>	<b>2000</b>	<b>2001</b>	<b>2004</b>	<b>2005</b>	<b>2006</b>
<b>Chiwawa – Natural Origin</b>	<b>1989</b>	-	-	-	-	-	*	*	*	*
	<b>1993</b>	-	-	-	*	*	*	HS	*	HS
	<b>1996</b>	-	-	-	-	-	-	-	-	-
	<b>1998</b>	-	*	-	-	*	*	HS	*	*
	<b>2000</b>	-	*	-	*	-	HS	-	HS	HS
	<b>2001</b>	*	*	-	*	HS	-	HS	*	HS
	<b>2004</b>	*	HS	-	HS	-	HS	-	HS	HS
	<b>2005</b>	*	*	-	*	HS	*	HS	-	*
	<b>2006</b>	*	HS	-	*	HS	HS	HS	*	-
<b>Nason</b>	<b>1996</b>	*	*	-	*	*	HS	HS	HS	HS
	<b>2000</b>	HS	HS	HS	HS	HS	HS	HS	HS	HS
	<b>2001</b>	HS	*	-	*	HS	HS	HS	HS	HS
	<b>2004</b>	HS	HS	-	HS	HS	HS	HS	HS	HS
	<b>2005</b>	*	*	-	*	HS	HS	HS	HS	HS
	<b>2006</b>	HS	HS	-	-	HS	HS	HS	HS	HS
<b>White</b>	<b>1989</b>	HS	HS	*	HS	HS	HS	HS	HS	HS
	<b>1991</b>	HS	HS	*	-	HS	HS	HS	HS	HS
	<b>1992</b>	HS	HS	-	*	HS	HS	HS	HS	HS
	<b>1993</b>	HS	*	-	*	HS	HS	HS	HS	HS
	<b>2005</b>	HS	*	*	*	HS	HS	HS	*	HS
	<b>2006</b>	HS	HS	*	HS	HS	HS	HS	HS	HS
<b>Other</b>	<b>Wen-M</b>	*	-	-	-	*	*	HS	*	*
	<b>Leaven</b>	HS	HS	*	*	HS	HS	HS	HS	HS
	<b>Entiat</b>	HS	HS	*	HS	HS	HS	HS	HS	HS



**Table 3 (con't)**

		Nason					
		1996	2000	2001	2004	2005	2006
Nason	1996		HS	-	HS	-	*
	2000	HS		HS	HS	HS	HS
	2001	-	HS		*	-	*
	2004	HS	HS	*		*	HS
	2005	-	HS	-	*		-
	2006	*	HS	*	HS	-	
White	1989	HS	HS	HS	HS	HS	HS
	1991	*	HS	HS	HS	*	*
	1992	HS	HS	HS	HS	HS	HS
	1993	*	HS	HS	HS	HS	HS
	2005	*	HS	HS	HS	HS	HS
	2006	HS	HS	HS	HS	HS	HS
Other	Wen-M	HS	HS	HS	HS	*	HS
	Leaven	HS	HS	HS	HS	HS	HS
	Entiat	HS	HS	HS	HS	HS	HS

**Table 3 (con't)**

		White					Other			
		1989	1991	1992	1993	2005	2006	Wen-M 2001	Leaven 2000	Entiat 1997
White	1989		-	*	-	HS	HS	HS	HS	HS
	1991	-		-	-	*	*	*	HS	HS
	1992	*	-		-	*	*	HS	HS	HS
	1993	-	-	-		*	*	HS	HS	HS
	2005	HS	*	*	*		*	HS	HS	HS
	2006	HS	*	*	*	*		HS	HS	HS
Other	Wen-M	HS	*	HS	HS	HS	HS		HS	HS
	Leaven	HS	HS	HS	HS	HS	HS	HS		HS
	Entiat	HS	HS	HS	HS	HS	HS	HS	HS	

**Table 4** Probabilities (above diagonal) and levels of significance (below diagonal) for pairwise tests of genic differentiation among all Chiwawa hatchery broodstock and Chiwawa natural spawner collections used in this analysis. HS = highly significant ( $P < 0.000476$ ; the Bonferroni corrected p-value for an  $\alpha = 0.05$ ); \* =  $P < 0.05$  (nominal critical value for most statistical test); - =  $P > 0.05$  (considered not significant). A significant result between pairs of populations indicates that the allele frequencies between the pair are significantly different. Pairwise comparisons between the hatchery broodstock and natural spawner collections from 2001, 2004, 2005, and 2006, respectively, are highlighted.

	Smolt		Hatchery Broodstock								Natural Spawners				
	1993	1994	1993	1996	1998	2000	2001	2004	2005	2006	1989	2001	2004	2005	2006
Smolt	1993	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
	1994	HS	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
Hatchery Broodstock	1993	HS	HS	0.9155	0.0000	0.0073	0.3647	0.0003	0.0694	0.0000	0.2220	0.0039	0.0008	0.0095	0.0000
	1996	HS	HS	-	0.0151	0.8388	0.0452	0.4916	0.3189	0.0716	0.5591	0.0759	0.8101	0.2364	0.0786
	1998	HS	HS	HS	*	0.0000	0.0000	0.0000	0.0000	0.0043	0.0000	0.0000	0.0000	0.0000	0.0005
	2000	HS	HS	*	-	HS	0.0000	0.4720	0.0000	0.0000	0.0036	0.0000	0.0712	0.0000	0.0000
	2001	HS	HS	-	*	HS	HS	0.0000	0.0059	0.0000	0.0003	0.0000	0.0000	0.0126	0.0000
	2004	HS	HS	*	-	HS	-	HS	0.0000	0.0000	0.0001	0.0000	0.0012	0.0000	0.0000
	2005	HS	HS	-	-	HS	HS	*	HS	0.0005	0.0024	0.0137	0.0025	0.7782	0.0018
	2006	HS	HS	HS	-	*	HS	HS	HS	*	0.0000	0.0000	0.0000	0.0000	0.5770
Natural Spawners	1989	HS	HS	-	-	HS	*	*	HS	*	HS	0.0023	0.0317	0.0000	0.0003
	2001	HS	HS	*	-	HS	HS	HS	HS	*	HS	*	0.0000	0.2641	0.0000
	2004	HS	HS	*	-	HS	-	HS	*	*	HS	*	HS	0.0000	0.0000
	2005	HS	HS	*	-	HS	HS	*	HS	-	HS	HS	-	HS	0.0000
	2006	HS	HS	HS	-	*	HS	HS	HS	*	-	*	HS	HS	HS

**Table 5** Analysis of molecular variance (AMOVA) for the Chiwawa collections, showing the partition of molecular variance into (1) within collections, (2) among collections but within group, and (3) among group components. Each column in the table represents a separate analysis testing for differences under a different spatial or temporal hypothesis. The different analyses are grouped together in a single table for comparisons. The values within the table are percentages and the parenthetical values are P-values, or probabilities, associated with that percentage. P-values greater than 0.05 indicate that the percentage is not significantly different from zero. For example, when collections are organized by hatchery- versus natural-origin (“Origin” – fourth column), 0.11% of the molecular variance is attributed to among group (i.e., hatchery- versus natural-origin), which is not significantly different from zero. No collections (first column) indicates no organization or grouping among all collections, and the among-group percentage is equal to the  $F_{ST}$  for the entire data set.

	No Structure	Collection Year	Spawning Location	Origin	Origin-Spawning Location
Among Groups	0.26 (0.00)	0.20 (0.43)	0.05 (0.48)	0.11 (0.15)	0.11 (0.06)
Among collections - Within groups	-	0.08 (0.003)	0.24 (0.00)	0.21 (0.00)	0.18 (0.06)
Within collections	99.74 (0.00)	99.72 (0.00)	99.71 (0.00)	99.68 (0.00)	99.71 (0.00)

**Table 6**  $F_{ST}$  values for all pairwise combinations of populations. Each  $F_{ST}$  is the median value for all pairwise combinations of collections within each population (the number of collections within each population is shown parenthetically next to each population name on each row). For example, the  $F_{ST}$  for the Chiwawa hatchery versus the White River (0.019) is the median value of 54 pairwise comparisons. The bold values along the center diagonal are the median  $F_{ST}$  values within each collection. For those populations with only one collection, the diagonal value was set at 0.000.

	Chiwawa-Hatchery	Chiwawa-Natural	Entiat	Leavenworth	Nason	Wenatchee-main	White	Little Wenatchee
Chiwawa-Hatchery (9)	<b>0.013</b>	0.008	0.016	0.012	0.011	0.005	0.019	0.111
Chiwawa-Natural (9)		<b>0.003</b>	0.012	0.011	0.007	0.003	0.014	0.105
Entiat (1)			<b>0.000</b>	0.005	0.010	0.008	0.019	0.078
Leavenworth (1)				<b>0.000</b>	0.007	0.008	0.014	0.092
Nason (6)					<b>0.006</b>	0.008	0.015	0.099
Wenatchee-main (1)						<b>0.000</b>	0.012	0.098
White (6)							<b>0.005</b>	0.113
Little Wenatchee (1)								<b>0.000</b>

**Table 7** As in Table 5, except data includes Chiwawa hatchery- and natural-origin, Nason Creek, and White River collections

	All Years	All Years	1989-1996	2005-2006	2005-2006
	No Structure	Origin	Origin	Origin	Collection Year
Among Groups	0.28 (0.00)	0.33 (0.00)	-0.07 (0.67)	0.43 (0.01)	-0.06 (0.57)
Among Collections - Within groups	-	0.04 (0.00)	0.22 (0.00)	0.25 (0.00)	0.64 (0.00)
Within Collections	99.72	99.63	99.85	99.32	99.41

**Table 8** Individual assignment results reported are the numbers of individuals assigned to each population using the partial Bayesian criteria of Rannala and Mountain (1997) and a “jack-knife” procedure (see Methods). The population with the highest posterior probability is considered the stock of origin (i.e., no unassigned individuals). Individuals from each population are assigned to specific populations (along rows). Bold values indicate correct assignment back to population of origin. Individuals assigned to a population are read down columns. For example, of the 595 individuals from Chiwawa hatchery origin, 134 individuals were assigned to Chiwawa natural origin (reading across). Of the 511 individuals assigned to Chiwawa natural origin (reading down), 60 were from Nason Creek.

Population	Total	Unassigned	1	2	3	4	5	6	7	8
1) Chiwawa Hatchery	595	0	<b>371</b>	134	2	16	0	45	15	12
2) Chiwawa Natural	501	0	156	<b>269</b>	4	5	0	42	9	16
3) Entiat	37	0	4	5	<b>13</b>	8	0	6	1	0
4) Leavenworth	73	0	9	8	3	<b>33</b>	0	17	0	3
5) Little Wenatchee	19	0	0	0	0	0	<b>19</b>	0	0	0
6) Nason	268	0	49	60	5	11	0	<b>131</b>	1	11
7) Wenatchee Mainstem	32	0	12	9	0	1	0	2	<b>6</b>	2
8) White	179	0	22	26	0	2	0	13	1	<b>115</b>
TOTAL	1704	0	623	511	27	76	19	256	33	159

**Table 9** As in Table 8, except the posterior probability from the partial Bayesian criteria of Rannala and Mountain (1997) must be 0.90 or greater, to be assigned to a population. Those individuals with posterior probabilities less than 0.90 are unassigned.

<b>Aggregate</b>	<b>Total</b>	<b>Unassigned</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>	<b>8</b>
1) Chiwawa Hatchery	595	332	<b>214</b>	31	1	4	0	10	3	0
2) Chiwawa Natural	501	375	30	<b>82</b>	0	1	0	5	2	6
3) Entiat	37	24	1	1	<b>5</b>	4	0	2	0	0
4) Leavenworth	73	51	0	1	1	<b>19</b>	0	1	0	0
5) Little Wenatchee	19	2	0	0	0	0	<b>17</b>	0	0	0
6) Nason	268	188	11	6	2	5	0	<b>53</b>	0	3
7) Wenatchee Mainstem	32	23	4	3	0	0	0	0	<b>2</b>	0
8) White	179	92	4	3	0	1	0	5	1	<b>73</b>
<b>TOTAL</b>	<b>1704</b>	<b>1087</b>	<b>264</b>	<b>127</b>	<b>9</b>	<b>34</b>	<b>17</b>	<b>76</b>	<b>8</b>	<b>82</b>

**Table 10** Estimates of  $N_e$  based on bias correction method of Waples (2006) implemented in LDNe (Do and Waples unpublished). Collections are categorized by spawning location. Sample size is the harmonic mean of the sample size, 95% CI is the confidence interval calculated using Waples' (2006) equation 12, and Major Cohort assumes that each collection is 100% four-year-olds.

	Sample size	Estimated $N_b$	95% CI	Major Cohort	Census	$N_e/N$
1993 Chiwawa Broodstock	58.4	103.1	77.0 - 149.7	1989	1392	0.30
1996 Chiwawa Broodstock	15.5	30.4	19.6 - 58.1	1992	1099	0.11
1998 Chiwawa Broodstock	33.4	37.7	29.8 - 49.7	1994	280	0.54
2000 Chiwawa Broodstock	77.8	48.4	41.4 - 57.2	1996	182	1.06
2001 Chiwawa Broodstock	80.4	49.6	42.2 - 59.2	1997	389	0.51
2004 Chiwawa Broodstock	56.6	48.1	39.0 - 60.9	2000	688	0.28
2005 Chiwawa Broodstock	73	274.3	148.9 - 1131.8	2001	4130	0.27
2006 Chiwawa Broodstock	88.4	198.3	136.1 - 340.5	2002	1613	0.49
1989 Chiwawa River	26.6	5.2	3.9 - 6.3	1985		
2001 Chiwawa River	46.7	38.6	31.0 - 49.3	1997	389	0.40
2004 Chiwawa River	88.5	82.6	67.3 - 104.4	2000	688	0.48
2005 Chiwawa River	104.2	231.5	161.8 - 382.7	2001	4130	0.22
2006 Chiwawa River	101.1	107.3	87.2 - 136	2002	1613	0.27



**Table 11** Summary of output from program SALMONNb and data for eight Chiwawa broodstock collections from Wenatchee River. For each pairwise comparison of samples  $i$  and  $j$ ,  $\tilde{S}$  is the harmonic mean sample size,  $n$  is the number of independent alleles used in the comparison,  $\hat{N}_{b(i,j)}$  are the pairwise estimates of  $N_b$ , and  $\text{Var} [\hat{N}_{b(i,j)}]$  is the variance of  $\hat{N}_{b(i,j)}$ .  $\tilde{N}_b$  is the harmonic mean of the  $\hat{N}_{b(i,j)}$ . Alleles with a frequency below 0.05 were excluded from the analysis to reduce potential bias.

Year	1993	1996	1998	2000	2001	2004	2005	2006
Pairwise $\tilde{S}$ (above diagonal) and $n$ (below diagonal):								
1993	-	24.5	42.5	66.4	67.2	57.2	64.6	70.3
1996	82	-	21.2	25.8	26.0	24.4	25.6	26.4
1998	80	81	-	46.7	47.2	42.0	45.8	48.4
2000	80	82	84	-	78.6	65.2	75.1	82.7
2001	73	77	81	76	-	66.0	76.2	84.2
2004	77	81	75	76	78	-	63.5	69.0
2005	71	75	82	73	73	69	-	80.0
2006	81	80	84	75	74	75	72	-
Pairwise $\hat{N}_{b(i,j)}$ (above diagonal) and $\text{Var} [\hat{N}_{b(i,j)}]$ (below diagonal):								
1993	-	-742.7	406.9	1240.8	-5432.0	829.8	808.9	729.0
1996	22491.2	-	110.4	-1786.5	765.9	162.8	824.7	382.7
1998	10910.4	67299.1	-	101.8	237.1	69.6	307.0	140.0
2000	6910.0	742895.8	19122.7	-	490.6	1498.2	706.9	201.6
2001	49318.3	21402.8	9754.2	6126.6	-	307.8	82.0	362.5
2004	8338.4	257267.7	24283.0	145043.4	7095.7	-	269.7	140.1
2005	31511.8	22242.5	10015.8	6596.6	114931.1	8240.4	-	599.6
2006	6223.8	43935.2	73518.7	10152.5	5885.3	12827.0	6370.8	-

$$\tilde{N}_b = 269.4$$

---

**Table 12** Summary of output from program SALMONNb and data for five Chiwawa in-river spawner collections from Wenatchee River. For each pairwise comparison of samples  $i$  and  $j$ ,  $\tilde{S}$  is the harmonic mean sample size,  $n$  is the number of independent alleles used in the comparison,  $\hat{N}_{b(i,j)}$  are the pairwise estimates of  $N_b$ , and  $\text{Var}[\hat{N}_{b(i,j)}]$  is the variance of  $\hat{N}_{b(i,j)}$ .  $\tilde{N}_b$  is the harmonic mean of the  $\hat{N}_{b(i,j)}$ . Alleles with a frequency below 0.05 were excluded from the analysis to reduce potential bias.

---



---

Year	1989	2001	2004	2005	2006
Pairwise $\tilde{S}$ (above diagonal) and $n$ (below diagonal):					
1989	-	33.3	40.2	41.7	42.2
2001	72	-	60.5	63.9	63.3
2004	72	77	-	95.3	94.0
2005	69	72	75	-	102.5
2006	76	76	77	78	-
Pairwise $\hat{N}_{b(i,j)}$ (above diagonal) and $\text{Var}[\hat{N}_{b(i,j)}]$ (below diagonal):					
1989	-	118.4	299.0	143.3	165.3
2001	40378.8	-	181.7	-1537.3	153.5
2004	10455.2	7265.5	-	387.1	329.4
2005	20923.6	68660.6	5040.7	-	356.8
2006	16227.2	8886.9	3802.0	4522.8	-
$\tilde{N}_b = 224.2$					

---

**Table 13** Summary of output from program SALMONNb and data for three brood years that combined Chiwawa natural- and hatchery-origin samples from Wenatchee River. For each pairwise comparison of samples  $i$  and  $j$ ,  $\tilde{S}$  is the harmonic mean sample size,  $n$  is the number of independent alleles used in the comparison,  $\hat{N}_{b(i,j)}$  are the pairwise estimates of  $N_b$ , and  $\text{Var}[\hat{N}_{b(i,j)}]$  is the variance of  $\hat{N}_{b(i,j)}$ .  $\tilde{N}_b$  is the harmonic mean of the  $\hat{N}_{b(i,j)}$ . Alleles with a frequency below 0.05 were excluded from the analysis to reduce potential bias.

---



---

Year	2004	2005	2006
Pairwise $\tilde{S}$ (above diagonal) and $n$ (below diagonal):			
2004	-	162	164.3
2005	77	-	188.2
2006	76	75	-
Pairwise $\hat{N}_{b(i,j)}$ (above diagonal) and $\text{Var}[\hat{N}_{b(i,j)}]$ (below diagonal):			
2004	-	611.3	210.8
2005	9351.5	-	727.5
2006	14965.5	8673.9	-
$\tilde{N}_b = 386.8$			

---

# Appendix K

**Fish Trapping at the Nason Creek Smolt Trap 2014**



# Population Estimates for Juvenile Salmonids in Nason Creek, WA

## 2014 Annual Final Report

Prepared by:  
Bryan Ishida  
Cory Kamphaus  
Keely Murdoch

YAKAMA NATION  
FISHERIES RESOURCE MANAGEMENT  
Toppenish, WA 98948



Prepared for:

Public Utility District No. 2 of Grant County  
Ephrata, Washington 98823

and

U.S Department of Energy  
Bonneville Power Administration  
Division of Fish and Wildlife  
Portland OR, 97208-3621

Project No. 1996-040-00

## ABSTRACT

In 2014, Yakama Nation Fisheries Resource Management (YN FRM) monitored emigration of naturally spawned juvenile coho salmon as well as Endangered Species Act (ESA) listed Upper Columbia River (UCR) spring Chinook salmon and summer steelhead in Nason Creek. This report summarizes juvenile abundance and freshwater survival estimates for each of these species. Fish were captured using a 1.5m rotary smolt trap between March 1 and November 30, 2014. We collected 2,693 spring Chinook salmon, 1,266 summer steelhead, 4 bull trout, and 24 coho; all of natural origin and varying age classes. Daily fish abundances for spring Chinook, steelhead, and coho were expanded by stream discharge-to-trap efficiency regression. All estimates were made with a 95% confidence interval (CI) with total emigration estimates for BY2012 spring Chinook juveniles and coho juveniles of 32,671 ( $\pm$  4,863) and 479 ( $\pm$  237), respectively. We estimated the total BY2011 summer steelhead emigration at the trap to be 13,605 ( $\pm$  3,525). Egg-to-emigrant survival rates for BY2012 Chinook and BY2012 coho were 1.9% and 0.8%, respectively. The egg-to-emigrant survival rate for BY2011 summer steelhead was 0.9%. Productivity, as measured by emigrants-per-redd, for spring Chinook, summer steelhead, and coho was 79, 58 and 23, respectively.



# CONTENTS

CONTENTS.....	v
LIST OF FIGURES .....	vii
LIST OF TABLES .....	ix
ACKNOWLEDGEMENTS .....	xi
1.0 INTRODUCTION .....	12
1.1 Watershed Description .....	12
2.1 Trapping Equipment and Operation.....	15
2.2 Biological Sampling.....	16
2.3 PIT Tagging.....	16
2.4 Mark-Recapture Trials .....	17
2.5 Data Analysis .....	17
2.5.1 <i>Estimate of Abundance</i> .....	17
2.5.2 <i>Emigration During The Non-Trapping Period</i> .....	20
2.5.3 <i>Production and Survival</i> .....	20
3.0 RESULTS .....	21
3.1 Dates of Operation .....	21
3.2 Daily Captures and Biological Sampling .....	21
3.2.1 <i>Spring Chinook Yearlings (BY2012)</i> .....	21
3.2.2 <i>Spring Chinook Subyearlings (BY2013)</i> .....	22
3.2.3 <i>Summer Steelhead</i> .....	23
3.2.4 <i>Hatchery Steelhead Smolts</i> .....	24
3.2.5 <i>Bull Trout</i> .....	25
3.2.6 <i>Coho Yearlings (BY2012)</i> .....	26
3.2.7 <i>Coho Subyearlings (BY2013)</i> .....	27
3.2.8 <i>Hatchery Coho Smolts (BY2012)</i> .....	27
3.3 Remote Parr Tagging (BY2013 Spring Chinook).....	28
3.4 Trap Efficiency Calibration and Population Estimates .....	29
3.4.1 <i>Spring Chinook Yearlings (BY2012)</i> .....	29
3.4.2 <i>Spring Chinook Subyearlings (BY2013)</i> .....	31
3.4.3 <i>Summer Steelhead</i> .....	32

3.4.4 Coho Yearlings (BY2012).....	35
3.4.5 Coho Subyearlings (BY2013) .....	37
3.5 PIT Tagging.....	37
3.6 Incidental Species.....	38
3.7 ESA Compliance.....	38
4.0 DISCUSSION.....	40
5.0 LITERATURE CITED.....	45
APPENDIX A. Daily Stream Discharge and Stream Temperature.....	46
APPENDIX B. Daily Trap Operation.....	51
APPENDIX C. Regression Models .....	55
APPENDIX D. Historical Morphometric Data .....	60

## LIST OF FIGURES

Figure 1. Map of Wenatchee River Subbasin with the Nason Creek rotary trap location.....	13
Figure 2. Mean daily stream discharge at the Nason Creek WDOE stream monitoring station in 2014.....	14
Figure 3. Mean daily water temperature at the Nason Creek DOE stream monitoring station in 2014.....	14
Figure 4. Daily catch of BY2012 spring Chinook yearlings with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014. ....	22
Figure 5. Daily catch of BY2013 spring Chinook subyearlings with mean daily stream discharge at the Nason Creek rotary trap, July 1 to November 30, 2014.....	23
Figure 6. Daily catch of wild summer steelhead with mean daily stream discharge at the Nason Creek rotary trap, March 1 to November 30, 2014. Estimates of fish passage during trap interruptions are not depicted.....	24
Figure 7. Daily catch of hatchery steelhead smolt with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014. ....	25
Figure 8. Daily catch of BY2012 naturally produced coho yearlings with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014. ....	26
Figure 9. Daily catch of BY2013 naturally produced coho subyearlings with mean daily stream discharge at the Nason Creek rotary trap, July 1 to November 30, 2014. ....	27
Figure 10. Daily catch of BY2012 hatchery coho smolt with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014. ....	28
Figure 11. Daily detections of remote-tagged BY2013 spring Chinook at the lower Nason Creek PIT tag antenna array (NAL) between October 2014 and March 2015.....	29

Figure 12. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for Nason Creek spring Chinook, BY 2003 to 2012. 31

Figure 13. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for Nason Creek summer Steelhead, BY 2003 to 2011.  
..... 35

Figure 14. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for Nason Creek naturally-produced coho, BY 2004 to 2012..... 37

Figure 15. Comparison of wild spring Chinook abundance estimates (BY2007-2012) made at the White River, Nason Creek, and Chiwawa River smolt traps..... 41

Figure 16. Comparison of egg-to-emigrant survival (BY 2007-2012) and egg deposition for Nason Creek, Chiwawa River, and White River spring Chinook..... 42

## LIST OF TABLES

Table 1. Summary of Nason Creek rotary trap operation. ....	21
Table 2. Summary of length and weight sampling of juvenile spring Chinook captured at the Nason Creek rotary trap. ....	22
Table 3. Summary of length, weight and condition factor by age class of wild summer steelhead emigrants and hatchery steelhead captured at the Nason Creek rotary trap. ....	24
Table 4. Summary of length, weight and condition factor for bull trout captured at the Nason Creek rotary trap. ....	25
Table 5. Summary of length and weight sampling of juvenile coho salmon captured at the Nason Creek rotary trap in 2014. ....	26
Table 6. Trap efficiency trials conducted with BY2012 wild spring Chinook yearlings. Note: trap efficiency is reported as the percentage of recaptures + 1 divided by the number of marked fish in the trial*. ....	29
Table 7. Estimated egg-to-emigrant survival and smolts-per-redd production for Nason Creek spring Chinook salmon. ....	30
Table 8. Trap efficiency trials conducted with BY2013 wild spring Chinook subyearlings. Note: trap efficiency is reported as the percentage of recaptures + 1 divided by the number of marked fish in the trial*. ....	32
Table 9. Efficiency trials conducted with wild summer steelhead juveniles. Note: trap efficiency is reported as the percentage of recaptures + 1 divided by the number of marked fish in the trial*. ....	33
Table 10. Estimated egg-to-emigrant survival and emigrants-per-redd production for Nason Creek summer steelhead. ....	33
Table 11. Estimated egg-to-emigrant survival and smolts-per-redd production for Nason Creek coho salmon. ....	35
Table 12. Number of PIT tagged coho, Chinook, and steelhead with shed rates at the Nason Creek rotary trap in 2014. ....	37

Table 13. Summary of length and weight sampling of incidental species captured at the Nason Creek rotary trap in 2014. .... 38

Table 14. Summary of ESA species and coho salmon mortality at the Nason Creek rotary trap. 38

## **ACKNOWLEDGEMENTS**

This project is part of a basin wide monitoring program requiring close coordination between multiple agencies and contractors. We greatly appreciate the hard work of the YN FRM crew members including Matthew Clubb, Jamie Hallman, Arlene Heemsah, Barry Hodges, Tim Jeffris, and Kevin Swager who maintained and operated the trap during all hours including nights/weekends and through challenging weather conditions. We would like to also thank the Wenatchee River Ranger District (U.S. Forest Service) and Mr. Duane Bolser for providing use of the trapping site and accommodating the needs of this project as well as to Peter Graf (Grant County PUD) for administering contracting and funding. Finally, thank you to Mike Hughes, Andrew Murdoch, Ben Truscott, and J.B. Walters (Washington Department of Fish and Wildlife), and Tracy Hillman (Bio Analysts, Inc.) for shared data and smolt trap methodologies.

## 1.0 INTRODUCTION

Beginning in the fall of 2004, YN began operating a rotary smolt trap in Nason Creek for nine months per year. Prior to 2004, the smolt trap was operated on a limited basis solely for hatchery coho predation studies. This project is a cost share between the YN's Mid-Columbia Coho Reintroduction and Grant County PUD's Hatchery Monitoring Plan. Trap operations were conducted in compliance with ESA consultation specifically to address abundance and productivity of spring Chinook, steelhead trout, and coho salmon in Nason Creek.

Within this document we will report:

- 1) Juvenile abundance and productivity of spring Chinook salmon (tkwínat) *Oncorhynchus tshawytscha*, steelhead trout (shúshaynsh) *Oncorhynchus mykiss* and coho salmon (súnx) *Oncorhynchus kisutch* in Nason Creek.
- 2) Emigration timing of spring Chinook salmon, steelhead trout and coho salmon emigrating from Nason Creek.

The data presented will be directly used to address Objective 2 in the Monitoring and Evaluation Plan for PUD Hatchery Programs (Hillman et al. 2013) on a 5-year analytic cycle:

**Objective 2: Determine if the proportion of hatchery fish on the spawning grounds affects the freshwater productivity of supplemented stocks (Hillman et al. 2013).**

### 1.1 Watershed Description

The Nason Creek watershed drains 65,600 acres of alpine glaciated landscape where high precipitation and moderate rain on snow recurrence controls the hydrology and aquatic communities. Nason Creek originates near the Cascade crest at Stevens Pass and flows east for approximately 37 river kilometers (rkm) until joining the Wenatchee River at rkm 86.3 just below Lake Wenatchee. Both smolt trap locations employed in 2014 (see section 2.1 Trapping Equipment and Operations) were downstream from the majority of spring Chinook and steelhead spawning grounds (Figure 1). There are 26.4 rkm along the mainstem accessible to anadromous fish in Nason Creek. Private land ownership comprises 52,300 acres (79.7%) of the watershed while 12,800 acres (19.5%) are federal and 480 acres (0.1%) are state owned (USFS et al. 1996).

The channel morphology of the lower 25 kilometers of Nason Creek has been impacted by development of highways, railroads, power lines, and residential development resulting in channel confinement and reduced side-channel habitat. The present condition is a low gradient (< 1.1%), low sinuosity (1:2 to 2:0 channel-to-valley length ratio) and depositional channel (USFS et al. 1996). Peak runoff typically occurs in May and June with occasional high water produced by rain on snow events in October and November.

In 2014, mean daily discharge for Nason Creek was 452 cfs with mean daily stream temperatures ranging from 0.0°C to 18.6°C (Figure 2 & 3). Spring and fall freshets included multiple high-



water events exceeding 12-year mean discharge levels. Most significant of these was a late November rain-on-snow event that pushed discharge levels to nearly 3,000 cfs. Water temperatures in Nason Creek were well below the 12-year mean values in the spring of 2014. Conversely, fall water temperatures were unseasonably high.

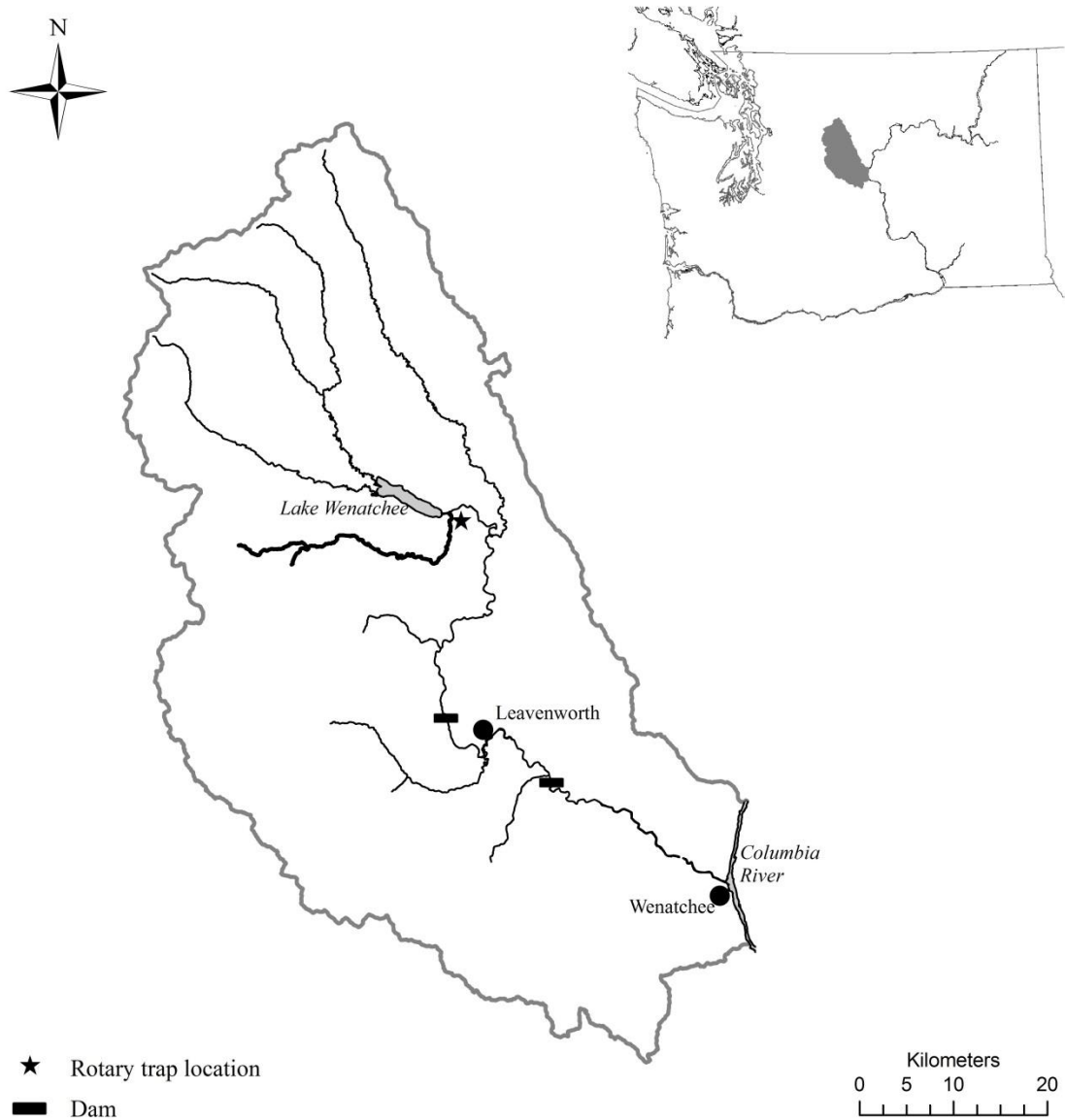


Figure 1. Map of Wenatchee River Subbasin with the Nason Creek rotary trap location.

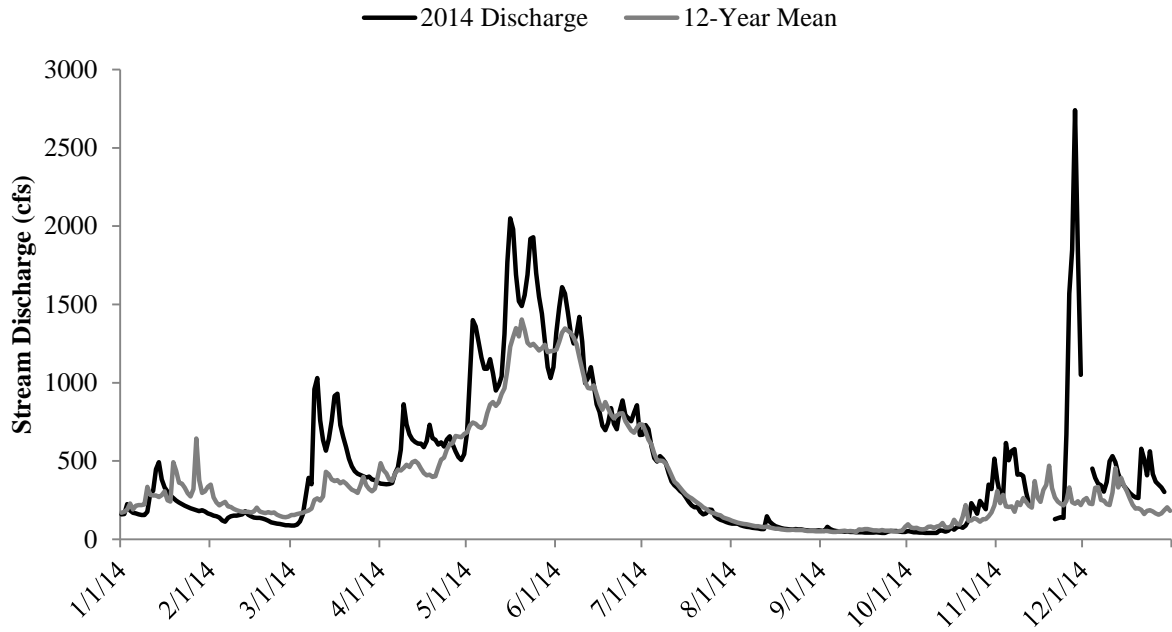


Figure 2. Mean daily stream discharge at the Nason Creek WDOE stream monitoring station in 2014.

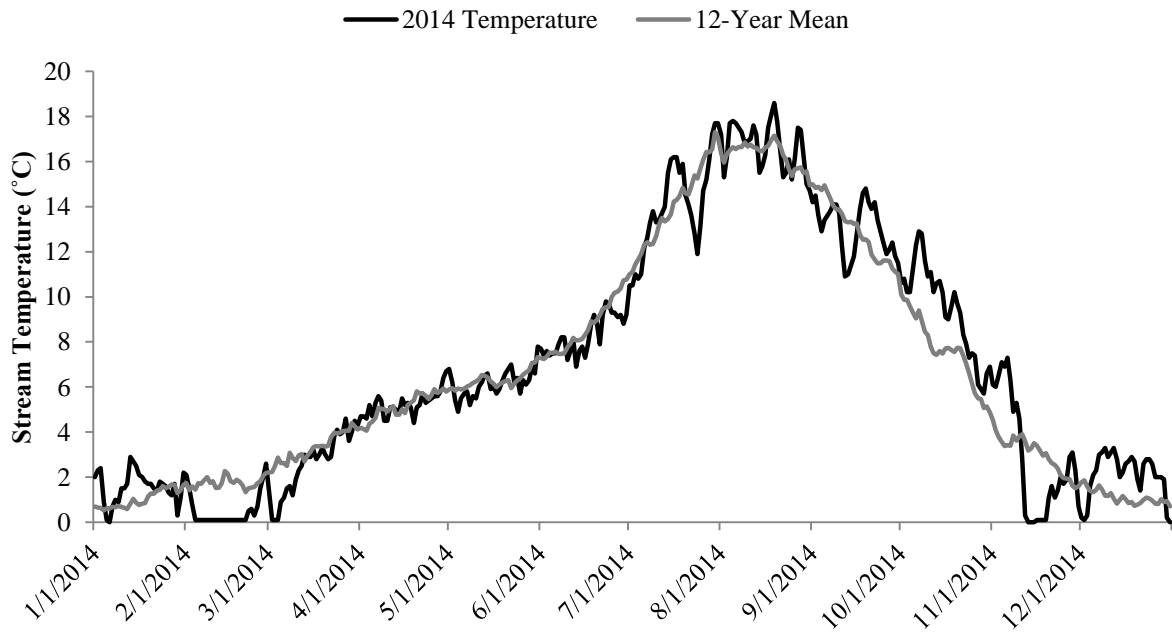


Figure 3. Mean daily water temperature at the Nason Creek DOE stream monitoring station in 2014.

## 2.0 METHODS

### 2.1 Trapping Equipment and Operation

In 2014, two different trap locations were employed; the originally used location herein referred to as “campground” (rkm 0.9), and new location herein referred to as “Bolser” (rkm 0.3). Initial trap operations began on March 1 at campground and extended until June 25. The trap was then relocated to Bolser, where it was operated from July 1 to November 30. Equipment at both sites included the same 1.5m rotary smolt trap and wire rope suspension system of rigging. Whereas campground required seasonal changes in the anchoring of the trap to accommodate campground traffic, cable configuration at Bolser remained static throughout operations. The move to Bolser is intended as a permanent relocation with campground no longer being utilized.

At both locations, the smolt trap was operated continually 24 hours per day, 7 days per week for the majority of their respective seasons. During spring snowmelt, operations at campground occurred only during hours of darkness in order to minimize trap damage and capture mortality, while retaining the ability to sample during periods of peak fish movement. Without the threat of vandalism posed during periods of peak use at the campground, summer operations at Bolser were not modified (daytime suspension) as required at the previous site.

On a daily basis, fish were removed from the primary collection box and retained in separate shore-anchored holding boxes until removed for efficiencies trials (up to 72 hours; Section 7 permit 2011/05645). A rotating drum-screen constantly removed small debris from the live box to avoid fish injury. All changes/modifications to the trap as well as periods of stoppage were noted. During periods when the trap was not operating (e.g. high discharge, high debris or mechanical malfunction), the number of target species captured was estimated. The estimated number of fish captured was calculated using the average number of fish captured three days prior and three days after the break in operation. This estimate of daily capture was incorporated into the overall emigration estimate.

## 2.2 Biological Sampling

Trap operating procedures and techniques followed a standardized basin-wide monitoring plan developed by the Upper Columbia Regional Technical Team (RTT) for the Upper Columbia Salmon Recovery Board (UCSRB; Hillman 2004), which was adapted from Murdoch and Petersen (2000).

All fish were enumerated by species and size class. Fish to be sampled were anesthetized in a solution of MS-222, weighed with an electronic scale and measured in a wetted trough-type measuring board. Anesthetized fish received oxygen through aquarium bubblers and were allowed to fully recover before being either released downstream of the trap or used in efficiency trials. Fork length (FL) and weight were recorded for all fish except when large numbers of fry or non-target species were collected; a sub-sample of 25 fish were measured and weighed while the remaining fish were tallied. Weight was measured to the nearest 0.1 gram and FL to the nearest millimeter. We used these data to calculate a Fulton-type condition factor (K-factor) using the formula:

$$K = (W/L^3) \times 100,000$$

Where K = Fulton-type condition metric, W = weight in grams, L = fork length in millimeters and 100,000 is a scaling constant.

Scale samples were collected from steelhead measuring  $\geq 60$  mm FL so that age and brood year could be assigned. Samples were collected according to the needs and protocols set by Washington Department of Fish and Wildlife (WDFW), who conducted the analysis and provided YN with results. Tissue samples were collected from spring Chinook and steelhead for DNA analysis. Samples from spring Chinook and steelhead were retained for reproductive success analyses conducted by WDFW and National Marine Fisheries Service (NMFS). All target salmonids were classified as either natural or hatchery origin by physical appearance, presence/absence of coded wire tags (CWTs), or post-orbital elastomer tags. Developmental stages were visually classified as fry, parr, transitional, or smolt. Fry were defined as newly emerged fish with or without a visible yolk sac and a FL measuring  $< 50$  mm. Age-0 coho and spring Chinook salmon captured before July 1 were considered 'fry' and were excluded from subyearling population estimates because of the uncertainty that these fish were actively migrating (UCRTT, 2001).

## 2.3 PIT Tagging

All natural origin Chinook, steelhead and coho measuring  $\geq 60$ mm were PIT tagged. Once anesthetized, each fish was examined for external wounds or descaling, then scanned for the presence of a previously implanted PIT tag. If a tag was not detected, a pre-loaded 12mm Digital Angel 134.2 kHz type TX 1411ST PIT tag was inserted into the body cavity using a Biomark MK-25 Rapid Implant Gun. Each unique tag code was electronically recorded along with date of tag implantation, date of fish release, tagging personnel, FL, weight, and anesthetic bath temperature. Data were entered using P3 software and submitted to the PIT Tag Information System (PTAGIS). PIT tagging methods were consistent with methodologies

described in the PIT Tag Marking Procedures Manual (CBFWA 1999) as well as in 2008 ISEMP protocols (Tussing 2008).

After marking and sampling, fish were held for a minimum of 24-hours in holding boxes at the trap to; a) ensure complete recovery, b) assess tagging mortality, and c) determine a PIT tag shed rate. Fish that were not used in mark-recapture trials were released downstream from the trap. Fish used in mark-recapture trials at campground were then transported in 5-gallon buckets 1.0 rkm upstream and released at nautical twilight from an automated release box. Mark groups released during operation at Bolser were released by hand 0.8 rkm above the trap at nautical twilight. Fish released by hand were distributed evenly along apposing banks in pools and other protected areas.

## 2.4 Mark-Recapture Trials

Groups of marked juveniles were released during a range of stream discharges in order to determine the trapping efficiency. PIT tags were the only method of marking used in 2014. These releases followed the protocols described in Hillman (2004), in which the author suggests a minimum sample size of 100 fish for each mark-recapture trial. Although 100 fish/trial represented the ideal mark group, low abundance of fish often required mark-recapture trials be completed with smaller sample sizes. To achieve the largest marked group possible, we combined catch over a maximum of 72 hours. Fish being held for mark-recapture trials were kept in auxiliary live boxes attached to the end of each pontoon or floating holding boxed anchored to the stream bank. A pre-season, minimum mark group size for each species/life stage was initially determined based on past regression models. In light of high abundance, minimum trial sizes could be raised to a more robust mark group with the intention of strengthening existing regression models.

Each mark-recapture trial was conducted over a three-day (72 hour) period to allow time for passage or capture. Completed trials were only considered invalid if an interruption to trapping occurred or proper pre-release procedures were not followed. Trials resulting in zero recaptures were included in the efficiency regression (if determined valid once vetted through release/recapture protocols) as allowed by the new method of observed trap efficiency calculation. The model used (Bailey) employs use of recaptures +1 in the calculation of efficiency as a mode of bias correction. As a result, even trials yeilding no recaptures can be included in regression modeling (See equation 3 in **2.5.1 Estimate of Abundance**).

## 2.5 Data Analysis

### 2.5.1 Estimate of Abundance

A recent WDFW review of smolt monitoring programs in the Wenatchee basin suggested that changes in the calculations for estimating abundance and its associated variance were necessary. Calculation of daily and seasonal smolt abundance changed only slightly. More significant changes were made to the variance estimator making the calculations more complex. The following describes the revised calculation of the point estimate, variance, and standard error of seasonal smolt abundances based on regression relationships.

Seasonal juvenile migration,  $N$ , was estimated as the sum of daily migrations,  $N_i$ , i.e.,  $N = \sum_i N_i$ , and daily migration was calculated from catch and efficiency:

$$\hat{N}_i = \frac{C_i}{\hat{e}_i}, \quad (1)$$

where  $C_i$  = number of fish caught in period  $i$ ;

$\hat{e}_i$  = trap efficiency estimated from the flow-efficiency relationship,  $\sin^2(b_0 + b_1 \text{flow}_i)$ ,

where  $b_0$  is estimated intercept and  $b_1$  is the estimated slope of the regression.

The regression parameters  $b_0$  and  $b_1$  are estimated using linear regression for the model:

$$\arcsin(\sqrt{e_k^{obs}}) = \beta_0 + \beta_1 \text{flow}_k + \varepsilon, \quad (2)$$

where  $e_k^{obs}$  = observed trap efficiency of Eq. 2 for trapping period  $k$ ;

$\beta_0$  = intercept of the regression model;

$\beta_1$  = slope parameter;

$\varepsilon$  = error with mean 0 and variance  $\sigma^2$ .

In Equation 2, the observed trap efficiency,  $e_k^{obs}$ , is calculated as follows,

$$e_k^{obs} = \frac{r_k + 1}{m}. \quad (3)$$

The estimated variance of seasonal migration is calculated from daily estimates as:

$$\text{Var}\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i \text{Var}(N_i)}_{\text{Part A}} + \underbrace{\sum_i \sum_j \text{Cov}(N_i, N_j)}_{\text{Part B}},$$

or,

$$\text{Var}\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i \text{Var}\left(\frac{(C_i + 1)}{\hat{e}_i}\right)}_{\text{Part A}} + \underbrace{\sum_i \sum_j \text{Cov}\left(\frac{(C_i + 1)}{\hat{e}_i}, \frac{(C_j + 1)}{\hat{e}_j}\right)}_{\text{Part B}}. \quad (4)$$

Part A of equation 4 is the variance of daily estimates. Part B is the between-day covariance. Note that the between-day covariance exists only for days that use the same trap efficiency model. If, for example, day 1 is estimated with one trap efficiency model, and day 2 estimated from a different model, then there is no covariance between day 1 and day 2. The full expression for the estimated variance:

$$\hat{V}ar\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i \hat{N}_i^2 \left( \frac{N_i \hat{e}_i (1 - \hat{e}_i)}{(C_i + 1)^2} + \frac{4(1 - \hat{e}_i)}{\hat{e}_i} \hat{V}ar(b_0 + b_1 flow_i) \right)}_{PartA} + \underbrace{\sum_i \sum_j 4(\hat{N}_i (1 - \hat{e}_i))(\hat{N}_j (1 - \hat{e}_j)) \cdot [\hat{V}ar(b_0) + flow_i flow_j \hat{V}ar(b_1)]}_{PartB}$$

where  $\hat{V}ar(b_0 + b_1 flow_i) = M\hat{S}E \left( 1 + \frac{1}{n} + \frac{(flow_i - \overline{flow})^2}{(n-1)s_{flow}^2} \right)$ , and  $\hat{V}ar(b_0)$  and  $\hat{V}ar(b_1)$  are

obtained from regression results. In Excel, the standard error (SE) of the coefficients is provided. The variance is calculated as the square of the standard error,  $SE^2$ .

In cases when there was no significant flow-efficiency relationship (i.e., low correlation), then a pooled, or average trap efficiency will suffice for the stratum. The estimator is calculated as follows:

$$\hat{e} = \frac{\sum_{j=1}^k r_j}{\sum_{j=1}^k m_j}$$

where  $\hat{e}$  = the average or pooled trap efficiency for the stratum;  
 $m_j$  = the number of smolts marked and released in efficiency trial  $j$  for the stratum;  
 $r_j$  = the number of smolts recaptured out of  $m_j$  marked fish in efficiency trial  $j$ .

Abundance for a trapping period is estimated as:

$$\hat{N}_i^{pooled} = \frac{C_i}{\hat{e}},$$

,and total stratum abundance is:

$$N^{pooled} = \sum_i \hat{N}_i^{pooled}.$$

The variance of seasonal abundance takes into account the variability in catch numbers that are a result of binomial sampling (Part A), the pooled variance of trap efficiency,  $\hat{e}$  (Part B), and the covariance in daily estimates that arises from using a common estimate of efficiency across all trapping days (Part C):

$$\hat{V}ar\left(\sum_{i=1}^n \hat{N}_i^{pooled}\right) = \underbrace{\left(\sum_i \frac{\hat{N}_i (1 - \hat{e})}{\hat{e}}\right)}_{PartA} + \underbrace{\frac{Var(\hat{e})}{\hat{e}^2} \sum_i \hat{N}_i^2}_{PartB} + \underbrace{\frac{Var(\hat{e})}{\hat{e}^2} \sum_i \sum_j \hat{N}_i \hat{N}_j}_{PartC}.$$

The Part B and Part C terms are combined in the calculation as a new Part B:

$$\text{Var}\left(\sum_{i=1}^n \hat{N}_i^{\text{pooled}}\right) = \underbrace{\left(\sum_i \frac{\hat{N}_i(1-\hat{e})}{\hat{e}}\right)}_{\text{PartA}} + \underbrace{\frac{\text{Var}(\hat{e})}{\hat{e}^2} \left[\sum_i \hat{N}_i^2 + \sum_i \sum_j \hat{N}_i \hat{N}_j\right]}_{\text{PartB}}.$$

The variance of  $\hat{e}$  is calculated as:

$$\text{Var}(\hat{e}) = \text{Var}\left(\frac{\sum_{k=1}^n r_k}{\sum_{k=1}^n m_k}\right) = \frac{\sum_{k=1}^n (r_k - \hat{e}_k m_k)^2}{\bar{m}^2 n(n-1)}$$

where  $\bar{m}$  is the average release size across all efficiency trial,  $\frac{\sum_{k=1}^n m_k}{n}$ .

Confidence intervals were calculated using the following formulas:

$$95\% \text{ confidence interval} = 1.96 \times \sqrt{\sum \text{var}[\hat{N}_i]}$$

The single M-R estimator of abundance carries a set of well documented assumptions (Everhart and Youngs 1981; Seber 1982),

1. The population is closed to mortality.
2. The probability of capturing a marked or unmarked fish is equal.
3. Marked fish were randomly dispersed in the population prior to recapture.
4. Marking does not affect probabilities of capture.
5. Marks were not lost between the time of release and recapture.
6. All marks are reported upon recapture.
7. The number of fish in the trap, C, is fully enumerated and known without error.

### 2.5.2 Emigration During The Non-Trapping Period

An estimate of spring chinook emmigration during the non-trapping period (December 1 through February 28) was calculated using remote-tagged spring chinook parr and the lower Nason Creek PIT tag array (NAL). A flow-detection efficiency regression was developed using mark-groups previously released to test the efficiency of the smolt trap. Daily spring Chinook detections at NAL and the developed regression were then applied to the Bailey estimator, as was performed with daily trap abundance data(See section **2.5.1 Estimate of Abundance**).

### 2.5.3 Production and Survival

Production estimates by age class were summed to produce a total emmigration estimate. For spring Chinook and coho, estimates of fall migrant parr were added to subsequent spring smolt estimates to generate a single brood year estimate. For steelhead, a single brood year may



require up to three years for emigration from Nason Creek to occur. Pending scale analysis, steelhead captured in 2014 were aged via an age-length histogram built upon previously analyzed scale samples. For all three species, egg-to-emigrant estimates were calculated by dividing estimated emigrants by approximated egg deposition during a spawning brood (average fecundity used to determine egg deposition derived from WDFW Chiwawa broodstock spawning). The number of emigrants-per-redd for each brood year was calculated by dividing the total emigrant estimate by the number of redds counted during spawning ground surveys.

### 3.0 RESULTS

#### 3.1 Dates of Operation

The Nason Creek trap was installed at campground on February 26, 2014 (started on March 1) and removed on June 25 for relocation to Bolser. Installation at Bolser occurred on June 26 and operations began on July 1. Removal of the trap occurred on December 2 (operations concluded November 30). The trap was operated continuously 24 hours a day, 7 days per week including periods of extreme high flows (>2,000cfs) associated with spring snowmelt. Interruptions to trapping at campground were mainly caused by ice formation in the cone at the onset of the season and relocation of the trap in late June (Table 1). Interruptions at Bolser were more frequent and caused primarily by debris and low discharge levels (Table 1).

**Table 1. Summary of Nason Creek rotary trap operation.**

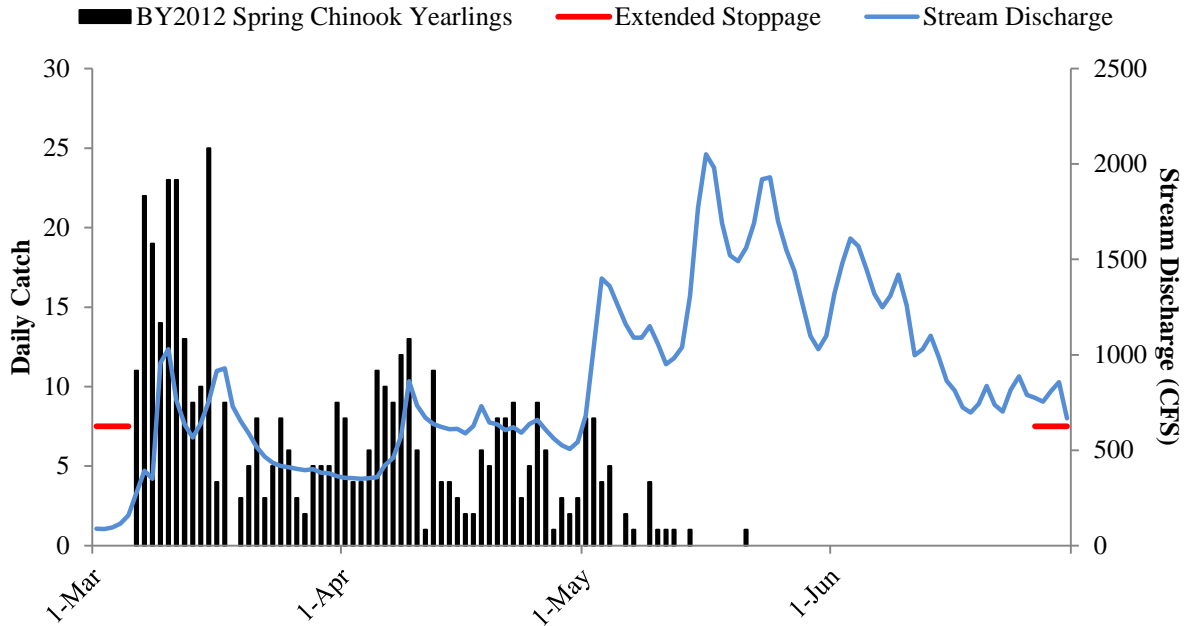
Date of Trap Operations	Trap Status	Description	Days
March 1 to June 30 (Campground)	Operating	Continuous data collection.	113
	Interrupted	Interrupted by debris, ice and/or low flows.	2
	Pulled	Intentionally pulled to prevent harm to fish or protect the trap during high flows.	7
July 1 to November 30 (Bolser)	Operating	Continuous data collection.	114
	Interrupted	Interrupted by debris and/or low flows.	27
	Pulled	Intentionally pulled due to low discharge levels or ice formation	12

#### 3.2 Daily Captures and Biological Sampling

##### 3.2.1 Spring Chinook Yearlings (BY2012)

Between March 1 and June 30, a total of 464 wild Chinook yearlings were captured at the trap (Figure 4). The majority of these fish were collected prior to the onset of major spring snowmelt, with peak catch occurring on March 25. Following a significant increase in stream discharge, capture numbers dropped substantially with the last emigrating Chinook yearling captured on May 21. Daily catch estimates during the initial trap stoppage in March (March 1-4) were not made due to a lack of pre-stoppage data and influence of coinciding drastically changing flows. Mean FL and weight for Chinook yearlings was 89.5mm ( $n = 464$ ;  $SD = 6.9$ )

and 7.5g ( $n = 464$ ;  $SD = 1.8$ ; Table 2), respectively. Tissue sample were collected from 456 fish for an ongoing, parental-based DNA analysis by WDFW. There were no yearling Chinook mortalities.



**Figure 4.** Daily catch of BY2012 spring Chinook yearlings with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014.

**Table 2.** Summary of length and weight sampling of juvenile spring Chinook captured at the Nason Creek rotary trap.

Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-Factor
		Mean	<i>n</i>	SD	Mean	<i>n</i>	SD	
2012	Wild Chinook Yearling Smolt	89.5	464	6.9	7.5	464	1.8	1.0
2013	Wild Chinook Subyearling Fry	40.1	677	5.2	0.9	221	0.5	1.4
2013	Wild Chinook Subyearling Parr	69.1	1,549	12.3	3.8	1,547	2.3	1.2

### 3.2.2 Spring Chinook Subyearlings (BY2013)

A total of 1,550 wild spring Chinook subyearling parr were captured between July 1 and November 30, with an additional 679 subyearling fry captured prior to July 1 (Figure 5). A peak daily capture of 93 subyearling Chinook parr occurred in July as snowmelt-driven high water subsided. The initial peak in subyearling parr catch decreased accordingly with discharge levels. A second major peak in parr catch occurred in late October at the onset of the first fall freshets. Mean FL and weight among fall subyearling parr was 69.1mm ( $n = 1,549$ ;  $SD = 12.3$ ) and 3.8g ( $n = 1,547$ ;  $SD = 2.3$ ), respectively. We estimate that an additional 349 Chinook subyearling parr would have been captured if the trap had been operated without interruption during this

period: 200 Chinook during short discreet stoppages ( $\leq 3$  days in duration) and 149 chinook during prolonged suspension due to low flow or ice accumulation. A total of 11 subyearling Chinook (3 fry and 8 parr) mortalities occurred in 2014. Causes of death included trapping mortality, tagging/handing mortality, and pre-existing fungal infection/poor condition.

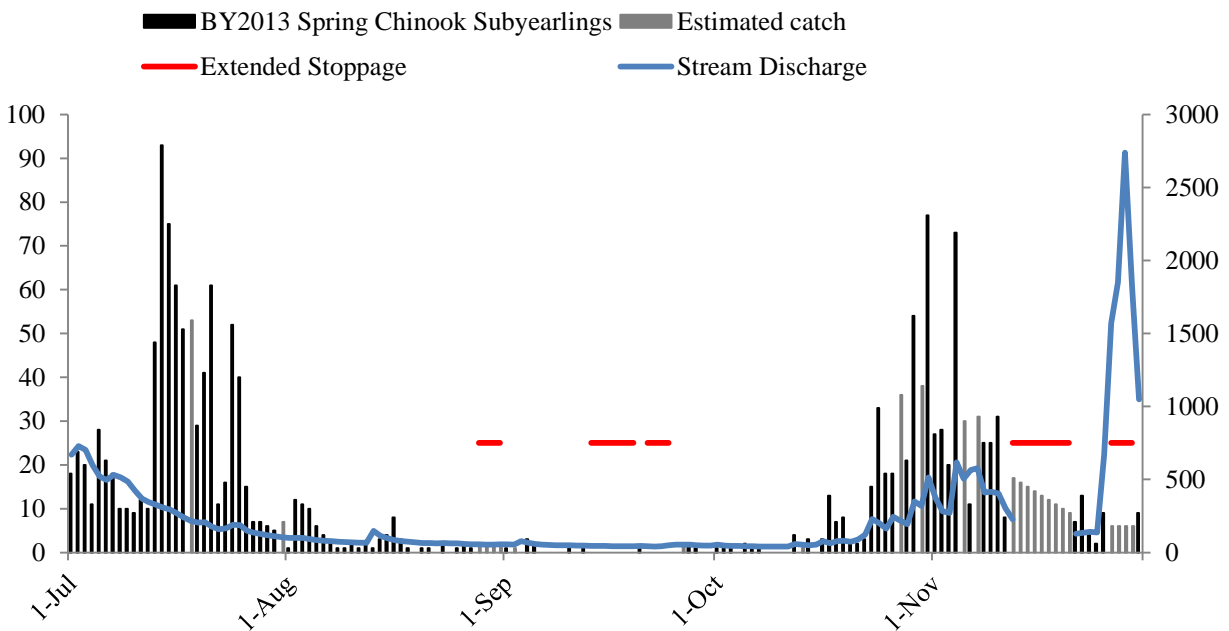


Figure 5. Daily catch of BY2013 spring Chinook subyearlings with mean daily stream discharge at the Nason Creek rotary trap, July 1 to November 30, 2014.

### 3.2.3 Summer Steelhead

A total of 1,267 wild summer steelhead juveniles were captured throughout the season from March 1 to November 30 with a peak catch of 40 fry on September 14 (Figure 6). We estimated that an additional 35 age-1 juveniles would have been captured if there had been no interruptions to trapping during the migratory period (Mar 1 to July 31). Histogram analysis of known steelhead ages sampled from 2005 to 2012 allowed us to estimate ages of fish captured in 2014 using FL. We estimate that of the total steelhead captured, 491 were young-of-the-year, 745 were age-1, and 30 were age-2. One steelhead did not have FL measurements taken and could not be aged. Subyearling steelhead had a mean FL of 50mm ( $n = 490$ ;  $SD = 12.8$ ), and a mean weight of 1.7g ( $n = 389$ ;  $SD = 1.1$ ). The majority of steelhead juveniles captured were age-1 parr emigrating past the trap in spring. Mean FL and weight of age-1 fish was 82mm ( $n = 745$ ;  $SD = 13.6$ ; Table 3) and 6.3g ( $n = 745$ ;  $SD = 3.5$ ), respectively. Age-2 steelhead were caught primarily in the spring, with only one fish being captured after July 31. Mean FL and weight of age-2 fish was 145mm ( $n = 30$ ;  $SD = 16.5$ ) and 33.0g ( $n = 30$ ;  $SD = 13.4$ ), respectively. Tissue samples were not taken from wild-origin in 2014 as per the request WDFW personnel. Scales were taken from a sub-sample ( $n = 852$ ) to be used for future age analyses. There was one steelhead fry trapping mortality (See 3.6 ESA Compliance).

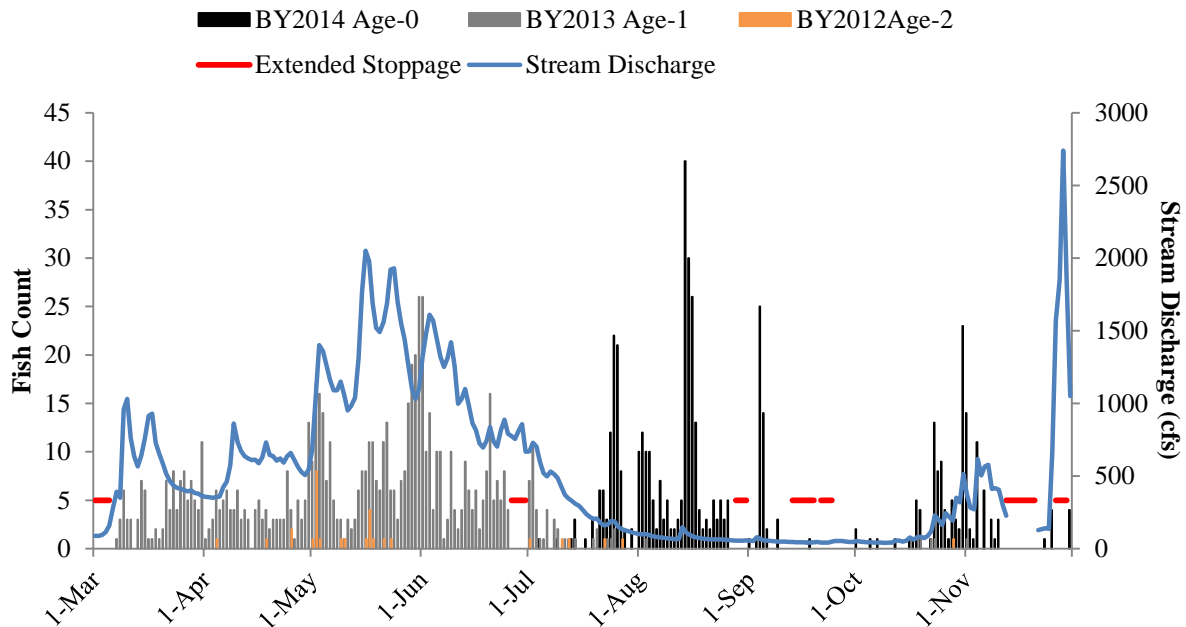


Figure 6. Daily catch of wild summer steelhead with mean daily stream discharge at the Nason Creek rotary trap, March 1 to November 30, 2014. Estimates of fish passage during trap interruptions are not depicted.

Table 3. Summary of length, weight and condition factor by age class of wild summer steelhead emigrants and hatchery steelhead captured at the Nason Creek rotary trap.

Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-Factor
		Mean	<i>n</i>	SD	Mean	<i>n</i>	SD	
2014	Wild Summer Steelhead (Age-0)	49.6	490	12.8	1.7	389	1.1	1.4
2013	Wild Summer Steelhead (Age-1)	82.2	745	13.6	6.3	745	3.5	1.1
2012	Wild Summer Steelhead (Age-2)	145.1	30	16.5	33	30	13.4	1.1
2011	Wild Summer Steelhead (Age-3)	—	—	—	—	—	—	—
2013	Hatch. Summer Steelhead Smolt	173.4	632	18.7	52.6	633	15.9	1.0

### 3.2.4 Hatchery Steelhead Smolts

During the months of April and May, WDFW released a total of 90,090 hatchery steelhead smolts into Nason Creek. Subsequently, a total of 1,571 hatchery steelhead were captured at the smolt trap with a mean FL and weight of 173mm ( $n = 632$ ;  $SD = 18.7$ ) and 52.6g ( $n = 633$ ;  $SD = 15.9$ ), respectively (Figure 7). The presence of hatchery-origin steelhead at the trap was limited one to two months after initial release, and did not continue into the summer. Hatchery origin was determined by the presence of coded wire tags (CWT). There were no hatchery steelhead mortalities.

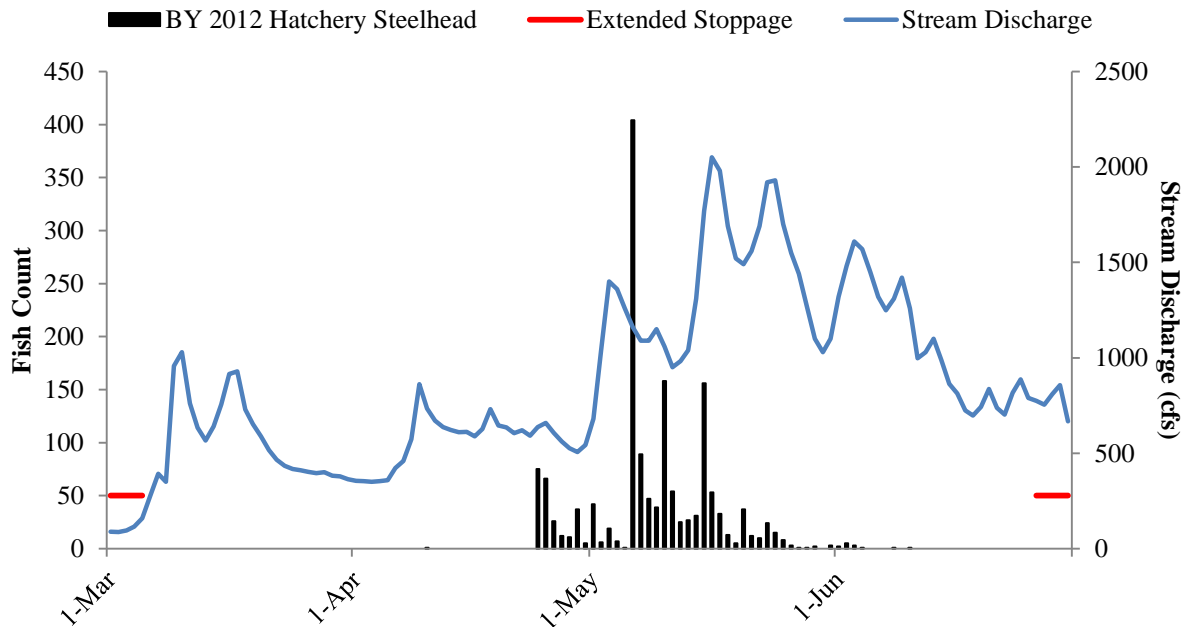


Figure 7. Daily catch of hatchery steelhead smolt with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014.

### 3.2.5 Bull Trout

A total of four bull trout were captured with a mean fork length of 150mm ( $n = 4$ ;  $SD = 12.8$ ; Table 4). There were no mortalities incurred.

Table 4. Summary of length, weight and condition factor for bull trout captured at the Nason Creek rotary trap.

Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-Factor
		Mean	<i>n</i>	SD	Mean	<i>n</i>	SD	
Unknown	Wild Bull Trout	150.3	4	12.8	33.4	4.9	9.8	1.0

### 3.2.6 Coho Yearlings (BY2012)

A total of 20 naturally produced coho yearlings were captured during spring emigration between March 1 and June 30 (Figure 8). Peak catch of three yearling smolts occurred on May 16 and 19 following an increase in flow associated with spring snowmelt. Mean FL and weight were 96mm ( $n = 20$ ;  $SD = 9.8$ ) and 9.9g ( $n = 20$ ;  $SD = 3.0$ ), respectively (Table 5). There were no coho yearling mortalities. Scale samples were collected from 19 fish to continue developing a baseline of freshwater growth patterns for naturally produced coho from Nason Creek. A subsample of 13 coho yearlings also had tissue samples taken from them for parental-based DNA analysis.

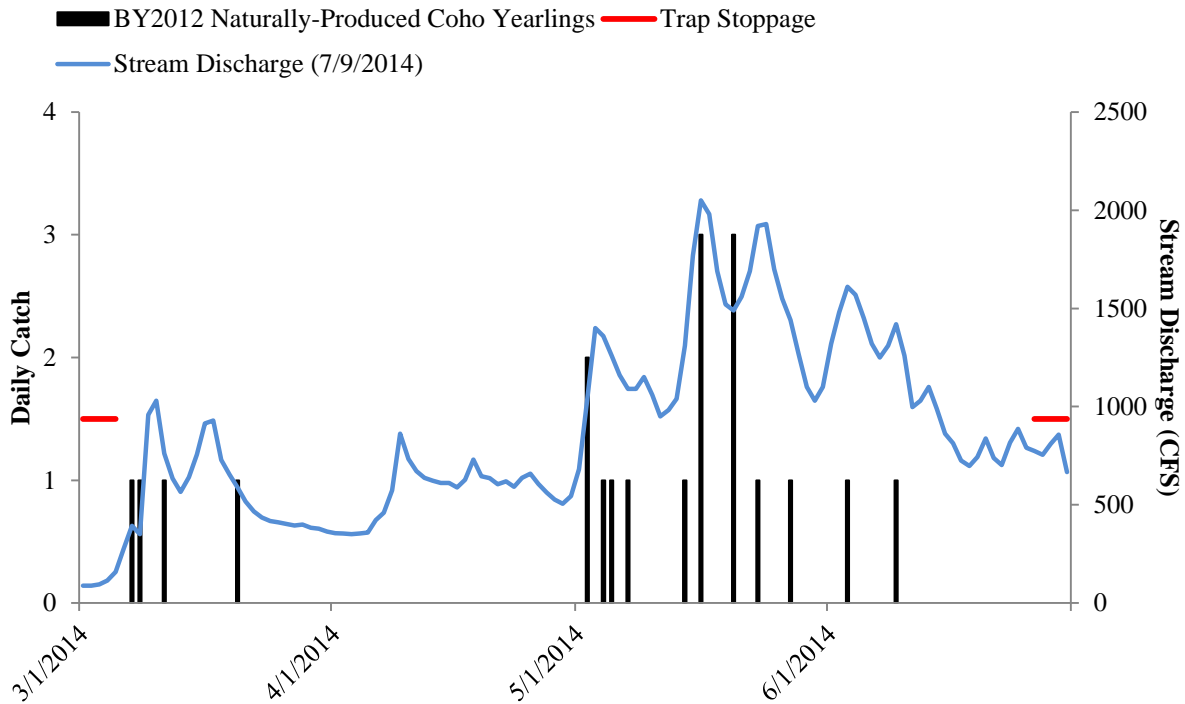


Figure 8. Daily catch of BY2012 naturally produced coho yearlings with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014.

Table 5. Summary of length and weight sampling of juvenile coho salmon captured at the Nason Creek rotary trap in 2014.

Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-Factor
		Mean	<i>n</i>	SD	Mean	<i>n</i>	SD	
2012	Naturally Produced Coho Yearling Smolt	96.3	20	9.8	9.9	20	3.0	1.1
2013	Naturally Produced Coho Subyearling Fry	36.0	1	—	—	—	—	—
2013	Naturally Produced Coho Subyearling Parr	73.0	3	22.5	5.9	3	4.7	1.5
2012	Hatchery Coho Yearling Smolt	127.0	1,203	9.7	21.7	1,201	5.0	1.1

### 3.2.7 Coho Subyearlings (BY2013)

A total of three naturally produced coho subyearling parr were captured during between July 1 and November 30 (Figure 9). Mean FL and weight were 73mm ( $n = 3$ ;  $SD = 22.5$ ) and 5.9g ( $n = 3$ ;  $SD = 4.7$ ), respectively. One additional subyearling coho fry was also captured with a mean FL of 47mm. Scale samples were taken from two coho parr. Collected scale samples will continue to develop a freshwater aging baseline mentioned previously. There was one parr mortality attributed to unknown causes (no signs of external injury).

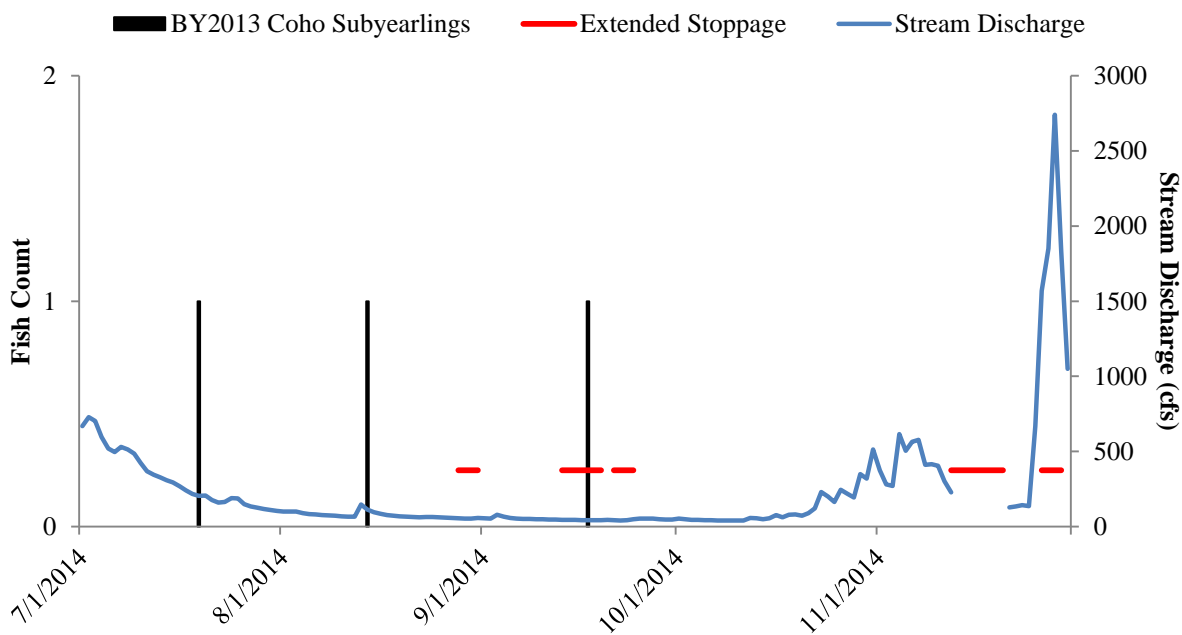
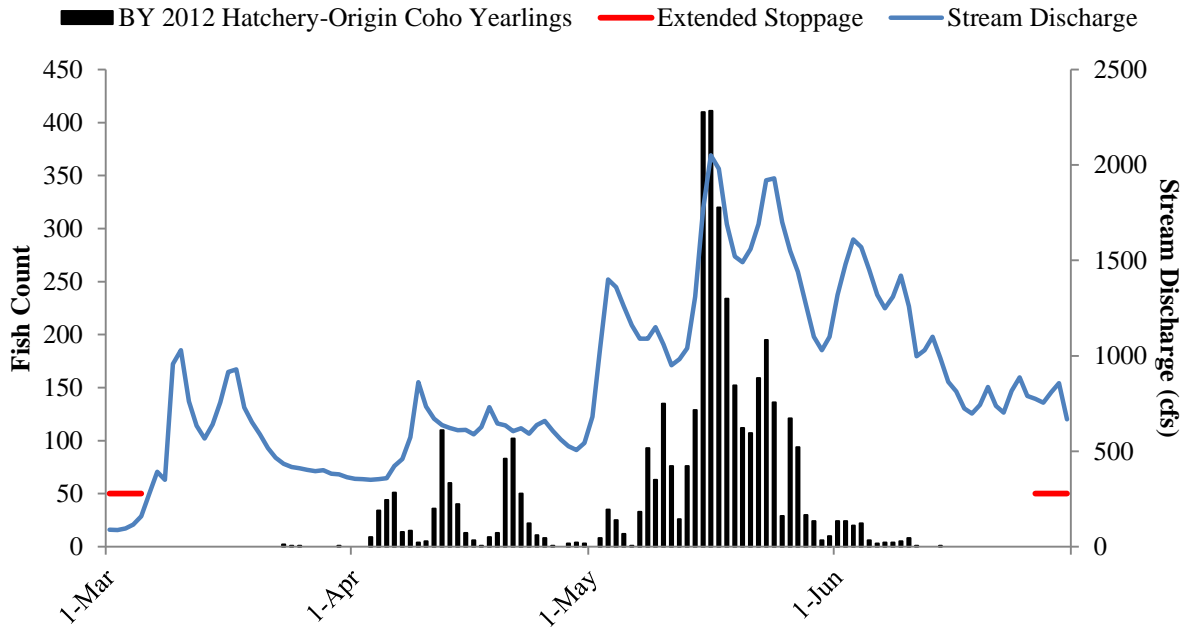


Figure 9. Daily catch of BY2013 naturally produced coho subyearlings with mean daily stream discharge at the Nason Creek rotary trap, July 1 to November 30, 2014.

### 3.2.8 Hatchery Coho Smolts (BY2012)

A total of 253,343 hatchery coho were released into Nason Creek above the trap in spring of 2014. All hatchery coho released were acclimated in natural ponds adjacent to Nason Creek and reared to smolt stage prior to volitional release. Between March 1 and June 30, a total of 4,410 hatchery coho were captured at the trap (Figure 10). Mean FL was 127mm ( $n = 1,203$ ;  $SD = 9.7$ ) and mean weight was 21.7g ( $n = 1,201$ ;  $SD = 5.0$ ; Table 2). Peak daily catch occurred on May 16 ( $n = 411$ ) following volitional release into Nason Creek. There were no hatchery coho smolt mortalities incurred during the 2014 trapping season. Hatchery coho emigration data at the Nason Creek trap assists MCCRP by providing size-at-emigration, emigration timing and duration of residence in Nason Creek.



**Figure 10. Daily catch of BY2012 hatchery coho smolt with mean daily stream discharge at the Nason Creek rotary trap, March 1 to June 30, 2014.**

### 3.3 Remote Parr Tagging (BY2013 Spring Chinook)

YNF and WDFW personnel PIT tagged and released a total of 1,821 BY2013 spring Chinook parr between September 22 and October 24. The total surveyed area included Nason Creek from rkm 0.78 to rkm 26.12. All collections were performed via backpack electrofisher.

Between October 1 and March 30, a total of 311 re-sights of the remote tagged Chinook were documented at NAL (Figure 11). Of these detections, only 13 were during the winter non-trapping period. PTAGIS event logs for NAL indicated that it operated continuously for the duration of this time with no alterations to the array.



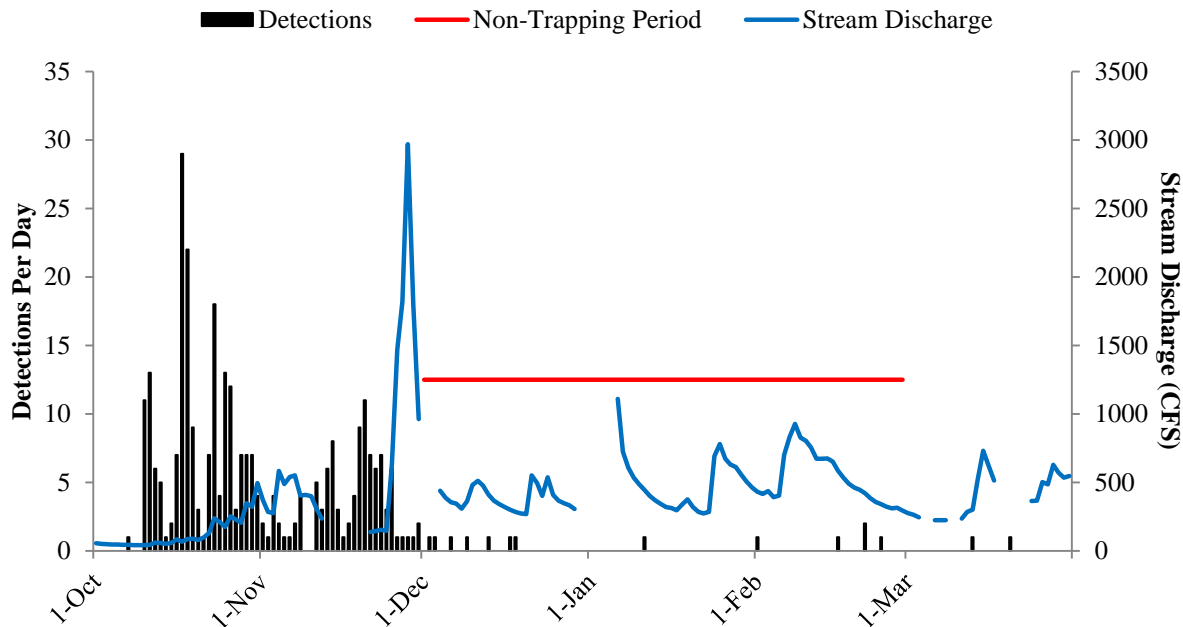


Figure 11. Daily detections of remote-tagged BY2013 spring Chinook at the lower Nason Creek PIT tag antenna array (NAL) between October 2014 and March 2015.

### 3.4 Trap Efficiency Calibration and Population Estimates

#### 3.4.1 Spring Chinook Yearlings (BY2012)

Low abundance of yearling Chinook allowed us to only conduct two efficiency trials in 2014 (Minimum mark group size = 40 smolts; Table 6). The multi-year, weighted flow-efficiency regression was statistically significant ( $r^2 = 0.15$ ,  $p = 0.03$ ; See Appendix C). We estimated a total of 4,561 ( $\pm 1,540$ ; 95% CI) BY2012 Chinook yearlings emigrated in spring of 2014 (Table 7). Combined with a recalculated BY2012 subyearling estimate of 28,110 ( $\pm 4,611$ ; 95% CI), we estimated that a total of 32,671 ( $\pm 4,863$ ; 95% CI) BY2012 spring Chinook juveniles emigrated from Nason Creek during the period of trap operation.

Table 6. Trap efficiency trials conducted with BY2012 wild spring Chinook yearlings. Note: trap efficiency is reported as the percentage of recaptures + 1 divided by the number of marked fish in the trial\*.

Origin/Species/Stage	Age	Date	Trap Position	Marked	Recaptured	Trap Efficiency	Discharge (cfs)
Wild Chinook Yearlings	1+	3/9/2014	Back	65	4	7.69%	958
Wild Chinook Yearlings	1+	3/13/2014	Back	67	9	14.93%	566

\*See equation 3 in 2.5.1 Estimate of Abundance

**Table 7. Estimated egg-to-emigrant survival and smolts-per-redd production for Nason Creek spring Chinook salmon.**

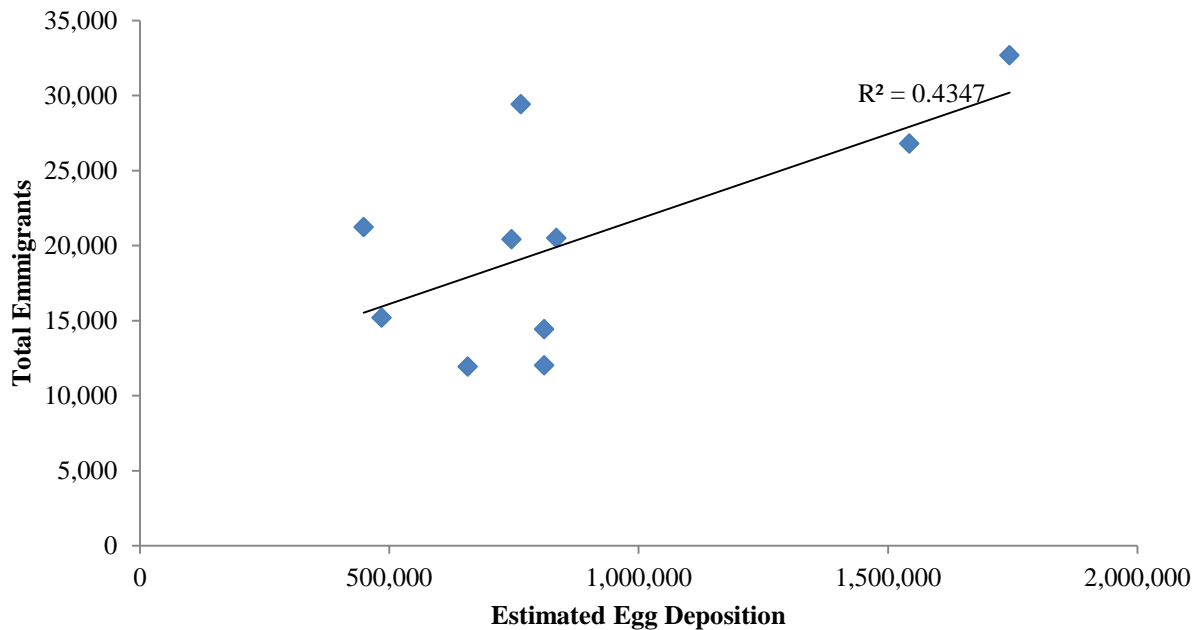
Brood Year	No. of Redds	Fecundity <sup>a</sup>	Est. Egg Deposition	No. of Emigrants			Egg-to-Emigrant	Emigrants per Redd
				Age-0 <sup>b</sup>	Non Trap <sup>d</sup>	Age-1		
2002	294	4,654	1,368,276	DNOT		4,683	—	—
2003	83	5,844	485,052	8,829		6,358	15,187 ± 1,605	3.10%
2004	169	4,799	811,031	11,822		2,597	14,419 ± 2,766	1.80%
2005	193	4,327	835,111	11,814		8,696	20,510 ± 5,018	2.50%
2006	152	4,324	657,248	4,144		7,798	11,942 ± 1,744	1.80%
2007	101	4,441	448,541	15,556		5,679	21,235 ± 2,864	4.70%
2008	336	4,592	1,542,912	23,182		3,611	26,793 ± 6,756	1.70%
2009	167	4,573	763,691	27,720		1,705	29,425 ± 12,777	3.90%
2010	188	4,314	811,032	8,491		3,535	12,026 ± 1,954	1.50%
2011	170	4,385	745,450	17,991		2,422	20,413 ± 3,889	2.70%
2012	413	4,223	1,744,099	28,110		4,561	32,671 ± 4,863	1.90%
2013	212	4,716	999,792	29,784	223	—	—	—
Avg. <sup>c</sup>	197	4,582	884,417	15,766	—	4,696	20,502	2.60%

<sup>a</sup> Data provided by Hillman et al. 2014.

<sup>b</sup> Does not include subyearling fry prior to July 1.

<sup>c</sup> 10-year average of complete brood data, BY2003-2012.

<sup>d</sup> Estimated emigration during the winter non-trapping period (December 1 – February 28).



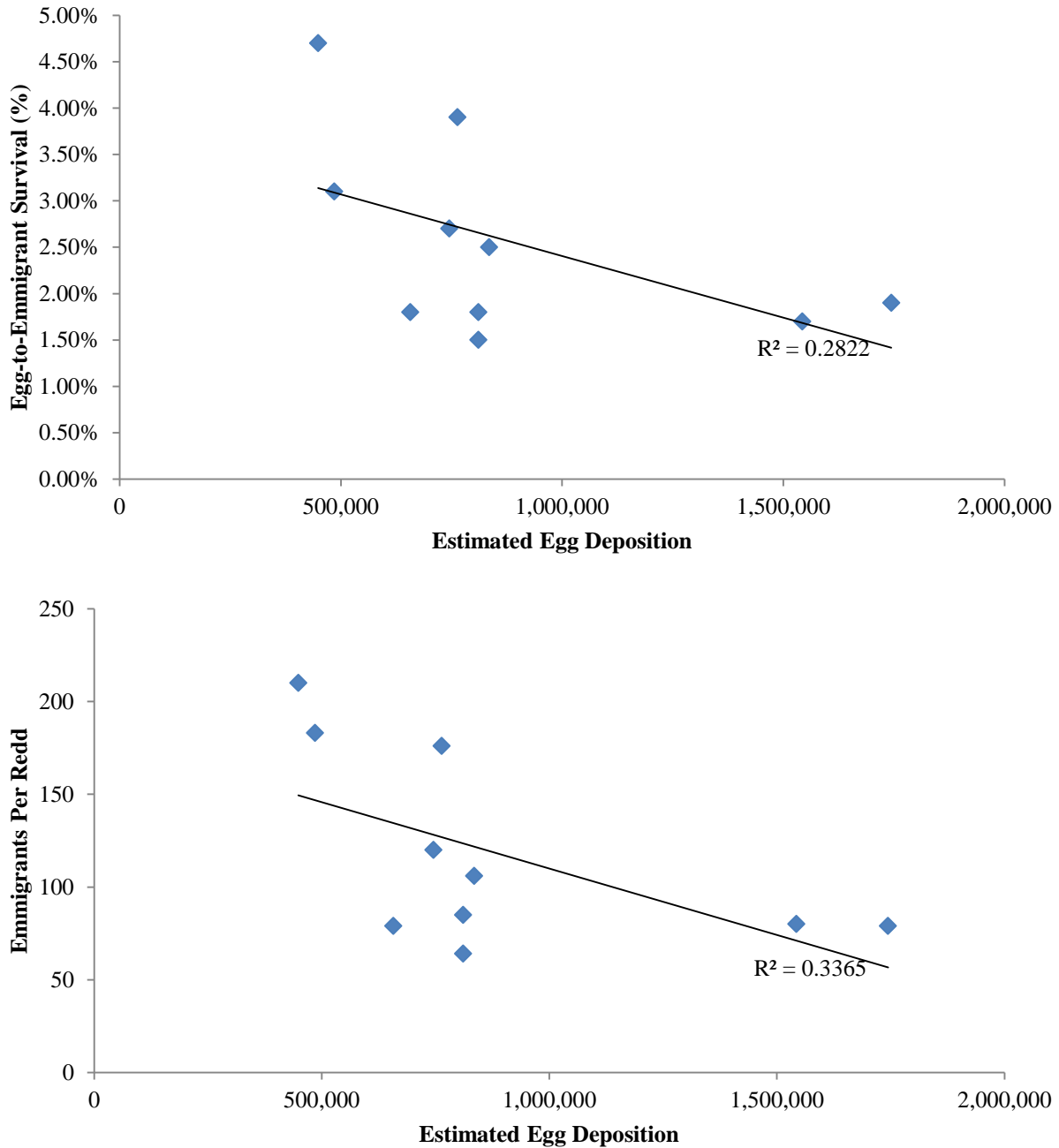


Figure 12. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for Nason Creek spring Chinook, BY 2003 to 2012.

### 3.4.2 Spring Chinook Subyearlings (BY2013)

Initially, attempts to create a flow-efficiency regression were made at Bolser (Table 8). Due to limited parr abundance at higher flows and trap stoppages due to debris and ice, a viable model could not be developed. Instead, a pooled efficiency using mark groups released at the new trapping site was used to calculate parr emigrant abundance (Minimum mark group size = 50 parr; Table 8). Using this model we estimated that a total of 29,784 ( $\pm 32,081$ ; 95% CI) BY2013

spring Chinook emigrated past the trap in the Fall of 2013 (Table 7). Use of this pooled efficiency is seen as only a temporary method of expansion in lieu of the regression model. Continued efforts will be made in 2015 to develop a regression model; at which time recalculation of the BY2013 parr estimate will be made.

**Table 8. Trap efficiency trials conducted with BY2013 wild spring Chinook subyearlings. Note: trap efficiency is reported as the percentage of recaptures + 1 divided by the number of marked fish in the trial\*.**

Origin/Species/Stage	Age	Date	Trap Position	Marked	Recaptured	Trap Efficiency	Discharge (cfs)
Wild Chinook Subyearlings	0	7/14/2014	Back	89	7	8.99%	309
Wild Chinook Subyearlings	0	7/21/2014	Back	74	4	6.76%	176
Wild Chinook Subyearlings	0	7/27/2014	Back	72	4	6.94%	135
Wild Chinook Subyearlings	0	10/27/2014	Back	71	3	5.63%	219
Wild Chinook Subyearlings	0	10/30/2014	Back	70	5	8.57%	320
Wild Chinook Subyearlings	0	11/1/2014	Back	96	6	7.29%	374

\*See equation 3 in **2.5.1 Estimate of Abundance**

Utilizing mark groups previously used to test the efficiency of the smolt trap, a viable flow-antenna efficiency regression was developed for NAL. In order to best describe emigration patterns during this period of no previous efficiency trials, mark-groups included in this regression were limited to those performed one month before and after (November and March) the winter non-trapping period. In March 2010, the NAL array was converted from a floating antenna system to its current flat plate configuration. Because of this change to the array's configuration, all trials performed prior to 2010 were also excluded from the model. The resulting regression showed statistical significance ( $r^2 = 0.64$ ;  $p = 0.0004$ ) and covered a discharge range of 127cfs to 880cfs (see **Appendix C: Regression Models**). Using the Baily Model, we estimate that a total of 233 ( $\pm 253$ ; 95% CI) BY2013 emigrants passed the antenna during the non-trapping period. This estimate of winter migratory movement will be included in the total BY2013 emigrant estimate made at the Nason Creek smolt trap.

### 3.4.3 Summer Steelhead

Summer steelhead efficiency trials were conducted on three separate occasions between March 1 and July 31 (Minimum mark group size = 50 parr/smolt; Table 9). Although an in-year model could not be achieved, M-R data from 2014 was used to strengthen our multi-year steelhead regression. The modified regression was used to produce estimates of abundance for 2014 summer steelhead emigrants as well as recalculations of previous estimates. We utilized a single steelhead model specific to the back position to estimate age 1+ smolt/parr abundance throughout the entire trapping period. Estimates of age-0 fry and parr were not made due to insufficient evidence that active migration is occurring at this young age. Previous attempts to build a model based on YOY steelhead parr in the fall have yielded weak flow-efficiency relationships; further suggesting that age-0 parr catch is the result of displacement rather than active migration. We estimated that 11,837 ( $\pm 3,611$ ; 95% CI) BY2013 age-1 and 813 ( $\pm 731$ ; 95% CI) BY2012 age-2 steelhead emigrated past the trap in 2014 (Table 10). There were no age-3 steelhead identified through age estimation (histogram). We estimate that total (age 1-3) BY2011 emigration to be 13,605 ( $\pm 3,525$ ; 95% CI).

**Table 9. Efficiency trials conducted with wild summer steelhead juveniles. Note: trap efficiency is reported as the percentage of recaptures + 1 divided by the number of marked fish in the trial\*.**

Origin/Species/Stage	Date	Trap Position	Marked	Recaptured	Trap Efficiency	Discharge (cfs)
Wild Steelhead Parr/Smolt	5/3/2014	Back	50	2	6.00%	1400
Wild Steelhead Parr/Smolt	5/30/2014	Back	57	0	1.75%	1030
Wild Steelhead Parr/Smolt	6/3/2014	Back	75	1	2.67%	1610

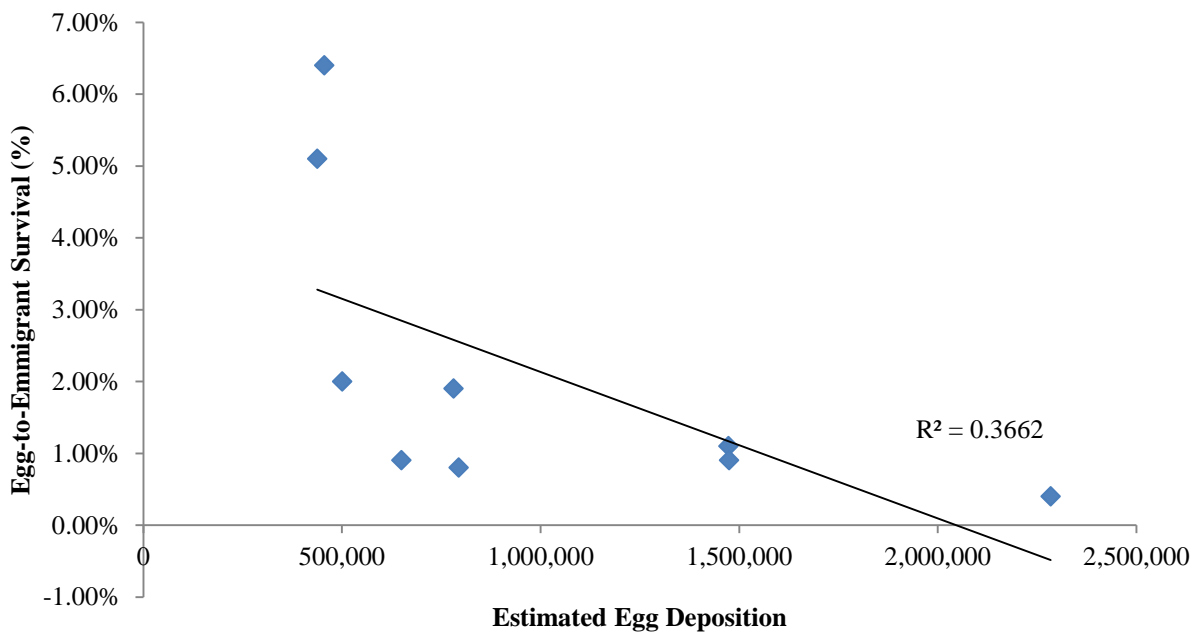
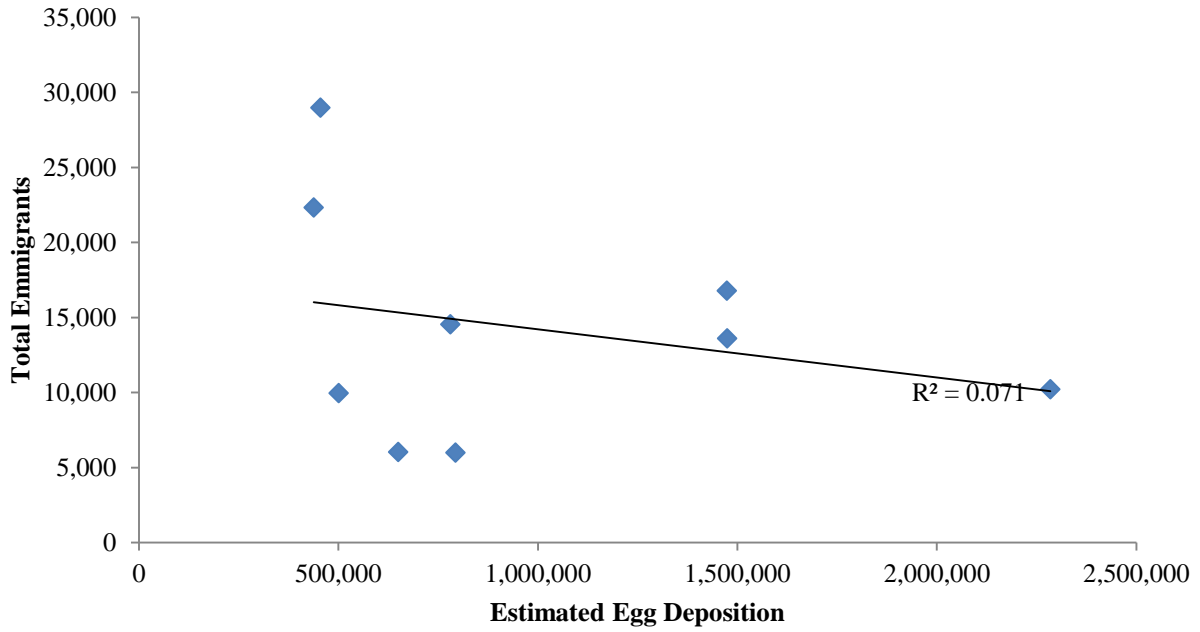
\*See equation 3 in **2.5.1 Estimate of Abundance**

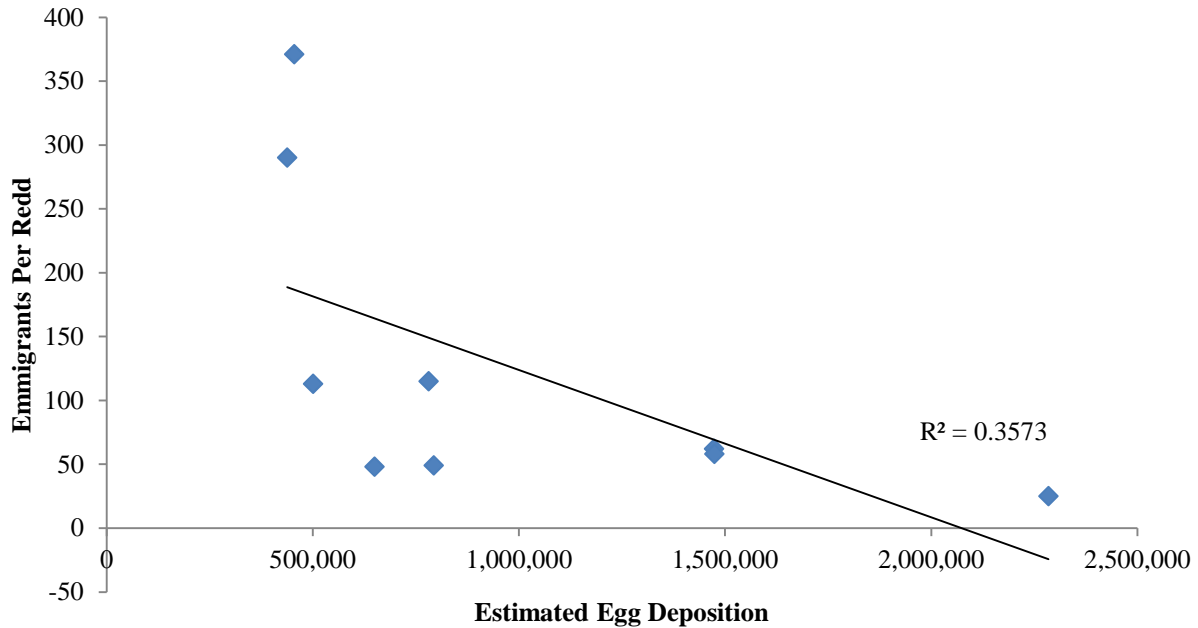
**Table 10. Estimated egg-to-emigrant survival and emigrants-per-redd production for Nason Creek summer steelhead.**

Brood Year	No. of Redds	Fecundity <sup>a</sup>	Est. Egg Deposition	No. of Emigrants				Egg-to-Emigrant	Emigrants per Redd
				1+	2+	3+	Total ± 95%CI		
2001	27	5,951	160,677	DNOT	DNOT	846	—	—	—
2002	80	5,776	462,080	DNOT	2,475	0	—	—	—
2003	121	6,561	793,881	4,906	1,054	27	5,987 ± 1,193	0.8%	49
2004	127	5,118	649,986	5,107	906	22	6,035 ± 885	0.9%	48
2005	412	5,545	2,284,540	7,416	2,502	298	10,216 ± 2,147	0.4%	25
2006	77	5,688	437,976	19,609	2,673	37	22,319 ± 5,722	5.1%	290
2007	78	5,840	455,520	26,518	2,325	117	28,960 ± 7,739	6.4%	371
2008	88	5,693	500,984	8,782	1,164	0	9,946 ± 2,382	2.0%	113
2009	126	6,199	781,074	13,606	608	312	14,526 ± 2,868	1.9%	115
2010	270	5,458	1,473,660	12,767	3,999	0	16,776 ± 3,885	1.1%	62
2011	235	6,276	1,474,860	13,109	482	0	13,605 ± 3,525	0.9%	58
2012	212	5,309	1,125,508	26,637	813	—	—	—	—
2013	135	5,761	777,735	11,837	—	—	—	—	—
Avg <sup>b</sup>	170	5,820	983,609	12,424	1,747	90	14,262	0	126

<sup>a</sup> Data provided by Hillman et al. 2014.

<sup>b</sup> 8-year average of complete brood estimates, BY2003-2011.





**Figure 13. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for Nason Creek summer Steelhead, BY 2003 to 2011.**

### 3.4.4 Coho Yearlings (BY2012)

Limited abundance of BY2012 coho yearlings did not provide any opportunities to perform any efficiency trials in the spring of 2014. As in previous years, a wild spring Chinook yearling model was applied to coho to provide a smolt estimate ( $r^2 = 0.15$ ,  $p = 0.03$ ; See Appendix C). In the spring of 2014, we estimated that 434 ( $\pm 235$ ; 95% CI) emigrated past the trap (Table 11). This gave us a total BY2012 emigrant estimate of 479 ( $\pm 237$ ; 95% CI).

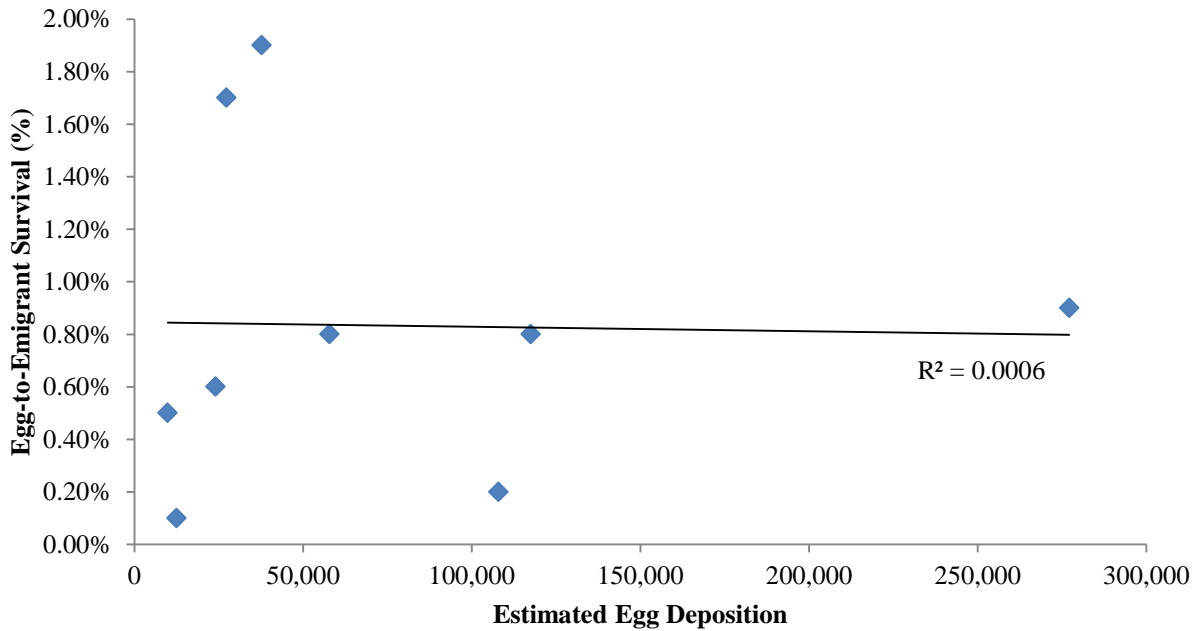
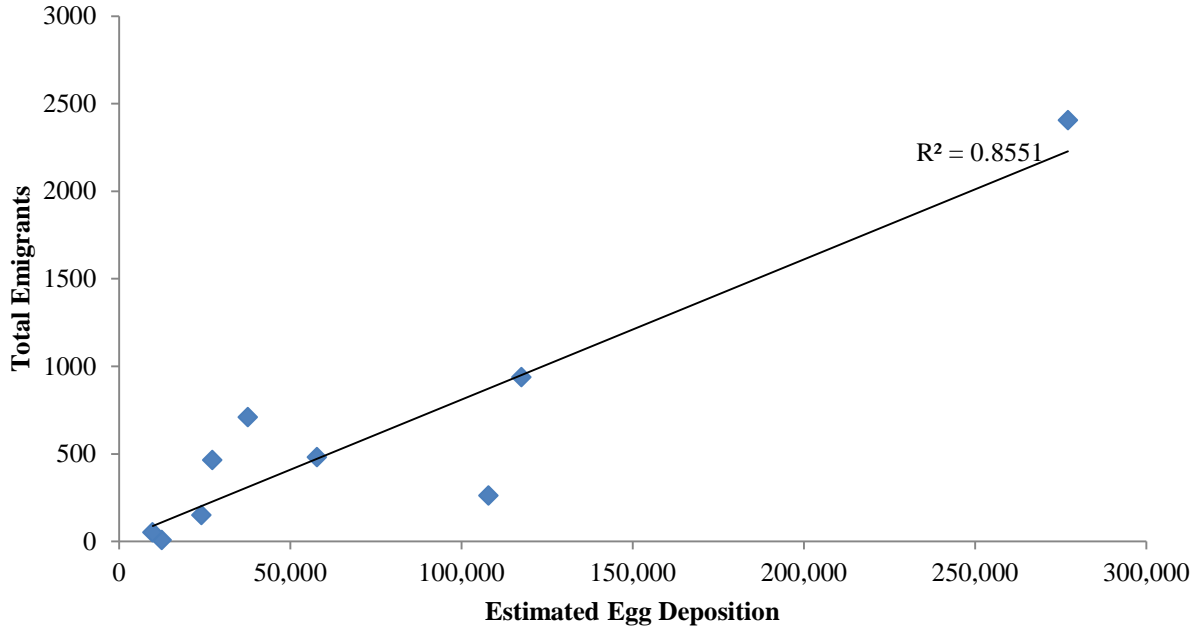
**Table 11. Estimated egg-to-emigrant survival and smolts-per-redd production for Nason Creek coho salmon.**

Brood Year	No. of Redds	Fecundity	Est. Egg Deposition	No. of Emigrants			Egg-to-Emigrant	Emigrants per Redd
				Age-0 <sup>a</sup>	Age-1	Total $\pm$ 95% CI		
2003	6	2,458	14,748	DNOT	394	—	—	—
2004	35	3,084	107,940	204	56	260 $\pm$ 155	0.2%	7
2005	41	2,866	117,506	27	910	937 $\pm$ 347	0.8%	23
2006	4	3,126	12,504	7	0	7 $\pm$ 10	0.1%	2
2007	10	2,406	24,060	14	136	150 $\pm$ 104	0.6%	15
2008	3	3,275	9,825	50	0	50 $\pm$ 57	0.5%	17
2009	14	2,691	37,674	471	237	708 $\pm$ 478	1.9%	51
2010	8	3,411	27,288	27	437	464 $\pm$ 231	1.7%	58
2011	89	3,114	277,146	1,018	1,387	2,405 $\pm$ 612	0.9%	27
2012	21	2,752	57,792	46	434	480 $\pm$ 237	0.8%	23
2013	0	—	—	49	—	—	—	—

Avg. <sup>b</sup>	25	2,969	74,637	207	400	607	0.8%	25
-------------------	----	-------	--------	-----	-----	-----	------	----

<sup>a</sup> Does not include subyearling fry prior to July 1.

<sup>b</sup> 9-year average of complete brood data, BY2004-2012.





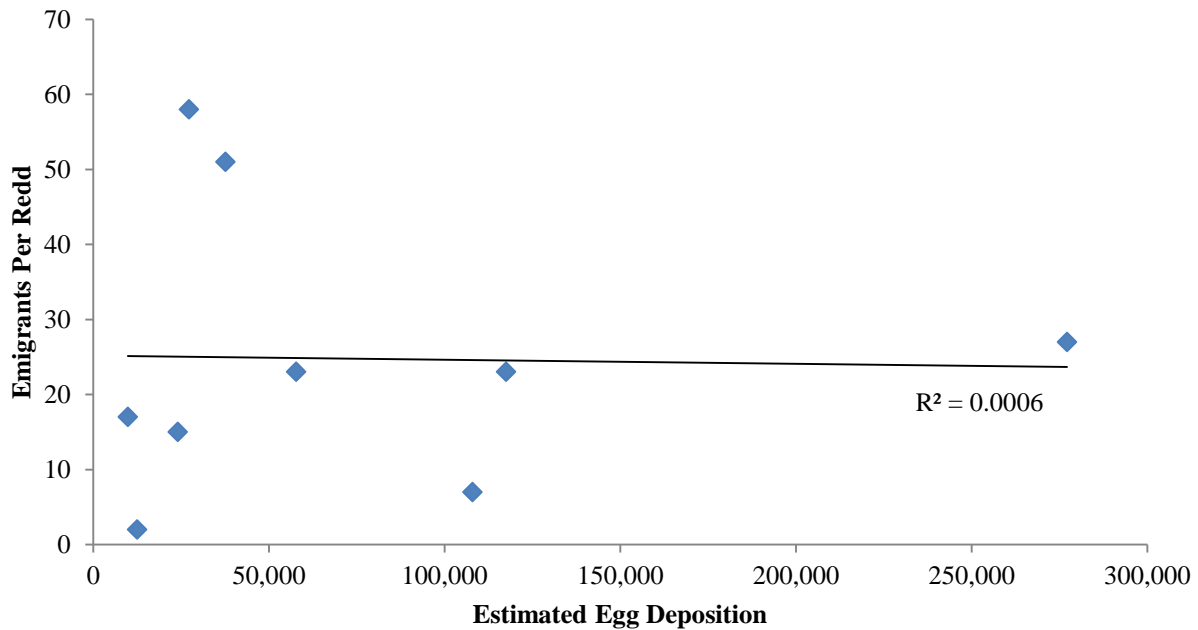


Figure 14. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for Nason Creek naturally-produced coho, BY 2004 to 2012.

### 3.4.5 Coho Subyearlings (BY2013)

A total of only three coho subyearling parr did not allow us to make any attempts to build an species/age specific a regression model at the new trap location. Because these subyearling emigrants were captured at the DB site prior to establishment of a viable flow-efficiency regression, a pooled estimate based on subyearling Chinook parr efficiency trials performed in the summer/fall of 2014 was used. Using the pooled efficiency, we estimated that 49 ( $\pm 82$  ; 95% CI) emigrated past the trap in the fall of 2014 (Table 11).

### 3.5 PIT Tagging

During the 2014 trapping season, we PIT tagged 1,578 wild spring Chinook, 838 steelhead, and 22 naturally produced coho (Table 12). All tagging files were submitted to the PTAGIS database. There were no shed PIT tags were recovered in holding boxes where fish had been held for 24-72 hours after tagging.

Table 12. Number of PIT tagged coho, Chinook, and steelhead with shed rates at the Nason Creek rotary trap in 2014.

Species/Stage	Year-to-date Catch	Year-to-date PIT Tagged	No. of Shed Tags	Percent Shed Tags
Chinook Yearling Smolt	464	456	0	0.00%
Chinook Subyearling Parr (Mar 1 to June 30)	62	4	0	0.00%
Chinook Subyearling Parr (July 1 to Nov 30)	1,502	1,109	0	0.00%

Steelhead Parr	991	820	0	0.00%
Steelhead Smolt	18	18	0	0.00%
Coho Yearling Smolt	20	20	0	0.00%
Coho Subyearling Parr	3	2	0	0.00%

\* Counts do not include fish with FL<50mm (fry).

During remote tagging efforts in the fall of 2014, 1,893 spring Chinook were PIT tagged by YN and WDFW personnel. Of the total tagged, 78% were held overnight to determine tag retention. Shed rate for this tagging effort was 0.07%.

### 3.6 Incidental Species

Along with wild spring Chinook, wild steelhead/rainbow trout, and naturally produced coho, other resident fish species captured at the Nason Creek rotary trap and included in Table 13 are: bull trout *Salvelinus confluentus*, cutthroat trout *Oncorhynchus clarki*, flathead minnow *Pimephales promelas*, longnose dace *Rhinichthys cataractae*, northern pikeminnow *Ptychocheilus oregonensis*, redbside shiner *Richardsonius balteatus*, sculpin *Cottus sp.*, sucker *Catostomus sp.*, and mountain whitefish *Prosopium williamsoni*.

**Table 13. Summary of length and weight sampling of incidental species captured at the Nason Creek rotary trap in 2014.**

Species	Total Count	Length (mm)			Weight (g)		
		Mean	N	SD	Mean	N	SD
Bull Trout	4	150.2	4	12.8	33.4	4	9.8
Cutthroat Trout	2	272.0	2	5.7	200.3	2	4.9
Fathead Minnow	26	58.6	26	5.3	2.5	26	0.7
Longnose Dace	144	63.9	144	29.0	6.7	114	6.8
Northern Pikeminnow	12	150.2	11	98.0	100.0	8	116.5
Redside Shiner	7	60.6	7	11.8	2.9	6	1.9
Sculpin	70	72.4	70	38.6	12.3	57	17.4
Sucker	24	116.8	24	90.6	18.3	21	28.1
Whitefish Fry	27	37.0	27	5.4	—	—	—
Whitefish Parr	42	79.6	42	47.7	9.8	41	33.0

### 3.7 ESA Compliance

The Nason Creek smolt trap was operated under consultation with NMFS and USFWS. Total numbers of UCR spring Chinook and UCR summer steelhead that were captured or handled (indirect take) at the trap were less than the maximum permitted (20%) for each species. Lethal take was well below the allowable level of 2% for spring Chinook (0.41%), summer steelhead (0.08%), and bull trout (0.0%; Table 14). Stream temperatures did not exceeded 18°C at any time in which fish were being handled.

**Table 14. Summary of ESA species and coho salmon mortality at the Nason Creek rotary trap.**

Species/Stage/Brood Year	Total Collected	Total Mortality	% Mortality
Spring Chinook Yearling (BY2012)	464	0	0.00%
Spring Chinook Subyearling (BY 2013)	2,229	11	0.49%
<b>Total Wild Spring Chinook</b>	<b>2,693</b>	<b>11</b>	<b>0.41%</b>
Steelhead Age-0 (BY2014)	491	1	0.20%
Steelhead Age-1 (BY2013)	745	0	0.00%
Steelhead Age-2 (BY2012)	30	0	0.00%
<b>Total Wild Summer Steelhead</b>	<b>1,266</b>	<b>1</b>	<b>0.08%</b>
<b>Total Hatchery Summer Steelhead</b>	<b>4,140</b>	<b>0</b>	<b>0.00%</b>
<b>Total Bull Trout</b>	<b>4</b>	<b>0</b>	<b>0.00%</b>
Coho Yearling (BY2012)	20	0	0.00%
Coho Subyearling (BY2013)	4	0	0.00%
<b>Total Coho</b>	<b>24</b>	<b>0</b>	<b>0.00%</b>

## 4.0 DISCUSSION

Operation of the Nason Creek smolt trap at campground endured several chronic problems during the past eleven trapping seasons. Most notable of these included both shifting channel morphology and the seasonal threat of vandalism. Thalweg positioning at campground varied drastically with changing discharge levels. This posed a significant problem in that our flow-efficiency models are assigned temporally to a static trap position/configuration. Because the trap could therefore not be moved readily within a specific migratory period without confounding the associated model, we periodically found ourselves trapping well outside the thalweg's bounds as flows fluctuated. We believe these problems warranted the relocation of the smolt trap. The alternate location (Bolser property) at ~rkm 0.3 was chosen for its ease of access, remote setting (to dissuade vandalism), and favorable channel morphology. Relocation of the trap offered the opportunity to improve our estimates while minimizing the threat to both fish and public safety.

In order to ensure that the move coincided with the onset of the subyearling spring Chinook (BY2013) migratory period (July 1), relocation occurred in late June of 2014. In doing so, we prevented splitting collections of the BY2012 brood between the two trap locations. The trap was positioned on the river-right bank of a slight bend in the river where the thalweg was pushed to the outer-edge of the channel. Once the position of the trap was established, an anchoring system was created that ensured a static location throughout the hydrograph. We intend to operate the smolt trap in this configuration for the duration of its lifetime at this location.

Initial trap operations at Bolser in the summer and fall of 2014 presented no significant challenges. The trap ran successfully until approximately 45cfs, at which time water velocity could no longer rotate the cone consistently. This is an improvement over campground, which could only be operated until approximately 60 cfs. We surmise that the low cone speeds experienced at extreme base flow is due in part to an unbalanced cone. A replacement cone will be installed in 2015, which we expect will allow us to operate the smolt trap at even lower discharge levels. There were no signs of tampering with the smolt trap or suspected intentional stoppages by the public at large.

### *Spring Chinook*

Returning adult spring Chinook in 2012 produced a record 413 redds in Nason Creek. Despite the relatively low average fecundity of these adults, high spawner recruitment in 2012 yielded an estimated egg deposition twice that of the ten-year average. Reflecting the spawner escapement, our BY2012 estimate of 32,671 spring Chinook was the largest that we have estimated since the trap was established. Historical brood estimates (2003-2012) indicate that an average of 76.9% of the brood leave prior to overwintering. The observed migratory timing of BY2012 spring Chinook juveniles was typical of Nason Creek with the majority (86.0%) of the cohort moving out of the system as subyearling parr. Our pooled BY2013 subyearling spring Chinook parr of 29,784 emigrants should be considered provisional until a viable regression model can be built. The pooled efficiency used to calculate this estimate was admittedly skewed toward the few efficiency trials that could be performed at lower flows. The resulting estimate would therefore inherently overestimate abundance. Combined with the non-trapping period estimate of 223 emigrants, we estimate that a total of 30,007 BY2013 spring Chinook emigrated out of Nason Creek by March 2015.

Comparison of spring Chinook estimates to those of other upper-Wenatchee tributary smolt traps suggests that egg-to-emigrant survival in Nason Creek is relatively low (Figure 15). With the exception of BY2009, Nason Creek egg-to-emigrant survival has consistently fallen below those of the Chiwawa River and White River, which is not surprising given the extent of development and associated habitat impacts in Nason Creek. Comparison with the other Wenatchee Basin tributaries also indicates that BY2012's low rate of egg-to-emigrant survival despite high spawner escapement is not a tributary-specific phenomenon (Figure 16). Strong 2012 adult returns to both the Chiwawa River and White River also produced low in-stream survival rates.

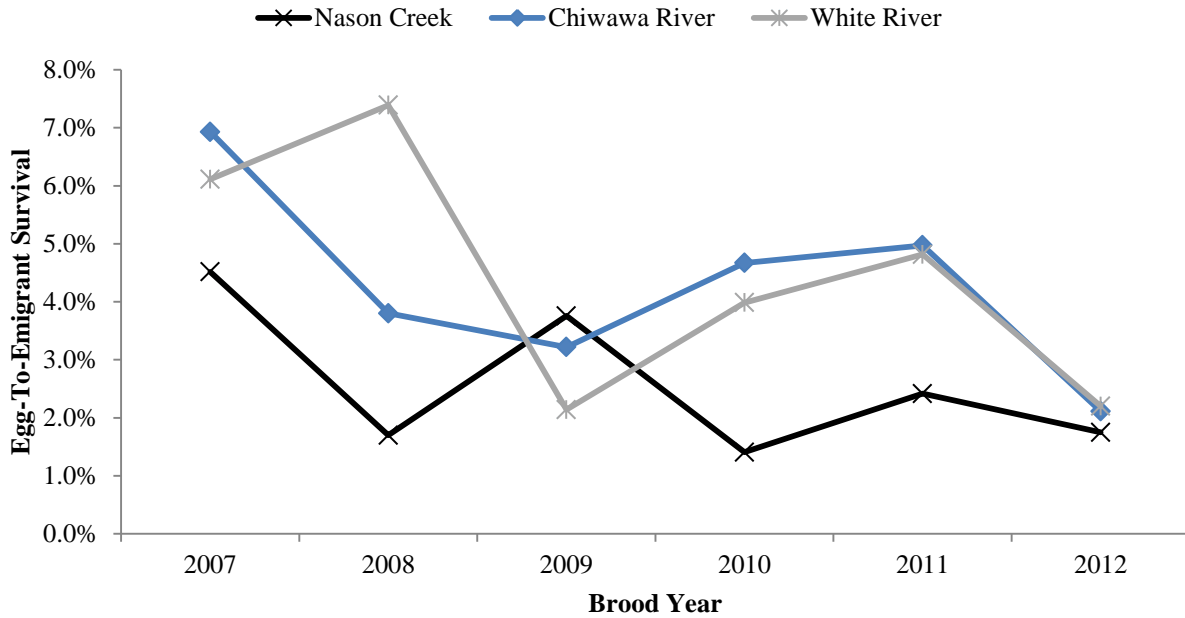


Figure 15. Comparison of wild spring Chinook abundance estimates (BY2007-2012) made at the White River, Nason Creek, and Chiwawa River smolt traps.

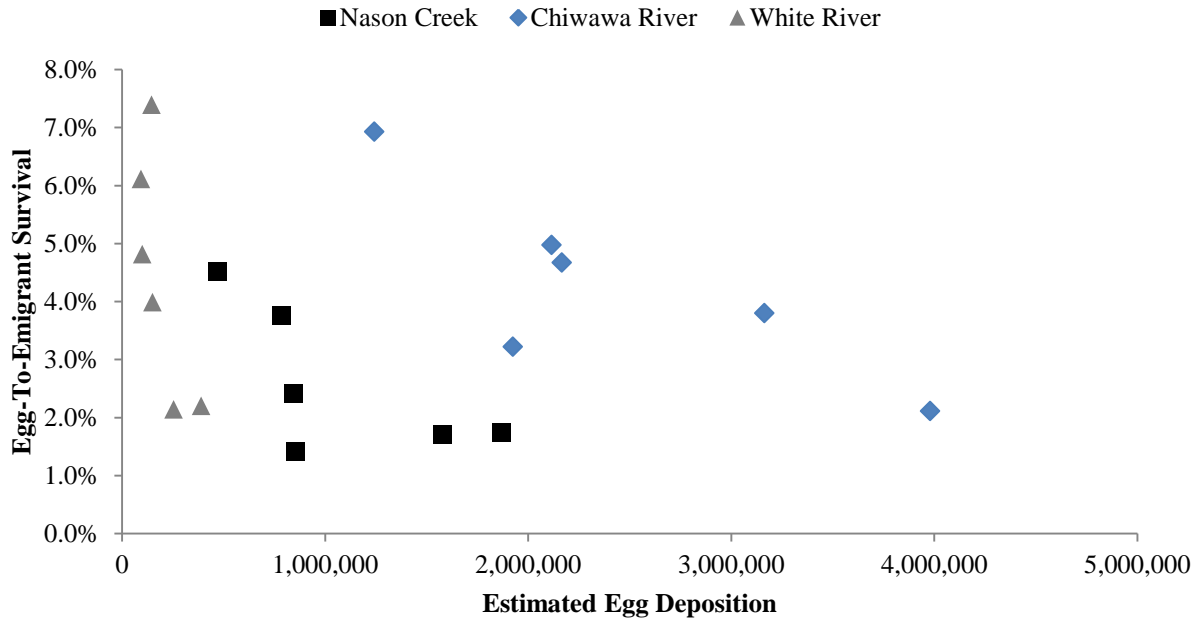


Figure 16. Comparison of egg-to-emigrant survival (BY 2007-2012) and egg deposition for Nason Creek, Chiwawa River, and White River spring Chinook. \*Chiwawa BY2012 spring Chinook data provided by WDFW and to be considered provisional until peer review

### Summer Steelhead

Unable to produce an in-year regression model as we did in 2013, estimates produced in 2014 were based on a multi-year flow-efficiency regression. All age 1+ steelhead captured between March 1 and June 30 were considered migratory, and subsequently included in the model. Steelhead catch numbers at Bolser (July1-November 30) were not expanded because a viable regression or alternate method of estimation could not be developed in the first year of operation. The impact of this exclusion was ultimately minimal as the overwhelming majority (98.5%) of steelhead captured in the summer/fall are age-0 young-of-the-year, and therefore excluded from all estimation. Age 1+ steelhead presence at Bolser was minimal throughout the duration of trapping ( $n = 6$ ). Expansions of daily age 1+ steelhead catch at the new trap location will be made and added to their respective brood years as soon as the necessary regression can be developed.

Steelhead estimates produced in 2014 concluded our final BY2011 emigrant estimate (age 1-3) with a total of 13,605 parr/smolt. Because there were no age-3 steelhead captured in 2014, the temporary inability to expand steelhead daily abundance at Bolser did not affect the validity of this brood estimate. Although total emigrants estimated was near the nine-year mean, BY2011 egg-to-emigrant survival was well below the running average. Consideration of the relatively strong 2011 spawner success suggests density dependent mortality may be a limiting factor. However, the ongoing (age 1-2) BY2012 estimate ( $n = 25,450$ ) demonstrates that a relatively large return can produce a brood with a high relative rate of survival. The initial BY2013 age-1

steelhead estimate of 11,837 emigrants fall close to the nine-year average, as does the number of redds observed in 2013.

The general migratory timing of the different steelhead brood years caught in 2014 was typical of Nason Creek. The overwhelming majority of migratory steelhead encountered at the trap were captured during spring run-off. During this exodus, age-1 parr represented approximately 85% of the total steelhead caught while only 3% of the steelhead captured were age-2 fish of the previous brood. Age-3 steelhead are infrequently captured in Nason Creek and completely absent at the smolt trap in 2014. As flows subsided, age-0 fry became the predominant age class. Age 1+ steelhead catch in the summer and fall is infrequent, suggesting that most steelhead rarely spend more than a year rearing in Nason Creek.

### ***Coho***

The BY2012 naturally-produced coho estimate of 480 emigrants falls below the nine-year average of 607 emigrants. While the yearling component of this emigrant estimate fell close to the nine-year smolt mean ( $n = 434$ ), subyearling representation appears lower than expected ( $n = 46$ ). A poor adult coho return in 2013 led to exhaustive measures by YN FRM to collect the necessary broodstock to supply its hatchery-supplementation efforts. Included in these measures was the retention of 100% of all coho collected at tumwater dam until late in the run. A total of only 32 adult coho were allowed to pass above the dam, none of which were observed spawning in Nason creek. Despite no observed redds in 2013, 49 subyearlings were estimated to have passed the trap in 2014.

Juvenile coho in Nason Creek primarily emigrate as yearling smolts. Exceptions to this pattern do exist however, with a greater proportion of subyearling emigrants in BY2004, BY2006, BY2008, and BY2009. Comparison of average egg-to-emigrant survival rates of coho to those of spring Chinook suggest that coho are surviving at lower rates than spring Chinook (0.8% vs 2.6%), this could be in part due to the coho stock used in the reintroduction program not being considered 'locally adapted'. Reported egg- to emigrant survival rates are also affected by the proportion of fish that leave in the fall as sub-yearlings (77% for spring Chinook compared to 34% for coho) since overwinter mortality would not be included in the estimate. While subsequent monitoring efforts (e.g. - remote PIT tagging, reliable trap estimates, etc.) have not been possible due to low adult escapements over most years, a future programmatic shift towards maximizing natural production in key tributaries (Natural Production Phases; YN Master Plan) should help alleviate uncertainties regarding migration timing and survival.

### ***2015 Trap Operations at Nason Creek***

Trap operations in 2015 will be performed entirely at Bolser. At the onset of the 2015 season, our main priority will be the development of our flow efficiency regressions. Part of this process is the adherence to a strict set of trap operation and fish-handling protocols. Steps have been taken to ensure that all efficiency trials are both necessary and adequately robust prior to release. We ultimately intend to produce year-specific flow-efficiency regressions annually for each species and age class estimated. Multi-year regressions will be continually improved as a failsafe in the event that low abundance prevents in-year modeling.

The additional PIT-tagging effort in the fall of 2013 allowed for an estimate of subyearling migration during the winter non-trapped period. Using the NAL PIT-array model, an estimated

233 out of 30,007 subyearlings (0.7%) migrated during the winter non-trapping period. These results support the assumption that the migration of juvenile spring Chinook during the winter period is minimal. Future Nason Creek spring Chinook winter migration will continue to be monitored by additional remote parr tagging. A joint effort by GCPUD, WDFW, and YN FRM has targeted up to 3,000 spring Chinook parr that will be systematically tagged throughout Nason Creek in the fall of 2015. Successful development of a viable regression and estimate of winter migrants in this report marks the first time in which Chinook movement in the non-trapping period could be quantified at Nason Creek. This tool contributes to our ability to test assumptions inherent in screw trap based estimates of migrant abundance.



## 5.0 LITERATURE CITED

CBFWA (Columbia Basin Fish and Wildlife Authority). 1999. PIT tag marking procedures manual, version 2.0. Columbia Basin Fish and Wildlife Authority, Portland OR.

Everhart, W.H. and W.D. Youngs. 1981. Principles of Fishery Science, second edition. Comstock Publishing Associates, a division of Cornell University Press, Ithica and London.

Hillman, T.W. 2004. Monitoring strategy for the Upper Columbia Basin: Draft report February 1, 2004. *Prepared for* Upper Columbia Regional Technical Team, Wenatchee, Washington.

Hillman, T.M., T. Kahler, G. Mackey, J. Murauskas, A. Murdoch, K. Murdoch, T. Pearsons, and M. Tonseth. 2013. Monitoring and Evaluation Plan for PUD Hatchery Programs: 2013 Update. *Prepared for:* HCP and PRCC Hatchery Committees.

Murdoch, A. T. Miller, B. L. Truscott, C. Snow, C. Frady, K. Ryding, J. Arteburn and D. Hathaway. 2012. Upper Columbia Spring Chinook Salmon and Steelhead Juvenile and Adult Abundance, Productivity and Spatial Structure Monitoring. BPA Project 2010-034-00.

Murdoch, A., and K. Petersen. 2000. Freshwater Production and Emigration of Juvenile Spring Chinook from the Chiwawa River in 2000. Washington State Department of Fish and Wildlife

PTAGIS (Columbia Basin PIT Tag Information System). 2014. Interrogation Site Metadata: <http://www.ptagis.org/sites/interrogation-site-metadata?IntSiteCode=NAL>

Seber, G.A.F. 1982. The Estimation of Animal Abundance and Related Parameters, 2<sup>nd</sup> edition. Edward Arnold: London

Tussing, S.P. 2008. A Field Manual of Scientific Protocols for Downstream Migrant Trapping within the Upper Columbia Monitoring Strategy: 2008 Working Version 1.0. Prepared for Bonneville Power Administration's Integrated Status and Effectiveness Monitoring Program.

UCRTT (Upper Columbia Regional Technical Team). 2001. A Strategy to Protect and Restore Salmonid Habitat in the Upper Columbia Region, a Discussion Draft Report. Upper Columbia Salmon Recovery Board.

USFS (United States Forest Service). 1996. Nason Creek Stream Survey Report.

WDOE (Washington State Department of Ecology). 2013. River and Stream Flow Monitoring: <https://fortress.wa.gov/ecy/wrx/wrx/flows/station.asp?sta=45J070>

YNFRM (Yakama Nation Fisheries Resource Management). 2010. Mid-Columbia Coho Restoration Master Plan. *Prepared for:* Northwest Power and Conservation Council, Portland OR.

## APPENDIX A. Daily Stream Discharge and Stream Temperature

Date	Stream Discharge (CFS)	Water Temperature (°C)			
			2/8/2014	148	0.1
			2/9/2014	151	0.1
			2/10/2014	151	0.1
1/1/2014	159	2	2/11/2014	156	0.1
1/2/2014	161	2.3	2/12/2014	159	0.1
1/3/2014	225	2.4	2/13/2014	179	0.1
1/4/2014	183	0.9	2/14/2014	157	0.1
1/5/2014	170	0.1	2/15/2014	148	0.1
1/6/2014	166	0	2/16/2014	139	0.1
1/7/2014	159	0.7	2/17/2014	137	0.1
1/8/2014	155	1	2/18/2014	137	0.1
1/9/2014	155	0.9	2/19/2014	132	0.1
1/10/2014	176	1.5	2/20/2014	127	0.1
1/11/2014	295	1.5	2/21/2014	119	0.1
1/12/2014	309	1.7	2/22/2014	108	0.5
1/13/2014	447	2.9	2/23/2014	104	0.6
1/14/2014	493	2.7	2/24/2014	101	0.3
1/15/2014	382	2.5	2/25/2014	97.5	0.7
1/16/2014	332	2.1	2/26/2014	93	1.5
1/17/2014	304	2	2/27/2014	90.8	2
1/18/2014	280	1.8	2/28/2014	90.1	2.6
1/19/2014	263	1.7	3/1/2014	88.4	1.4
1/20/2014	247	1.7	3/2/2014	87.6	0.1
1/21/2014	235	1.5	3/3/2014	94.9	0.1
1/22/2014	225	1.4	3/4/2014	115	0.1
1/23/2014	215	1.8	3/5/2014	159	0.9
1/24/2014	206	1.7	3/6/2014	276	1.1
1/25/2014	198	1.6	3/7/2014	393	1.5
1/26/2014	192	1.3	3/8/2014	351	1.6
1/27/2014	185	1.2	3/9/2014	958	1.2
1/28/2014	179	1.7	3/10/2014	1030	1.9
1/29/2014	185	0.3	3/11/2014	761	2.3
1/30/2014	177	1.1	3/12/2014	634	2.5
1/31/2014	166	2.2	3/13/2014	566	3
2/1/2014	159	2.1	3/14/2014	640	2.9
2/2/2014	152	1.5	3/15/2014	756	2.9
2/3/2014	148	0.8	3/16/2014	915	3.2
2/4/2014	138	0.1	3/17/2014	929	2.8
2/5/2014	120	0.1	3/18/2014	729	3
2/6/2014	112	0.1	3/19/2014	653	3.3
2/7/2014	137	0.1	3/20/2014	588	3

3/21/2014	516	2.8	5/3/2014	1400	5.4
3/22/2014	466	2.9	5/4/2014	1360	4.9
3/23/2014	435	3.8	5/5/2014	1260	5.5
3/24/2014	418	4.1	5/6/2014	1160	5.7
3/25/2014	411	3.9	5/7/2014	1090	5.8
3/26/2014	403	4	5/8/2014	1090	5.2
3/27/2014	395	4.6	5/9/2014	1150	5.6
3/28/2014	400	3.6	5/10/2014	1060	5.5
3/29/2014	383	4	5/11/2014	951	6
3/30/2014	379	4.5	5/12/2014	982	6.2
3/31/2014	364	4.3	5/13/2014	1040	6.5
4/1/2014	356	4.7	5/14/2014	1310	6.6
4/2/2014	354	4.7	5/15/2014	1770	5.9
4/3/2014	351	4.6	5/16/2014	2050	6
4/4/2014	354	5.2	5/17/2014	1980	5.7
4/5/2014	359	4.7	5/18/2014	1690	5.9
4/6/2014	422	5.3	5/19/2014	1520	6.3
4/7/2014	460	5.6	5/20/2014	1490	6.6
4/8/2014	573	5.4	5/21/2014	1560	6.8
4/9/2014	862	4.5	5/22/2014	1690	7
4/10/2014	734	4.5	5/23/2014	1920	6.2
4/11/2014	671	5.1	5/24/2014	1930	6.4
4/12/2014	637	5.1	5/25/2014	1700	5.7
4/13/2014	622	5	5/26/2014	1550	6.3
4/14/2014	611	4.9	5/27/2014	1440	6.1
4/15/2014	612	5.5	5/28/2014	1270	6.3
4/16/2014	589	5.2	5/29/2014	1100	6.9
4/17/2014	627	5.3	5/30/2014	1030	6.6
4/18/2014	731	5.1	5/31/2014	1100	7.8
4/19/2014	645	4.4	6/1/2014	1320	7.7
4/20/2014	636	5.1	6/2/2014	1480	7.4
4/21/2014	605	5.2	6/3/2014	1610	7.6
4/22/2014	620	5.7	6/4/2014	1570	7.4
4/23/2014	592	5.3	6/5/2014	1450	7.5
4/24/2014	637	5.4	6/6/2014	1320	7.5
4/25/2014	659	5.5	6/7/2014	1250	7.9
4/26/2014	607	5.6	6/8/2014	1310	8.2
4/27/2014	562	5.6	6/9/2014	1420	8.2
4/28/2014	526	5.8	6/10/2014	1260	7.2
4/29/2014	506	6.4	6/11/2014	998	7.6
4/30/2014	544	6.7	6/12/2014	1030	8.1
5/1/2014	681	6.8	6/13/2014	1100	6.9
5/2/2014	1040	6.2	6/14/2014	990	7.6

6/15/2014	863	7.8	7/28/2014	125	16.1
6/16/2014	814	7.3	7/29/2014	118	17.2
6/17/2014	725	7.9	7/30/2014	111	17.7
6/18/2014	697	8.8	7/31/2014	105	17.7
6/19/2014	743	9.2	8/1/2014	99.2	17.2
6/20/2014	837	8.8	8/2/2014	99.9	15.3
6/21/2014	738	7.9	8/3/2014	99.6	16.2
6/22/2014	702	9.2	8/4/2014	90.1	17.7
6/23/2014	817	9.8	8/5/2014	83.9	17.8
6/24/2014	887	9.7	8/6/2014	80	17.7
6/25/2014	790	9.3	8/7/2014	77.5	17.5
6/26/2014	775	9.3	8/8/2014	74.1	17.3
6/27/2014	755	9.1	8/9/2014	71.5	16.8
6/28/2014	810	9.2	8/10/2014	68.5	16.9
6/29/2014	857	8.8	8/11/2014	66.1	17
6/30/2014	667	9.2	8/12/2014	66.2	17.6
7/1/2014	669	10.5	8/13/2014	147	17.2
7/2/2014	729	10.5	8/14/2014	112	15.5
7/3/2014	702	11	8/15/2014	96.8	15.8
7/4/2014	597	10.8	8/16/2014	84.3	16.3
7/5/2014	520	11	8/17/2014	77.4	17.5
7/6/2014	497	12.2	8/18/2014	72.5	18.1
7/7/2014	531	12.6	8/19/2014	68	18.6
7/8/2014	513	13.3	8/20/2014	65.3	17.8
7/9/2014	486	13.8	8/21/2014	63.7	16.6
7/10/2014	424	13.3	8/22/2014	62.8	15.3
7/11/2014	369	13.4	8/23/2014	64.5	15.5
7/12/2014	345	13.7	8/24/2014	63.1	16.1
7/13/2014	328	14	8/25/2014	62.7	15.2
7/14/2014	309	15.5	8/26/2014	59	16.1
7/15/2014	294	16.1	8/27/2014	56.9	17.5
7/16/2014	268	16.2	8/28/2014	55.3	17.4
7/17/2014	240	16.2	8/29/2014	54.3	16.2
7/18/2014	217	15.5	8/30/2014	54.2	15
7/19/2014	204	15.9	8/31/2014	56.9	14.7
7/20/2014	207	14.5	9/1/2014	55.4	14.2
7/21/2014	176	14.1	9/2/2014	53.6	14.5
7/22/2014	160	13.6	9/3/2014	78.8	13.6
7/23/2014	165	12.8	9/4/2014	66.2	12.9
7/24/2014	190	11.9	9/5/2014	57.7	13.4
7/25/2014	187	13.1	9/6/2014	54	13.6
7/26/2014	149	14.7	9/7/2014	51.8	13.8
7/27/2014	135	15.2	9/8/2014	50.2	14.1

9/9/2014	49.1	14.1	10/22/2014	121	8.3
9/10/2014	49.5	13.7	10/23/2014	230	7.9
9/11/2014	47.7	12.2	10/24/2014	201	7.3
9/12/2014	46.5	10.9	10/25/2014	164	7.5
9/13/2014	45.7	11	10/26/2014	245	7.4
9/14/2014	45.1	11.4	10/27/2014	219	6.1
9/15/2014	44.3	11.8	10/28/2014	193	5.9
9/16/2014	43.4	12.8	10/29/2014	350	5.7
9/17/2014	42.8	13.9	10/30/2014	320	6.6
9/18/2014	42.1	14.6	10/31/2014	514	6.9
9/19/2014	43.5	14.8	11/1/2014	374	6.1
9/20/2014	45.5	14.2	11/2/2014	281	6
9/21/2014	42.6	13.9	11/3/2014	270	6.5
9/22/2014	41.5	14.2	11/4/2014	616	7.1
9/23/2014	41.8	13.4	11/5/2014	505	6.9
9/24/2014	48.9	12.9	11/6/2014	565	7.3
9/25/2014	53.8	12.4	11/7/2014	577	6.3
9/26/2014	53.8	11.9	11/8/2014	412	4.9
9/27/2014	53.7	12.1	11/9/2014	416	5.3
9/28/2014	48.8	12.4	11/10/2014	405	4.6
9/29/2014	46.3	11.8	11/11/2014	302	2.8
9/30/2014	46.6	11.5	11/12/2014	227	0.3
10/1/2014	52.8	10.6	11/13/2014		0
10/2/2014	48.2	10.8	11/14/2014		0
10/3/2014	45.4	10.2	11/15/2014		0
10/4/2014	44.3	10.2	11/16/2014		0.1
10/5/2014	43.5	11.3	11/17/2014		0.1
10/6/2014	42.1	12.3	11/18/2014		0.1
10/7/2014	41.4	12.9	11/19/2014		0.1
10/8/2014	40.9	12.8	11/20/2014		1.1
10/9/2014	40.6	11.6	11/21/2014	128	1.6
10/10/2014	40.9	10.9	11/22/2014	135	1.1
10/11/2014	41.4	11.1	11/23/2014	142	1.4
10/12/2014	57.1	10.2	11/24/2014	137	2
10/13/2014	54.8	10.6	11/25/2014	669	1.7
10/14/2014	48.9	10.7	11/26/2014	1570	1.9
10/15/2014	54.5	10.2	11/27/2014	1850	2.9
10/16/2014	77.2	9.1	11/28/2014	2740	3.1
10/17/2014	62.2	9	11/29/2014	1830	2.3
10/18/2014	78.3	9.6	11/30/2014	1050	0.7
10/19/2014	80.8	10.2	12/1/2014		0.2
10/20/2014	72.7	9.7	12/2/2014		0.1
10/21/2014	88.8	9.3	12/3/2014		0.3

12/4/2014	451	1.6
12/5/2014	392	2.1
12/6/2014	355	2.3
12/7/2014	345	3
12/8/2014	305	3.1
12/9/2014	365	3.3
12/10/2014	498	2.9
12/11/2014	531	3.1
12/12/2014	493	3.3
12/13/2014	418	2.8
12/14/2014	369	2
12/15/2014	341	2.2
12/16/2014	318	2.6
12/17/2014	297	2.7
12/18/2014	280	2.9
12/19/2014	268	2.7
12/20/2014	263	1.9
12/21/2014	579	1.4
12/22/2014	513	2.6
12/23/2014	408	2.8
12/24/2014	561	2.8
12/25/2014	417	2.6
12/26/2014	368	2
12/27/2014	349	2
12/28/2014	332	2
12/29/2014	303	1.9
12/30/2014		0.2
12/31/2014		0

## APPENDIX B. Daily Trap Operation

Date	Trap Status	Comments		
			4/8/2014	Op.
			4/9/2014	Op.
3/1/2014	Op.		4/10/2014	Op.
3/2/2014	No Op.	Stopped - ice	4/11/2014	Op.
3/3/2014	No Op.	Pulled - ice	4/12/2014	Op.
3/4/2014	No Op.	Pulled - ice	4/13/2014	Op.
3/5/2014	Op.		4/14/2014	Op.
3/6/2014	Op.		4/15/2014	Op.
3/7/2014	Op.		4/16/2014	Op.
3/8/2014	Op.		4/17/2014	Op.
3/9/2014	Op.		4/18/2014	Op.
3/10/2014	Op.		4/19/2014	Op.
3/11/2014	Op.		4/20/2014	Op.
3/12/2014	Op.		4/21/2014	Op.
3/13/2014	Op.		4/22/2014	Op.
3/14/2014	Op.		4/23/2014	Op.
3/15/2014	Op.		4/24/2014	Op.
3/16/2014	Op.		4/25/2014	Op.
3/17/2014	Op.		4/26/2014	Op.
3/18/2014	Op.		4/27/2014	Op.
3/19/2014	Op.		4/28/2014	Op.
3/20/2014	Op.		4/29/2014	Op.
3/21/2014	Op.		4/30/2014	Op.
3/22/2014	Op.		5/1/2014	Op.
3/23/2014	Op.		5/2/2014	Op.
3/24/2014	Op.		5/3/2014	Op.
3/25/2014	Op.		5/4/2014	Op.
3/26/2014	Op.		5/5/2014	Op.
3/27/2014	Op.		5/6/2014	Op.
3/28/2014	Op.		5/7/2014	Op.
3/29/2014	Op.		5/8/2014	Op.
3/30/2014	Op.		5/9/2014	Op.
3/31/2014	Op.		5/10/2014	Op.
4/1/2014	Op.		5/11/2014	Op.
4/2/2014	Op.		5/12/2014	Op.
4/3/2014	Op.		5/13/2014	Op.
4/4/2014	Op.		5/14/2014	Op.
4/5/2014	Op.		5/15/2014	Op.
4/6/2014	Op.		5/16/2014	No Op.
4/7/2014	Op.		5/17/2014	Op.
				Stopped - woody debris

5/18/2014	Op.		6/30/2014	No Op.	Trap set at new site
5/19/2014	Op.		7/1/2014	Op.	
5/20/2014	Op.		7/2/2014	Op.	
5/21/2014	Op.		7/3/2014	Op.	
5/22/2014	Op.		7/4/2014	Op.	
5/23/2014	Op.		7/5/2014	Op.	
5/24/2014	Op.		7/6/2014	Op.	
5/25/2014	Op.		7/7/2014	Op.	
5/26/2014	Op.		7/8/2014	Op.	
5/27/2014	Op.		7/9/2014	Op.	
5/28/2014	Op.		7/10/2014	Op.	
5/29/2014	Op.		7/11/2014	Op.	
5/30/2014	Op.		7/12/2014	Op.	
5/31/2014	Op.		7/13/2014	Op.	
6/1/2014	Op.		7/14/2014	Op.	
6/2/2014	Op.		7/15/2014	Op.	
6/3/2014	Op.		7/16/2014	Op.	
6/4/2014	Op.		7/17/2014	Op.	
6/5/2014	Op.		7/18/2014	No Op.	Pulled - fire activity
6/6/2014	Op.		7/19/2014	Op.	
6/7/2014	Op.		7/20/2014	Op.	
6/8/2014	Op.		7/21/2014	Op.	
6/9/2014	Op.		7/22/2014	Op.	
6/10/2014	Op.		7/23/2014	Op.	
6/11/2014	Op.		7/24/2014	Op.	
6/12/2014	Op.		7/25/2014	Op.	
6/13/2014	Op.		7/26/2014	Op.	
6/14/2014	Op.		7/27/2014	Op.	
6/15/2014	Op.		7/28/2014	Op.	
6/16/2014	Op.		7/29/2014	Op.	
6/17/2014	Op.		7/30/2014	Op.	
6/18/2014	Op.		7/31/2014	No Op.	Stopped - low water
6/19/2014	Op.		8/1/2014	Op.	
6/20/2014	Op.		8/2/2014	Op.	
6/21/2014	Op.		8/3/2014	Op.	
6/22/2014	Op.		8/4/2014	Op.	
6/23/2014	Op.		8/5/2014	Op.	
6/24/2014	Op.		8/6/2014	Op.	
6/25/2014	Op.		8/7/2014	Op.	
6/26/2014	No Op.	Pulled - relocation	8/8/2014	Op.	
6/27/2014	No Op.	Pulled	8/9/2014	Op.	
6/28/2014	No Op.	Pulled	8/10/2014	Op.	
6/29/2014	No Op.	Pulled	8/11/2014	Op.	



8/12/2014	Op.		9/24/2014	No Op.	Stopped - low water
8/13/2014	Op.		9/25/2014	Op.	
8/14/2014	Op.		9/26/2014	No Op.	Stopped - low water
8/15/2014	Op.		9/27/2014	Op.	
8/16/2014	Op.		9/28/2014	Op.	
8/17/2014	Op.		9/29/2014	Op.	
8/18/2014	Op.		9/30/2014	Op.	
8/19/2014	Op.		10/1/2014	Op.	
8/20/2014	Op.		10/2/2014	Op.	
8/21/2014	Op.		10/3/2014	Op.	
8/22/2014	Op.		10/4/2014	Op.	
8/23/2014	Op.		10/5/2014	Op.	
8/24/2014	Op.		10/6/2014	Op.	
8/25/2014	Op.		10/7/2014	Op.	
8/26/2014	Op.		10/8/2014	Op.	
8/27/2014	Op.		10/9/2014	No Op.	Stopped - low water
8/28/2014	No Op.	Stopped - low water	10/10/2014	Op.	
8/29/2014	No Op.	Stopped - low water	10/11/2014	Op.	
8/30/2014	No Op.	Stopped - low water	10/12/2014	Op.	
8/31/2014	No Op.	Stopped - low water	10/13/2014	No Op.	Stopped - woody debris
9/1/2014	Op.		10/14/2014	Op.	
9/2/2014	No Op.	Stopped - low water	10/15/2014	Op.	
9/3/2014	Op.		10/16/2014	Op.	
9/4/2014	Op.		10/17/2014	Op.	
9/5/2014	Op.		10/18/2014	Op.	
9/6/2014	Op.		10/19/2014	Op.	
9/7/2014	No Op.	Stopped - low water	10/20/2014	Op.	
9/8/2014	Op.		10/21/2014	Op.	
9/9/2014	Op.		10/22/2014	Op.	
9/10/2014	Op.		10/23/2014	Op.	
9/11/2014	Op.		10/24/2014	Op.	
9/12/2014	Op.		10/25/2014	Op.	
9/13/2014	No Op.	Stopped - low water	10/26/2014	Op.	
9/14/2014	No Op.	Stopped - low water	10/27/2014	No Op.	Stopped - woody debris
9/15/2014	No Op.	Stopped - low water	10/28/2014	Op.	
9/16/2014	No Op.	Stopped - low water	10/29/2014	Op.	
9/17/2014	No Op.	Stopped - low water	10/30/2014	No Op.	Stopped - woody debris
9/18/2014	No Op.	Stopped - low water	10/31/2014	Op.	
9/19/2014	No Op.	Stopped - low water	11/1/2014	Op.	
9/20/2014	Op.		11/2/2014	Op.	
9/21/2014	No Op.	Stopped - low water	11/3/2014	Op.	
9/22/2014	No Op.	Stopped - low water	11/4/2014	Op.	
9/23/2014	No Op.	Stopped - low water	11/5/2014	No Op.	Stopped - woody debris

11/6/2014	Op.	
11/7/2014	No Op.	Stopped - woody debris
11/8/2014	Op.	
11/9/2014	Op.	
11/10/2014	Op.	
11/11/2014	Op.	
11/12/2014	No Op.	Stopped - ice
11/13/2014	No Op.	Pulled - ice
11/14/2014	No Op.	Pulled - ice
11/15/2014	No Op.	Pulled - ice
11/16/2014	No Op.	Pulled - ice
11/17/2014	No Op.	Pulled - ice
11/18/2014	No Op.	Pulled - ice
11/19/2014	No Op.	Pulled - ice
11/20/2014	No Op.	Pulled - ice
11/21/2014	Op.	
11/22/2014	Op.	
11/23/2014	Op.	
11/24/2014	Op.	
11/25/2014	Op.	
11/26/2014	No Op.	Stopped - ice
11/27/2014	No Op.	Pulled - ice
11/28/2014	No Op.	Pulled - ice
11/29/2014	No Op.	Pulled - ice
11/30/2014	Op.	Last day of trapping

## APPENDIX C. Regression Models

Model: Chinook Yearlings (Spring '06-'14) Back Position, ( $r^2 = 0.15$ ;  $p = 0.03$ )

Origin/Species/Stage	Age	Date	Trap Position	Mark	Recap	Trap Efficiency (R+1) / M	ASIN Transform	Discharge
Wild Chinook Smolt	1+	3/31/2007	Back	40	2	0.08	0.28	869
Wild Chinook Smolt	1+	4/6/2006	Back	42	9	0.24	0.51	264
Wild Chinook Smolt	1+	4/14/2010	Back	42	4	0.12	0.35	173
Wild Chinook Smolt	1+	3/31/2012	Back	43	5	0.14	0.38	250
Wild Chinook Smolt	1+	4/3/2007	Back	46	1	0.04	0.21	656
Wild Chinook Smolt	1+	4/19/2012	Back	48	7	0.17	0.42	434
Wild Chinook Smolt	1+	4/10/2007	Back	53	4	0.09	0.31	966
Wild Chinook Smolt	1+	4/21/2009	Back	53	0	0.02	0.14	732
Wild Chinook Smolt	1+	4/13/2012	Back	53	4	0.09	0.31	358
Wild Chinook Smolt	1+	4/16/2012	Back	53	7	0.15	0.40	443
Wild Chinook Smolt	1+	4/24/2008	Back	57	8	0.158	0.409	210
Wild Chinook Smolt	1+	4/23/2012	Back	58	1	0.034	0.187	1380
Wild Chinook Smolt	1+	4/24/2006	Back	59	3	0.068	0.263	368
Wild Chinook Smolt	1+	3/23/2007	Back	59	7	0.136	0.377	876
Wild Chinook Smolt	1+	3/17/2007	Back	64	7	0.125	0.361	936
Wild Chinook Smolt	1+	4/18/2010	Back	67	2	0.045	0.213	330
Wild Chinook Smolt	1+	4/17/2008	Back	72	13	0.194	0.457	274
Wild Chinook Smolt	1+	4/3/2006	Back	81	10	0.136	0.377	188
Wild Chinook Smolt	1+	3/20/2007	Back	91	13	0.154	0.403	1230
Wild Chinook Smolt	1+	5/1/2008	Back	102	16	0.167	0.421	315
Wild Chinook Smolt	1+	4/28/2008	Back	127	19	0.157	0.408	271
Wild Chinook Smolt	1+	4/14/2008	Back	195	40	0.21	0.476	327
Wild Chinook Smolt	1+	3/9/2014	Back	65	4	0.077	0.281	958
Wild Chinook Smolt	1+	3/13/2014	Back	67	9	0.149	0.397	566

Model: Chinook Subyearling (Fall '06-'13) Back Position, ( $r^2 = 0.55$ ;  $p = 0.001$ )

Origin/Species/Stage	Age	Date	Trap Position	Mark	Recap	Trap Efficiency (R+1) / M	ASIN Transform	Discharge
Wild Chinook Parr	0	10/26/2006	Back	183	50	0.28	0.56	51
Wild Chinook Parr	0	10/30/2006	Back	168	52	0.32	0.60	63
Wild Chinook Parr	0	11/1/2010	Back	254	42	0.17	0.42	198
Wild Chinook Parr	0	11/4/2010	Back	287	49	0.17	0.43	215
Wild Chinook Parr	0	11/7/2010	Back	168	32	0.20	0.46	241
Wild Chinook Parr	0	11/13/2010	Back	185	35	0.19	0.46	131

Wild Chinook Parr	0	11/3/2012	Back	201	25	0.13	0.37	402
Wild Chinook Parr	0	11/7/2012	Back	233	27	0.12	0.35	394
Wild Chinook Parr	0	11/11/2012	Back	328	87	0.27	0.54	217
Wild Chinook Parr	0	11/15/2012	Back	195	34	0.18	0.44	213
Wild Chinook Parr	0	9/30/2013	Back	171	12	0.08	0.28	542
Wild Chinook Parr	0	10/2/2013	Back	213	43	0.21	0.47	328
Wild Chinook Parr	0	10/3/2013	Back	181	41	0.23	0.50	296
Wild Chinook Parr	0	10/7/2013	Back	242	31	0.13	0.37	233
Wild Chinook Parr	0	10/9/2013	Back	203	40	0.20	0.47	303
Wild Chinook Parr	0	11/27/2013	Back	241	55	0.23	0.50	182

Model: Chinook Subyearling (Fall '06-'13) Forward Position, ( $r^2 = 0.16$ ;  $p = 0.02$ )

Origin/Species/Stage	Age	Date	Trap Position	Mark	Recap	Trap Efficiency (R+1) / M	ASIN Transform	Discharge
Wild Chinook Parr	0	7/13/2006	Back	52	8	0.17	0.43	171
Wild Chinook Parr	0	7/17/2006	Back	138	15	0.12	0.35	129
Wild Chinook Parr	0	7/20/2006	Back	74	5	0.08	0.29	113
Wild Chinook Parr	0	7/28/2006	Back	54	5	0.11	0.34	91
Wild Chinook Parr	0	7/31/2006	Back	99	7	0.08	0.29	79
Wild Chinook Parr	0	9/18/2006	Back	55	10	0.20	0.46	46
Wild Chinook Parr	0	7/31/2008	Back	60	15	0.27	0.54	121
Wild Chinook Parr	0	8/12/2008	Back	103	2	0.03	0.17	85.6
Wild Chinook Parr	0	8/22/2008	Back	75	11	0.16	0.41	97
Wild Chinook Parr	0	8/28/2008	Back	72	7	0.11	0.34	81.9
Wild Chinook Parr	0	10/9/2008	Back	110	22	0.21	0.48	63.5
Wild Chinook Parr	0	10/27/2008	Back	51	12	0.26	0.53	56.1
Wild Chinook Parr	0	10/30/2008	Back	84	15	0.19	0.45	53
Wild Chinook Parr	0	11/6/2008	Back	78	8	0.12	0.35	77.7
Wild Chinook Parr	0	11/10/2008	Back	88	0	0.01	0.11	309
Wild Chinook Parr	0	7/14/2009	Back	86	2	0.04	0.19	193
Wild Chinook Parr	0	7/15/2009	Back	105	4	0.05	0.22	179
Wild Chinook Parr	0	7/17/2009	Back	122	8	0.07	0.28	157
Wild Chinook Parr	0	7/20/2009	Back	89	2	0.03	0.19	135
Wild Chinook Parr	0	8/17/2009	Back	73	1	0.03	0.17	58
Wild Chinook Parr	0	9/10/2009	Back	56	7	0.14	0.39	60
Wild Chinook Parr	0	8/8/2010	Back	58	1	0.03	0.19	85
Wild Chinook Parr	0	8/11/2010	Back	114	8	0.08	0.29	77
Wild Chinook Parr	0	9/11/2010	Back	68	9	0.15	0.39	75
Wild Chinook Parr	0	10/12/2010	Back	216	42	0.20	0.46	126
Wild Chinook Parr	0	10/15/2010	Back	192	37	0.20	0.46	95
Wild Chinook Parr	0	10/18/2010	Back	193	36	0.19	0.45	81
Wild Chinook Parr	0	10/22/2010	Back	92	18	0.21	0.47	69
Wild Chinook Parr	0	10/25/2010	Back	60	7	0.13	0.37	78
Wild Chinook Parr	0	10/29/2010	Back	127	0	0.01	0.09	95.1
Wild Chinook Parr	0	8/19/2011	Back	106	5	0.06	0.24	123

Model: Summer Steelhead Back Position ('07-'14), ( $r^2 = 0.35$ ;  $p = 2.90E-05$ )

Origin/Species/Stage	Age	Date	Trap Position	Mark	Recap	Trap Efficiency (R+1) / M	ASIN Transform	Discharge
Wild Steelhead Parr/Smolt	1+	3/20/2007	Back	55	1	0.04	0.19	1230
Wild Steelhead Parr/Smolt	1+	3/31/2007	Back	56	4	0.09	0.30	869
Wild Steelhead Parr/Smolt	1+	4/10/2007	Back	60	8	0.15	0.40	966
Wild Steelhead Parr/Smolt	1+	5/1/2007	Back	52	2	0.06	0.24	783
Wild Steelhead Parr/Smolt	1+	6/9/2007	Back	71	9	0.14	0.38	842
Wild Steelhead Parr/Smolt	1+	6/12/2007	Back	65	8	0.14	0.38	704
Wild Steelhead Parr/Smolt	1+	6/14/2007	Back	61	5	0.10	0.32	687
Wild Steelhead Parr/Smolt	1+	6/21/2007	Back	67	4	0.07	0.28	751
Wild Steelhead Parr/Smolt	1+	4/14/2008	Back	149	46	0.32	0.60	327
Wild Steelhead Parr/Smolt	1+	4/17/2008	Back	75	3	0.05	0.23	274
Wild Steelhead Parr/Smolt	1+	4/28/2008	Back	74	11	0.16	0.41	271
Wild Steelhead Parr/Smolt	1+	5/1/2008	Back	176	29	0.17	0.43	315
Wild Steelhead Parr/Smolt	1+	5/12/2008	Back	55	8	0.16	0.42	663
Wild Steelhead Parr/Smolt	1+	5/15/2008	Back	57	1	0.04	0.19	1390
Wild Steelhead Parr/Smolt	1+	6/9/2008	Back	142	20	0.15	0.39	938
Wild Steelhead Parr/Smolt	1+	6/12/2008	Back	83	10	0.13	0.37	823
Wild Steelhead Parr/Smolt	1+	6/16/2008	Back	81	8	0.11	0.34	1140
Wild Steelhead Parr/Smolt	1+	4/20/2010	Back	121	11	0.10	0.32	675
Wild Steelhead Parr/Smolt	1+	4/22/2010	Back	121	10	0.09	0.31	726
Wild Steelhead Parr/Smolt	1+	6/20/2010	Back	128	11	0.09	0.31	926
Wild Steelhead Parr/Smolt	1+	4/5/2011	Back	52	1	0.04	0.20	761
Wild Steelhead Parr/Smolt	1+	5/22/2011	Back	84	3	0.05	0.22	1540
Wild Steelhead Parr/Smolt	1+	6/12/2012	Back	69	5	0.09	0.30	1170
Wild Steelhead Parr/Smolt	1+	7/26/2012	Back	63	4	0.08	0.29	278
Wild Steelhead Parr/Smolt	1+	4/22/2013	Back	66	6	0.11	0.33	520
Wild Steelhead Parr/Smolt	1+	4/26/2013	Back	50	2	0.06	0.25	642
Wild Steelhead Parr/Smolt	1+	4/30/2013	Back	54	2	0.06	0.24	778
Wild Steelhead Parr/Smolt	1+	5/8/2013	Back	62	0	0.02	0.13	2170
Wild Steelhead Parr/Smolt	1+	5/19/2013	Back	122	15	0.13	0.37	1130
Wild Steelhead Parr/Smolt	1+	5/22/2013	Back	58	4	0.09	0.30	1080
Wild Steelhead Parr/Smolt	1+	5/26/2013	Back	79	3	0.05	0.23	724
Wild Steelhead Parr/Smolt	1+	5/30/2013	Back	92	7	0.09	0.30	849
Wild Steelhead Parr/Smolt	1+	6/3/2013	Back	71	6	0.10	0.32	962
Wild Steelhead Parr/Smolt	1+	6/7/2013	Back	94	4	0.05	0.23	1420
Wild Steelhead Parr/Smolt	1+	6/13/2013	Back	64	2	0.05	0.22	745
Wild Steelhead Parr/Smolt	1+	6/17/2013	Back	115	5	0.05	0.23	883
Wild Steelhead Parr/Smolt	1+	6/29/2013	Back	60	12	0.22	0.48	730

Wild Steelhead Parr/Smolt	1+	7/7/2013	Back	75	9	0.13	0.37	325
Wild Steelhead Parr/Smolt	1+	5/5/2014	Back	55	3	0.07	0.27	1260
Wild Steelhead Parr/Smolt	1+	5/20/2014	Back	57	0	0.02	0.13	1490
Wild Steelhead Parr/Smolt	1+	6/3/2014	Back	75	1	0.03	0.16	1610

Model: 2013 Summer Steelhead Back Position (In-yr.), ( $r^2 = 0.15$ ;  $p = 0.05$ )

Origin/Species/Stage	Age	Date	Trap Position	Mark	Recap	Trap Efficiency (R+1) / M	ASIN Transform	Discharge
Wild Chinook Smolt	1+	3/31/2007	Back	40	2	0.08	0.28	869
Wild Chinook Smolt	1+	4/6/2006	Back	42	9	0.24	0.51	264
Wild Chinook Smolt	1+	4/14/2010	Back	42	4	0.12	0.35	173
Wild Chinook Smolt	1+	3/31/2012	Back	43	5	0.14	0.38	250
Wild Chinook Smolt	1+	4/3/2007	Back	46	1	0.04	0.21	656
Wild Chinook Smolt	1+	4/19/2012	Back	48	7	0.17	0.42	434
Wild Chinook Smolt	1+	4/10/2007	Back	53	4	0.09	0.31	966
Wild Chinook Smolt	1+	4/21/2009	Back	53	0	0.02	0.14	732
Wild Chinook Smolt	1+	4/13/2012	Back	53	4	0.09	0.31	358
Wild Chinook Smolt	1+	4/16/2012	Back	53	7	0.15	0.40	443
Wild Chinook Smolt	1+	4/24/2008	Back	57	8	0.158	0.409	210
Wild Chinook Smolt	1+	4/23/2012	Back	58	1	0.034	0.187	1380
Wild Chinook Smolt	1+	4/24/2006	Back	59	3	0.068	0.263	368
Wild Chinook Smolt	1+	3/23/2007	Back	59	7	0.136	0.377	876
Wild Chinook Smolt	1+	3/17/2007	Back	64	7	0.125	0.361	936
Wild Chinook Smolt	1+	4/18/2010	Back	67	2	0.045	0.213	330
Wild Chinook Smolt	1+	4/17/2008	Back	72	13	0.194	0.457	274
Wild Chinook Smolt	1+	4/3/2006	Back	81	10	0.136	0.377	188
Wild Chinook Smolt	1+	3/20/2007	Back	91	13	0.154	0.403	1230
Wild Chinook Smolt	1+	5/1/2008	Back	102	16	0.167	0.421	315
Wild Chinook Smolt	1+	4/28/2008	Back	127	19	0.157	0.408	271
Wild Chinook Smolt	1+	4/14/2008	Back	195	40	0.21	0.476	327
Wild Chinook Smolt	1+	3/9/2014	Back	65	4	0.077	0.281	958
Wild Chinook Smolt	1+	3/13/2014	Back	67	9	0.149	0.397	566

Model: Spring Chinook 2010-2014 Non-Trapping Period Array (NAL) Efficiency, ( $r^2 = 0.64$ ;  $p = 0.0004$ )

Origin/Species/Stage	Age	Date	Mark	Detections	Trap Efficiency (R+1) / M	ASIN Transform	Discharge
Wild Chinook Parr	0	11/4/2010	254	95	0.38	0.66	224
Wild Chinook Parr	0	11/7/2010	287	70	0.25	0.52	248

Wild Chinook Parr	0	11/10/2010	168	74	0.45	0.73	169
Wild Chinook Parr	0	11/18/2010	185	22	0.12	0.36	278
Wild Chinook Parr	0	11/3/2012	201	21	0.11	0.34	384
Wild Chinook Parr	0	11/7/2012	233	31	0.14	0.38	394
Wild Chinook Parr	0	11/11/2012	328	66	0.20	0.47	378
Wild Chinook Parr	0	11/15/2012	195	68	0.35	0.64	219
Wild Chinook Parr	0	11/4/2013	130	51	0.40	0.68	127
Wild Chinook Parr	0	11/8/2013	106	39	0.38	0.66	146
Wild Chinook Parr	0	11/4/2014	114	5	0.05	0.23	583
Wild Chinook Parr	0	3/9/2014	65	4	0.08	0.28	880
Wild Chinook Parr	0	3/13/2014	67	5	0.09	0.30	541
Wild Chinook Parr	0	11/13/2010	74	41	0.57	0.85	140

---

## APPENDIX D. Historical Morphometric Data

### Spring Chinook (2004-2014)

Trap Year	Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-factor
			Mean	n	SD	Mean	n	SD	
2004	2002	Wild Chinook Yearling Smolt	93.4	336	12.4	9.0	337	5.0	1.1
2004	2003	Wild Chinook Subyearling Fry	39.5	82	5.1	0.6	79	0.3	1.0
2004	2003	Wild Chinook Subyearling Parr	82.4	792	7.9	6.1	702	2.7	1.1
2005	2003	Wild Chinook Yearling Smolt	93.6	278	7.9	8.7	276	2.1	1.1
2005	2004	Wild Chinook Subyearling Fry	42.1	107	5.6	0.7	102	0.4	0.9
2005	2004	Wild Chinook Subyearling Parr	75.9	924	9.6	4.9	890	3.8	1.1
2006	2004	Wild Chinook Yearling Smolt	91.2	363	7.1	7.5	362	1.8	1.0
2006	2005	Wild Chinook Subyearling Fry	—	—	—	—	—	—	—
2006	2005	Wild Chinook Subyearling Parr	72.9	1,428	9.6	3.9	1,428	2.3	1.0
2007	2005	Wild Chinook Yearling Smolt	89.0	676	8.2	8.0	675	6.1	1.1
2007	2006	Wild Chinook Subyearling Fry	39.0	24	3.7	0.6	24	0.5	1.0
2007	2006	Wild Chinook Subyearling Parr	79.5	686	13.8	6.1	685	2.6	1.2
2008	2006	Wild Chinook Yearling Smolt	96.1	904	6.6	9.5	904	2.1	1.1
2008	2007	Wild Chinook Subyearling Fry	42.8	127	4.6	0.8	127	0.4	1.0
2008	2007	Wild Chinook Subyearling Parr	75.8	2,049	12.5	5.2	2,049	2.4	1.2
2009	2007	Wild Chinook Yearling Smolt	94.4	198	8.9	9.2	198	2.5	1.1
2009	2008	Wild Chinook Subyearling Fry	44.8	82	4.8	0.9	82	0.6	1.0
2009	2008	Wild Chinook Subyearling Parr	70.1	2,333	12.0	4.2	2,333	2.0	1.2
2010	2008	Wild Chinook Yearling Smolt	96.9	366	7.3	10.2	366	2.3	1.1
2010	2009	Wild Chinook Subyearling Fry	41.8	30	5.0	1.3	8	0.2	1.8
2010	2009	Wild Chinook Subyearling Parr	80.7	3,021	10.7	6.2	3,021	2.3	1.2
2011	2009	Wild Chinook Yearling Smolt	89.1	152	9.9	7.7	152	1.8	1.1
2011	2010	Wild Chinook Subyearling Fry	39.8	217	6.6	0.6	217	0.5	1.0
2011	2010	Wild Chinook Subyearling Parr	73.4	1,046	13.1	4.9	1,046	2.5	1.2
2012	2010	Wild Chinook Yearling Smolt	93.3	368	7.0	9.2	368	2.2	1.1
2012	2011	Wild Chinook Subyearling Fry	42.7	48	9.1	0.9	48	0.6	1.2
2012	2011	Wild Chinook Subyearling Parr	77.9	2,160	10.7	5.3	2,160	1.9	1.1
2013	2011	Wild Chinook Yearling Smolt	90.6	239	75.0	7.9	239	2.1	1.1
2013	2012	Wild Chinook Subyearling Fry	45.6	1,824	6.8	1.0	1,803	0.6	1.1
2013	2012	Wild Chinook Subyearling Parr	70.0	4,422	11.4	3.8	4,409	1.7	1.1
2014	2012	Wild Chinook Yearling Smolt	89.5	464	6.9	7.5	464	1.8	1.0
2014	2013	Wild Chinook Subyearling Fry	40.1	677	5.2	0.9	221	0.5	1.4
2014	2013	Wild Chinook Subyearling Parr	69.1	1,549	12.3	3.8	1,547	2.3	1.2



Summer Steelhead (2004-2014)

Trap Year	Brood Year	Age	Origin/Species	Fork Length (mm)			Weight (g)			K-factor
				Mean	n	SD	Mean	n	SD	
2004	2004	0	Wild Summer Steelhead	67.0	358	10.0	3.5	279	1.5	1.2
2004	2003	1	Wild Summer Steelhead	101.7	394	23.2	13.2	366	27.3	1.3
2004	2002	2	Wild Summer Steelhead	161.6	146	19.8	43.4	141	15.5	1.0
2004	2001	3	Wild Summer Steelhead	201.6	43	11.2	76.0	43	21.2	0.9
2004	2003	1	Hat. Summer Steelhead	182.8	523	22.4	62.1	497	21.2	1.0
2005	2005	0	Wild Summer Steelhead	54.1	649	15.7	2.2	616	3.2	1.4
2005	2004	1	Wild Summer Steelhead	93.6	585	25.6	10.8	575	10.1	1.3
2005	2003	2	Wild Summer Steelhead	153.5	103	21.2	38.1	102	16.4	1.1
2005	2002	3	Wild Summer Steelhead	144	1	—	43.2	1	—	1.4
2005	2004	1	Hat. Summer Steelhead	188.2	343	21.2	66.0	343	24.0	1.0
2006	2006	0	Wild Summer Steelhead	66.3	180	5.8	2.5	180	1.0	0.9
2006	2005	1	Wild Summer Steelhead	85.2	877	18.7	6.7	877	6.6	1.1
2006	2004	2	Wild Summer Steelhead	155.9	106	26.8	36.1	105	13.5	1.0
2006	2003	3	Wild Summer Steelhead	197	2	—	73.5	2	—	1.0
2006	2005	1	Hat. Summer Steelhead	—	—	—	—	—	—	—
2007	2007	0	Wild Summer Steelhead	54.2	329	11.7	2.0	328	1.4	1.3
2007	2006	1	Wild Summer Steelhead	82.7	1,330	16.8	7.2	1,329	6.3	1.3
2007	2005	2	Wild Summer Steelhead	143.8	102	20.6	31.4	102	11.9	1.1
2007	2004	3	Wild Summer Steelhead	143	1	—	26.8	1	—	0.9
2007	2006	1	Hat. Summer Steelhead	149.3	3	47	33.1	3	29.1	1.0
2008	2008	0	Wild Summer Steelhead	52.9	930	11.1	1.7	930	1.2	1.1
2008	2007	1	Wild Summer Steelhead	84.5	1,876	17.1	7.4	1,874	6.6	1.2
2008	2006	2	Wild Summer Steelhead	149.9	122	22.9	36.0	122	15.5	1.1
2008	2005	3	Wild Summer Steelhead	180.3	13	18.9	57.4	13	16.4	1.0
2008	2007	1	Hat. Summer Steelhead	179.4	389	16.5	55.9	388	14.8	1.0
2009	2009	0	Wild Summer Steelhead	55.6	843	10.5	2.2	688	1.1	1.3
2009	2008	1	Wild Summer Steelhead	82.6	452	18.6	7.1	447	5.5	1.3
2009	2007	2	Wild Summer Steelhead	156.9	72	22.0	40.9	72	15.5	1.1
2009	2006	3	Wild Summer Steelhead	195.0	3	5.0	73.0	3	6.7	1.0
2009	2008	1	Hat. Summer Steelhead	183.1	280	16.7	60.8	280	18.2	1.0
2010	2010	0	Wild Summer Steelhead	55.0	1,287	11.1	2.5	917	1.3	1.5
2010	2009	1	Wild Summer Steelhead	89.8	1,079	19.1	9.0	1,072	7.1	1.2
2010	2008	2	Wild Summer Steelhead	144.9	87	25.1	35.0	87	17.4	1.2
2010	2007	3	Wild Summer Steelhead	184.0	8	12.2	61.9	8	10.2	1.0
2010	2009	1	Hat. Summer Steelhead	183.5	531	19.5	61.3	526	19.6	1.0
2011	2011	0	Wild Summer Steelhead	43.5	1,093	10.1	1.1	783	0.9	1.3
2011	2010	1	Wild Summer Steelhead	75.7	818	18.5	5.5	811	5.7	1.3
2011	2009	2	Wild Summer Steelhead	144.8	27	41.3	42.1	27	62.1	1.4
2011	2008	3	Wild Summer Steelhead	—	—	—	—	—	—	—

2011	2010	1	Hat. Summer Steelhead	180.7	464	17.0	59.1	464	17.6	1.0
2012	2012	0	Wild Summer Steelhead	55.1	589	14.2	2.6	402	1.2	1.6
2012	2011	1	Wild Summer Steelhead	84.7	747	17.4	7.6	741	5.7	1.3
2012	2010	2	Wild Summer Steelhead	127.1	132	27.0	23.7	132	14.5	1.2
2012	2009	3	Wild Summer Steelhead	161.0	4.0	32.0	40.5	4.0	15.6	1.0
2012	2011	1	Hat. Summer Steelhead	154.8	318	20.9	37.7	318	14.0	1.0
2013	2013	0	Wild Summer Steelhead	56.1	878	11.3	2.1	777	1.1	1.2
2013	2012	1	Wild Summer Steelhead	44.5	1,777	14.7	5.4	1,772	4.2	1.2
2013	2011	2	Wild Summer Steelhead	144.7	21	15.7	36.1	21	10.2	1.0
2013	2010	3	Wild Summer Steelhead	—	—	—	—	—	—	—
2013	2012	1	Hat. Summer Steelhead	166.2	365	21.4	49.2	363	18.2	1.1
2014	2014	0	Wild Summer Steelhead	49.6	490	12.8	1.7	389	1.1	1.4
2014	2013	1	Wild Summer Steelhead	82.2	745	13.6	6.3	745	3.5	1.1
2014	2012	2	Wild Summer Steelhead	145.1	30	16.5	33.0	30	13.4	1.1
2014	2011	3	Wild Summer Steelhead	—	—	—	—	—	—	—
2014	2013	1	Hat. Summer Steelhead	173.4	632	18.7	52.6	633	15.9	1.0

#### Coho (2007-2014)

Trap Year	Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-factor
			Mean	n	SD	Mean	n	SD	
2004	2002	Nat. Orig. Coho Yearling Smolt	—	—	—	—	—	—	—
2004	2003	Nat. Orig. Coho Subyearling Fry	—	—	—	—	—	—	—
2004	2003	Nat. Orig. Coho Subyearling Parr	—	—	—	—	—	—	—
2004	2002	Hatchery Coho Yearling Smolt	136.6	847	12.8	27.4	820	7.5	1.1
2005	2003	Nat. Orig. Coho Yearling Smolt	114.4	17	8.8	16.2	17	3.6	1.1
2005	2004	Nat. Orig. Coho Subyearling Fry	49.1	9	10.4	1.3	9	0.8	1.1
2005	2004	Nat. Orig. Coho Subyearling Parr	76.7	9	12.8	4.9	9	2.7	1.1
2005	2003	Hatchery Coho Yearling Smolt	137.3	689	11.3	28.6	690	7.2	1.1
2006	2004	Nat. Orig. Coho Yearling Smolt	—	—	—	—	—	—	—
2006	2005	Nat. Orig. Coho Subyearling Fry	—	—	—	—	—	—	—
2006	2005	Nat. Orig. Coho Subyearling Parr	71.0	4	13.6	3.8	4	2.9	1.1
2006	2004	Hatchery Coho Yearling Smolt	—	—	—	—	—	—	—
2007	2005	Nat. Orig. Coho Yearling Smolt	92.9	36	12.5	8.7	36	4.0	1.1
2007	2006	Nat. Orig. Coho Subyearling Fry	—	—	—	—	—	—	—
2007	2006	Nat. Orig. Coho Subyearling Parr	83.0	1	—	6.2	1	—	1.1
2007	2005	Hatchery Coho Yearling Smolt	116.0	2	—	16.8	2	—	1.1
2008	2006	Nat. Orig. Coho Yearling Smolt	—	—	—	—	—	—	—
2008	2007	Nat. Orig. Coho Subyearling Fry	—	—	—	—	—	—	—
2008	2007	Nat. Orig. Coho Subyearling Parr	87.0	1	—	6.4	1	—	1.0

2008	2006	Hatchery Coho Yearling Smolt	130.2	843	10.4	23.6	843	6.2	1.1
2009	2007	Nat. Orig. Coho Yearling Smolt	103.0	4	9.7	11.7	4	3.4	1.1
2009	2008	Nat. Orig. Coho Subyearling Fry	—	—	—	—	—	—	—
2009	2008	Nat. Orig. Coho Subyearling Parr	79.6	5	20.1	6.6	5	4.8	1.3
2009	2007	Hatchery Coho Yearling Smolt	135.3	625	8.9	26.2	579	5.2	1.1
2010	2008	Nat. Orig. Coho Yearling Smolt	—	—	—	—	—	—	—
2010	2009	Nat. Orig. Coho Subyearling Fry	48.0	2	—	1.3	2	—	1.2
2010	2009	Nat. Orig. Coho Subyearling Parr	83.6	27	8.6	6.7	27	2.4	1.1
2010	2008	Hatchery Coho Yearling Smolt	130.0	1,051	10.1	23.8	1,049	5.3	1.1
2011	2009	Nat. Orig. Coho Yearling Smolt	100.2	14	12.7	11.3	14	3.9	1.1
2011	2010	Nat. Orig. Coho Subyearling Fry	—	—	—	—	—	—	—
2011	2010	Nat. Orig. Coho Subyearling Parr	64.7	3	10.8	3.0	3	1.5	1.1
2011	2009	Hatchery Coho Yearling Smolt	124.6	969	8.6	21.0	969	4.8	1.1
2012	2010	Nat. Orig. Coho Yearling Smolt	102.1	17	9.1	11.9	17	3.0	1.1
2012	2011	Nat. Orig. Coho Subyearling Fry	36.0	1	—	—	—	—	—
2012	2011	Nat. Orig. Coho Subyearling Parr	78.4	84	9.3	5.0	84	2.1	1.0
2012	2010	Hatchery Coho Yearling Smolt	126.2	1,684	7.6	21.5	1,684	5.5	1.1
2013	2011	Nat. Orig. Coho Yearling Smolt	97.0	81	10.0	10.0	81	3.1	1.1
2013	2012	Nat. Orig. Coho Subyearling Fry	47.3	3	1.0	1.0	3	1.0	0.9
2013	2012	Nat. Orig. Coho Subyearling Parr	87.8	4	3.8	6.6	4	1.0	1.0
2013	2011	Hatchery Coho Yearling Smolt	130.1	982	8.5	23.3	977	4.9	1.1
2014	2012	Nat. Orig. Coho Yearling Smolt	96.3	20	9.8	9.9	20	3.0	1.1
2014	2013	Nat. Orig. Coho Subyearling Fry	36.0	1	—	—	—	—	—
2014	2013	Nat. Orig. Coho Subyearling Parr	73.0	3	22.5	5.9	3	4.7	1.5
2014	2012	Hatchery Coho Yearling Smolt	127.0	1,203	9.7	21.7	1,207	5.0	1.1



# Appendix L

**Fish Trapping at the White River Smolt Trap during 2014**



# Population Estimates for Juvenile Spring Chinook Salmon in White River, WA

## 2014 Annual Final Report

Prepared by:  
Bryan Ishida  
Cory Kamphaus  
Keely Murdoch

YAKAMA NATION  
FISHERIES RESOURCE MANAGEMENT  
Toppenish, WA 98948



Prepared for:

Public Utility District No. 2 of Grant County  
Ephrata, Washington 98823

## ABSTRACT

In 2007, Yakama Nation Fisheries Resource Management began monitoring emigration of Endangered Species Act (ESA) listed Upper Columbia River (UCR) spring Chinook salmon in the White River to provide abundance and freshwater survival estimates. This report summarizes data collected between March 1 and November 30, 2014. We used a 1.5 m rotary screw trap to collect 261 juvenile spring Chinook; 7 precocial parr, 27 fry, 185 subyearling parr, and 43 yearling smolts. Daily counts caught at the trap were expanded via regression analysis derived from mark and recapture trials. We estimated that 3,995 ( $\pm 3,616$ ; 95% CI) BY2012 wild spring Chinook smolt and 2,482 ( $\pm 851$ ; 95% CI) BY2013 wild spring Chinook parr emigrated past the White River trap. Combined with data collected in 2013, this gives us a total estimate of 7,900 ( $\pm 3,898$ ; 95% CI) BY2012 emigrants. Using spring Chinook spawning ground data collected by Washington Department of Fish and Wildlife (WDFW) in 2012, we estimated egg-to-emigrant survival of BY2012 spring Chinook to be 2.2% (92 smolts-per-redd).



# CONTENTS

LIST OF FIGURES .....	iv
LIST OF TABLES .....	v
ACKNOWLEDGEMENTS .....	vi
1.0 INTRODUCTION.....	1
1.1 Watershed Description .....	1
2.0 METHODS.....	5
2.1 Trapping Equipment and Operation .....	5
2.2 Biological Sampling .....	5
2.3 Mark-Recapture Trials.....	6
2.3.1 Marking and PIT tagging.....	7
2.4 Data Analysis .....	7
2.4.1 Estimate of Abundance.....	7
3.0 RESULTS.....	12
3.1 Dates of Operation.....	12
3.2 Daily Captures and Biological Sampling .....	12
3.2.1 Wild Spring Chinook Yearlings (BY2012).....	12
3.2.2 Wild Spring Chinook Subyearlings (BY2013).....	12
3.2.3 Hatchery Spring Chinook Yearlings (BY2012) .....	14
3.3 Trap Efficiency Calibration and Population Estimates .....	14
3.3.1 Wild Spring Chinook Yearlings (BY 2012).....	14
3.3.2 Wild Spring Chinook Subyearling (BY 2013) .....	14
3.4 PIT Tagging.....	16
3.5 Incidental Species.....	17
3.6 ESA Compliance .....	17
4.0 DISCUSSION .....	19
5.0 LITERATURE CITED.....	21
APPENDIX A: White River Temperature and Discharge Data.....	23
APPENDIX B: Daily Trap Operation Status .....	27
APPENDIX C: Regression Models.....	31
Appendix D. Historical Morphometric Data .....	32

## LIST OF FIGURES

Figure 1. Map of the Wenatchee River subbasin with White River rotary trap location. ..	2
Figure 2. Mean daily stream discharge at the White River DOE stream monitoring station at Sears Creek Bridge in 2014. ....	3
Figure 3. Mean daily water temperatures at the White River DOE stream monitoring station at Sears Creek Bridge in 2014. ....	4
Figure 4. Daily catch of yearling spring Chinook smolt with mean daily stream discharge at the White River rotary trap, March 1 to June 30, 2014. ....	13
Figure 5. Daily catch of wild subyearling spring Chinook with mean daily stream discharge at the White River rotary trap, July 1 to November 30, 2014 .....	13
Figure 6. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for White River spring Chinook, BY 2006 to 2012.....	16
Figure 7. Comparison of wild spring Chinook abundance estimates (BY2007-2012) made at the White R., Nason Cr., and Chiwawa R. smolt traps. Chiwawa R. data provided by Hillman et al. (2014).....	20

## LIST OF TABLES

Table 1. Summary of White River smolt trap operation, 2014.....	12
Table 2. Summary of length and weight sampling of juvenile spring Chinook captured at the White River rotary trap in 2014.....	14
Table 3. Estimated egg-to-emigrant survival and emigrants per redd for White River spring Chinook. ....	15
Table 4. Number of PIT tagged spring Chinook and steelhead with shed rates at the White River rotary trap in 2014.....	17
Table 5. Summary of length and weight sampling of incidental species captured at the White River rotary trap in 2014.....	17
Table 6. Summary of White River ESA listed species catch and mortality in 2014. ....	18

## **ACKNOWLEDGEMENTS**

This project is part of a basin-wide monitoring program requiring close coordination between multiple agencies and contractors. We greatly appreciate the hard work of the Yakama Nation FRM crew members including Matthew Clubb, Jamie Hallman, Barry Hodges, Tim Jeffris and Kevin Swager who maintained and operated the trap during all hours including nights/weekends through challenging weather conditions. Also thank you to Peter Graf (Grant County PUD) for administering contracting and funding as well as John Walters and Mike Hughes (WDFW) for data sharing and collaborating on smolt trap methodologies.

## 1.0 INTRODUCTION

White River spring Chinook salmon (tkwínat) *Oncorhynchus tshawytscha* are part of the Upper Columbia River (UCR) spring Chinook salmon Evolutionarily Significant Unit (ESU) which was listed as endangered under the Endangered Species Act (ESA) in 1999. Due to critically low abundance, a captive broodstock program has operated in the White River since 1997 as a risk aversion measure. Determining freshwater productivity of spring Chinook salmon in the White River is an essential component to overall population monitoring and will help contribute to the body of knowledge needed to evaluate if further supplementation in the White River is warranted. In 2007, Public Utility District No. 2 of Grant County (GCPUD) contracted the Yakama Nation (YN) to operate a rotary trap in the White River. Fish trap operations were conducted in compliance with ESA consultation specifically to address abundance and productivity of spring Chinook salmon in the White River.

Within this document, we will report:

- 1) Juvenile abundance and productivity of spring Chinook salmon in the White River.
- 2) Emigration timing of spring Chinook salmon emigrating from the White River.

Data presented will be directly used to address Objective 2 in the Monitoring and Evaluation Plan for PUD Hatchery Programs (Hillman et al. 2013) on a 5-year analytic cycle:

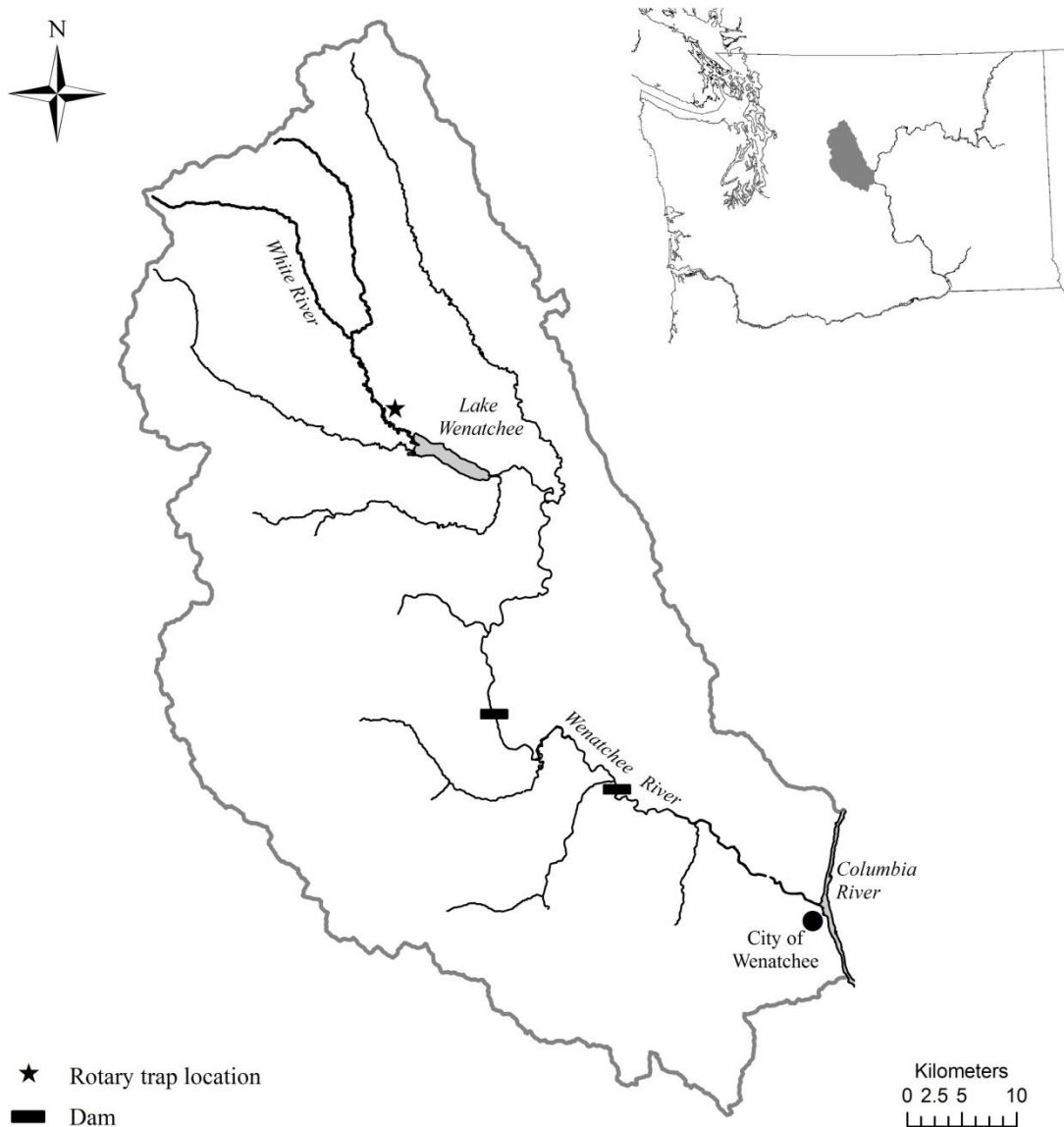
**Objective 2: Determine if the proportion of hatchery fish on the spawning grounds affects the freshwater productivity of supplemented stocks (Hillman et al. 2013).**

In the fall of 2005, Washington State Department of Fish and Wildlife (WDFW) began trapping in the lower reaches of the White River in order to provide an estimate of juvenile spring Chinook salmon production. No trapping was conducted in 2006 as there was a transition between trap operators. In 2007, YN resumed trap operations on the White River for nine months of the year. This document reports data collected between March 1 and November 30, 2014 and provides emigration estimates for spring Chinook salmon yearlings (BY2012) and subyearlings (BY2013) during that time period. Data generated from this project was used to calculate annual egg-to-emigrant survival.

## 1.1 Watershed Description

The White River drainage encompasses 99,956 acres and originates in alpine glaciers and perennial snow fields (Figure 1; USFS 2004). Elevations in the drainage vary from 1,868 ft. at the Lake Wenatchee surface to 8,575 ft. at Clark Mountain (Andonaegui 2001). As one of two primary tributaries to Lake Wenatchee, the White River flows in a south-easterly direction for 42.9 river kilometers (RK) before emptying into the lake. Precipitation ranges from 79 cm at the

mouth to more than 356 cm in the head waters (Andonaegui 2001). Due to its glacial origins, peak runoff for the White River typically occurs between April and July with occasional high

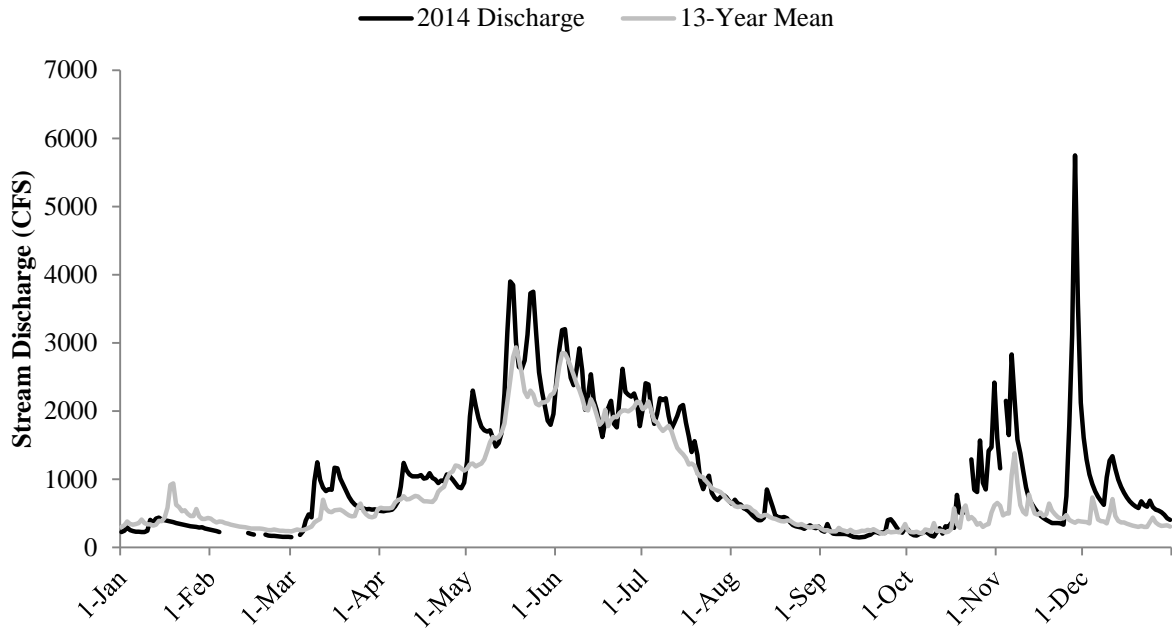


**Figure 1. Map of the Wenatchee River subbasin with White River rotary trap location.**

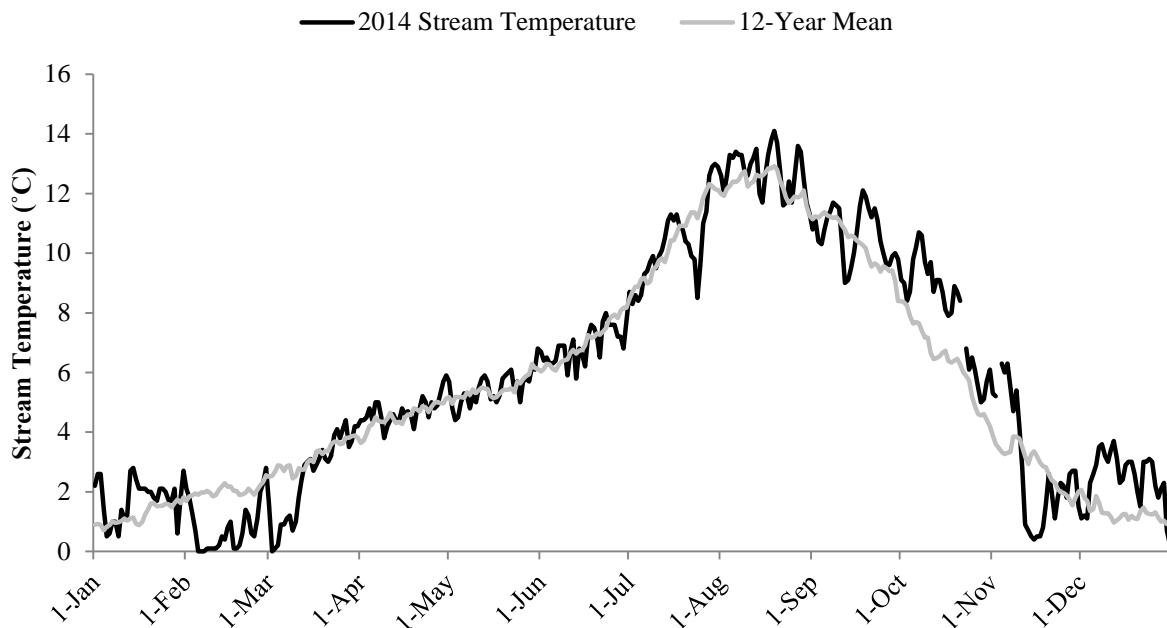
flows caused by rain-on-snow events in the fall and winter months. Water temperatures in this watershed tend to be cooler than other tributaries to the upper Wenatchee River subbasin. As of September 2002, Washington State Department of Ecology (WDOE) began operating a stream monitoring station at RK 9.9 of White River. Operation of this station by WDOE is currently maintained with funding provided by GCPUD. In 2014, daily mean stream discharge ranged from 146cfs to 5,750cfs (Figure 2) while mean daily stream temperatures ranged from 0.0°C to

14.1°C (Figure 3). Discharge and temperature data provided by WDOE should be considered provisional and are presented in **Appendix A**.

The White River drainage has had minimal riparian harvest from the 1950's to the present on federally owned land. Turn of the century settlement and land clearing have impacted the



**Figure 2. Mean daily stream discharge at the White River DOE stream monitoring station at Sears Creek Bridge in 2014.**



**Figure 3. Mean daily water temperatures at the White River DOE stream monitoring station at Sears Creek Bridge in 2014.**

riparian reserve network up to the Napeequa confluence, yet, riparian areas in the mainstem below Panther Creek remain in fair condition (USFS 2004). In the remainder of the watershed woody debris recruitment, shade, aquatic habitat connectivity, and riparian vegetation appear to be in good condition. Current habitat concerns pertaining to the development of homes and vacation retreats on private lands do exist. Rip-rapping, channel constriction, and stream degradation are considered minor in the watershed. Public ownership comprises 78% of the drainage area; more than half of public land is located within the Glacier Peak Wilderness. The remaining 22% of the drainage is in private ownership (USFS 2004).

Downstream of White River Falls are key spawning grounds for spring Chinook salmon (tkwínat) *Oncorhynchus tshawytscha*, sockeye salmon (kálux) *O. nerka*, and bull trout *Salvelinus confluentus*. Two large tributaries to the White River, Napeequa River and Panther Creek, are also known to support populations of anadromous salmonids (Mullen et al. 1992). For a complete list of known fish species encountered in the White River see (3.4 Incidental Species).



## 2.0 METHODS

### 2.1 Trapping Equipment and Operation

In 2014, a 1.5m diameter cone rotary trap was operated in a single position at all discharge levels. This revised trapping regime was implemented in 2013 to simplify data analysis by eliminating obsolete trap positions that generated very little data. Past attempts at developing a high flow position generated very few efficiency trials resulting in limited trap efficiency data. Operating season long at a single position, the trap was suspended from a river-spanning cable and its position could be adjusted perpendicular to stream flow by hand powered winches anchored on a tree on the river-right bank.

The trap was operated 24 hours per day, seven days per week for the majority of the season. During spring snowmelt, operations only occurred during hours of darkness to minimize trap damage and subsequent capture mortality; still enabling sampling during the hours of peak fish movement. When trap operations were suspended, the cone was raised to avoid damage by debris.

During all ranges of river discharge, fish were removed daily. Additional trap checks were necessary during periods of high discharge in the spring and in the autumn due to increased fall foliage. Debris in the live-box was removed continually by a rotating drum screen, located at the rear of the holding box and hydraulically powered by the cone. A record of daily trap operations is provided in **Appendix B**.

### 2.2 Biological Sampling

Trap operating procedures and techniques followed a standardized, basin-wide monitoring plan developed by the Upper Columbia Regional Technical Team (UCRTT) for the Upper Columbia Salmon Recovery Board (UCSRB; Hillman 2004), which was adapted from Murdoch & Petersen (2000).

Captured fish were transferred from the rotary trap's live box using five-gallon plastic buckets with lids to a stream-side, portable sampling station. Fish were anesthetized in a solution of MS-222 to facilitate sampling and reduce handling stress. Fork length (FL) and weight were recorded for all fish, except large numbers of sockeye (*Oncorhynchus nerka*) fry. For these fish, a representative sample of 25 individuals was measured and weighed while the remaining fish were enumerated and released. Weight was measured to the nearest 0.1 g with a portable digital scale while FL was recorded to the nearest 1.0 mm using a trough-type measuring board. These data were used to calculate a Fulton-type condition factor (K-factor) for each target species using the formula:

$$K = (W/L^3) \times 100,000$$

Where K = Fulton-type condition metric, W = weight in grams, L = fork length in millimeters and 100,000 is a scaling constant.

Portable aerators were used to oxygenate holding water during sampling. All fish were allowed to fully recover from anesthesia before being released. Spring Chinook salmon were classified as either natural or hatchery origin by the presence/absence of coded wire tags (CWT's). Developmental stages (fry, parr, transitional or smolt) were visually identified and assigned to each individual sampled. Transitional juveniles were identified as having both parr and smolt characteristics; visible parr marks, semi-transparent fin coloration along with silvery coloration throughout body. Smolts were identified by a strong silvery coloration over entire body and faint or absent parr marks. Fry were defined as newly emerged fish with or without a visible yolk sac and a FL measuring < 50 mm. Age-0 spring Chinook salmon captured before July 1 were considered 'fry' and excluded from population estimates due to the inconclusive nature of their movement (i.e. active emigration or local distribution in-stream). Age-0 spring Chinook salmon captured after 1 July were considered subyearling emigrants and included in the population estimate (UCRTT, 2001).

Tissue samples were taken from spring Chinook salmon and steelhead (small, upper lobe caudal fin clip) and applied to blotter sheets. Samples from both species were provided to WDFW for reproductive success analysis. Scale samples were also collected from all steelhead captured. Scale samples were submitted to WDFW for age analysis. Bull trout tissue samples were not collected in 2014.

During periods when the trap operations were suspended (e.g. - high discharge, high debris and/or mechanical problems), passage estimates were generated to account for emigrants during these time periods. This estimate was calculated using the average number of fish captured three days prior and three days after the break in operation (Hillman et al., 2013; Snow et al., 2013).

### 2.3 Mark-Recapture Trials

Groups of marked spring Chinook salmon were used for trap efficiency trials. Fish were marked by insertion of a passive integrated transponder (PIT) tag into the abdominal cavity. Ideally, marked groups of fish would be released over a broad range of stream discharges in order to determine a trap efficiency-discharge relationship. (See **2.4 Data Analysis**). However, due to low abundance and limited holding time of ESA listed species (reducing the ability to meet trials size requirements on a more consistent basis), marked groups were released whenever the minimum sample size ( $\geq 20$ ) was obtained. Mark-recapture (M-R) trials followed the protocol described in Hillman (2004). Although the protocol suggests a minimum sample size of 100 fish for each mark-group, the limited abundance of juvenile emigrants from the White River required that efficiency trials be completed with much smaller sample sizes. YN's continued goal is to increase individual mark-group sizes, when possible, to meet the standard described above.

Number of wild fish included in a marked group was maximized by combining catches from three days of trapping. Fish were held up to 72 hours prior to release and retained in holding boxes attached to the end of each pontoon. Fish to be used in efficiency trials were then transported in five gallon buckets ~1.0 RK upstream to the release location at Sears Creek Bridge (RK 10.3). Marked groups used in efficiency trials were released at nautical twilight via an automatic release box. Visual observation concluded that all fish exit the box shortly after release using this automated method.

Each M-R trial was conducted over a three-day (72 hour) period to allow time for passage or capture. Completed trials were only considered invalid if an interruption to trapping occurred or proper pre-release procedures were not followed. Trials resulting in zero recaptures were included in the efficiency regression as allowed by the new method of observed trap efficiency calculation (See equation 3 in **2.5.1 Estimate of Abundance**).

### 2.3.1 Marking and PIT tagging

All spring Chinook and summer steelhead juveniles with FL of  $\geq 60$ mm were PIT tagged unless the health of an individual was in question (e.g.- fungus). Once anesthetized, each fish was examined for external wounds or descaling and scanned for the presence of a previously implanted PIT tag. If a tag was not detected, a pre-loaded 12mm Digital Angel 134.2 kHz type TX 1411ST PIT tag was inserted into the body cavity using a Biomark MK-25 Rapid Implant Gun. Each unique tag code was electronically recorded with an appropriate tagging date, release date, tagging personnel and biological data. These data were entered into P<sub>3</sub> and submitted to the PIT Tag Information System (PTAGIS) at the end of each month. Tagging methods were consistent with methodology described in the PIT Tag Marking Procedures Manual (CBFWA 1999) as well as with 2008 ISEMP protocols (Tussing 2008).

After marking and/or PIT tagging, fish were held for a minimum of 24-hours to a) ensure complete recovery, b) assess tagging mortality and c) determine tag-shed rate. Fish that were not to be used in an efficiency trial were released downstream of the smolt trap.

## 2.4 Data Analysis

### 2.4.1 Estimate of Abundance

A recent WDFW review of smolt monitoring programs in the Wenatchee basin suggested that changes in the calculations for estimating abundance and its associated variance were necessary. Calculation of daily and seasonal smolt abundance changed only slightly. More significant changes were made to the variance estimator making the calculations more complex. The following describes the revised calculation of the point estimate, variance, and standard error of seasonal smolt abundances based on regression relationships.

Seasonal juvenile migration,  $N$ , was estimated as the sum of daily migrations,  $N_i$ , i.e.,

$N = \sum_i N_i$ , and daily migration was calculated from catch and efficiency:

$$\hat{N}_i = \frac{C_i}{\hat{e}_i}, \quad (1)$$

where  $C_i$  = number of fish caught in period  $I$ ;

$\hat{e}_i$  = trap efficiency estimated from the flow-efficiency relationship,  $\sin^2(b_0 + b_1 \text{flow}_i)$ ,

where  $b_0$  is estimated intercept and  $b_1$  is the estimated slope of the regression.

The regression parameters  $b_0$  and  $b_1$  are estimated using linear regression for the model:

$$\arcsin\left(\sqrt{e_k^{obs}}\right) = \beta_0 + \beta_1 flow_k + \varepsilon, \quad (2)$$

where  $e_k^{obs}$  = observed trap efficiency of Eq. 2 for trapping period  $k$ ;

$\beta_0$  = intercept of the regression model;

$\beta_1$  = slope parameter;

$\varepsilon$  = error with mean 0 and variance  $\sigma^2$ .

In Equation 2, the observed trap efficiency,  $e_k^{obs}$ , is calculated as follows,

$$e_k^{obs} = \frac{r_k + 1}{m}. \quad (3)$$

The estimated variance of seasonal migration is calculated from daily estimates as:

$$Var\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i Var(N_i)}_{Part A} + \underbrace{\sum_i \sum_j Cov(N_i, N_j)}_{Part B},$$

or,

$$Var\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i Var\left(\frac{(C_i + 1)}{\hat{e}_i}\right)}_{Part A} + \underbrace{\sum_i \sum_j Cov\left(\frac{(C_i + 1)}{\hat{e}_i}, \frac{(C_j + 1)}{\hat{e}_j}\right)}_{Part B}. \quad (4)$$

Part A of equation 4 is the variance of daily estimates. Part B is the between-day covariance. Note that the between-day covariance exists only for days that use the same trap efficiency model. If, for example, day 1 is estimated with one trap efficiency model, and day 2 estimated from a different model, then there is no covariance between day 1 and day 2. The full expression for the estimated variance:

$$\hat{V}ar\left(\sum_{i=1}^n \hat{N}_i\right) = \underbrace{\sum_i \hat{N}_i^2 \left( \frac{N_i \hat{e}_i (1 - \hat{e}_i)}{(C_i + 1)^2} + \frac{4(1 - \hat{e}_i)}{\hat{e}_i} \hat{V}ar(b_0 + b_1 flow_i) \right)}_{PartA} + \underbrace{\sum_i \sum_j 4(\hat{N}_i (1 - \hat{e}_i))(\hat{N}_j (1 - \hat{e}_j)) \cdot [\hat{V}ar(b_0) + flow_i flow_j \hat{V}ar(b_1)]}_{PartB}$$

where  $\hat{V}ar(b_0 + b_1 flow_i) = M\hat{S}E\left(1 + \frac{1}{n} + \frac{(flow_i - \overline{flow})^2}{(n-1)s_{flow}^2}\right)$ , and  $\hat{V}ar(b_0)$  and  $\hat{V}ar(b_1)$  are

obtained from regression results. In Excel, the standard error (SE) of the coefficients is provided. The variance is calculated as the square of the standard error,  $SE^2$ .

In cases when there was no significant flow-efficiency relationship (i.e., low correlation), then a pooled, or average trap efficiency will suffice for the stratum. The estimator is calculated as follows:

$$\hat{e} = \frac{\sum_{j=1}^k r_j}{\sum_{j=1}^k m_j}$$

where  $\hat{e}$  = the average or pooled trap efficiency for the stratum;

$m_j$  = the number of smolts marked and released in efficiency trial  $j$  for the stratum;

$r_j$  = the number of smolts recaptured out of  $m_j$  marked fish in efficiency trial  $j$ .

Abundance for a trapping period is estimated as:

$$\hat{N}_i^{pooled} = \frac{C_i}{\hat{e}},$$

,and total stratum abundance is:

$$N^{pooled} = \sum_i \hat{N}_i^{pooled}.$$

The variance of seasonal abundance takes into account the variability in catch numbers that are a result of binomial sampling (Part A), the pooled variance of trap efficiency,  $\hat{e}$  (Part B), and the covariance in daily estimates that arises from using a common estimate of efficiency across all trapping days (Part C):

$$V\hat{a}r\left(\sum_{i=1}^n \hat{N}_i^{pooled}\right) = \underbrace{\left(\sum_i \frac{\hat{N}_i(1-\hat{e})}{\hat{e}}\right)}_{PartA} + \underbrace{\frac{Var(\hat{e})}{\hat{e}^2} \sum_i \hat{N}_i^2}_{PartB} + \underbrace{\frac{Var(\hat{e})}{\hat{e}^2} \sum_i \sum_j \hat{N}_i \hat{N}_j}_{PartC}.$$

The Part B and Part C terms are combined in the calculation as a new Part B:

$$V\hat{a}r\left(\sum_{i=1}^n \hat{N}_i^{pooled}\right) = \underbrace{\left(\sum_i \frac{\hat{N}_i(1-\hat{e})}{\hat{e}}\right)}_{PartA} + \underbrace{\frac{Var(\hat{e})}{\hat{e}^2} \left[ \sum_i \hat{N}_i^2 + \sum_i \sum_j \hat{N}_i \hat{N}_j \right]}_{PartB}.$$

The variance of  $\hat{e}$  is calculated as:

$$V\hat{a}r(\hat{e}) = V\hat{a}r\left(\frac{\sum_{k=1}^n r_k}{\sum_{k=1}^n m_k}\right) = \frac{\sum_{k=1}^n (r_k - \hat{e} m_k)^2}{\bar{m}^2 n(n-1)}$$

where  $\bar{m}$  is the average release size across all efficiency trial,  $\frac{\sum_{k=1}^n m_k}{n}$ .

Confidence intervals were calculated using the following formulas:

$$95\% \text{ confidence interval} = 1.96 \times \sqrt{\sum \text{var}[\hat{N}_i]}$$

The single M-R estimator of abundance carries a set of well documented assumptions (Everhart and Youngs 1981; Seber 1982),

1. The population is closed to mortality.
2. The probability of capturing a marked or unmarked fish is equal.
3. Marked fish were randomly dispersed in the population prior to recapture.
4. Marking does not affect probabilities of capture.

5. Marks were not lost between the time of release and recapture.
6. All marks are reported upon recapture.
7. The number of fish in the trap, C, is fully enumerated and known without error.

## 3.0 RESULTS

### 3.1 Dates of Operation

In 2014, we operated a 1.5m rotary trap between March 1 and November 30. During this period, the trap operated 24 hours per day, 7 days per week barring inoperable environmental conditions (i.e. heavy debris loads or high discharge) or mechanical malfunctions. Trapping was interrupted or intentionally suspended a total of 20 times during the season due to: a) debris/ice jams, b) unsafe working conditions, or c) maintenance and repairs (Table 1).

**Table 1. Summary of White River smolt trap operation, 2014.**

Trap Status	Description	Days
Operating	Continuous data collection	255
Interrupted	Interrupted by debris, ice, and/or flows	13
Pulled	Intentionally pulled to prevent harm to fish or protect the trap during high flows	7

### 3.2 Daily Captures and Biological Sampling

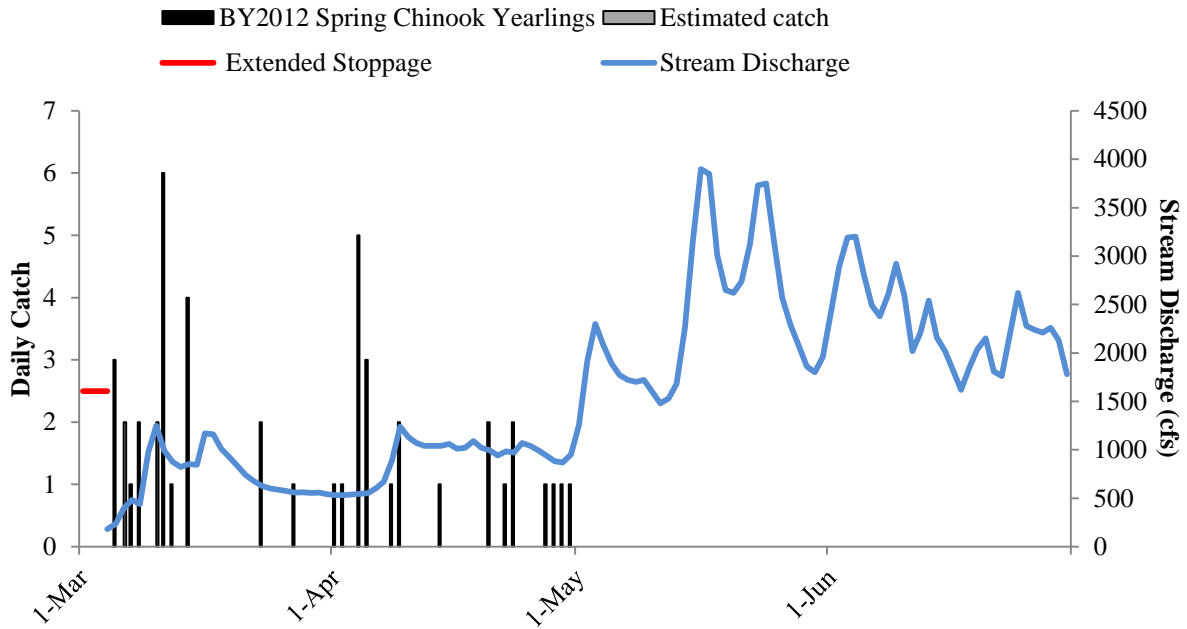
#### 3.2.1 Wild Spring Chinook Yearlings (BY2012)

A total of 43 wild yearling Chinook smolts were collected between March 1 and June 30, with peak catch occurring on March 11 ( $n = 6$ ; Figure 4). Mean FL was 92.3mm ( $n = 43$ ;  $SD = 7.0$ ) and mean weight was 9.4g ( $n = 43$ ;  $SD = 2.2$ ); see Table 2. All 43 smolts were PIT tagged. Genetic samples were also taken from 40 fish. An additional seven yearling Chinook were captured after July 1. Mean FL for these yearlings was 126.9mm ( $n = 7$ ;  $SD = 13.5$ ) and mean weight was 23.2 g ( $n = 7$ ;  $SD = 7.4$ ); see Table 2. These fish were identified as precocial parr by their large size, timing of capture, and release of milt during handling. All precocial parr were excluded from emigration estimates. There were no yearling Chinook mortalities (See **3.4 ESA Compliance**).

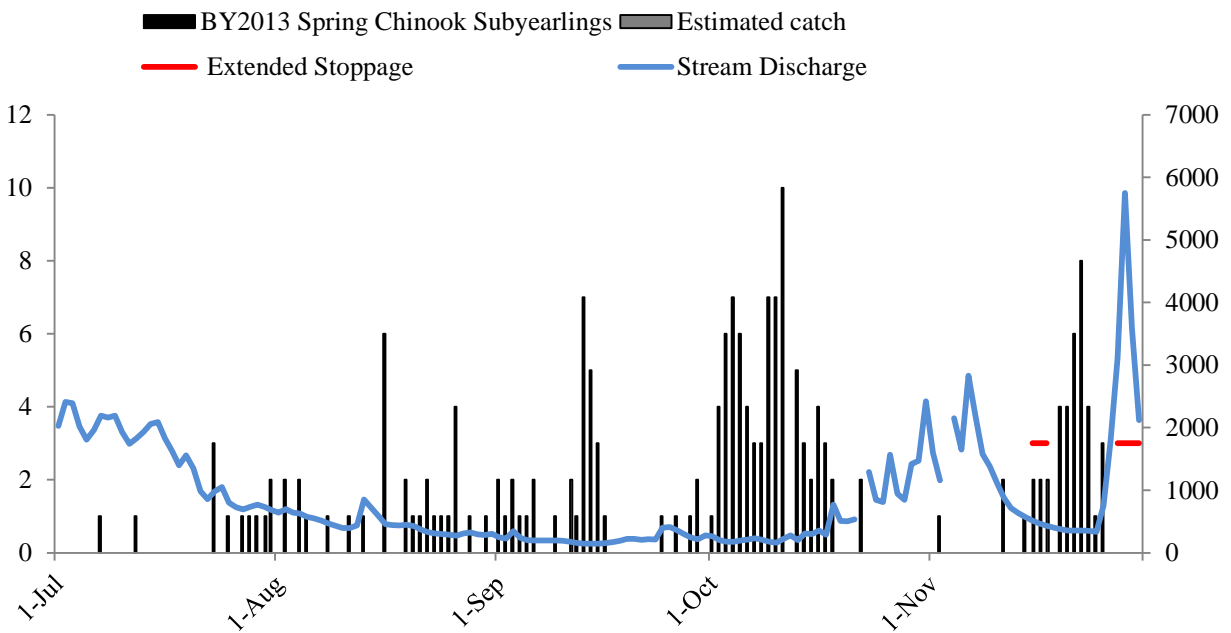
#### 3.2.2 Wild Spring Chinook Subyearlings (BY2013)

Spring Chinook fry were captured at the trap between March 21 and June 29 ( $n = 27$ ). During this period one fry trapping mortality was incurred. A total of 185 wild subyearling Chinook parr were collected between July 7 and November 25, with peak catch occurring on October 11 ( $n = 10$ ; Figure 5). The mean FL for subyearling parr was 86.0mm ( $n = 185$ ;  $SD = 14.1$ ) and the mean weight was 7.5g ( $n = 185$ ;  $SD = 3.3$ ); see Table 2. PIT tags were implanted into a total of 156 subyearling Chinook parr. Genetic samples were also taken from all 156 of the parr tagged (Table 4). Tags were not implanted into 14 parr showing signs of parasitic and/or fungal infection while 1 parr was not tagged due to visible injuries or poor condition. An additional 12 parr did not meet the minimum tagging length and were subsequently released without marking. Mortality of Chinook subyearlings during the 2014 trapping season totaled two fish (0.9%; See **3.4 ESA Compliance**).





**Figure 4. Daily catch of yearling spring Chinook smolt with mean daily stream discharge at the White River rotary trap, March 1 to June 30, 2014.**



**Figure 5. Daily catch of wild subyearling spring Chinook with mean daily stream discharge at the White River rotary trap, July 1 to November 30, 2014**

### 3.2.3 Hatchery Spring Chinook Yearlings (BY2012)

Hatchery-origin yearling Chinook released downstream of the smolt trap are sometimes caught in the summer months as precocial parr. Direct releases of BY2012 spring Chinook were not performed in the White River or in close proximity to its confluence with Lake Wenatchee (netpen-rearing). There were no hatchery-origin spring Chinook captured at the smolt trap in 2014. Hatchery fish captured at the trap are identified by the presence of CWT tags.

**Table 2. Summary of length and weight sampling of juvenile spring Chinook captured at the White River rotary trap in 2014.**

Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-factor
		Mean	n	SD	Mean	n	SD	
2012	Wild Yearling Smolt	94.3	43	7.0	9.4	43	2.2	1.1
2012	Wild Yearling Precocial Parr	126.9	7	13.0	23.2	7	7.4	1.1
2013	Wild Subyearling Fry	40.0	22	3.8	—	—	—	—
2013	Wild Subyearling Parr	86.0	185	14.1	7.5	185	3.3	1.2

## 3.3 Trap Efficiency Calibration and Population Estimates

### 3.3.1 Wild Spring Chinook Yearlings (BY 2012)

Due to low abundance, no BY2012 natural yearling Chinook efficiency trials were performed in 2014. A composite regression model using previous year's (2008-2012) efficiency trials showed statistical significance ( $r^2 = 0.56$ ;  $p = 0.001$ ) for a flow-efficiency relationship and was used to calculate yearling abundance. Use of a single spring trapping position allowed this regression to be applied to all yearling Chinook captured in 2014. Weighting of this regression via an R script (provided by WDFW) did not affect calculation parameters greatly and yielded the same r-square and  $p$ -values. In the fall of 2013, we estimated that 3,905 ( $\pm 1,456$ ; 95% CI) BY2012 subyearlings emigrated past the trap. In the spring of 2014, we estimated that 3,995 ( $\pm 3,616$ ; 95% CI) emigrated past the trap. Combining the two estimates, total BY2012 wild spring Chinook emigrants was 7,900 ( $\pm 3,898$ ; 95% CI; Table 3).

### 3.3.2 Wild Spring Chinook Subyearling (BY 2013)

Low parr abundance presented few opportunities to perform mark-group releases in 2014. Our current strategy to improve the flow-efficiency model includes targeting mark-group releases at discharge levels where data is currently lacking. For all other discharge levels only large mark-groups will be released ( $n \geq 50$ ); continuing to release small groups ( $n \leq 50$ ) of fish at discharge levels where sufficient data exists will only serve to increase variability in the model. A multi-year composite regression was applied to BY2013 subyearling emigrants. The regression was comprised of all trials conducted fulfilling the minimum number marked ( $n \geq 20$ ) including efforts in which zero recaptured were made (Appendix C). Mark-groups in which validity of the trial could be called into question (suspected trap stoppage or improper pre-release handling of the mark group) were removed. The weighted regression was not significant ( $r^2 = 0.14$ ;  $p = 0.079$ ) at our accepted limit ( $\alpha = 0.05$ ). However, after comparison with a pooled method and

considerations of the pooled estimate limitations, we decided to use the regression model despite its slightly higher *p*-value (for more explanation please see Section 4.0 Discussion). This single regression was the only model required to estimate total subyearling migration due to the fact only one fall trapping position was used in 2014. We estimated that in 2014, 2,482 ( $\pm$  851; 95% CI) spring Chinook subyearling parr moved past the trap (Table 3).

**Table 3. Estimated egg-to-emigrant survival and emigrants per redd for White River spring Chinook.**

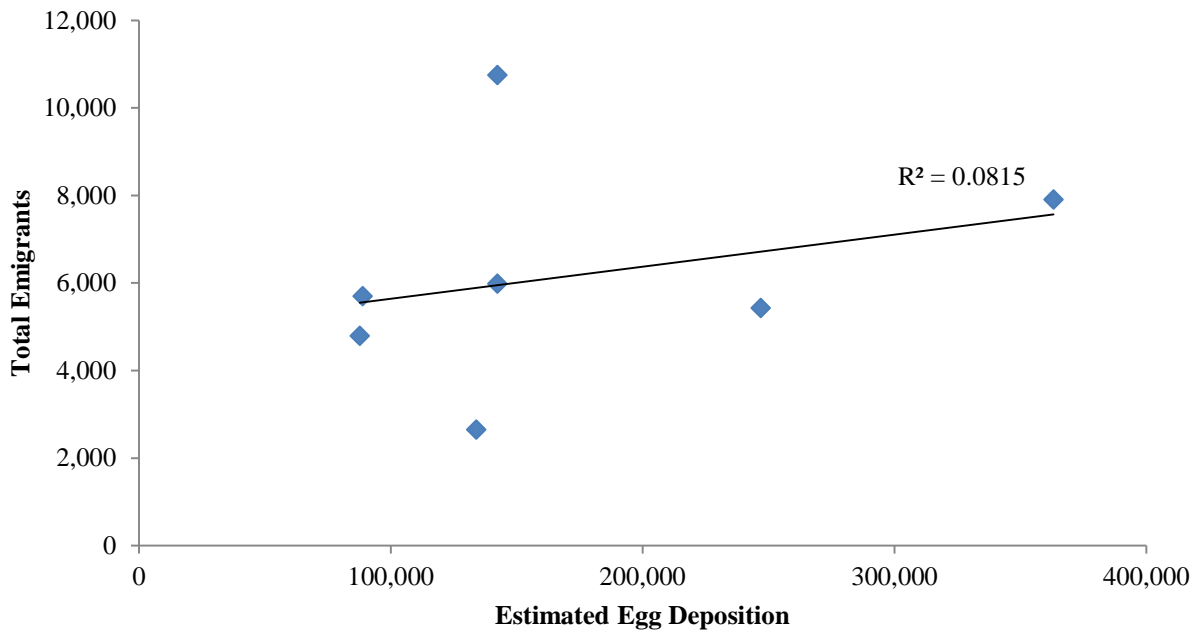
Brood Year	No. of Redds <sup>a</sup>	Fecundity <sup>b</sup>	No. of Eggs	No. of Emigrants			Egg-to Emigrant	Emigrants per Redd
				Age-0 <sup>c</sup>	Age-1	Total $\pm$ 95% CI		
2005	86	4,327	372,122	DNOT <sup>d</sup>	4,856	—	—	—
2006	31	4,324	134,044	642	2,004	2,646 $\pm$ 1,597	2.0%	85
2007	20	4,441	88,820	2,293	3,399	5,692 $\pm$ 2,214	6.4%	285
2008	31	4,592	142,352	5,552	5,193	10,745 $\pm$ 3,837	7.5%	347
2009	54	4,573	246,942	2,485	2,939	5,424 $\pm$ 2,522	2.2%	100
2010	33	4,314	142,362	1,859	4,121	5,980 $\pm$ 3,455	4.2%	181
2011	20	4,385	87,700	3,128	1,659	4,787 $\pm$ 2,022	5.5%	239
2012	86	4,223	363,178	3,905	3,995	7,900 $\pm$ 3,898	2.2%	92
2013	54	4,716	254,664	2,482	—	—	—	—
Avg	39	4,407	172,200	2,838	3,330	6,168	4.3%	190

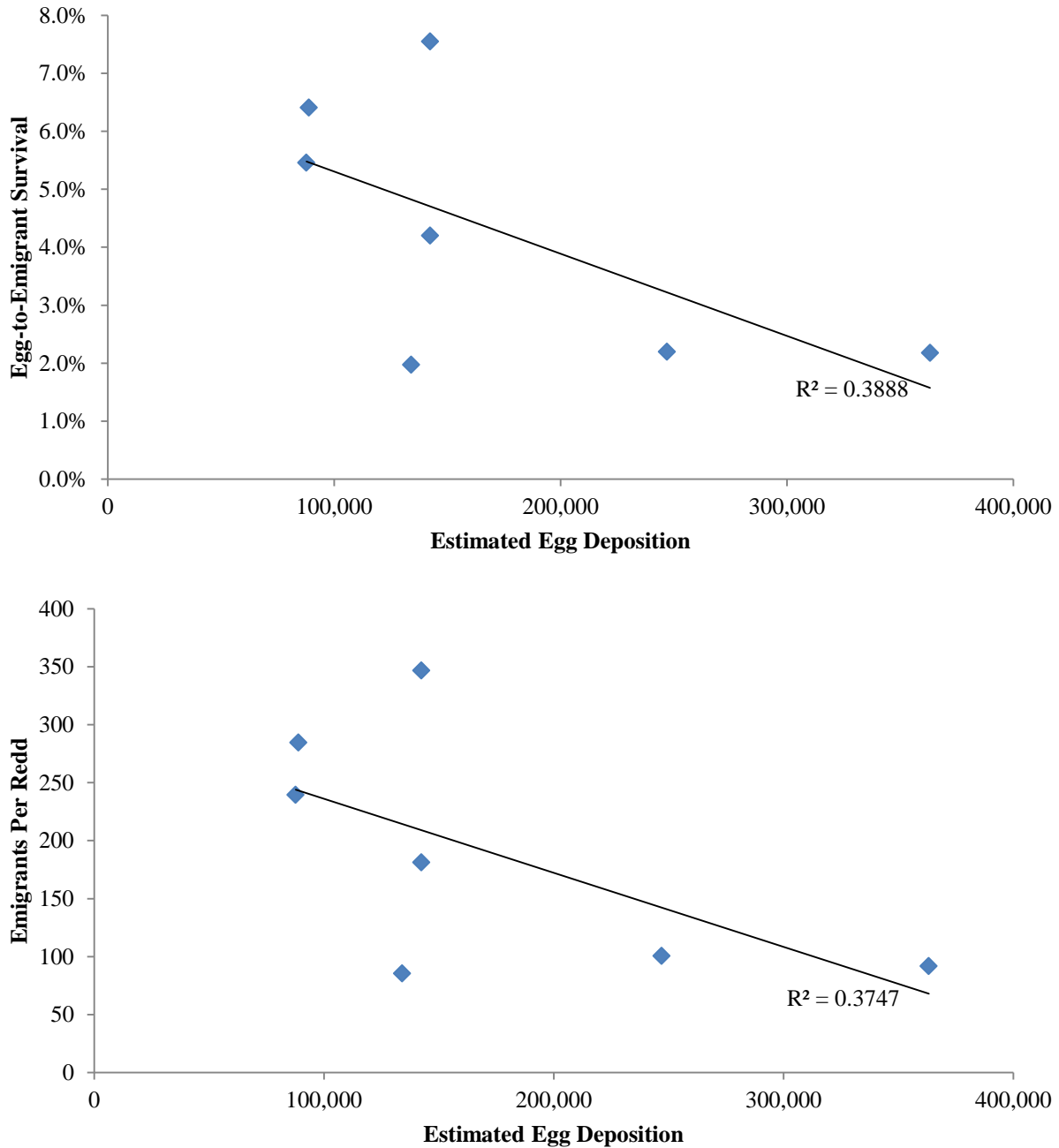
<sup>a</sup> Number of complete redds in White River (Hillman et al. 2014)

<sup>b</sup> Mean annual fecundity of wild spring Chinook broodstock at Chiwawa River Hatchery

<sup>c</sup> Estimate is based on capture of parr collected during summer/fall and does not include fry captured prior to July 1.

<sup>d</sup> Did not operate trap; no production estimates were made.





**Figure 6. Relationships between estimated egg deposition and total emigrants produced, egg-to-emigrant survival, and emigrants per redd for White River spring Chinook, BY 2006 to 2012.**

### 3.4 PIT Tagging

In 2014, a total of 234 spring Chinook and 5 steelhead were PIT tagged at the trap. PIT tag retention after 24 hours of observation was 100% (Table 4). There no tagging mortalities (Table 6).

**Table 4. Number of PIT tagged spring Chinook and steelhead with shed rates at the White River rotary trap in 2014.**

Brood Year	Species/Stage	Total Catch	Total PIT Tagged	Percent Tagged	Percent Tags Shed
2012	Yearling Chinook smolt	43	43	100.0%	0.0%
2012	Yearling Chinook Precocial Parr	7	6	85.7%	0.0%
2013	Subyearling Chinook parr	185	156	84.3%	0.0%
*	Steelhead parr	5	5	100.0%	0.0%

\* Brood year unknown

### 3.5 Incidental Species

Incidental species were enumerated and sampled for length and weight (Table 5). Incidental species included: bull trout *Salvelinus confluentus*, steelhead/rainbow trout (shúshaynsh) *Oncorhynchus mykiss*, sockeye salmon *Oncorhynchus nerka*, mountain whitefish *Prosopium williamsoni*, sculpin *Cottus sp.*, northern pikeminnow *Ptychocheilus oregonensis*, redbside shiner *Richardsonius balteatus*, longnose dace *Rhinichthys cataractae*, westslope cutthroat *Oncorhynchus clarkii lewisi*, and sucker *Catostomus sp.*.

**Table 5. Summary of length and weight sampling of incidental species captured at the White River rotary trap in 2014.**

Species	Total Count	Fork Length (mm)			Weight (g)		
		Mean	<i>n</i>	SD	Mean	<i>n</i>	SD
<b>Bull Trout Fry</b>	11	30.7	11	5.7	-	-	-
<b>Bull Trout Parr</b>	2	140.0	2	89.7	64.5	2	89.7
<b>Mountain Whitefish</b>	64	69.8	64	21.0	4.3	61	6.6
<b>Sculpin</b>	59	57.7	59	19.8	3.5	47	2.2
<b>Northern Pikeminnow</b>	3	95.3	3	28.3	9.7	3	9.5
<b>Redside Shiner</b>	13	66.2	13	16.0	3.8	13	14.2
<b>Dace</b>	7	70.4	7	23.2	4.2	7	3.7
<b>Sucker</b>	9	174.4	9	128.8	109.2	7	139.0
<b>Cutthroat</b>	9	224.0	9	53.3	118.9	8	49.1
<b>Sockeye Fry</b>	1,355	27.8	770	1.6	-	-	-
<b>Sockeye Parr</b>	6	60.8	6	9.5	2.1	6	1.1
<b>Rainbow Trout/Steelhead Parr</b>	5	164.4	5	50.4	56.9	5	40.4

### 3.6 ESA Compliance

The total combined mortality rate for spring Chinook captured at the trap in 2014 was 1.1% (Table 6). Two parr were found dead in the holding box after handling and were therefore deemed as trapping mortalities. A single fry was found crushed in the live box and considered a trapping mortality. Fungus observed has previously been identified by a USFWS fish pathologist as a common *sp. Saprolegnia*. For the past several years, trap personnel have

observed fungal infections on and around the pectoral fins of juvenile Chinook. Closer inspection of infected fish has recently shown that the fungus is associated with parasitic copepods attaching at the base of the pectoral fins. Generally, live fish with visible infections are immediately released from the trap and not otherwise handled or sampled. At times, discovery of infection did not occur until the fish were anesthetized for sampling. Such fish were not PIT tagged and immediately released downstream of the trap.

**Table 6. Summary of White River ESA listed species catch and mortality in 2014.**

Species/Stage	Total Catch	Fungal Mortality	Trap Mortality	Tagging Mortality	Total % Mortality
Yearling Chinook Smolt	43	0	0	0	0.0%
Yearling Chinook Precocial Parr	7	0	0	0	0.0%
Subyearling Chinook Parr	189	0	2	0	1.1%
Subyearling Chinook Fry	23	0	1	0	4.3%
<b>Total Wild Spring Chinook</b>	<b>262</b>	<b>0</b>	<b>3</b>	<b>0</b>	<b>1.1%</b>
Bull Trout	13	0	0	0	0.0%
Steelhead/Rainbow Trout	5	0	0	0	0.0%

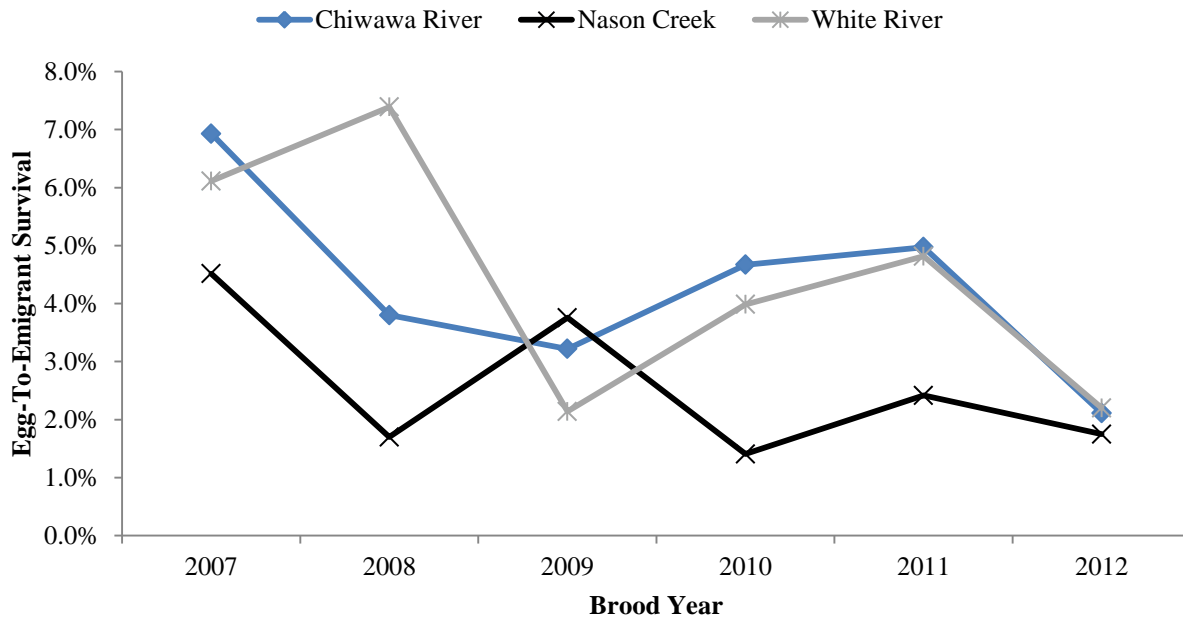
## 4.0 DISCUSSION

Expansions of yearling (BY2012) and subyearling (BY2013) spring Chinook catch data collected in 2014 were performed using the same multi-year compounded regressions used in previous years. Based on the assumption that channel morphology and the flow-efficiency relationship remain constant in time, regression analysis provides the ability to make estimates when a year-specific relationship cannot be achieved. This attribute is especially important at the White River smolt trap, where in 2014 low abundance provided limited opportunities to perform efficiency trials.

Using these regression analysis techniques, we estimated that 3,995 BY2012 spring Chinook yearlings emigrated past the trap in the spring of 2014. This above-average estimate of yearling emigrants falls in line with the high spawner escapement of the 2012 adult spring Chinook return. Including the previous estimate of 3,905 BY2012 subyearling parr captured in the fall of 2013, the total expansion of 7,900 BY2012 emigrants is the second largest in the past seven broods. Despite the high relative abundance of BY2012 emigrants, the estimated egg-to-emigrant survival rate (2.2%) is below the seven-brood average (4.3%).

With no subyearling efficiency trials performed in 2014, we used our existing model to generate a population estimate (despite lack of significance at  $\alpha = .05$ ). Using the existing flow-efficiency relationship we estimated that a total of 2,482 BY2013 spring Chinook parr emigrated past the trap in the fall of 2014. Our strategy for building a statistically robust flow-efficiency model includes targeting discharge levels where efficiency data is currently lacking. For discharge ranges where sufficient data exists, only large release groups (sample sizes larger than those used to build the current model) will be used. Frequent small release groups often increase the variability in the model results and do not contribute to improving the multi-year model. Although sometimes considered an alternative to the flow-efficiency regression, a pooled estimate relies on many in-year marked-group releases which were not possible given the low abundance of spring Chinook at the White River trap in 2014. We believe the most effective method of estimation will be to continue development of the current flow-efficiency model recalculating previous estimates as updates to the model are made.

Historically, the high-water trapping regime at the White River smolt trap has varied in cone size and trap location. Driven mainly by the need to capture more fish at elevated flows, three different position/trap size combinations have been used at elevated flows since 2007. With little success using any of the different high-water positions, we were ultimately left with a fragmented data set requiring different methods of data expansion. In 2013, the decision was made to relinquish the use of high-water trapping techniques and instead use a single cone size and trap location throughout the season. This change was conducive to the use of flow-efficiency regression modeling because it minimized the number of age-specific regressions required. Unencumbered by multiple trapping iterations of varying success, single regressions for yearling and subyearling emigrants will ultimately improve our precision in tracking population trends by eliminating unreliable and sporadically-used methodologies. The White River's spring Chinook egg-to-emigrant rates generally comport well with those of the nearby Chiwawa River and Nason Creek (Figure 7). Although unclear how the changes implemented at the White River will affect estimates into the future, it appears that rates of juvenile in-stream survivals are trending similarly to data from the Chiwawa River smolt trap (trap size and location constant).



**Figure 7. Comparison of wild spring Chinook abundance estimates (BY2007-2012) made at the White R., Nason Cr., and Chiwawa R. smolt traps. Chiwawa R. data provided by Hillman et al. (2014).**

Our goals in 2015 will again include the improvement of our spring Chinook flow-efficiency regressions via the expansions of flows tested and larger mark-group sizes. As previously mentioned, replication of trials will only occur in an attempt to raise the minimum mark-group size by establishing a viable regression based on more substantial releases. In the coming season we will look to set this minimum at  $\geq 50$  chinook juveniles. Although ultimately limited by low Chinook abundance in the White River, our refined methodologies place us in the position to only improve on both future and past estimates.



## 5.0 LITERATURE CITED

- Andonaegui, C. 2001. Salmon, Steelhead, and Bull Trout Habitat Limiting Factors for the Wenatchee Subbasin (Water Resource Inventory Area 45) and Portions of WRIA 40 within Chelan County (Squilchuck, Stemilt and Colockum drainages). Final draft report. WSCC.
- CBFWA (Columbia Basin Fish and Wildlife Authority). 1999. PIT Tag Marking Procedures Manual, Version 2.0. Columbia Basin Fish and Wildlife Authority, Portland OR.
- Everhart, W.H. and W.D. Youngs. 1953. Principles of Fishery Science, second edition. Comstock Publishing Associates, *a division of* Cornell University Press, Ithica and London.
- Hillman, T.W. 2004. Monitoring strategy for the Upper Columbia Basin: Draft report February 1, 2004. *Prepared for* Upper Columbia Regional Technical Team, Wenatchee, Washington.
- Hillman, T.W., M. Miller, T. Miller, M. Tonseth, M. Hughes and A. Murdoch. 2013. Monitoring and Evaluation of the Chelan County PUD Hatchery Programs: 2012 Annual Report. *Prepared for* Habitat Conservation Plan Hatchery Committee. Wenatchee, WA.
- Marshall, A. R., and S. Young. 1994. Genetic Analysis of Upper Columbia Spring and Summer Chinook Salmon for the Rock Island Hatchery Evaluation Program. Final report, Washington Department of Fisheries, Olympia.
- Mullan, J.W., K.R. Williams, G. Rhodus, T.W. Hillman and J.D. McIntyre. 1992. Production and Habitat of Salmonids in Mid Columbia River Tributaries. Monograph 1, USFWS, Leavenworth, Washington.
- Murdoch, A., and K. Petersen. 2000. Freshwater Production and Emigration of Juvenile Spring Chinook from the Chiwawa River in 2000. Washington State Department of Fish and Wildlife.
- Pearsons, T.N., and R.B. Langshaw. 2009. Monitoring and Evaluation Plan for Grant County PUD's Salmon and Steelhead Supplementation Programs. Grant County Public Utility District, Ephrata, WA.
- Snow, C., C. Frady, and A. Repp. 2013. Monitoring and Evaluation of Wells and Methow Hatchery Programs: 2012 Annual Report. *Prepared for* Douglas County Public Utility District and Wells Habitat Conservation Plan Hatchery Committee.
- Tussing, S.P. 2008. A Field Manual of Scientific Protocols for Downstream Migrant Trapping within the Upper Columbia Monitoring Strategy: 2008 Working Version 1.0. Prepared for Bonneville Power Administration's Integrated Status and Effectiveness Monitoring Program.
- Upper Columbia RTT. 2001. A Strategy to Protect and Restore Salmonid Habitat in the Upper Columbia Region, a Discussion Draft Report. Upper Columbia Salmon Recovery Board.
- United States Forest Service. 2004. White River Survey Report.

Washington Department of Ecology (WDOE). 2012. River and Stream Flow Monitoring.  
<https://fortress.wa.gov/ecy/wrx/wrx/flows/station.asp?sta=45J0>

## APPENDIX A: White River Temperature and Discharge Data

<b>Date</b>	<b>Stream Discharge (CFS)</b>	<b>Water Temperature (°C)</b>			
			4/6/2014	600	5.0
			4/7/2014	672	5.0
			4/8/2014	894	4.5
3/1/2014	152	1.6	4/9/2014	1240	3.8
3/2/2014		0.0	4/10/2014	1130	4.2
3/3/2014		0.1	4/11/2014	1070	4.4
3/4/2014	181	0.2	4/12/2014	1040	4.6
3/5/2014	235	0.9	4/13/2014	1040	4.4
3/6/2014	393	0.9	4/14/2014	1040	4.4
3/7/2014	483	1.1	4/15/2014	1060	4.8
3/8/2014	440	1.2	4/16/2014	1010	4.6
3/9/2014	973	0.7	4/17/2014	1020	4.7
3/10/2014	1250	1.0	4/18/2014	1090	4.6
3/11/2014	992	1.8	4/19/2014	1020	4.1
3/12/2014	876	2.4	4/20/2014	993	4.7
3/13/2014	824	2.9	4/21/2014	943	4.8
3/14/2014	853	3.0	4/22/2014	983	5.2
3/15/2014	846	3.1	4/23/2014	970	5.0
3/16/2014	1170	2.7	4/24/2014	1070	4.5
3/17/2014	1160	2.9	4/25/2014	1040	5.0
3/18/2014	1010	3.1	4/26/2014	994	4.8
3/19/2014	922	3.4	4/27/2014	937	4.9
3/20/2014	832	3.1	4/28/2014	881	5.3
3/21/2014	741	3.0	4/29/2014	870	5.7
3/22/2014	676	3.2	4/30/2014	950	5.9
3/23/2014	628	3.9	5/1/2014	1260	5.7
3/24/2014	602	4.1	5/2/2014	1920	4.8
3/25/2014	588	3.8	5/3/2014	2300	4.4
3/26/2014	574	4.1	5/4/2014	2080	4.5
3/27/2014	558	4.4	5/5/2014	1890	5.0
3/28/2014	561	3.5	5/6/2014	1770	5.3
3/29/2014	555	3.7	5/7/2014	1720	5.2
3/30/2014	559	4.2	5/8/2014	1700	4.8
3/31/2014	541	4.2	5/9/2014	1720	5.2
4/1/2014	533	4.4	5/10/2014	1600	5.0
4/2/2014	532	4.4	5/11/2014	1480	5.5
4/3/2014	538	4.5	5/12/2014	1530	5.8
4/4/2014	544	4.8	5/13/2014	1680	5.9
4/5/2014	553	4.4			

5/14/2014	2260	5.7	6/26/2014	2240	7.6
5/15/2014	3160	5.1	6/27/2014	2210	7.2
5/16/2014	3900	5.2	6/28/2014	2260	7.2
5/17/2014	3850	5.0	6/29/2014	2130	6.8
5/18/2014	3010	5.2	6/30/2014	1780	7.7
5/19/2014	2650	5.8	7/1/2014	2030	8.7
5/20/2014	2620	5.9	7/2/2014	2410	8.3
5/21/2014	2740	6.0	7/3/2014	2390	8.6
5/22/2014	3120	6.1	7/4/2014	2020	8.4
5/23/2014	3730	5.4	7/5/2014	1810	8.6
5/24/2014	3750	5.7	7/6/2014	1960	9.3
5/25/2014	3140	5.0	7/7/2014	2190	9.4
5/26/2014	2570	5.7	7/8/2014	2160	9.7
5/27/2014	2290	5.8	7/9/2014	2190	9.9
5/28/2014	2080	5.7	7/10/2014	1930	9.5
5/29/2014	1860	6.2	7/11/2014	1740	9.9
5/30/2014	1800	6.1	7/12/2014	1830	10.1
5/31/2014	1960	6.8	7/13/2014	1930	10.5
6/1/2014	2430	6.7	7/14/2014	2060	11.1
6/2/2014	2890	6.4	7/15/2014	2090	11.3
6/3/2014	3190	6.5	7/16/2014	1830	11.1
6/4/2014	3200	6.3	7/17/2014	1630	11.3
6/5/2014	2810	6.3	7/18/2014	1400	10.9
6/6/2014	2490	6.4	7/19/2014	1560	10.8
6/7/2014	2380	6.9	7/20/2014	1350	10.4
6/8/2014	2600	6.9	7/21/2014	983	10.3
6/9/2014	2920	6.9	7/22/2014	856	9.9
6/10/2014	2590	5.9	7/23/2014	977	9.8
6/11/2014	2020	6.7	7/24/2014	1050	8.5
6/12/2014	2210	7.1	7/25/2014	799	9.6
6/13/2014	2540	5.8	7/26/2014	727	11.0
6/14/2014	2160	6.8	7/27/2014	693	11.4
6/15/2014	2020	6.6	7/28/2014	736	12.6
6/16/2014	1820	6.2	7/29/2014	771	12.9
6/17/2014	1620	7.2	7/30/2014	735	13.0
6/18/2014	1850	7.6	7/31/2014	683	12.9
6/19/2014	2040	7.5	8/1/2014	640	12.6
6/20/2014	2150	7.2	8/2/2014	698	12.0
6/21/2014	1810	6.5	8/3/2014	643	12.6
6/22/2014	1760	7.7	8/4/2014	628	13.3
6/23/2014	2200	8.0	8/5/2014	573	13.2
6/24/2014	2620	7.6	8/6/2014	552	13.4
6/25/2014	2280	7.6	8/7/2014	516	13.3

8/8/2014	469	13.3	9/20/2014	225	11.5
8/9/2014	433	12.7	9/21/2014	205	11.2
8/10/2014	397	12.6	9/22/2014	220	11.5
8/11/2014	398	13.0	9/23/2014	212	11.1
8/12/2014	436	13.2	9/24/2014	401	10.4
8/13/2014	852	13.5	9/25/2014	412	10.0
8/14/2014	723	12.0	9/26/2014	362	9.6
8/15/2014	594	11.7	9/27/2014	298	9.6
8/16/2014	460	12.6	9/28/2014	241	9.9
8/17/2014	444	13.3	9/29/2014	215	10.0
8/18/2014	438	13.8	9/30/2014	281	9.8
8/19/2014	448	14.1	10/1/2014	263	9.1
8/20/2014	427	13.7	10/2/2014	203	9.0
8/21/2014	372	12.8	10/3/2014	176	8.4
8/22/2014	330	11.6	10/4/2014	174	8.7
8/23/2014	308	11.7	10/5/2014	197	9.8
8/24/2014	300	12.4	10/6/2014	211	10.2
8/25/2014	289	11.7	10/7/2014	233	10.7
8/26/2014	274	12.6	10/8/2014	206	10.6
8/27/2014	307	13.6	10/9/2014	174	9.7
8/28/2014	325	13.4	10/10/2014	161	9.3
8/29/2014	295	12.5	10/11/2014	224	9.7
8/30/2014	282	11.7	10/12/2014	281	8.7
8/31/2014	304	11.3	10/13/2014	203	9.1
9/1/2014	248	10.8	10/14/2014	308	9.1
9/2/2014	230	11.1	10/15/2014	301	8.7
9/3/2014	343	10.4	10/16/2014	357	8.1
9/4/2014	237	10.3	10/17/2014	287	7.9
9/5/2014	203	10.8	10/18/2014	772	8.0
9/6/2014	198	11.2	10/19/2014	509	8.9
9/7/2014	197	11.4	10/20/2014	503	8.7
9/8/2014	195	11.7	10/21/2014	534	8.4
9/9/2014	195	11.6	10/22/2014		
9/10/2014	193	11.5	10/23/2014	1290	6.8
9/11/2014	175	10.3	10/24/2014	846	6.1
9/12/2014	156	9.0	10/25/2014	811	6.5
9/13/2014	148	9.1	10/26/2014	1570	6.1
9/14/2014	146	9.5	10/27/2014	946	5.5
9/15/2014	148	10.0	10/28/2014	848	5.0
9/16/2014	155	10.7	10/29/2014	1420	5.1
9/17/2014	172	11.6	10/30/2014	1470	5.7
9/18/2014	190	12.1	10/31/2014	2420	6.1
9/19/2014	223	11.9	11/1/2014	1600	5.3

11/2/2014	1160	5.2
11/3/2014		
11/4/2014	2150	6.3
11/5/2014	1650	6.0
11/6/2014	2830	6.3
11/7/2014	2180	5.5
11/8/2014	1580	4.7
11/9/2014	1380	5.4
11/10/2014	1120	4.2
11/11/2014	877	2.8
11/12/2014	712	0.9
11/13/2014	638	0.7
11/14/2014	571	0.5
11/15/2014	516	0.4
11/16/2014	467	0.5
11/17/2014	431	0.5
11/18/2014	403	0.8
11/19/2014	378	1.6
11/20/2014	357	2.6
11/21/2014	356	2.0
11/22/2014	356	1.1
11/23/2014	353	1.8
11/24/2014	333	2.3
11/25/2014	753	2.2
11/26/2014	1790	1.8
11/27/2014	3120	2.6
11/28/2014	5750	2.7
11/29/2014	3580	2.7
11/30/2014	2120	1.5

## APPENDIX B: Daily Trap Operation Status

<b>Date</b>	<b>Trap Status</b>	<b>Comments</b>		
			4/8/2014	Op.
3/1/2014	No Op.	Pulled - ice	4/9/2014	Op.
3/2/2014	No Op.	Pulled - ice	4/10/2014	Op.
3/3/2014	Op.		4/11/2014	Op.
3/4/2014	No Op.	Stopped - ice	4/12/2014	Op.
3/5/2014	Op.		4/13/2014	Op.
3/6/2014	No Op.	Stopped - woody debris	4/14/2014	Op.
3/7/2014	Op.		4/15/2014	Op.
3/8/2014	Op.		4/16/2014	Op.
3/9/2014	Op.		4/17/2014	Op.
3/10/2014	No Op.	Stopped - woody debris	4/18/2014	Op.
3/11/2014	Op.		4/19/2014	Op.
3/12/2014	Op.		4/20/2014	Op.
3/13/2014	Op.		4/21/2014	Op.
3/14/2014	Op.		4/22/2014	Op.
3/15/2014	Op.		4/23/2014	Op.
3/16/2014	Op.		4/24/2014	Op.
3/17/2014	Op.		4/25/2014	Op.
3/18/2014	Op.		4/26/2014	Op.
3/19/2014	Op.		4/27/2014	Op.
3/20/2014	Op.		4/28/2014	Op.
3/21/2014	Op.		4/29/2014	Op.
3/22/2014	Op.		4/30/2014	Op.
3/23/2014	Op.		5/1/2014	Op.
3/24/2014	Op.		5/2/2014	Op.
3/25/2014	Op.		5/3/2014	Op.
3/26/2014	Op.		5/4/2014	Op.
3/27/2014	Op.		5/5/2014	Op.
3/28/2014	Op.		5/6/2014	Op.
3/29/2014	Op.		5/7/2014	Op.
3/30/2014	Op.		5/8/2014	Op.
3/31/2014	Op.		5/9/2014	Op.
4/1/2014	Op.		5/10/2014	Op.
4/2/2014	Op.		5/11/2014	Op.
4/3/2014	Op.		5/12/2014	Op.
4/4/2014	Op.		5/13/2014	Op.
4/5/2014	Op.		5/14/2014	Op.
4/6/2014	Op.		5/15/2014	Op.
4/7/2014	Op.		5/16/2014	Op.
			5/17/2014	Op.

5/18/2014	Op.		6/30/2014	Op.	
5/19/2014	Op.		7/1/2014	Op.	
5/20/2014	Op.		7/2/2014	Op.	
5/21/2014	Op.		7/3/2014	Op.	
5/22/2014	Op.		7/4/2014	Op.	
5/23/2014	No Op.	Stopped - woody debris	7/5/2014	Op.	
5/24/2014	Op.		7/6/2014	Op.	
5/25/2014	Op.		7/7/2014	Op.	
5/26/2014	Op.		7/8/2014	Op.	
5/27/2014	Op.		7/9/2014	Op.	
5/28/2014	No Op.	Stopped - woody debris	7/10/2014	Op.	
5/29/2014	Op.		7/11/2014	Op.	
5/30/2014	Op.		7/12/2014	Op.	
5/31/2014	Op.		7/13/2014	Op.	
6/1/2014	Op.		7/14/2014	Op.	
6/2/2014	Op.		7/15/2014	Op.	
6/3/2014	Op.		7/16/2014	Op.	
6/4/2014	Op.		7/17/2014	Op.	
6/5/2014	Op.		7/18/2014	No Op.	Pulled due to fire activity
6/6/2014	Op.		7/19/2014	Op.	
6/7/2014	Op.		7/20/2014	Op.	
6/8/2014	Op.		7/21/2014	Op.	
6/9/2014	Op.		7/22/2014	Op.	
6/10/2014	Op.		7/23/2014	Op.	
6/11/2014	Op.		7/24/2014	Op.	
6/12/2014	Op.		7/25/2014	Op.	
6/13/2014	Op.		7/26/2014	Op.	
6/14/2014	Op.		7/27/2014	Op.	
6/15/2014	Op.		7/28/2014	Op.	
6/16/2014	Op.		7/29/2014	Op.	
6/17/2014	Op.		7/30/2014	No Op.	Stopped - woody debris
6/18/2014	Op.		7/31/2014	Op.	
6/19/2014	Op.		8/1/2014	Op.	
6/20/2014	Op.		8/2/2014	Op.	
6/21/2014	Op.		8/3/2014	Op.	
6/22/2014	Op.		8/4/2014	Op.	
6/23/2014	Op.		8/5/2014	Op.	
6/24/2014	Op.		8/6/2014	Op.	
6/25/2014	Op.		8/7/2014	Op.	
6/26/2014	Op.		8/8/2014	Op.	
6/27/2014	Op.		8/9/2014	Op.	
6/28/2014	Op.		8/10/2014	Op.	
6/29/2014	Op.				



8/11/2014	Op.		9/23/2014	Op.	
8/12/2014	Op.		9/24/2014	Op.	
8/13/2014	Op.		9/25/2014	Op.	
8/14/2014	Op.		9/26/2014	Op.	
8/15/2014	Op.		9/27/2014	Op.	
8/16/2014	Op.		9/28/2014	Op.	
8/17/2014	Op.		9/29/2014	Op.	
8/18/2014	Op.		9/30/2014	Op.	
8/19/2014	Op.		10/1/2014	Op.	
8/20/2014	Op.		10/2/2014	Op.	
8/21/2014	Op.		10/3/2014	Op.	
8/22/2014	Op.		10/4/2014	Op.	
8/23/2014	Op.		10/5/2014	Op.	
8/24/2014	Op.		10/6/2014	Op.	
8/25/2014	Op.		10/7/2014	Op.	
8/26/2014	Op.		10/8/2014	Op.	
8/27/2014	Op.		10/9/2014	Op.	
8/28/2014	Op.		10/10/2014	Op.	
8/29/2014	Op.		10/11/2014	Op.	
8/30/2014	No Op.	Stopped - woody debris	10/12/2014	Op.	
8/31/2014	Op.		10/13/2014	Op.	
9/1/2014	Op.		10/14/2014	Op.	
9/2/2014	Op.		10/15/2014	Op.	
9/3/2014	Op.		10/16/2014	Op.	
9/4/2014	Op.		10/17/2014	Op.	
9/5/2014	Op.		10/18/2014	Op.	
9/6/2014	Op.		10/19/2014	Op.	
9/7/2014	Op.		10/20/2014	Op.	
9/8/2014	Op.		10/21/2014	Op.	
9/9/2014	Op.		10/22/2014	Op.	
9/10/2014	Op.		10/23/2014	Op.	
9/11/2014	Op.		10/24/2014	Op.	
9/12/2014	Op.		10/25/2014	Op.	
9/13/2014	Op.		10/26/2014	No Op.	Stopped - woody debris
9/14/2014	Op.		10/27/2014	Op.	
9/15/2014	Op.		10/28/2014	Op.	
9/16/2014	Op.		10/29/2014	Op.	
9/17/2014	Op.		10/30/2014	No Op.	Stopped - woody debris
9/18/2014	Op.		10/31/2014	No Op.	Stopped - woody debris
9/19/2014	Op.		11/1/2014		
9/20/2014	Op.		11/2/2014		
9/21/2014	Op.		11/3/2014		
9/22/2014	Op.		11/4/2014		

11/5/2014		
11/6/2014		
11/7/2014		
11/8/2014		
11/9/2014		
11/10/2014		
11/11/2014		
11/12/2014		
11/13/2014		
11/14/2014		
11/15/2014	No Op.	Stopped - ice
11/16/2014	No Op.	Pulled - ice
11/17/2014	No Op.	Pulled - ice
11/18/2014		
11/19/2014		
11/20/2014		
11/21/2014		
11/22/2014		
11/23/2014		
11/24/2014		
11/25/2014		
11/26/2014		
11/27/2014	No Op.	Stopped - woody debris
11/28/2014	No Op.	Stopped - woody debris
11/29/2014	No Op.	Pulled - flooding
11/30/2014	No Op.	Pulled - flooding

## APPENDIX C: Regression Models

Model: Chinook Yearlings (Spring '08-'14) Back Position, ( $r^2=0.569$ ;  $p = 0.001$ )

Origin/Species/Stage	Date	Marked	Recaptured	Trap Efficiency	ASIN Transform	Discharge (cfs)
Wild Chinook Yearlings	4/10/2008	25	2	0.120	0.354	229
Wild Chinook Yearlings	3/26/2009	24	5	0.250	0.524	191
Wild Chinook Yearlings	3/30/2009	34	4	0.147	0.394	193
Wild Chinook Yearlings	4/2/2009	37	10	0.297	0.577	206
Wild Chinook Yearlings	4/5/2009	59	15	0.271	0.548	205
Wild Chinook Yearlings	4/10/2009	36	3	0.111	0.340	385
Wild Chinook Yearlings	3/12/2010	25	1	0.080	0.287	300
Wild Chinook Yearlings	3/16/2010	30	5	0.200	0.464	278
Wild Chinook Yearlings	3/20/2010	21	1	0.095	0.314	283
Wild Chinook Yearlings	4/5/2010	37	1	0.054	0.235	340
Wild Chinook Yearlings	4/9/2010	31	4	0.161	0.413	310
Wild Chinook Yearlings	4/12/2010	58	4	0.086	0.298	288
Wild Chinook Yearlings	4/16/2010	73	2	0.041	0.204	381
Wild Chinook Yearlings	4/14/2012	48	1	0.042	0.206	527

Model: Chinook Subyearlings (Fall '09-'14) Back Position, ( $r^2=0.147$ ;  $p = 0.079$ )

Origin/Species/Stage	Date	Marked	Recaptured	Trap Efficiency	ASIN Transform	Discharge (cfs)
Wild Chinook Subyearlings	8/20/2009	20	2	15.00%	0.398	311
Wild Chinook Subyearlings	8/29/2009	34	4	14.71%	0.394	227
Wild Chinook Subyearlings	10/7/2009	22	2	13.64%	0.378	95
Wild Chinook Subyearlings	10/16/2009	34	6	20.59%	0.471	134
Wild Chinook Subyearlings	11/17/2009	35	3	11.43%	0.345	375
Wild Chinook Subyearlings	11/23/2009	21	0	4.76%	0.220	313
Wild Chinook Subyearlings	11/21/2011	39	2	7.69%	0.281	172
Wild Chinook Subyearlings	10/4/2012	33	5	18.18%	0.441	140
Wild Chinook Subyearlings	10/24/2012	87	6	8.05%	0.288	268
Wild Chinook Subyearlings	10/28/2012	36	1	5.56%	0.238	711
Wild Chinook Subyearlings	10/31/2013	46	7	17.39%	0.430	258
Wild Chinook Subyearlings	11/6/2013	38	9	26.32%	0.539	248
Wild Chinook Subyearlings	11/9/2013	40	6	17.50%	0.432	251
Wild Chinook Subyearlings	11/13/2013	29	2	10.34%	0.327	422
Wild Chinook Subyearlings	11/23/2013	25	3	16.00%	0.412	406
Wild Chinook Subyearlings	11/27/2013	24	0	4.17%	0.206	335

## Appendix D. Historical Morphometric Data

### Spring Chinook (2007-2014)

Trap Year	Brood Year	Origin/Species/Stage	Fork Length (mm)			Weight (g)			K-factor
			Mean	n	SD	Mean	n	SD	
2007	2005	Wild Yearling Smolt	93.4	173	8.5	8.6	173	2.2	1.1
2007	2005	Wild Yearling Precocial Parr	122.5	4	7.2	22.2	4	5.8	1.2
2007	2005	Hatchery Yearling Smolt <sup>a</sup>	76.0	208	17.9	5.4	203	4.2	1.2
2007	2005	Hatchery Yearling Precocial Parr	97.8	20	8.7	11.1	19	2.2	1.2
2007	2006	Wild Subyearling Fry	35.4	7	1.6	—	—	—	—
2007	2006	Wild Subyearling Parr	95.3	33	12.4	9.8	33	4.1	1.1
2008	2006	Wild Yearling Smolt	100.2	105	12.3	12.5	105	13.5	1.2
2008	2006	Wild Yearling Precocial Parr	125.9	9	8.4	22.8	9	4.1	1.1
2008	2006	Hatchery Yearling Smolt	116.5	229	12.7	18.7	228	9.8	1.2
2008	2006	Hatchery Yearling Precocial Parr	155.0	2	15.6	47.6	2	12.6	1.3
2008	2007	Wild Subyearling Fry	41.4	10	4.4	—	—	—	—
2008	2007	Wild Subyearling Parr	94.6	202	9.1	9.4	202	2.5	1.1
2009	2007	Wild Yearling Smolt	103.9	275	6.4	12.5	274	2.6	1.1
2009	2007	Wild Yearling Precocial Parr	134.0	5	7.0	28.5	2	2.7	1.2
2009	2007	Hatchery Yearling Precocial Parr	187.5	2	17.7	81.9	2	27.1	1.2
2009	2008	Wild Subyearling Fry	37.5	13	2.1	—	—	—	—
2009	2008	Wild Subyearling Parr	85.0	507	11.8	7.2	499	2.7	1.2
2010	2008	Wild Yearling Smolt	95.5	345	7.1	11.2	345	2.4	1.3
2010	2008	Wild Yearling Precocial Parr	129.7	15	10.3	26.4	15	6.6	1.2
2010	2009	Wild Subyearling Fry	40.1	31	3.6	—	—	—	—
2010	2009	Wild Subyearling Parr	87.0	166	12.6	7.7	166	3.0	1.2
2011	2009	Wild Yearling Smolt	99.0	64	7.7	11.3	64	2.8	1.2
2011	2009	Wild Yearling Precocial Parr	137.0	1	—	32.3	1	—	1.3
2011	2009	Hatchery Yearling Smolt	127.3	46	10.6	24.3	46	6.5	1.2
2011	2010	Wild Subyearling Fry	36.7	26	2.5	—	—	—	—
2011	2010	Wild Subyearling Parr	90.6	159	13.0	9.2	159	7.1	1.2
2012	2010	Wild Yearling Smolt	97.9	182	7.9	10.9	179	2.8	1.2
2012	2010	Wild Yearling Precocial Parr	122.8	13	12.7	22.4	13	6.5	1.2
2012	2011	Hatchery Subyearling Fry <sup>b</sup>	83.7	29	4.4	6.5	2	2.3	1.1
2012	2011	Hatchery Subyearling Parr	109.6	25	7.4	14.6	25	3.3	1.1
2012	2011	Wild Subyearling Fry	35.2	18	2.7	—	—	—	—
2012	2011	Wild Subyearling Parr	91.0	315	10.1	8.8	288	2.8	1.2
2013	2011	Wild Yearling Smolt	102.8	20	7.0	12.3	20	3.0	1.1
2013	2011	Wild Yearling Precocial Parr	110.5	2	0.7	13.5	2	3.0	1.0
2013	2011	Hatchery Yearling Precocial Parr	155.0	4	17.4	43.4	4	17.8	1.2
2013	2012	Wild Subyearling Fry	40.3	77	8.1	—	—	—	—
2013	2012	Wild Subyearling Parr	83.8	445	12.3	6.7	444	4.7	1.1

2014	2012	Wild Yearling Smolt	94.3	43	7.0	9.4	43	2.2	1.1
2014	2012	Wild Yearling Precocial Parr	126.9	7	13.0	23.2	7	7.4	1.1
2014	2013	Wild Subyearling Fry	40.0	22	3.8	—	—	—	—
2014	2013	Wild Subyearling Parr	86.0	185	14.1	7.5	185	3.3	1.2

<sup>a</sup> Includes residualized non-precocial smolts caught after June 30

<sup>b</sup> “Fry” classification based on age despite  $FL \leq 50\text{mm}$



# Appendix M

## **Genetic Diversity of Upper Columbia River Summer Chinook Salmon**





Genetic Structure of upper Columbia River Summer Chinook and  
Evaluation of the Effects of Supplementation Programs

by

Todd W. Kassler and Scott Blankenship  
Washington Department of Fish and Wildlife  
Molecular Genetics Laboratory  
600 Capitol Way N  
Olympia, WA 98501

and

Andrew R. Murdoch  
Washington Department of Fish and Wildlife  
Hatchery/Wild Interactions  
3515 State Highway 97A  
Wenatchee, WA 98801

February 2011

## Abstract

We investigated genetic relationships among temporally replicated collections of summer Chinook from the Wenatchee River, Methow River, and Okanogan River in the upper Columbia River basin. Samples from the Eastbank Hatchery – Wenatchee stock, Eastbank Hatchery – MEOK stock, and Wells Hatchery were also included in the analysis. Samples of natural- and hatchery-origin summer Chinook were analyzed and compared to determine if the supplementation program has had any impacts to the genetic structure of these populations. We also calculated the effective number of breeders for collection locations of natural- and hatchery-origin summer Chinook from 1993 and 2008. In general, population differentiation was not observed among the temporally replicated collection locations. A single collection from the Okanogan River (1993) was the only collection showing statistically significant differences. The effective number of breeders was not statistically different from the early collection in 1993 in comparison to the late collection in 2008. Overall, these analyses revealed a lack of differentiation among the temporal replicates from the same locations and among the collection from different locations, suggesting the populations have been homogenized or that there has been substantial gene flow among populations. Additional comparisons among summer-run and fall-run Chinook populations in the upper Columbia River were conducted to determine if there was any differentiation between Chinook with different run timing. These analyses revealed pairwise  $F_{ST}$  values that were less than 0.01 for the collections of summer Chinook to collections of fall Chinook from Hanford Reach, lower Yakima River, Priest Rapids, and Umatilla. Collections of fall Chinook from Crab Creek, Lyons Ferry Hatchery, Marion Drain, and Snake River had pairwise  $F_{ST}$  values that were higher in comparison to the collections of summer Chinook. The consensus clustering analysis did not provide good statistical support to the groupings, but did show relationships among collections based on geographic proximity. Overall the summer and fall run Chinook that have historically been

spawned together were not differentiated while fall Chinook from greater geographic distances were differentiated.

## **Introduction**

The National Marine Fisheries Service (NMFS) recognizes 15 Evolutionary Significant Units (ESU) for Chinook salmon (*Oncorhynchus tshawytscha*) (Myers et al. 1998). The summer Chinook from the upper Columbia River are included in the Upper Columbia River Summer- and Fall-Run ESU, which encompasses all late-run (summer and fall), ocean-type Chinook salmon from the mainstem Columbia River and its tributaries (excluding the Snake River) between Chief Joseph and McNary Dams (Waknitz et al. 1995). Waknitz et al. (1995) concluded that due to high total abundance this ESU was not likely to become at risk from extinction. Yet, a majority of natural spawning activity was in the vicinity of Hanford Reach, and it was unclear whether natural production was self-sustaining given the vast summer Chinook artificial propagation efforts (Waknitz et al. 1995). Additionally, the Biological Review Team expressed concern about potential consequences to genetic and life-history traits from an increasing contribution of hatchery fish to total spawning escapement (Waknitz et al. 1995).

Artificial propagation of ocean-type Chinook from the middle/upper Columbia has been continuous since the implementation of the Grand Coulee Fish Maintenance Project (GCFMP) in 1939 (Myers et al. 1998). The US Fish and Wildlife Service established three hatchery programs for summer/fall Chinook during the GCFMP, Leavenworth NFH, Entiat NFH, and Winthrop NFH. The Washington Department of Fisheries (now Washington Department of Fish and Wildlife) followed with hatchery programs at Rocky Reach (1964), Wells Dam (1967), Priest Rapids (1974), and Eastbank (1990) facilities. Currently, only Leavenworth NFH and Winthrop NFH are not producing summer/fall Chinook. Entiat NFH has resumed production of summer/fall Chinook (Wells FH Stock) in 2009 and released their first yearling summer Chinook smolts in 2010. Since

1941, over 200 million ocean-type Chinook salmon have been released into the middle Columbia River Basin (Myers et al. 1998). Initially, the hatchery programs differentiated between early returning fish (i.e., stream-type) and later returning fish (i.e., ocean-type), but no distinction was made regarding the “summer” and “fall” components of the ocean-type stocks (Waknitz et al. 1995). Therefore, all Chinook salmon now migrating above Rock Island Dam descend from not only a mixture between different stocks from the basin, but also a mixture between the endemic summer and fall life histories. While hatchery protocols have been modified of late to maintain discreet summer and fall Chinook hatchery stocks (Utter et al. 1995; see also HGMP), physical evidence and genetic data suggests that summer and fall Chinook may have become homogenized. During the 1970’s and 80’s, given coded-wire tag recoveries, summer-run Chinook originating from above Rock Island Dam were believed to have spawned extensively with Hanford Reach and Priest Rapids Hatchery fish (Chapman 1994). Stuehrenberg et al. (1995) reported that 10% of their radio tagged summer Chinook were occupying typical fall-run spawning habitat on the mainstem Columbia river, and 25% of fall fish released from Priest Rapids were recovered as summers at (or above) Wells Hatchery. Genetic data reported by Marshall et al. (1995) and Waknitz et al. (1995) corroborate these observations, as genetic distances observed between summer and fall Chinook within the Upper Columbia River Summer- and Fall-Run ESU were essentially zero.

In response to the need for evaluation of the supplementation hatchery programs, both a monitoring and evaluation plan (DCPUD 2005; Murdoch and Peven 2005) and the associated analytical framework (Hays et al. 2006) were developed for the Habitat Conservation Plan’s Hatchery Committee through the joint effort of the fishery co-managers (CCT, NMFS, USFWS, WDFW, and YN) and Chelan County and Douglas County PUDs. These reports outline 10 objectives to be applied to various species assessing the impacts of hatchery operations mitigating the operation of Wells, Rocky Reach, and Rock Island hydroelectric projects. The present monitoring and evaluation study plan differs

in scope from previous monitoring and evaluation projects proposed by WDFW Molecular Genetics Lab, in that it does not investigate a single watershed, but instead will encompass all summer Chinook stocks from the upper Columbia River including the three supplementation (Wenatchee, Methow, and Okanogan) and the harvest augmentation program (Wells summer Chinook). The objectives of this study were to determine if genetic diversity, population structure, and effective population size have changed in natural spawning populations as a result of the hatchery programs.

## ***Materials and Methods***

### **Collections**

A total of 2,416 summer Chinook were collected from tributaries in the upper Columbia River basin and were analyzed (Table 1). Two collections of natural-origin summer Chinook from 1993 (prior to the supplementation program) were taken from the Wenatchee River Basin and were compared to collections of hatchery and natural-origin from 2006 and 2008 that were post-supplementation. Two pre-supplementation collections from the Methow River (1991 and 1993) were compared to post-supplementation collections from 2006 and 2008. Three pre-supplementation collections from the Okanogan River Basin (1991, 1992, and 1993) were compared with post-supplementation collections from 2006 and 2008. A collection of natural-origin summer Chinook from the Chelan River was also analyzed. Additionally, hatchery collections from Eastbank Hatchery (Wenatchee and MEOK stock) and Wells Hatchery were analyzed and compared to the in-river collections. Summer Chinook data (provided by the USFWS) from the Entiat River was also used for comparison. Lastly, data from eight collections of fall Chinook was compared to the collections of summer Chinook.

## **Laboratory Analyses**

All laboratory analyses were conducted at the WDFW Genetics Laboratory in Olympia, Washington. Genomic DNA was extracted by digesting a small piece of fin tissue using the nucleospin tissue kits obtained from Macherey-Nagel following the recommended conditions in the user manual. Extracted DNA was eluted with a final volume of 100  $\mu$ L.

Genotype information was generated using thirteen microsatellite markers following standard laboratory protocols and analysis methods. Descriptions of the loci assessed in this study and polymerase chain reaction (PCR) conditions are given in Table 2. PCR reactions were run with a thermal profile consisting of: denaturation at 95°C for 3 min, denaturation at 95°C for 15 sec, anneal for 30 sec at the appropriate temperature for each locus (Table 2), extension at 72°C for 1 min, repeat cycle (steps 2-4), final extension at 72°C for 30 minutes. PCR products were then processed with an ABI-3730 DNA Analyzer. Genotypes were visualized with a known size standard (GS500LIZ 3730) using GENEMAPPER 3.7 software. Alleles were binned in GENEMAPPER using the standardized allele sizes established for the Chinook GAPS dataset (Seeb et al. 2007).

## **Within-collection Statistical Analyses**

Allele frequencies were calculated with CONVERT (version 1.3, Glaubitz 2003). Hardy-Weinberg proportions for all loci within each collection were calculated using GENEPOP (version 3.4, Raymond and Rousset 1995). Heterozygosity (observed and expected) was computed for each collection group using GDA (Lewis and Zaykin 2001).

Allelic richness and  $F_{IS}$  (Weir and Cockerham 1984) inbreeding coefficient were calculated using FSTAT (version 2.9.3.2, Goudet 2001). Linkage disequilibrium for each pair of loci in each collection was calculated using GENEPOP v 3.4 (10,000 dememorizations, 100 batches, and 5,000 iterations per batch).

Pairwise estimates of genetic differentiation between collection groups were

calculated using GENEPOP (version 3.4, Raymond and Rousset 1995). Statistical significance for the tests of Hardy-Weinberg proportions, linkage disequilibrium, and genotypic differentiation was evaluated using a Bonferroni correction of p-values to account for multiple, simultaneous tests (Rice 1989).

### **Between-collection Statistical Analyses**

Pairwise  $F_{ST}$  estimates were computed to examine population structure among collections using GENETIX (version 4.03, Belkhir et al. 2001). This estimate uses allelic frequency data and departures from expected heterozygosity to assess differences between pairs of populations.

We used PHYLIP (version 3.5c, Felsenstein 1993) to calculate Cavalli-Sforza and Edwards (1967) pairwise chord distances between collections. Bootstrap calculations were performed using SEQBOOT followed by calculations of genetic distance using GENDIST. The NEIGHBOR-JOINING method of Saitou and Nei (1987) was used to generate the dendrograms and CONSENSE to generate a final consensus tree from the 1,000 replicates. The dendrogram generated in PHYLIP was plotted as an unrooted radial tree using TREEVIEW (version 1.6.6, Page 1996).

### **Effective Number of Breeders**

The effective number of breeders ( $N_b$ ) was estimated for pre- and post-supplementation program collections (where possible) to investigate whether hatchery programs had affected that genetic metric over the operational period. Wang (2009) derived an equation for effective size ( $N_e$ ) as a function of the frequency of nested full-sib and half-sib families in a random collection of individuals.

$$\frac{1}{N_e} = \frac{1+3\alpha}{4} (Q_1 + Q_2 + 2Q_3) - \frac{\alpha}{2} \left( \frac{1}{N_1} + \frac{1}{N_2} \right) \quad (\text{equation 10})$$

Where  $\alpha$  is a measure of the deviation of genotype frequencies from Hardy-Weinberg expectation (equivalent to Wright's (1969)  $F_{IS}$ ),  $Q_i$  are the probabilities that a pair of offspring are paternal half sibs, maternal half sibs, or full sibs, respectively, and  $N_1$  and  $N_2$  are the number of male and female parents that generation, respectively. Genetic parameters (i.e., sibship distributions) were estimated for summer Chinook collections using algorithms implemented in COLONY (Jones and Wang 2009). To be clear, Wang's (2009) method as implemented here will estimate  $N_b$ , given multi-locus genotypes from each collection were partitioned by brood year for this analysis. To obtain an estimate of  $N_e$  each  $N_b$  value must be multiplied by the mean generation time of that population.

## Results

### Collections

A total of 2,350 individuals from 32 collections of temporally replicated samples (six locations) were analyzed (Table 1). Temporally replicated collections of hatchery and natural-origin samples were from the Wenatchee, Methow, and Okanogan Rivers. Temporally replicated hatchery-origin summer Chinook were from Wells Hatchery, Eastbank Hatchery - Wenatchee stock, and Eastbank Hatchery - Methow/Okanogan (MEOK) stock. A total of 232 of those individuals were excluded from any analyses because they failed to amplify at nine or more loci. Data for remaining 2,118 individuals were analyzed to assess differences between temporally replicated natural- and hatchery-origin summer Chinook for each location and to compare the differences among the different collection locations. Summer Chinook data from the temporally replicated collection locations were then combined and compared to fall Chinook data from the GAPS v.3.0 dataset.

### Statistical Analyses



The population statistics (Hardy-Weinberg equilibrium and  $F_{IS}$ ) calculated for each of the 32 temporally replicated collection locations were consistent with neutral expectations (i.e., no associations among alleles). Three collections did have a single locus that did not meet expectations (Wenatchee hatchery-origin 2006, Wells hatchery 2006, and Okanogan hatchery-origin 2009). Based on these results we suggest the collections represented randomly breeding groups and were not comprised of mixtures of individuals from different genetic source populations.

Population differentiation was assessed for each of the temporally replicated collections from within each location (Table 3). This analysis revealed the only significant difference observed within a collection location pertained to the collection from 1993 Okanogan River natural-origin samples. Because of the significant difference of this collection to the other temporal replicates it was not included in further analyses.

Given the absence of genetic differentiation observed among the temporally replicated collections, the 32 collections from the Wenatchee, Methow, and Okanogan River were combined to form three location-specific collections for analysis. Population differentiation metrics were compared among the composite Wenatchee, Methow, and Okanogan collections and eight other location-specific collections (11 locations total). Comparing all collections, there were a total of 39 significant genic test comparisons out of a total 496 (Table 4). Thirty-eight of the 39 statistically significant pairwise differences pertained to the Okanogan River and 2006 Wells Hatchery collections (Table 4).  $F_{ST}$  results are described further below.

Within-collection genetic metrics were estimated for the 11 location-specific collections of summer Chinook from the upper Columbia River, in addition to eight collections of fall Chinook (Table 1). The population statistics (Hardy-Weinberg equilibrium and  $F_{IS}$ ) calculated for these collections of summer and fall

Chinook were also consistent with neutral expectations. The collection from Lyons Ferry Hatchery had one locus that did not meet expectations and the collections from Crab Creek and Marion Drain both had three loci that did not meet expectations.

The hatchery collections in general had a higher percentage of significantly linked loci; however the observed genetic diversity were similar for the natural and hatchery-origin collections. Analysis of allelic richness was based on 11 individuals per collection, the minimum number of individuals across all collections with complete multilocus genotypes. The largest number of linked loci occurred in the Crab Creek, Entiat River, and Okanogan natural-origin collections. Allelic richness was on average lower in the collections of summer Chinook (10.7) collections in comparison to the collections of fall Chinook (11.0).

Pairwise  $F_{ST}$  (Table 4) estimates revealed low levels of differentiation, where all observed  $F_{ST}$  values between the collections of summer Chinook were lower than 0.0096. There were 15 out of 28 comparisons between collections of summer Chinook that were significantly different from zero and occurred primarily from comparisons of the Okanogan River (hatchery and natural-origin) and Wells Hatchery to all other collections. The collection of Eastbank Hatchery – MEOK stock was differentiated from the Wenatchee River natural-origin and Entiat River collections. The collection from the Chelan River had a small sample size of 23 individuals and only differentiated from the Eastbank Hatchery – MEOK stock.  $F_{ST}$  estimates regarding pairwise comparisons between each of four fall Chinook collection locations (Crab Creek, Lyons Ferry Hatchery, Marion Drain, and Snake River) to all other collections were significantly different from zero (Table 5). Pairwise comparisons for three other fall Chinook collections (Hanford Reach, lower Yakima River, and Umatilla River) to the collections of summer Chinook were significantly different from zero (Table 6). The only fall Chinook collection that was not significantly differentiated from all of the summer Chinook was Priest Rapids.

The relative genetic relationships among the test groups were assessed using the consensus clustering analysis (Figure 1). Statistical support for the dendrogram topology (i.e., tree shape) was low regarding the branching that separated the collections of summer Chinook from the upper Columbia River. The collections of fall Chinook; however were supported with bootstrap support over 76% with the exception of three collections (lower Yakima River, Crab Creek, and Umatilla River). In other words, 760 of the 1000 bootstrap replicates supported the placement of the node separating summer and fall collections. The collection from the Chelan River had bootstrap support of 68%; however the sample size for that collections was small ( $N = 23$ ). Even though the bootstrap support was low among the collections of summer Chinook there was concordance between geography and genetic distance.

Where comparisons were possible between pre- and post-supplementation program collections, the effective number of breeders ( $N_b$ ) estimated to have comprised those collections were slightly lower for contemporary (2008) collections; however in all cases the 95% confidence intervals overlapped between historical and contemporary collections, suggesting statistical equivalency. Regarding Wenatchee River collections, the point estimates of  $N_b$  ranged from 134 (08FU) to 190 (93DD), where all collections had overlapping confidence intervals (Table 7). The upper bound of the 1989 brood year for collection 93DD was very large, suggesting the sample size was insufficient for properly inferring the sibship distribution within the collection. Comparing the Okanogan natural collections 93ED and 08GA, the estimated  $N_b$  were 142 (CI 102 – 203) and 127 (CI 92 – 180), respectively. For the Eastbank Hatchery MEOK stock comparisons, the  $N_b$  estimated for the 93DF collection was 171 (CI 129 – 229), as compared to the 166 (CI 126 – 226) estimated for collection 08MO. In all cases, the estimated  $N_b$  can be converted to effective population size ( $N_e$ ) by multiplying the estimate by the mean generation time.

## Discussion

The collections of summer Chinook populations from the upper Columbia River are of interest because census sizes are reduced below historic levels and are the subject of mitigation and supplementation hatchery programs. Concern over the impacts of hatchery supplementation programs on the genetic integrity of natural-origin populations led to our primary objective, which was to evaluate genetic metrics for temporally replicated collections of summer Chinook in the upper Columbia River pre and post hatchery supplementation. A similar analysis by Kassler and Dean (2010) was conducted on spring Chinook in the Tucannon River to evaluate the effects of a supplementation and captive brood program on natural-origin stocks. Additionally, upper Columbia River spring Chinook supplementation programs (Blankenship et al. 2007; Small et al. 2007), spring and fall Chinook populations in the Yakima Basin (Kassler et al. 2008), and a potentially unique population of fall Chinook in Crab Creek (Small et al. 2010) have been evaluated. In the present analysis of summer Chinook populations, collections of pre- and post- supplementation summer Chinook were collected from the Wenatchee River, Methow River, and Okanogan River Basins and analyzed to determine if the genetic profile has changed as a result of the supplementation program. Analysis was then conducted on the collections of summer run to compare the fall run Chinook collections in the upper Columbia River basin.

Allozyme analyses of these three summer run Chinook stocks in the upper Columbia River have identified that each stock was distinct, with a closer relationship detected between the Wenatchee and Methow Rivers (WDF and WDW 1993, Marshall 2002). Wenatchee summer Chinook are thought to be a mixture of native summer Chinook and Chinook from the Grand Coulee Fish Maintenance Project (GCFMP). The goal of the GCFMP project between 1939 and 1943 was to trap migrating Chinook salmon at Rock Island dam (75 miles below Grand Coulee) and homogenize the populations, which reduced the

genetic uniqueness of the distinct tributary populations present in the upper Columbia River.

We found allele frequencies for individual temporally replicated hatchery- and natural-origin collection locations of adult summer Chinook were not significantly different from that expected of a single underlying population, except for one collection (1993 Okanogan natural-origin; Table 3). This collection was differentiated to the Okanogan collections in 2006 and 2008; however it was not differentiated from the collection in 1992. The Okanogan collection from 1992 was also not differentiated to any other collection; therefore the difference in the collection from Okanogan 1993 was likely not an indication of genetic change from pre supplementation to post supplementation. The collection was however dropped from further analyses so as to not confuse interpretation of results. The lack of allelic differentiation observed among the temporally replicated collections was interpreted as the genetic metrics from each location in the early 1990's did not differ from the samples collected in 2008. Spanning a few generations, allele frequencies are not expected to change for large populations at genetic equilibrium. In contrast, changes in allele frequencies of small populations may occur due to the stochastic sampling of genes from one generation to the next (i.e., genetic drift).

A second round of analyses was conducted to evaluate the genetic relationships of the summer run collections (temporal collections were combined) with data from the Entiat River, Chelan River, and eight collections of fall Chinook. Assessment of the relationship between the summer run collections in comparison to each other provided very little evidence of genetic differentiation between these collections. While population differentiation did show some significant differences between the Okanogan River and Wells Hatchery collections, all of the pairwise  $F_{ST}$  values were below 0.003. Meaning that a very small proportion of the observed genetic variation could be attributed to restrictions in gene flow (i.e., population structure)

The comparison of the hatchery-origin collections revealed a lack of differentiation between the Eastbank Hatchery – Wenatchee stock, Eastbank Hatchery – MEOK stock, and the Wells Hatchery (with exception of the 2006 collection). The genetic similarity or low level of genetic differentiation among these stocks suggests that there has been an integration of natural- and hatchery-origin summer Chinook in the upper Columbia River or a lack of ancestral genetic difference. The difference of the 2006 Wells Hatchery collection to the other collections is most likely a result of sampling effect because of the lack of differentiation among the stocks in the basin. If the 2006 collection had been mixed from different sources of summer Chinook there would not be a detectable level of differentiation as was seen with the 2006 sample.

The analyses to compare summer and fall Chinook collections provided some understanding on the genetic relationships of Chinook with different run timings in the upper Columbia River basin. Historically, the hatchery programs in the upper Columbia River were separated into groups of the early returning fish (i.e., stream-type) and later returning fish (i.e., ocean-type), but the programs did not sort individuals identified as “summer” or “fall” stocks (Waknitz et al. 1995). Now all Chinook salmon that are migrating above Rock Island Dam descend from a mixture of different stocks from the upper Columbia River basin, but also a mixture between the endemic summer and fall life histories.

Small et al. (2010) conducted an analysis on summer run and fall run Chinook in the upper Columbia River and concluded that Crab Creek Chinook in the upper Columbia River were genetically distinct to all other fall and summer run Chinook stocks that were analyzed. They did note a departure from Hardy Weinberg expectation as a result of a null allele at the microsatellite locus *Ogo-4* and a higher linkage disequilibrium value due to the inclusion of family groups in one of their samples. Kassler et al. (2008) found differentiation among spring and fall Chinook populations in the Yakima River.

The tests of pairwise  $F_{ST}$  indicated a very low level of genetic differentiation (less than one percent difference) between collections of summer-run Chinook and fall-run Chinook. The range of pairwise  $F_{ST}$  values for comparisons between the summer run and fall run collections was 0.0016 – 0.0248. The larger values from the range were associated to the collections from Crab Creek, Lyons Ferry Hatchery, and Marion Drain. Studies by Kassler et al. (2008) and Small et al. (2010) have documented differences among the populations of these collections to others within the upper Columbia River basin. The low pairwise  $F_{ST}$  values between Priest Rapids and Hanford Reach collections and the summer run collections were not surprising because summer-run Chinook originating from above Rock Island Dam were believed to have spawned extensively with Hanford Reach and Priest Rapids Hatchery fish during the 1970's and 80's (Chapman 1994). The lack of differentiation among the summer and fall stocks in the Columbia River was also identified by Utter et al. (1995) and the HGMP where they state physical evidence and genetic data suggests that summer and fall Chinook may have become homogenized.

Despite low levels of statistical bootstrap support for dendrogram topology (i.e., tree shape), there was concordance observed between geographic location and the genetic relationships among the summer and fall Chinook populations. The collections from the Okanogan (hatchery and natural-origin) did separate out with collections from Wells Dam Hatchery, Entiat River, and Eastbank Hatchery – MEOK stock, and were next to a group of the Methow and Wenatchee collections. The fall Chinook populations are also separated to the summer collections and the position of all but three of these collections (lower Yakima River, Crab Creek, and Umatilla River) were statistically supported. The geographic proximity of the fall collections seemed to follow the observed pattern in this dendrogram. The relationship of the Snake River and Lyons Ferry Hatchery in proximity to the collection from Marion Drain was not surprising while

the relationship between Priest Rapids and Hanford Reach was easily a result of the stocking practices of fall Chinook in the 1970 and 1980's.

A secondary objective of this study was to determine if the effective population size of upper Columbia River summer Chinook populations had changed over time due to supplementation efforts. We observed that the number of effective breeders in the collections from 1993 and 2008 has not changed thus providing reason to believe that the genetic diversity of summer Chinook in the upper Columbia River has not been altered through the supplementation program.

### **Acknowledgements**

Chelan County PUD, Grant County PUD, and Washington State General Funds provided funding for this project. Cherril Bowman (WDFW – Molecular Genetics Laboratory) processed samples in the laboratory. Maureen (Mo) Small provided data for some collections and discussion of the analyses. USFWS provided data from the Entiat River.



## References

- Banks, M.A., M.S. Blouin, B.A. Baldwin, V.K. Rashbrook, H.A. Fitzgerald, S.M. Blankenship, and D. Hedgecock. 1999. Isolation and inheritance of novel microsatellites in Chinook salmon (*Oncorhynchus tshawytscha*). *Journal of Heredity* 90:281-288.
- Belkhir, K., P. Borsa, L. Chikhi, N. Raufaste, and F. Bonhomme. 2001. *Genetix, logiciel sous Windows TM pour la genetique des populations*. Laboratoire Genome, Populations, Interactions: CNRS UMR 5000, Universite de Montpellier II, Montpellier, France.
- Blankenship, S.M., J.F. VonBargen, K.I. Warheit, and A.R. Murdoch. 2007. Assessing the Genetic Diversity of Natural Chiwawa River Spring Chinook Salmon and Evaluating the Effectiveness of its Supportive Hatchery Supplementation Program. . Final Report. Unpublished Washington Department of Fish and Wildlife Molecular Genetics Laboratory Report submitted to Chelan County PUD.
- Cairney, M., J.B. Taggart, and B. Hoyheim. 2000. Characterization of microsatellite and minisatellite loci in Atlantic salmon (*Salmo salar L.*) and cross-species amplification in other salmonids. *Molecular Ecology* 9:2175–2178.
- Cavalli-Sforza, L.L. and A.W.F. Edwards. 1967. Phylogenetic analysis: models and estimation procedures. *Evolution* 32:550-570.
- Chapman, D., A. Giorgi, T. Hillman, D. Deppert, M. Erho, S. Hays, C. Peven, B. Suzumoto, and R. Klinge. 1994. Status of summer/fall chinook salmon in the mid-Columbia region. Report for Chelan, Douglas, and Grant County PUDs. 412 p. + app. (Available from Don Chapman Consultants, 3653 Rickenbacker, Ste. 200, Boise, ID 83705.)
- DCPUD. 2005. Conceptual approach for monitoring and evaluating the Douglas County Public Utility District hatchery programs. Douglas County Public Utility District, Wenatchee, Washington. 105 p.
- Felsenstein, J. 1993. PHYLIP (Phylogeny Inference Package) version 3.5c. Distributed by the author. Department of Genetics, University of Washington, Seattle, WA.
- Glaubitz, J.C. 2003. CONVERT (version 1.2): A user-friendly program to reformat diploid genotypic data for commonly used population genetic software packages.  
<http://www.agriculture.purdue.edu/fnr/html/faculty/Rhodes/Students%20and%20Staff/glaubitz/software.htm>.

- Goudet, J. 2001. FSTAT, a program to estimate and test gene diversities and fixation indices (version 293). Updated from Goudet (1995). Available from <http://www.unilch/izea/software/fstat.html>.
- Greig, C., J.P. Jacobson, and M.A. Banks. 2003. New tetranucleotide microsatellites for fine-scale discrimination among endangered Chinook salmon (*Oncorhynchus tshawytscha*). *Molecular Ecology Notes* 3:376-379.
- Hays, S., T. Hillman, T. Kahler, R. Klinge, R. Langshaw, B. Lenz, A. Murdoch, K. Murdoch, and C. Peven. 2006. Decision rules for monitoring and evaluating district hatchery programs. Draft study plan. 27 p.
- HGMP. Draft Hatchery and Genetic Management Plans (2005) for Wenatchee, Methow, and Okanogan River summer Chinook. Available at Washington Department of Fish and Wildlife, 600 Capitol Way N, Olympia, WA, 98501.
- Kassler, T.W., J.F. VonBargen, and D. Hawkins. 2008. DNA-based population of-Origin Assignments of Chinook Salmon Smolts Outmigrating Past Chandler Trap at Prosser Dam (Yakima River) in 2007. Final Report. Unpublished Washington Department of Fish and Wildlife Molecular Genetics Laboratory Report submitted to Bonneville Power Administration (BPA).
- Kassler, T.W. and C.A. Dean. 2010. Genetic analysis of natural-origin spring Chinook and comparison to spring Chinook from an integrated supplementation program and captive broodstock program in the Tucannon River. Final Report. Unpublished Washington Department of Fish and Wildlife Molecular Genetics Laboratory Report submitted to Mike Gallinat, WDFW - Snake River Laboratory, Dayton, WA.
- Jones, O. and J. Wang. 2009. COLONY: a program for parentage and sibship inference from multilocus genotype data. *Molecular Ecology Resources* 10: 551–555.
- Lewis, P. O. and D. Zaykin. 2001. Genetic Data Analysis: Computer program for the analysis of allelic data. Version 1.0 (d16c). Free program distributed by the authors over the internet from <http://lewis.eeb.uconn.edu/lewishome/software.html>

- Marshall, A.R., C. Smith, R. Brix, W. Dammers, J. Hymer, and L. LaVoy. 1995. Genetic diversity units and major ancestral lineages for chinook salmon in Washington. *In* C. Busack and J. B. Shaklee (eds.), Genetic diversity units and major ancestral lineages of salmonid fishes in Washington, p. 111-173. Wash. Dep. Fish Wildl. Tech. Rep. RAD 95-02. (Available from Washington Department of Fish and Wildlife, 600 Capital Way N., Olympia WA 98501-1091.)
- Marshall, A. 2002. 16 August memo to Ann Blakley (WDFW) and Amilee Wilson (WDFW) regarding genetic analyses of selected Washington Chinook stocks. WDFW. Olympia.
- Mobrand, L. (chair), J. Barr, L. Blankenship, D. Campton, T. Evelyn, C. Mahnken, P. Seidel, L. Seeb and B. Smoker. 2004. Hatchery Scientific Review Group (HSRG) – March 2004. Hatchery Reform Recommendations for the Puget Sound and Coastal Washington Hatchery Reform Project. Long Live the Kings, 1305 Fourth Avenue, Suite 810, Seattle, WA 98101 (available from [www.hatcheryreform.org](http://www.hatcheryreform.org)).
- Murdoch, A. and C. Peven. 2005. Conceptual approach for monitoring and evaluating the Chelan County Public Utility District hatchery programs. Chelan County Public Utility District, Wenatchee, Washington. 105 p.
- Myers, J.M., R.G. Kope, G.J. Bryant, D. Teel, L.J. Lierheimer, T.C. Wainwright, W.S. Grand, F.W. Waknitz, K. Neely, S.T. Lindley, and R.S. Waples. 1998. Status review of chinook salmon from Washington, Idaho, Oregon, and California. U.S. Dept. Commer., NOAA Tech. Memo. NMFS-NWFSC-35, 443 p.
- Olsen, J.B., P. Bentzen, and J.E. Seeb. 1998. Characterization of seven microsatellite loci derived from pink salmon. *Molecular Ecology* 7(8):1087-1089.
- Page, R.D.M. 1996. TREEVIEW: an application to display phylogenetic trees on personal computers. *Computer Application Biosciences* 12:351-358.
- Raymond, M. and F. Rousset. 1995. GENEPOP (Version 3.3): Population genetics software for exact tests and ecumenicism. *Journal of Heredity* 86:248-249.
- Rexroad, C.E., III, R.L. Coleman, A.M. Martin, W.K. Hershberger, and J. Killefer. 2001. Thirty-five polymorphic microsatellite markers for rainbow trout (*Oncorhynchus mykiss*). *Animal Genetics* 32:317-319.
- Rice, W.R. 1989. Analyzing tables of statistical tests. *Evolution* 43:223-225.

- Saitou, N. and M. Nei. 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4:406-425.
- Seeb, L.W., A. Antonovich, M.A. Banks, *et al.* 2007. Development of a standardized DNA database for Chinook salmon. *Fisheries* 32:11.
- Small, M.P., K.I. Warheit, C.A. Dean, and A.R. Murdoch. 2007. Methow spring Chinook genetic monitoring. Final Report. Unpublished Washington Department of Fish and Wildlife Molecular Genetics Laboratory Report.
- Small, M.P., D. Burgess, C. Dean, and K. Warheit. 2010. Does Lower Crab Creek in the Eastern WA desert have a native population of Chinook salmon? Submitted to special edition of American Fisheries Society, Proceedings from the Coastwide Salmonid Genetics Meeting, Boise, ID.
- Stuehrenberg, L.C., G.A. Swan, L.K. Timme, P.A. Ocker, M.B. Eppard, R.N. Iwamoto, B.L. Iverson, and B.P. Sanford. 1995. Migrational characteristics of adult spring, summer, and fall chinook salmon passing through reservoirs and dams of the mid-Columbia River. Final report. CZES Division, NWFSC, NMFS, Seattle, WA, 115 p.
- Utter, F.M., D.W. Chapman, and A.R. Marshall. 1995. Genetic population structure and history of chinook salmon of the Upper Columbia River. *American Fisheries Society Symposium* 17:149-165.
- Waknitz, F.W., G.M. Matthews, T. Wainwright, and G.A. Winans. 1995. Status review for Mid-Columbia River summer chinook salmon. NOAA Tech. Mem. NMFS-NWFSC-22, 80 p. (Available from Natl. Mar. Fish. Serv., Northwest Fisheries Science Center, Coastal Zone and Estuarine Studies Division, 2725 Montlake Blvd. E., Seattle, WA 98112-2097.)
- Wang, J. 2009. A new method for estimating effective population sizes from a single sample of multilocus genotypes. *Molecular Ecology* 18:2148-2164
- Wang, J. and A.W. Santure. 2009. Parentage and sibship inference from multilocus genotype data under polygamy. *Genetics* 181: 1579-1594.
- Waples R.S. 1990. Conservation genetics of Pacific salmon. III. Estimating effective population size. *Journal of Heredity* 81:277-289
- Waples R.S., M. Masuda, and J. Pella. 2007. SALMONNb: a program for computing cohort-specific effective population sizes ( $N_b$ ) in Pacific salmon and other semelparous species using the temporal method. *Molecular Ecology Notes* 7, 21-24

WDF (Washington Department of Fisheries) and WDW (Washington Department of Wildlife). 1993. 1992 Washington state salmon and steelhead stock inventory. Appendix Three. Columbia River stocks. WDF. Olympia, WA.

Weir, B.S. and C.C. Cockerham. 1984. Estimating F-statistics for the analysis of population structure. *Evolution* 38:1358-1370.

Williamson, K.S., J.F. Cordes, and B.P. May. 2002. Characterization of microsatellite loci in chinook salmon (*Oncorhynchus tshawytscha*) and cross-species amplification in other salmonids. *Molecular Ecology Notes* 2 (1):17-19.

Wright, S. 1969. *Evolution and the Genetics of Populations, Vol. 2, The Theory of Gene Frequencies*. The University of Chicago Press, Chicago, Illinois.

Table 1. Samples of adult hatchery- and natural-origin summer and fall Chinook that were analyzed from the upper Columbia River. Total number of individuals that were analyzed / individuals with data for 9 or more loci that were included in the analysis. Collection statistics (allelic richness, linkage disequilibrium (before and after Bonferroni correction),  $F_{IS}$ , heterozygosity ( $H_O$  and  $H_E$ )) and p-values for deviations from Hardy-Weinberg equilibrium (HWE). P-values were defined as significant after implementation of Bonferroni correction for multiple tests (Rice 1989).

WDFW GSI code <sup>a</sup>	Collection location	N =	Allelic Richness <sup>b</sup>	Linkage Disequilibrium <sup>c</sup>	$F_{IS}$ (p-value) <sup>d</sup>	$H_O$	$H_E$
93DD	Wenatchee River upstream of Tumwater Dam - natural origin	51 / 45					
93DE	Wenatchee River downstream of Tumwater Dam - natural origin	88 / 88					
06CQ	Wenatchee River upstream of Tumwater Dam - natural origin	95 / 86					
06CR	Wenatchee River downstream of Tumwater Dam - natural origin	95 / 82					
08FV	Wenatchee River upstream of Tumwater Dam - natural origin	95 / 82					
08FW	Wenatchee River downstream of Tumwater Dam - natural origin	95 / 87					
	<b>Wenatchee River - Natural origin combined</b>	<b>519 / 470</b>	<b>10.7</b>	<b>17 / 4</b>	<b>0.001 (0.403)</b>	<b>0.8504</b>	<b>0.8513</b>
06CP	Wenatchee River - hatchery origin	95 / 70					
08FU	Wenatchee River - hatchery origin	95 / 83					
	<b>Wenatchee River - Hatchery origin combined</b>	<b>190 / 153</b>	<b>10.6</b>	<b>18 / 6</b>	<b>0.018 (0.013)</b>	<b>0.8409</b>	<b>0.8561</b>
93EC	Methow River - natural origin	27 / 27					
06CT	Methow River - natural origin	95 / 90					
08FY	Methow River - natural origin	95 / 88					
09CO	Methow River - natural origin	91 / 80					
	<b>Methow River - Natural origin combined</b>	<b>308 / 285</b>	<b>10.7</b>	<b>4 / 1</b>	<b>0.006 (0.160)</b>	<b>0.8506</b>	<b>0.8554</b>
06CS	Methow River - hatchery origin	14 / 8					
08FX	Methow River - hatchery origin	21 / 18					
09CP	Methow River - hatchery origin	19 / 18					
	<b>Methow River - Hatchery origin combined</b>	<b>54 / 44</b>	<b>10.8</b>	<b>11 / 2</b>	<b>-0.003 (0.593)</b>	<b>0.8553</b>	<b>0.8523</b>

Table 1 continued.							
92FM	Okanogan River - natural origin	49 / 46					
93ED*	Okanogan River - natural origin	103 / 87					
06CV	Okanogan River - natural origin	95 / 88					
08GA	Okanogan River - natural origin	95 / 92					
09CN	Okanogan River - natural origin	133 / 126					
	<b>Okanogan River - Natural origin combined</b>	<b>475 / 439</b>	<b>10.8</b>	<b>9 / 4</b>	<b>0.003 (0.304)</b>	<b>0.8563</b>	<b>0.8596</b>
* - not included in the combined dataset							
06CU	Okanogan River - hatchery origin	58 / 49					
08FZ	Okanogan River - hatchery origin	19 / 18					
09CM	Okanogan River - hatchery origin	117 / 107					
	<b>Okanogan River - hatchery origin combined</b>	<b>194 / 174</b>	<b>10.8</b>	<b>31 / 10</b>	<b>-0.011 (0.920)</b>	<b>0.8678</b>	<b>0.8586</b>
91FL	Wells Hatchery	68 / 42					
92FK	Wells Hatchery	25 / 23					
93DG	Wells Hatchery	11 / 9					
06DM	Wells Hatchery	95 / 91					
08HY	Wells Hatchery	95 / 91					
	<b>Wells Hatchery combined</b>	<b>294 / 256</b>	<b>10.7</b>	<b>8 / 3</b>	<b>-0.001 (0.529)</b>	<b>0.8670</b>	<b>0.8665</b>
08MN	<b>Eastbank Hatchery - Wenatchee River stock</b>	<b>95 / 90</b>	<b>10.7</b>	<b>6 / 1</b>	<b>0.020 (0.024)</b>	<b>0.8326</b>	<b>0.8498</b>
92FO	Eastbank Hatchery - Methow / Okanogan (MEOK) stock	36 / 33					
93DF	Eastbank Hatchery - Methow / Okanogan (MEOK) stock	90 / 86					
08MO	Eastbank Hatchery - Methow / Okanogan (MEOK) stock	95 / 88					
	<b>Eastbank Hatchery - MEOK stock combined</b>	<b>221 / 207</b>	<b>10.7</b>	<b>2 / 0</b>	<b>-0.005 (0.782)</b>	<b>0.8647</b>	<b>0.8604</b>
		<b>2,350 / 2,118</b>					

Table 1 continued.							
06KN	<b>Chelan River</b>	<b>70 / 23</b>	<b>10.3</b>	<b>11 / 0</b>	<b>0.027 (0.118)</b>	<b>0.8334</b>	<b>0.8556</b>
Data provided by USFWS							
	<b>Entiat River - summer Chinook</b>	<b>190</b>	<b>10.9</b>	<b>33 / 10</b>	<b>0.008 (0.119)</b>	<b>0.8553</b>	<b>0.8625</b>
Data from Small et al. (2010)							
08EH	Crab Creek	108					
09AZ	Crab Creek	291					
	<b>Crab Creek</b>	<b>399</b>	<b>10.5</b>	<b>35 / 14</b>	<b>0.018 (0.000)</b>	<b>0.8519</b>	<b>0.8676</b>
GAPS v.3.0 data							
	<b>Priest Rapids Hatchery - fall Chinook</b>	<b>81</b>	<b>11.1</b>	<b>3 / 2</b>	<b>0.015 (0.079)</b>	<b>0.8591</b>	<b>0.8723</b>
	<b>Hanford Reach - fall Chinook</b>	<b>220</b>	<b>11.3</b>	<b>4 / 0</b>	<b>0.010 (0.068)</b>	<b>0.8661</b>	<b>0.8746</b>
	<b>Umatilla - fall Chinook</b>	<b>96</b>	<b>11.2</b>	<b>17 / 6</b>	<b>-0.003 (0.623)</b>	<b>0.8719</b>	<b>0.8693</b>
	<b>lower Yakima River - fall Chinook</b>	<b>103</b>	<b>11.0</b>	<b>3 / 1</b>	<b>0.000 (0.511)</b>	<b>0.8724</b>	<b>0.8721</b>
	<b>Marion Drain - fall Chinook</b>	<b>190</b>	<b>10.8</b>	<b>9 / 4</b>	<b>0.022 (0.001)</b>	<b>0.8586</b>	<b>0.8782</b>
	<b>Lyons Ferry Hatchery - fall Chinook</b>	<b>186</b>	<b>10.6</b>	<b>7 / 4</b>	<b>0.013 (0.033)</b>	<b>0.8527</b>	<b>0.8641</b>
	<b>Snake River - fall Chinook</b>	<b>521</b>	<b>11.1</b>	<b>0 / 0</b>	<b>-0.001 (0.634)</b>	<b>0.8720</b>	<b>0.8708</b>
		<b>NA / 2,009</b>					
<sup>a</sup> - Year that samples were collected is identified by the two numbers in the WDFW GSI code <sup>b</sup> - based on a minimum of 11 diploid individuals <sup>c</sup> - adjusted alpha p-value = 0.0006 <sup>d</sup> - adjusted alpha p-value = 0.0002							



Table 2. PCR conditions and microsatellite locus information (number alleles/locus and allele size range) for multiplexed loci used for the analysis of Chinook. Also included are the observed and expected heterozygosity ( $H_o$  and  $H_e$ ) for each locus.

PCR Conditions			Locus statistics		Heterozygosity		
Poolplex	Locus	Dye Label	# Alleles/ Locus	Allele Size Range (bp)	$H_o$	$H_e$	References
Ots-M	<i>Ots-201b</i>	blue	49	137 - 334	0.9474	0.9544	Unpublished
	<i>Ots-208b</i>	yellow	56	154 - 378	0.9523	0.9672	Greig et al. 2003
	<i>Ssa-408</i>	red	32	184 - 308	0.9177	0.9214	Cairney et al. 2000
Ots-N	<i>Ogo-2</i>	red	22	206 - 260	0.8526	0.8673	Olsen et al. 1998
Ots-O	<i>Ogo-4</i>	blue	20	128 - 170	0.6694	0.7028	Olsen et al. 1998
	<i>Ots-213</i>	yellow	45	178 - 370	0.9430	0.9525	Greig et al. 2003
	<i>Ots-G474</i>	red	16	152 - 212	0.6816	0.6838	Williamson et al. 2002
Ots-R	<i>Ots-3M</i>	blue	15	128 - 158	0.7854	0.7938	Banks et al. 1999
	<i>Omm-1080</i>	green	54	162 - 374	0.9517	0.9670	Rexroad et al. 2001
Ots-S	<i>Ots-9</i>	red	9	99 - 115	0.6531	0.6543	Banks et al. 1999
	<i>Ots-212</i>	blue	33	123 - 251	0.9205	0.9360	Greig et al. 2003
Ots-T	<i>Oki-100</i>	blue	50	164 - 361	0.9500	0.9567	Unpublished
	<i>Ots-211</i>	red	34	188 - 327	0.9325	0.9414	Greig et al. 2003

Table 3. Tests of population differentiation for temporal collections of summer Chinook from natural and hatchery-origin populations in the upper Columbia River. P-values that are highlighted grey are significantly different after Bonferroni correction (Rice 1989). Adjusted alpha p-value was 0.0001 . The H and W in the collection identifier is for wild or hatchery-origin and the two digit number identifies the year samples were collected.

<b>Wenatchee River</b>								
	WenW93U	WenW93D	WenH06	WenW06U	WenW06D	WenH08	WenW08U	WenW08D
WenW93U	****							
WenW93D	0.0162	****						
WenH06	0.0033	0.0102	****					
WenW06U	0.3039	0.1642	0.4795	****				
WenW06D	0.0261	0.0160	0.0678	0.5300	****			
WenH08	0.1126	0.0708	0.0073	0.4359	0.0893	****		
WenW08U	0.2115	0.1148	0.4191	0.7243	0.3830	0.8856	****	
WenW08D	0.1915	0.0014	0.7047	0.4928	0.1671	0.7755	0.7665	****
D - collection was downstream of Tumwater Dam; U - collection was upstream of Tumwater Dam								
<b>Methow River</b>								
	MetW93	MetH06	MetW06	MetH08	MetW08	MetW09	MetH09	
MetW93	****							
MetH06	0.3962	****						
MetW06	0.5481	0.4688	****					
MetH08	0.1408	0.1192	0.2052	****				
MetW08	0.8219	0.8937	0.6156	0.3779	****			
MetW09	0.2564	0.4282	0.2502	0.0328	0.7309	****		
MetH09	0.1543	0.5678	0.0547	0.0017	0.0098	0.0073	****	
<b>Okanogan River</b>								
	OkanW92	OkanW93	OkanH06	OkanW06	OkanH08	OkanW08	OkanH09	OkanW09
OkanW92	****							
OkanW93	0.0066	****						
OkanH06	0.0193	0.0000	****					
OkanW06	0.2843	0.0082	0.0031	****				
OkanH08	0.1290	0.1106	0.0652	0.7329	****			
OkanW08	0.0106	0.0029	0.0082	0.4075	0.7396	****		
OkanH09	0.0187	0.0001	0.0094	0.0551	0.2214	0.0281	****	
OkanW09	0.0527	0.0000	0.0024	0.7130	0.0262	0.0065	0.0002	****

Table 3 continued.					
<b>Wells Dam Hatchery</b>					
	Wells91	Wells92	Wells93	Wells06	Wells08
Wells91	****				
Wells92	0.5863	****			
Wells93	0.0490	0.0784	****		
Wells06	0.0089	0.0100	0.0542	****	
Wells08	0.0819	0.1088	0.2552	0.0256	****
<b>Eastbank Hatchery - Wenatchee and MEOK stocks</b>					
	EBHWen08	EBHME92	EBHME93	EBHME08	
EBHWen08	****				
EBHME92	0.8681	****			
EBHME93	0.0251	0.8661	****		
EBHME08	0.0086	0.9563	0.1895	****	

Table 4.  $F_{ST}$  pairwise comparisons and genotypic tests of differentiation for hatchery- and natural-origin summer Chinook from the upper Columbia River. Above the diagonal are the  $F_{ST}$  values and below are p-values for the test of genotypic differentiation. Non-significant p-values for the result of the genotypic differentiation test are in bold type and  $F_{ST}$  values that are not significantly different from zero are in bold type.

	Wenatchee Hatchery	Wenatchee Natural	Methow Hatchery	Methow Natural	Okanogan Hatchery	Okanogan Natural	Wells Hatchery	Eastbank Wenatchee stock	Eastbank MEOK stock	Entiat River	Chelan River
Wenatchee Hatchery	****	<b>0.0000</b>	<b>0.0011</b>	<b>0.0000</b>	<b>0.0013</b>	<b>0.0010</b>	0.0015	<b>0.0004</b>	<b>0.0007</b>	<b>0.0004</b>	<b>0.0072</b>
Wenatchee Natural	<b>0.4351</b>	****	<b>0.0016</b>	<b>0.0000</b>	0.0014	0.0016	0.0024	<b>0.0006</b>	0.0012	0.0009	<b>0.0068</b>
Methow Hatchery	<b>0.3800</b>	<b>0.0205</b>	****	<b>0.0012</b>	0.0029	<b>0.0008</b>	<b>0.0027</b>	<b>0.0014</b>	<b>0.0022</b>	<b>0.0019</b>	<b>0.0078</b>
Methow Natural	<b>0.2237</b>	<b>0.6566</b>	<b>0.1502</b>	****	<b>0.0011</b>	0.0011	0.0013	<b>0.0007</b>	<b>0.0007</b>	<b>0.0008</b>	<b>0.0053</b>
Okanogan Hatchery	0.0001	0.0000	<b>0.0364</b>	<b>0.0008</b>	****	<b>0.0010</b>	<b>0.0014</b>	0.0029	<b>0.0000</b>	<b>0.0007</b>	<b>0.0055</b>
Okanogan Natural	0.0000	0.0000	<b>0.1755</b>	0.0000	<b>0.0003</b>	****	0.0016	0.0023	<b>0.0005</b>	<b>0.0008</b>	<b>0.0049</b>
Wells Hatchery	0.0000	0.0000	<b>0.0129</b>	0.0000	0.0000	0.0000	****	0.0036	<b>0.0006</b>	<b>0.0008</b>	<b>0.0041</b>
Eastbank Wenatchee	<b>0.5261</b>	<b>0.4102</b>	<b>0.1215</b>	<b>0.8404</b>	<b>0.0015</b>	0.0000	0.0000	****	<b>0.0018</b>	0.0030	0.0096
Eastbank MEOK stock	<b>0.0485</b>	0.0000	<b>0.4246</b>	<b>0.0009</b>	<b>0.5786</b>	<b>0.0051</b>	0.0000	<b>0.0065</b>	****	<b>0.0005</b>	<b>0.0039</b>
Entiat River	<b>0.0565</b>	0.0000	<b>0.1795</b>	<b>0.0044</b>	<b>0.0005</b>	0.0000	<b>0.0032</b>	<b>0.0039</b>	<b>0.0042</b>	****	<b>0.0052</b>
Chelan River	<b>0.0091</b>	<b>0.0026</b>	<b>0.0182</b>	<b>0.0156</b>	<b>0.0048</b>	<b>0.0030</b>	<b>0.0066</b>	<b>0.0059</b>	<b>0.0493</b>	<b>0.0617</b>	****

Table 5.  $F_{ST}$  pairwise comparisons and genotypic tests of differentiation for fall Chinook. Above the diagonal are the  $F_{ST}$  values and below are p-values for the test of genotypic differentiation. Non-significant p-values for the result of the genotypic differentiation test are in bold type and  $F_{ST}$  values that are not significantly different from zero are in bold type.

	Crab Creek	Hanford Reach Fall	Lyons Ferry Hatchery Fall	lower Yakima River Fall	Marion Drain Fall	Priest Rapids Fall	Umatilla River Fall	Snake River Fall		
Crab Creek	****	0.0087	0.0134	0.0079	0.0143	0.0107	0.0073	0.0097		
Hanford Reach Fall	0.0000	****	0.0077	<b>0.0000</b>	0.0064	<b>0.0000</b>	<b>0.0000</b>	0.0022		
Lyons Ferry Hatchery Fall	0.0000	0.0000	****	0.0063	0.0074	0.0092	0.0062	0.0029		
lower Yakima River Fall	0.0000	<b>0.4140</b>	0.0000	****	0.0054	<b>0.0000</b>	<b>0.0000</b>	0.0018		
Marion Drain Fall	0.0000	0.0000	0.0000	0.0000	****	0.0067	0.0061	0.0060		
Priest Rapids Fall	0.0000	<b>0.0695</b>	0.0000	<b>0.0083</b>	0.0000	****	<b>0.0000</b>	0.0027		
Umatilla River Fall	0.0000	<b>0.4879</b>	0.0000	<b>0.4896</b>	0.0000	<b>0.2539</b>	****	0.0011		
Snake River Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	****		

Table 6.  $F_{ST}$  pairwise comparisons and genotypic tests of differentiation for hatchery- and natural-origin summer Chinook from the upper Columbia River and fall Chinook. Above the diagonal are the  $F_{ST}$  values and below are p-values for the test of genotypic differentiation. Non-significant p-values for the result of the genotypic differentiation test are in bold type and  $F_{ST}$  values that are not significantly different from zero are in bold type.

Population Differentiation											
	Wenatchee Hatchery	Wenatchee Natural	Methow Hatchery	Methow Natural	Okanogan Hatchery	Okanogan Natural	Wells Hatchery	Eastbank Wenatchee stock	Eastbank MEOK stock	Entiat River	Chelan River
Crab Creek	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
Hanford Reach Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	<b>0.0349</b>
Lyons Ferry Hatchery Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
lower Yakima River Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	<b>0.0074</b>
Marion Drain Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
Priest Rapids Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	<b>0.0642</b>
Umatilla River Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	<b>0.0579</b>
Snake River Fall	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000

Table 6 continued.								
Pairwise $F_{ST}$								
	Crab Creek	Hanford Reach Fall	Ferry Hatchery	Yakima River	Marion Drain Fall	Priest Rapids Fall	Umatilla River Fall	Snake River Fall
Wenatchee Hatchery	0.0158	0.0054	0.0180	0.0056	0.0153	<b>0.0025</b>	0.0053	0.0103
Wenatchee Natural	0.0162	0.0059	0.0185	0.0063	0.0157	0.0030	0.0059	0.0102
Methow Hatchery	0.0191	0.0104	0.0248	0.0095	0.0220	0.0069	0.0107	0.0165
Methow Natural	0.0148	0.0057	0.0182	0.0051	0.0148	0.0033	0.0055	0.0101
Okanogan Hatchery	0.0146	0.0041	0.0166	0.0042	0.0151	<b>0.0016</b>	0.0041	0.0082
Okanogan Natural	0.0163	0.0064	0.0187	0.0062	0.0170	0.0035	0.0068	0.0113
Wells Hatchery	0.0120	0.0051	0.0135	0.0044	0.0120	0.0028	0.0046	0.0077
Wenatchee stock	0.0184	0.0073	0.0203	0.0074	0.0167	0.0047	0.0084	0.0128
Eastbank MEOK stock	0.0128	0.0036	0.0143	0.0038	0.0135	<b>0.0019</b>	0.0038	0.0079
Entiat River	0.0147	0.0059	0.0176	0.0057	0.0156	0.0028	0.0056	0.0100
Chelan River	0.0074	<b>0.0046</b>	0.0110	<b>0.0040</b>	0.0160	<b>0.0047</b>	<b>0.0035</b>	0.0072

Table 7. Effective number of breeders per brood year with the largest number of samples of summer Chinook in the upper Columbia River. Brood years with sample size less than 19 individuals (shown in bold type) were not analyzed with exception of the 2008 Wells Hatchery collection. A comparison could not be made between an early and late collection from Wells Hatchery.

WDFW Code	Collection Location	Sample Size	Nb =	CI95(L) =	CI95(U) =
93DD <sup>A</sup>	Wenatchee Natural - upstream	23 / 19	152 / 190	77 / 87	616 / 2,147,483,647
08FV	Wenatchee Natural - upstream	56	162	112	249
93DE <sup>A</sup>	Wenatchee Natural - downstream	39 / 34	145 / 152	94 / 95	256 / 302
08FW	Wenatchee Natural - downstream	67	140	105	199
08FU	Wenatchee Hatchery	60	134	90	213
93EC <sup>A</sup>	Methow Natural	<b>10 / 15</b>	---	---	---
08FY	Methow Natural	62	150	106	218
08FX	Methow Hatchery	<b>9</b>	---	---	---
93ED	Okanogan Natural	69	142	102	203
08GA	Okanogan Natural	59	127	92	180
08FZ	Okanogan Hatchery	<b>16</b>	---	---	---
93DG	Wells Hatchery	<b>6</b>	---	---	---
08HY <sup>B</sup>	Wells Hatchery	24 / 39	---	---	---
08MN	Eastbank Hatchery - Wenatchee	88	190	144	263
93DF	Eastbank Hatchery - MEOK	84	171	129	229
08MO	Eastbank Hatchery - MEOK	88	166	126	226
<sup>A</sup> - calculations were made for samples from brood year 1988 / brood year 1989					
<sup>B</sup> - samples were collected from brood year 2003 / brood year 2004					



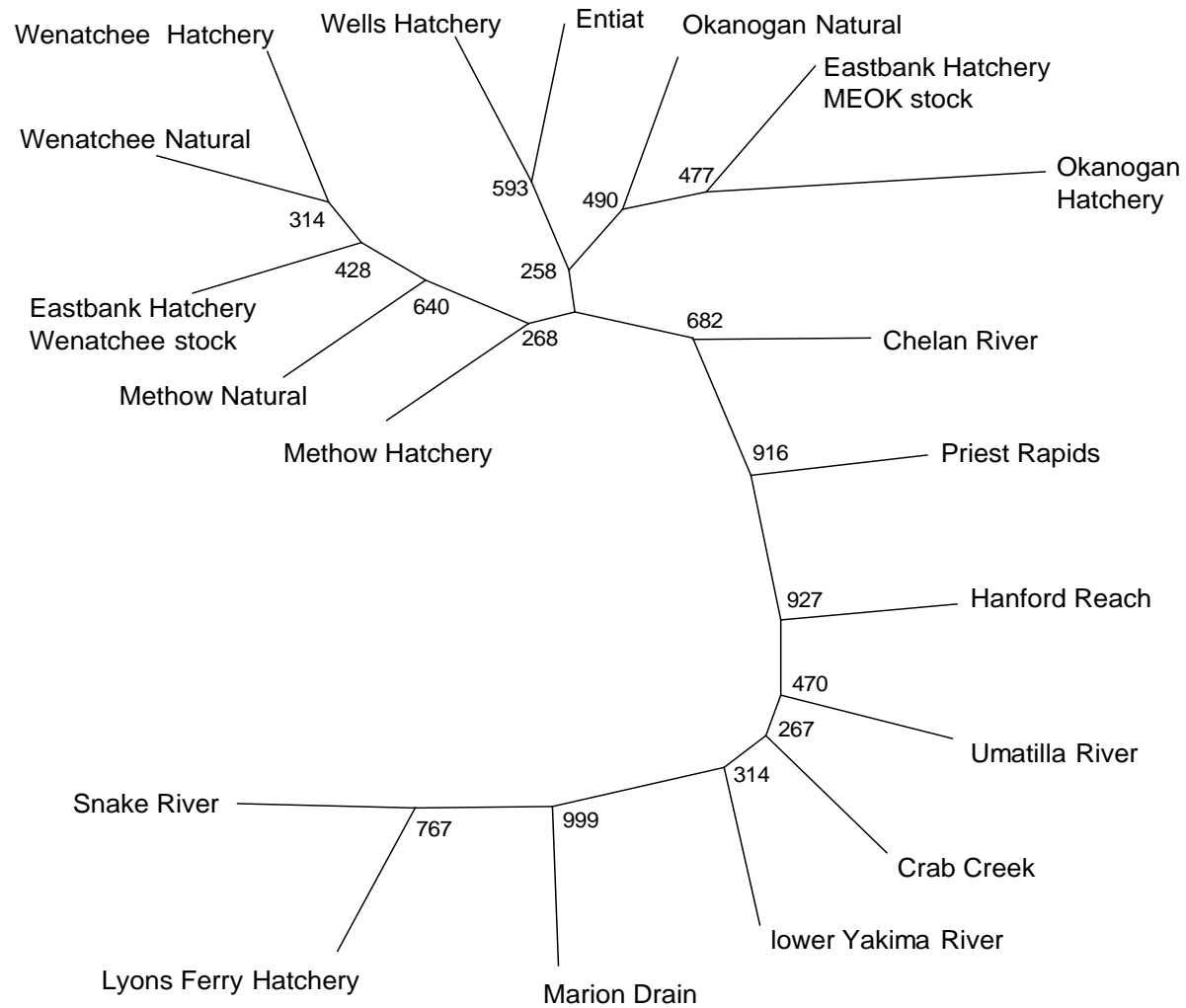


Figure 1. Relationship of natural- and hatchery-origin Chinook collections from the upper Columbia River basin using Cavalli-Sforza and Edwards (1967) chord distance. Bootstrap values are shown at each node.



# Appendix N

**Summer Chinook Spawning Ground Surveys in the Methow  
River Basin and Chelan River, 2014**





March 9, 2015

To: Chelan and Grant Public Utility Districts

From: Denny Snyder, Keith Watson, and Mark Miller

Re: 2014 Summer Chinook spawning ground surveys in the Methow Basin and Chelan River.

The purpose of this memo is to provide information on the spawning population of summer Chinook in the Methow and Chelan River basins. This work is part of a larger effort focused on monitoring and evaluating Chelan and Grant Public Utility Districts (PUDs) hatchery supplementation programs. The larger effort is presented in Hillman et al. (2015). The tasks and objectives associated with implementing the PUDs hatchery M&E plan for 2014 are outlined in Hillman et al. (2013). Figures and tables are presented at the end of this memo. In 2014, The Okanogan River basin was surveyed by the Confederated Tribes of the Colville Reservation. That work will be prepared in a Bonneville Power Administration report.

## **METHODS**

Spawning ground surveys were conducted by foot and raft beginning the last week of September and ending late November. We did not use aerial surveys on the Methow River because past work has demonstrated that ground counts were more accurate than aerial surveys (Miller and Hillman 1997). Ground surveys were used to provide more accurate counts and a complete census of Chinook redds within their spawning distribution. Observers floated or walked through sampling reaches and recorded the location and numbers of redds each week. Observers recorded the date, water temperature, and river mile, and constructed a drawing of the area where redds were located. A different symbol was used each week to record the number of new and incomplete redds.

To maintain consistency, at least one observer surveyed the same stream reach on successive dates. In areas where numerous summer Chinook spawn, we constructed detailed maps of the river and used the cell-area method (Hamilton and Bergersen 1984) to identify the number of redds within each cell. Cells were bound by noticeable landmarks along the banks (e.g., bridges or trees) or at stream habitat boundaries (e.g., transitions between pools and riffles). The number of redds were then recorded in the corresponding grid on the map. When possible, observers estimated the number of redds in a large disturbed area by counting females that defended redds. We assumed that the area or territory defended by a female was one redd.

Carcasses of summer Chinook were sampled to describe the spawning population. Biological data included collection of scale samples for age analysis, length measurements (POH and FKL),

sex, and egg voidance. All fish were scanned for the presence of a PIT tag and snouts were collected on all ad-clipped fish. The field data collected will be used to assess length-at-age, size-at-age, egg voidance, origin (hatchery or naturally produced), and stray rates. No DNA samples were collected on summer Chinook this year. Summer Chinook spawning surveys were also conducted on the Chelan River and those results are presented in this report. The only variables reported for the Okanogan and Similkameen rivers were escapement and number of redds.

## **RESULTS**

### **Methow**

There were 591 summer Chinook redds counted within seven reaches of the Methow River (Table 1). No summer Chinook redds were counted in the Chewuch and Twisp rivers. Spawning began the last week of September, peaked in mid-October, and ended the last week of October (Figure 1). Stream temperatures in the Methow River when spawning began varied from 9.5-12.0°C. Spawning peaked the first week of October in reach (M7). Peak spawning occurred in reaches (M3-M6) of the Methow River during the second week of October. Spawning peaked the third week of October in reaches (M1 and M2). Most redds (80%) were located in reaches (M3-M5) upstream of the town of Carlton to the Winthrop Bridge (Table 1). An increase in summer Chinook spawning occurred (12%) upstream from the Winthrop Bridge in reaches M6 and M7 compared to 2013 (1%). In 2014, Reach (M1) experienced a drastic decline in spawning with only nine redds observed (1%) compared to the average of 228 redds (20%) for the past five years. Estimated escapement based on expansion of redd counts from the sex-ratio observed at Wells Dam during broodstock collection indicates that 1,625 summer Chinook (591 redds x 2.75 fish/redd) escaped to the Methow River.

The Carlton Complex Fires and associated landslides that occurred in 2014 appeared to result in significant sedimentation in the M1 and M2 reaches of the Methow River. Extreme sedimentation was noticeable in the Methow River beginning at the confluence of Beaver Creek continuing downstream to Leecher Canyon and Benson creeks. High turbidity affected visibility and resulted in extremely poor survey conditions. There was virtually no visual evidence of any adult Summer Chinook digging in traditional spawning locations in most of the lower two reaches. Conditions were exacerbated further when heavy precipitation in late October and early November decreased visibility. Virtually no Chinook redds were found after heavy precipitation occurred. Late October and early November is typically when summer Chinook spawning increases in the lower Methow.

There were 485 summer Chinook salmon carcasses sampled within the seven reaches of the Methow River (Table 2). The presence or absence of an adipose fin could not be determined on three of those fish. About 30% of the fish returning to the Methow River were sampled based on the estimated escapement of 1,625 summer Chinook. Females made up 42% and males 58% of the carcasses examined. Mean percent egg voidance assessed from 207 female carcasses was 99%. One females died before spawning (i.e., she retained all her eggs). Ad-clipped hatchery fish made up 9% and naturally produced fish (adipose fin present) made up 91% of the sample collected (Table 2). The distribution of ad-clipped hatchery and naturally produced fish showed that more than half (52%) of the ad-clipped hatchery fish were located in reach (M3), while naturally produced fish were more evenly distributed with (47%) in the upper three reaches (M5-M7) (Figure 2). There were five PIT-tagged summer Chinook recovered during carcass surveys.

## **Chelan River**

There were 400 redds counted in the Chelan River. Spawning activity began the last week of September and peaked five weeks later (Table 1). Spawning continued into late November when spawning was considered complete. The majority of spawning occurred in the powerhouse and Columbia River tailrace (Table 1). Combined, redds in the habitat channel and upstream pool made up about 20% of the total redds observed. Estimated escapement based on expansion of redd counts indicates that 1,100 summer Chinook (400 redds x 2.75 fish/redd) escaped to the Chelan River.

There were 309 summer Chinook carcasses sampled (Table 2). About 28% of the summer Chinook returning to the Chelan River were sampled based on the estimated escapement of 1,100 fish. Females made up 71% and males 29% of the carcasses examined. Mean percent egg voidance from 221 female carcasses was 88%. Fifteen females (7%) died before spawning. Ad-clipped hatchery fish made up 30% and naturally produced fish made up 70% of the fish examined.

We counted five coho redds in the Chelan River. There was one redd in the pool, two within the habitat channel, one in the powerhouse tailrace, and one in the Columbia River tailrace. Seventeen coho salmon were sampled within the Chelan River and data from those fish were submitted to Yakama Nation (Peshastin office).

## **Okanogan Basin**

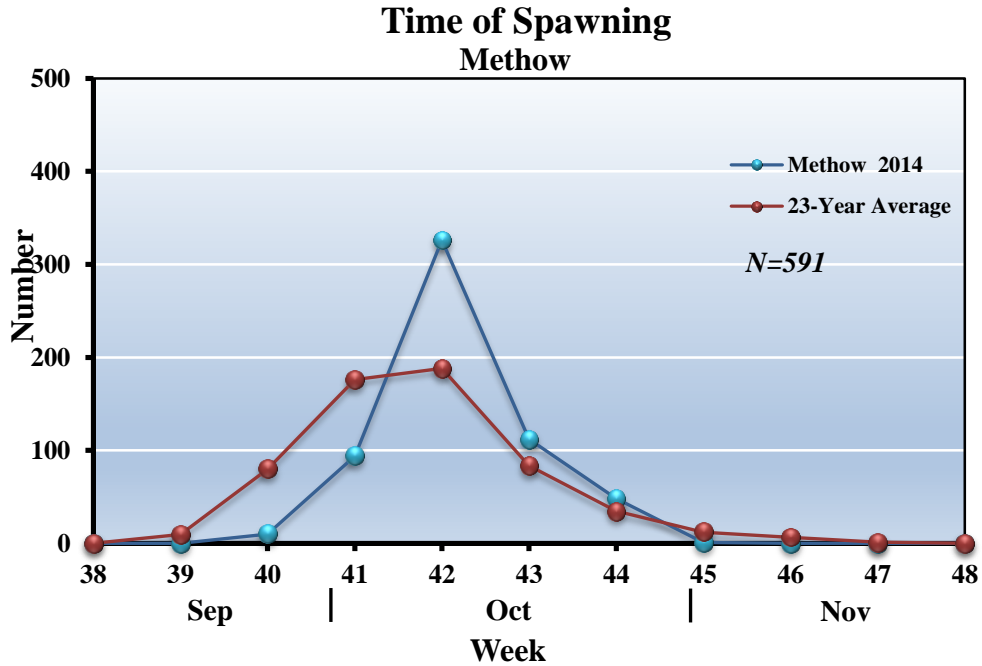
In 2014, the Colville Tribes conducted summer Chinook surveys on the Okanogan and Similkameen Rivers. A total of 2,231 redds were counted in the Okanogan River and 2,022 redds were counted in the Similkameen River. Based on expanded redd counts, about 6,381 summer Chinook spawned in the Okanogan River and 5,783 summer Chinook spawned in the Similkameen River (Andrea Pearl, CCT, personal communication).

## **REFERENCES**

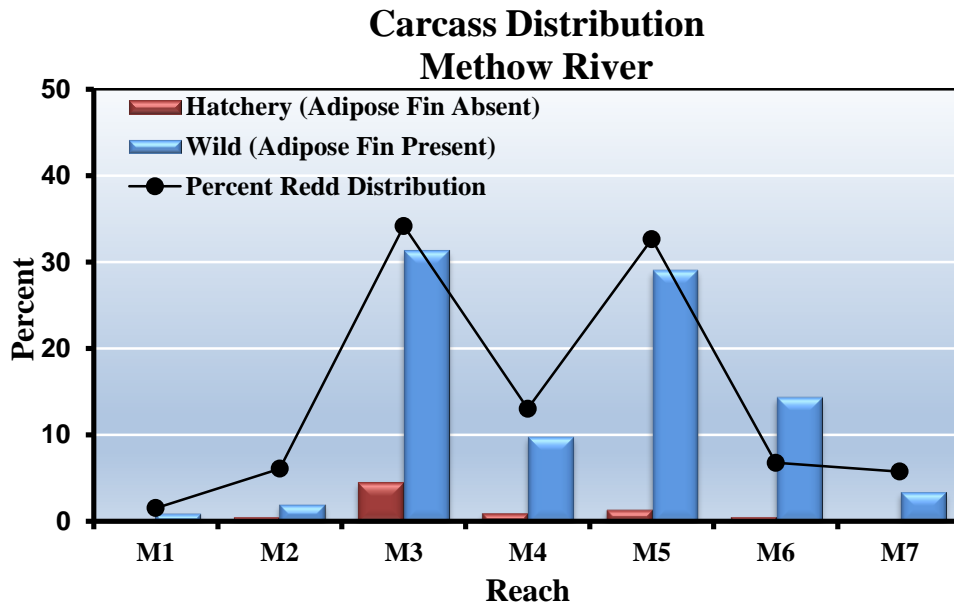
- Hamilton, K. and E. P. Bergersen. 1984. Methods to estimate aquatic habitat variables. Report for Bureau of Reclamation, Division of Planning and Technical Services, Denver, Colorado. Colorado Cooperative Fishery Research Unit, Colorado State University, Fort Collins, CO.
- Hillman, T., T. Kahler, G. Mackey, J. Murauskas, A. Murdoch, K. Murdoch, T. Pearsons, and M. Tonseth. 2013. Updated monitoring and evaluation plan for PUD hatchery programs. Report to the Hatchery Committees, Wenatchee, East Wenatchee, and Ephrata, WA.
- Hillman, T., M. Miller, M. Johnson, C. Moran, M. Tonseth, A. Murdoch, C. Willard, L. Keller, B. Ishida, C. Kamphaus, T. Pearsons, and P. Graf. 2015. Monitoring and evaluation of the Chelan and Grant County PUDs hatchery programs: 2014 annual report. Report to the HCP and PRCC Hatchery Committees, Wenatchee and Ephrata, WA.

Miller, M. D. and T. W. Hillman. 1997. Summer/fall Chinook salmon spawning ground surveys in the Methow and Okanogan river basins, 1997. Report to Chelan County PUD. Don Chapman Consultants, Inc. Boise, ID.





**Figure 1.** Number of new redds counted each week from mid-September to mid-November. The figure displays the beginning, peak, and end of spawning for summer Chinook in the Methow River in 2014 compared to a 23-year average (1991-2013).



**Figure 2.** Percent distribution of ad-clipped hatchery and naturally produced fish plotted against the percent distribution of redds observed in reaches on the Methow River, 2014.

**Table 1.** Number of summer Chinook redds observed each week within the Methow and Chelan rivers, 2014. Dashes indicate that no survey occurred.

Reach	Location (Rkm)	Sep		Oct				Nov				Total	Percent
		21-27	28-4	5-11	12-18	19-25	26-1	2-8	9-15	16-22	23-29		
<b>Methow River</b>													
M1	0.0-23.8	0	0	2	6	1	0	-	-	-	-	9	1.5
M2	23.8-43.8	0	3	12	18	2	1	-	-	-	-	36	6.1
M3	43.8-63.7	0	23	101	47	31	0	-	-	-	-	202	34.2
M4	63.7-72.3	0	11	45	15	6	0	-	-	-	-	77	13.0
M5	72.3-80.1	0	28	140	19	6	0	-	-	-	-	193	32.7
M6	80.1-83.0	0	10	21	9	0	0	-	-	-	-	40	6.8
M7	83.0-96.1	10	19	2	3	0	0	-	-	-	-	34	5.8
<b>Total:</b>		<b>10</b>	<b>94</b>	<b>323</b>	<b>117</b>	<b>46</b>	<b>1</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>0</b>	<b>591</b>	<b>100</b>
<b>Chelan River</b>													
P.H. Tailrace		0	3	0	4	31	89	101	15	2	1	246	61.5
Col. R. Tailrace		0	0	0	1	9	7	33	20	6	0	76	19
Habitat Channel		0	0	0	2	10	26	15	5	4	0	62	15.5
Pool		0	0	3	2	4	3	4	0	0	0	16	4
<b>Total:</b>		<b>0</b>	<b>3</b>	<b>3</b>	<b>9</b>	<b>54</b>	<b>125</b>	<b>153</b>	<b>40</b>	<b>12</b>	<b>1</b>	<b>400</b>	<b>100</b>

**Table 2.** Number and percent of hatchery (ad-clipped) and naturally produced (not ad-clipped) summer Chinook collected in Methow and Chelan rivers, 2014. The origin of three fish sampled could not be determined in the Methow River.

Reach	Location (Rkm)	Ad-Clipped Hatchery				Naturally Produced				Reach Total
		Male	Female	Total	Percent	Male	Female	Total	Percent	
<b>Methow River</b>										
<b>M1</b>	0.0-23.8	2	0	2	0.4	4	1	5	1.0	<b>7</b>
<b>M2</b>	23.8-43.8	1	2	3	0.6	6	4	10	2.1	<b>13</b>
<b>M3</b>	43.8-63.7	10	12	22	4.5	85	67	152	31.3	<b>174</b>
<b>M4</b>	63.7-72.3	4	1	5	1.0	27	21	48	9.9	<b>53</b>
<b>M5</b>	72.3-80.1	3	4	7	1.4	82	59	141	29.1	<b>148</b>
<b>M6</b>	80.1-83.0	2	1	3	0.6	43	27	70	14.4	<b>73</b>
<b>M7</b>	83.0-96.1	0	0	0	0.0	11	6	17	3.5	<b>17</b>
<b>Total</b>		<b>22</b>	<b>20</b>	<b>42</b>	<b>8.7</b>	<b>258</b>	<b>185</b>	<b>443</b>	<b>91.3</b>	<b>485</b>
<b>Chelan River</b>										
Powerhouse Tailrace		3	14	17	5.5	7	39	46	14.9	<b>63</b>
Columbia R. Tailrace		18	31	49	15.9	43	77	120	38.8	<b>169</b>
Pool		0	3	3	1.0	2	1	3	1.0	<b>6</b>
Habitat Channel		7	16	23	7.4	8	40	48	15.5	<b>71</b>
<b>Total</b>		<b>28</b>	<b>64</b>	<b>92</b>	<b>29.8</b>	<b>60</b>	<b>157</b>	<b>217</b>	<b>70.2</b>	<b>309</b>

**Table 3.** Historical aerial and ground redd counts of summer Chinook in the Methow, Chelan, Okanogan, and Similkameen rivers, 1956-2014.

Year	Methow		Okanogan		Similkameen		Chelan	
	Aerial	Ground	Aerial	Ground	Aerial	Ground	Aerial	Ground
1956	109	--	37	--	30	--	--	--
1957	451	--	53	--	30	--	--	--
1958	335	--	94	--	31	--	--	--
1959	130	--	50	--	23	--	--	--
1960	194	--	29	--	--	--	--	--
1961	120	--	--	--	--	--	--	--
1962	678	--	--	--	17	--	--	--
1963	298	--	9	--	51	--	--	--
1964	795	--	112	--	67	--	--	--
1965	562	--	109	--	154	--	--	--
1966	1,275	--	389	--	77	--	--	--
1967	733	--	149	--	107	--	--	--
1968	659	--	232	--	83	--	--	--
1969	329	--	103	--	357	--	--	--
1970	705	--	656	--	210	--	--	--
1971	562	--	310	--	55	--	--	--
1972	325	--	182	--	64	--	--	--
1973	366	--	138	--	130	--	--	--
1974	223	--	112	--	201	--	--	--
1975	432	--	273	--	184	--	--	--
1976	191	--	107	--	139	--	--	--
1977	365	--	276	--	268	--	--	--
1978	507	--	195	--	268	--	--	--
1979	622	--	173	--	138	--	--	--
1980	345	--	118	--	172	--	--	--
1981	195	--	55	--	121	--	--	--
1982	142	--	23	--	56	--	--	--
1983	65	--	36	--	57	--	--	--
1984	162	--	235	--	301	--	--	--
1985	164	--	138	--	309	--	--	--
1986	169	--	197	--	300	--	--	--
1987	211	--	201	--	164	--	--	--
1988	123	--	113	--	191	--	--	--
1989	126	--	134	--	221	370	--	--
1990	229	--	88	47	94	147	--	--
1991	--	153	55	64	68	91	--	--
1992	--	107	35	53	48	57	--	--
1993	--	154	144	162	152	288	--	--
1994	--	310	372	375	463	777	--	--
1995	--	357	260	267	337	616	--	--

Year	Methow		Okanogan		Similkameen		Chelan	
	Aerial	Ground	Aerial	Ground	Aerial	Ground	Aerial	Ground
1996	--	181	100	116	252	419	--	--
1997	--	205	149	158	297	486	--	--
1998	--	225	75	88	238	276	--	--
1999	--	448	222	369	903	1,275	--	--
2000	--	500	384	549	549	993	--	196
2001	--	675	883	1,108	865	1,540	--	240
2002	--	2,013	1,958	2,667	2,000	3,358	--	253
2003	--	1,624	1,099	1,035	103	378	--	173
2004	--	973	1,310	1,327	2,127	1,660	--	185
2005	--	874	1,084	1,611	1,111	1,423	--	179
2006	--	1,353	1,857	2,592	1,337	1,666	--	208
2007	--	620	1,265	1,301	523	707	--	86
2008	--	599	1,019	1,146	673	1,000	--	153
2009	--	692	1,109	1,672	907	1,298	--	246
2010	--	887	688	1,011	642	1,107	--	398
2011	--	941	1,203	1,714	1,047	1,409	--	413
2012	--	960	1,170	1,613	762	1,066	--	426
2013	--	1,551	NA	2,267	NA	1,280	--	729
2014	--	591	NA	2,231	NA	2,022	--	400